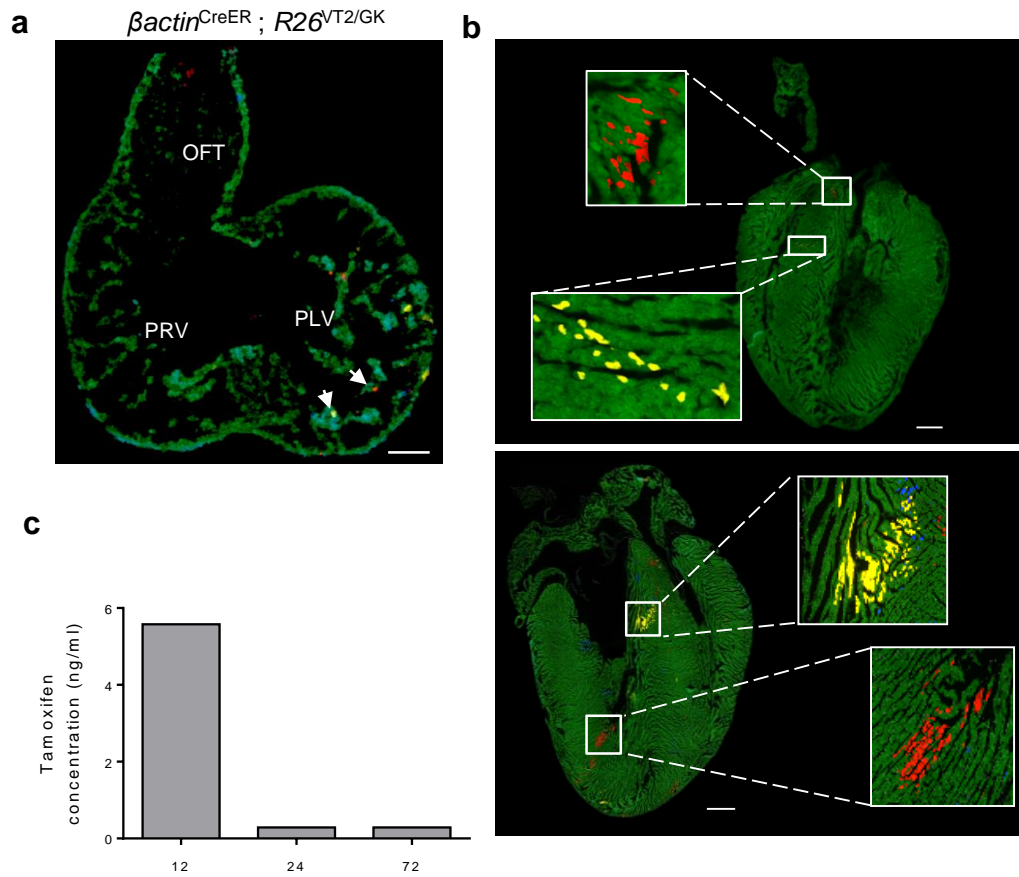
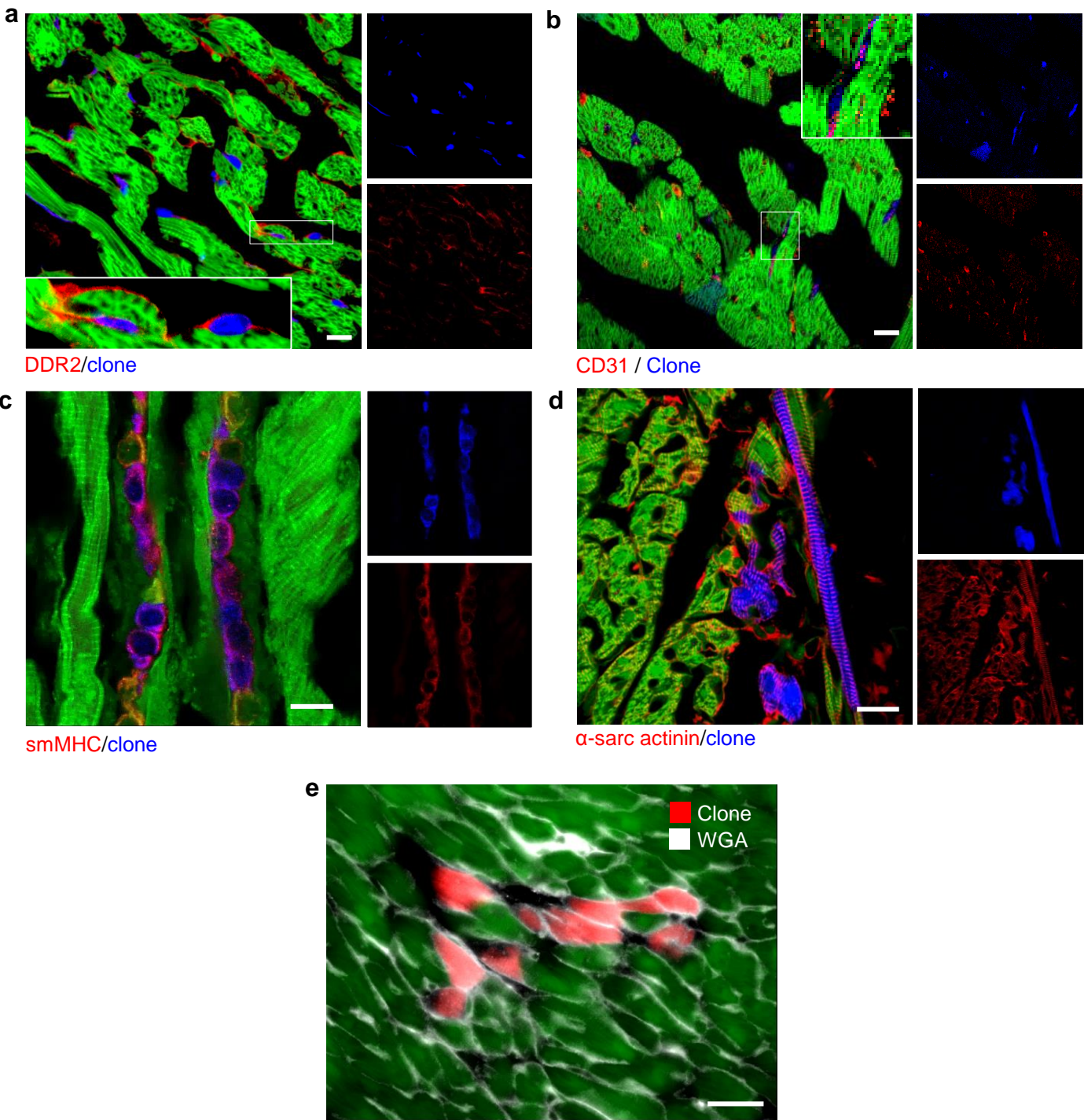


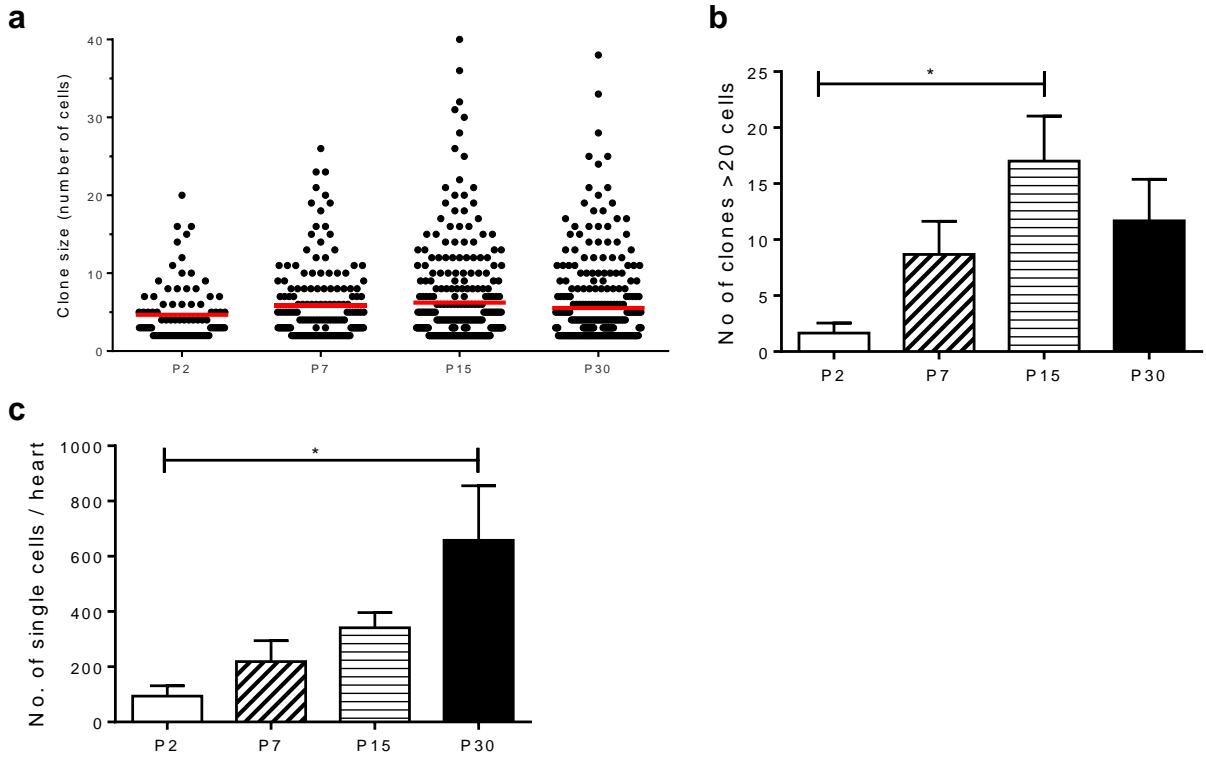
Supplementary Figure 1: The Rainbow model provides a precise, direct method to study clonal expansion of cardiac progenitors and CMs within the developing heart. a, Schematic representation of the Rainbow reporter system. Expression of Cre induces random recombination between mutated paired lox P sites (black, grey and white triangles) leading to expression of Cerulean, mOrange and mCherry respectively. **b-c**, Quantification of the number of labeled clones per heart from 2D sections of P1 hearts (n=11) (**b**) and 3D imaging of a $\alpha MHC^{Cre}; R26^{VT2/GK}$ P1 heart (**c**) demonstrates equal representation of Cerulean, mOrange and mCherry. All measurements shown are depicted as mean \pm s.e.m.



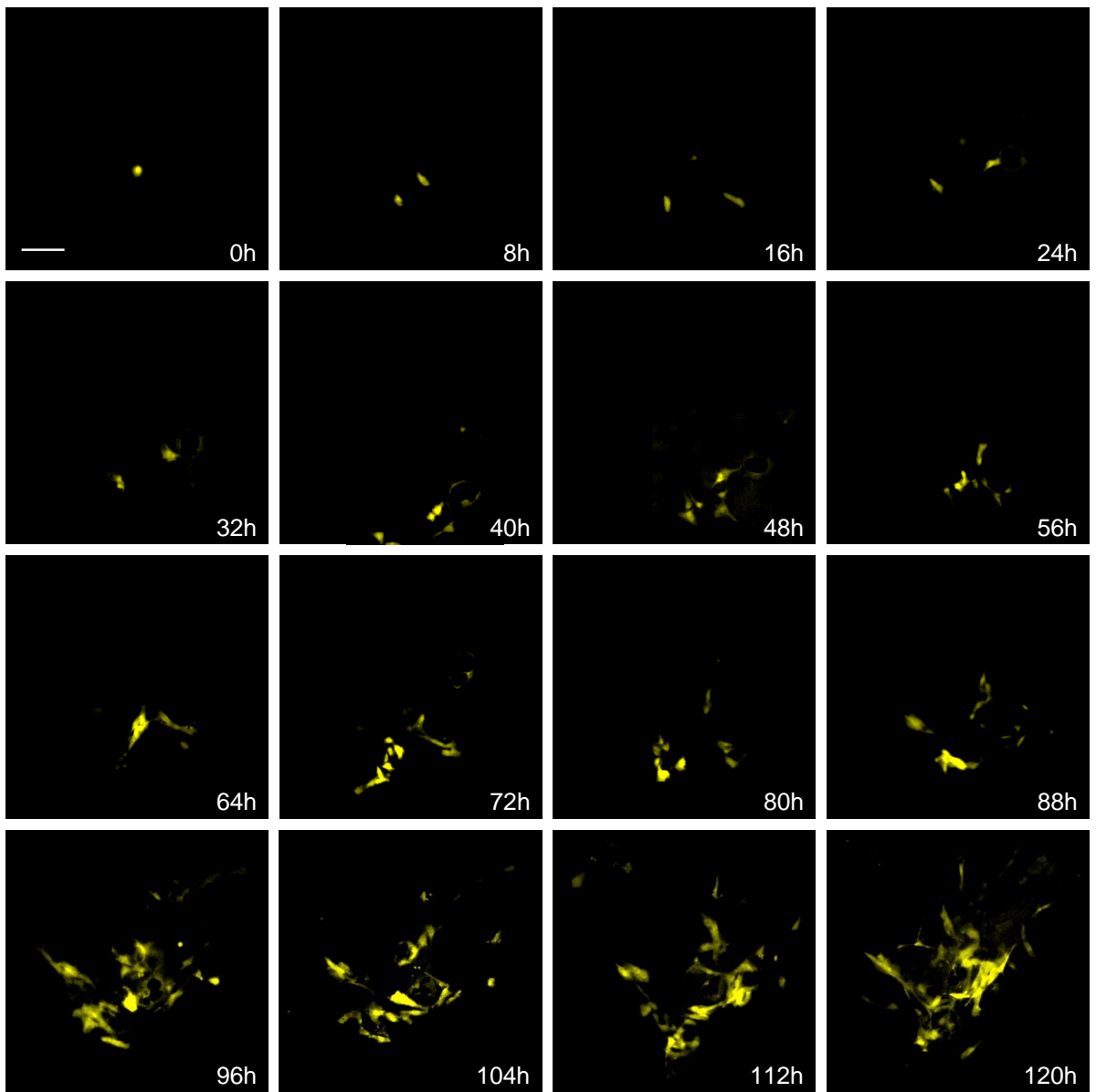
Supplementary Figure 2: Single low dose of tamoxifen induces rare recombination events. **a**, Representative fluorescent microscope image of $\beta actin^{CreER}; R26^{VT2/GK}$ E10.5 hearts 24hrs post tamoxifen administration. Arrows indicate single recombination events. Scale bar 100 μm . **b**, Representative images of adult hearts with sparse clones. Recombination was induced at E9.5. Insets show magnification of boxed areas. Scale bar 500 μm . **c**, Tamoxifen is removed from the circulation within 24 hours after treatment. Blood serum levels of tamoxifen measured 12, 24, and 72 hours following administration. Tam, tamoxifen; OFT, outflow tract; PRV, primitive right ventricle; PLV, primitive left ventricle.



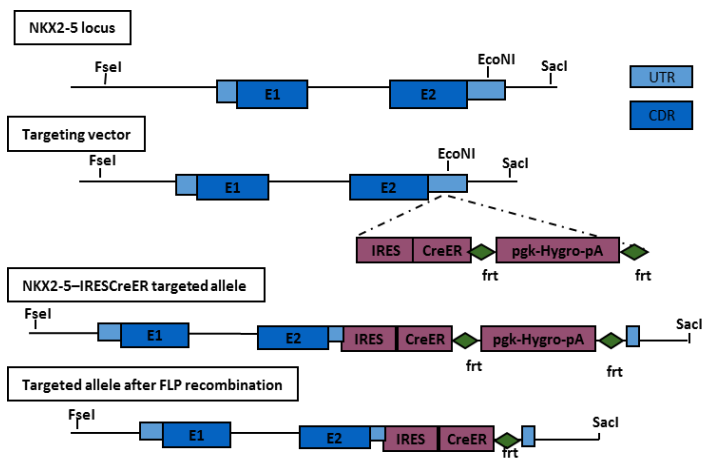
Supplementary Figure 3: Immunohistochemical staining of clones. Immunohistochemical staining of β actin^{CreER}; R26^{VT2/GK} clones positive for **a**, the fibroblast marker, DDR2, **b**, endothelial cell marker, CD31, **c**, smooth muscle myosin heavy chain, smMHC (scale bar 50 μ m), and **d**, CM, α -sarcomeric actinin. Insets show close ups of the co-localization. **e**, WGA (white) was used to delineate individual cells within a clone (red pseudocolor) for counting purposes. Scale bar 100 μ m unless otherwise denoted.



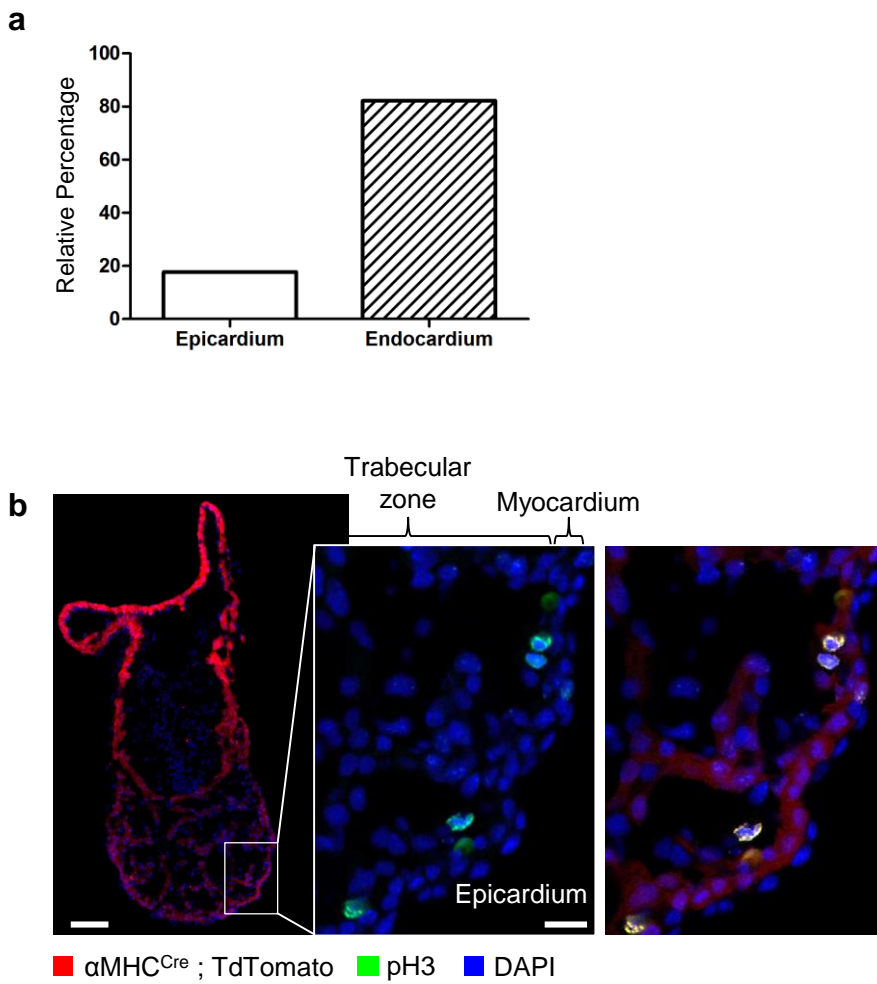
Supplementary Figure 4: Clonal expansion occurs mainly during embryonic development. **a**, Dot plot analysis of clonal sizes in $\beta actin^{CreER}; R26^{VT2/GK}$ hearts labeled at E9.5 and analyzed at P2, P7, P15, and P30 (n=3). **b**, Number of clones > 20 cells progressively increase up to P15 and declines by P30. **c**, Number of single cells increases over time (n=3). All measurements shown are depicted as mean \pm s.e.m.



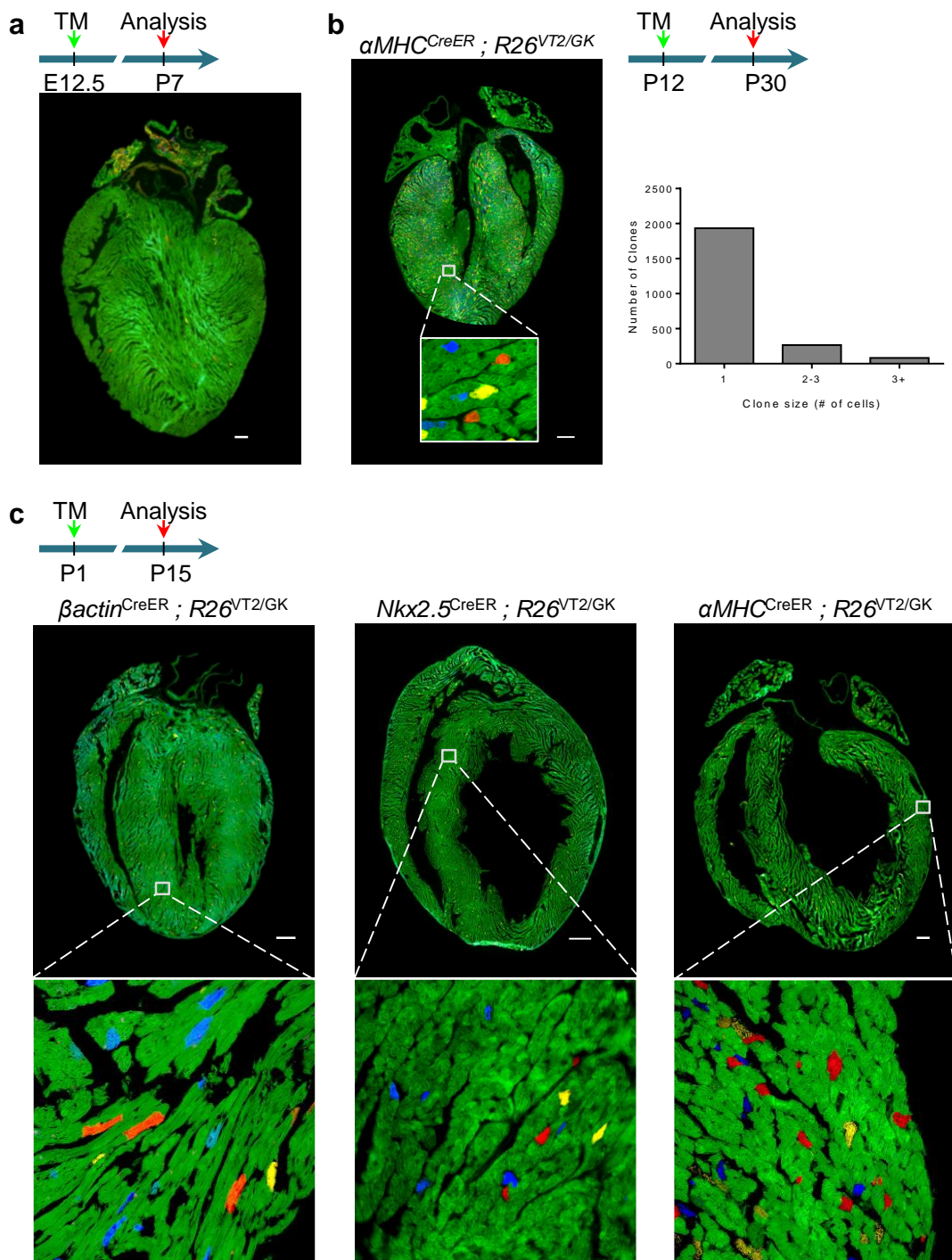
Supplementary Figure 5: “Rainbow-labeled” cardiac cells expand clonally *in vitro*. Time-lapse imaging of a single rainbow-labeled cell (yellow pseudocolor) forming a clone *in vitro*. Monitoring was performed 0-120 hours after plating. Scale bar 100 μ m.



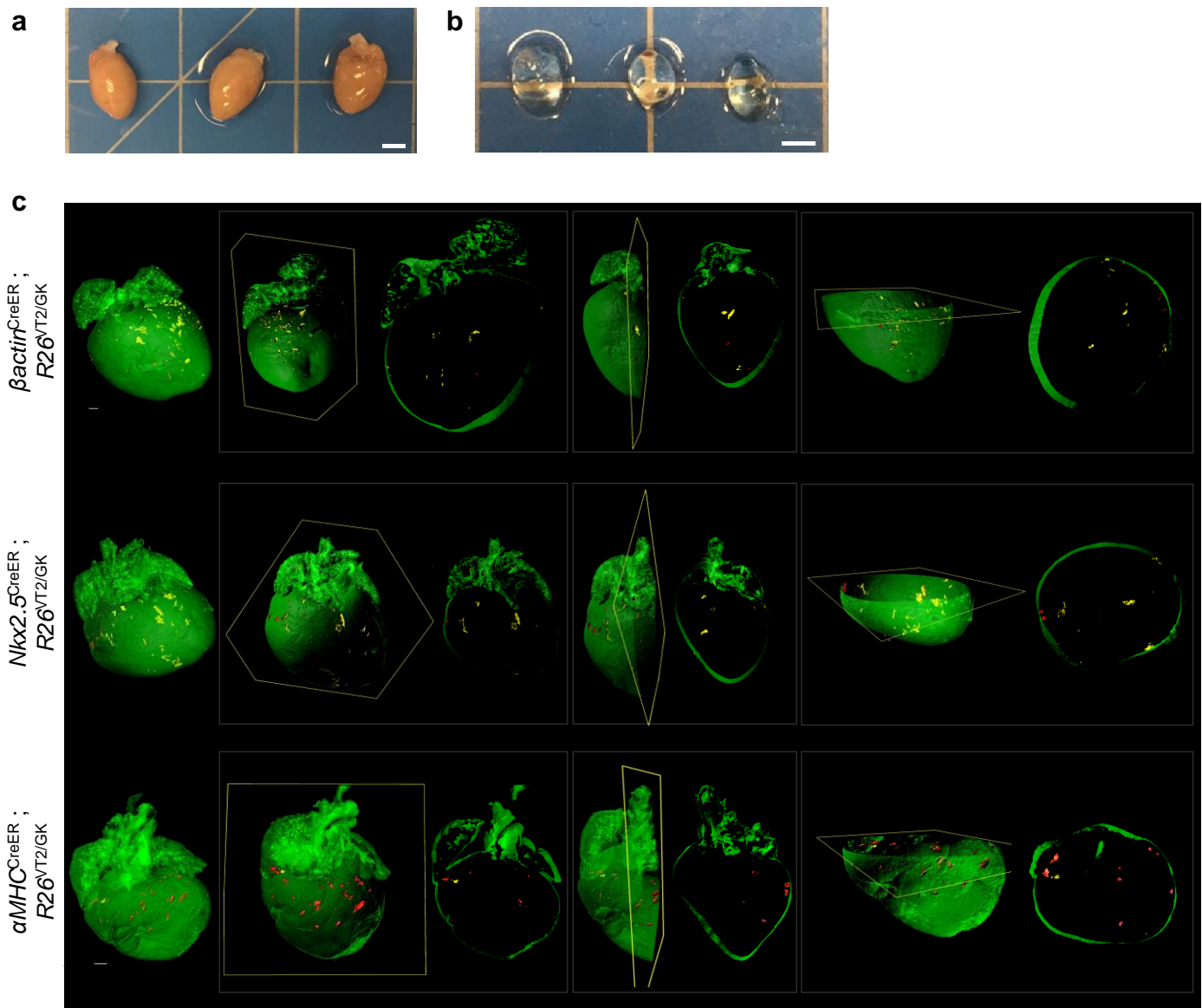
Supplementary Figure 6: Generation of the Nkx2.5-CreER construct. Maps of the Nkx2.5 wild-type allele, gene targeting construct and resultant mutant alleles before (Nkx2.5IRES-CreER-pgkHYGRO) and after (Nkx2.5IRES-CreER) flp-mediated deletion. Several stable clones were generated and founders of this new strain (Nkx2.5-CreER) were backcrossed onto C57BL/6 mice.



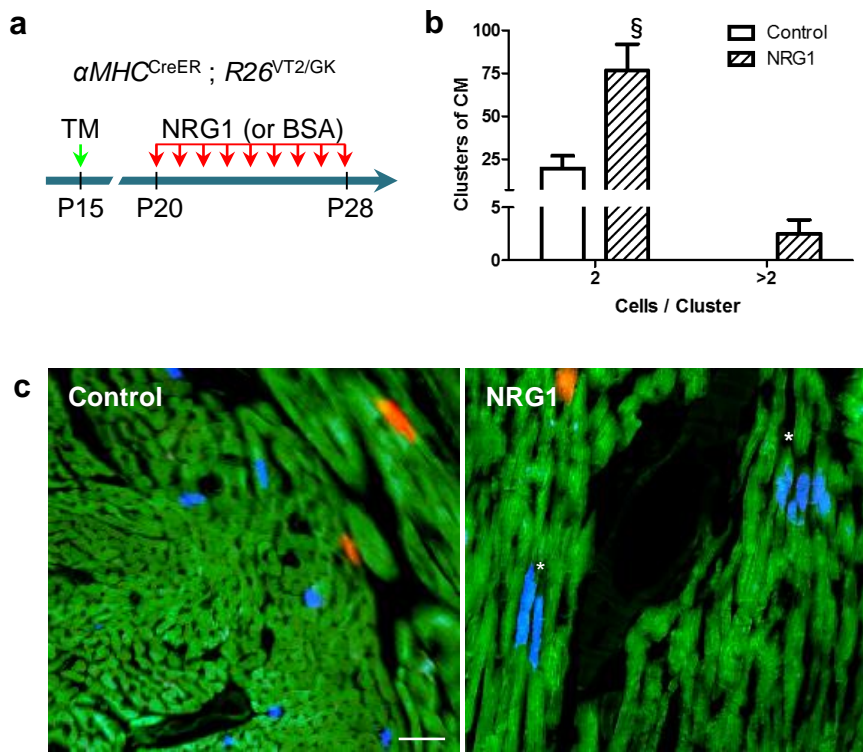
Supplementary Figure 7: CM clones are mainly localized near the endocardium. **a**, Quantification of CM clones based on their proximity to either the epicardium or the endocardium. Tamoxifen was administered at E9.5 and hearts were analyzed at P2. **b**, Immunohistochemical staining for phospho-Histone H3 (pH3 in green) of a E9.5 α MHC^{Cre} ; TdTomato heart. Scale bar 100 μ m. Inset shows magnification of boxed area, scale bar 20 μ m.



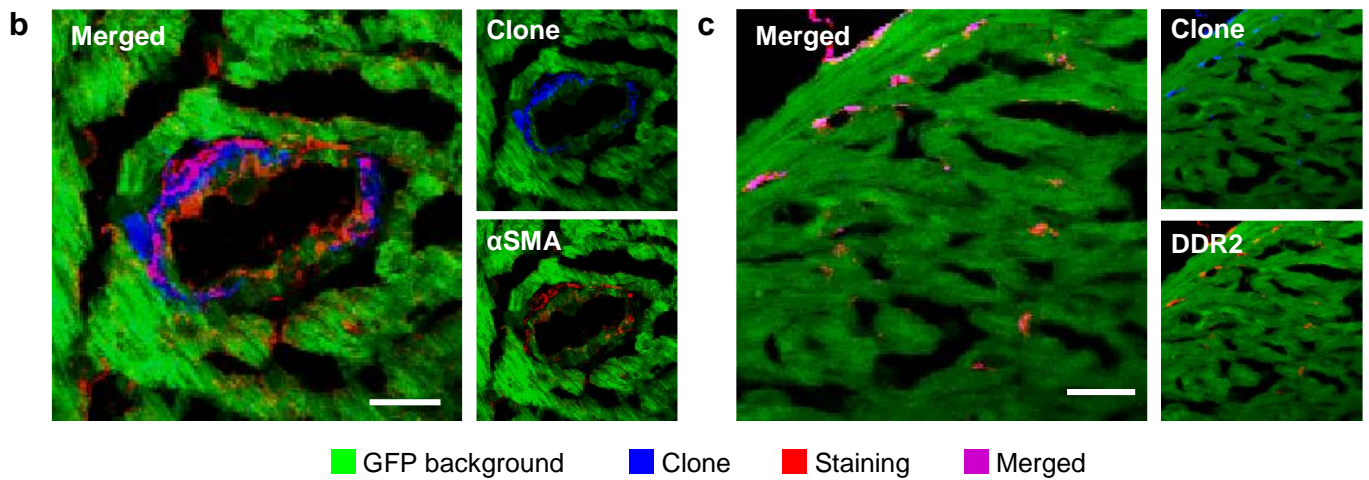
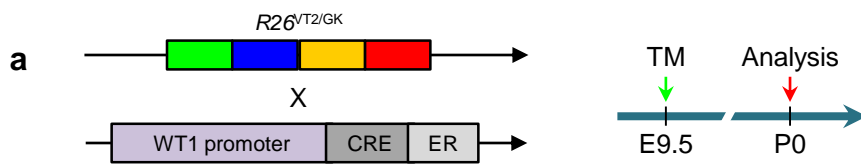
Supplementary Figure 8: Postnatal CMs exhibit limited proliferative capacity in the absence of injury. **a**, $\alpha MHC^{CreER}; R26^{VT2/GK}$ mouse heart labeled at E12.5 and analyzed P7 showing the distribution of αMHC primarily in the atria and to a lesser extent in the ventricles, scale bar = 200 μm . **(b, left)** Representative image of $\alpha MHC^{CreER}; R26^{VT2/GK}$ P30 heart labeled at P12 showing the dominance of single Rainbow-labeled cells, scale bar 500 μm . **(b, right)** Quantitation of clone size and number within $\alpha MHC^{CreER}; R26^{VT2/GK}$ mouse hearts labeled at P12 and analyzed P30 (n=4, 8 images at 40X magnification analyzed per section, 5 sections/heart). **c**, Representative confocal microscope images of $\beta actin^{CreER}; R26^{VT2/GK}$, $Nkx2.5^{CreER}; R26^{VT2/GK}$, and $\alpha MHC^{CreER}; R26^{VT2/GK}$ P15 hearts labeled at P1. Insets show up close view of boxed areas. Scale bar 500 μm .



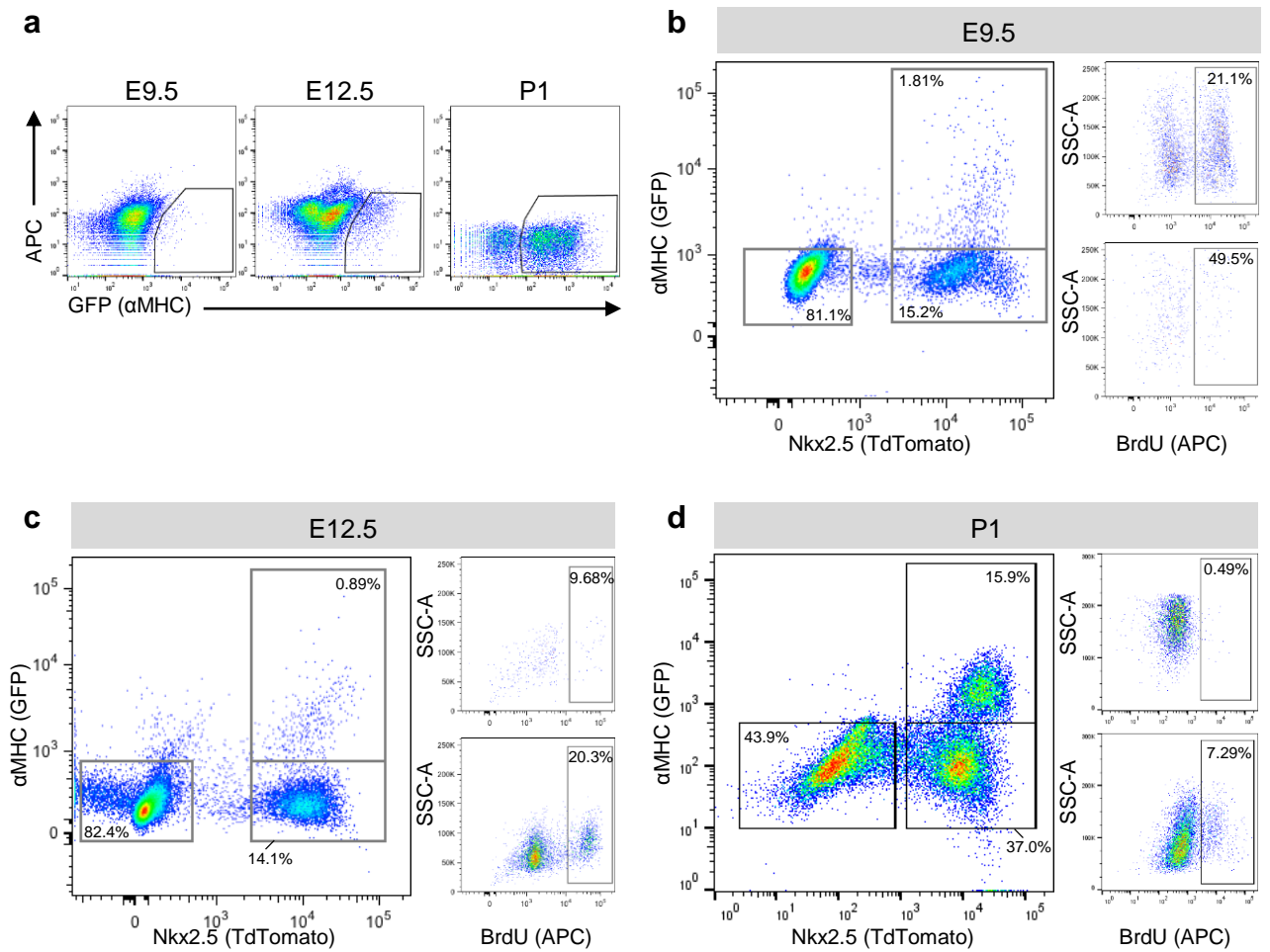
Supplementary Figure 9: Three dimensional imaging of cleared whole neonatal mouse hearts using the CLARITY method. P2 hearts prior to CLARITY (**a**) and post CLARITY (**b**), scale bars = 2 mm. **c**, 3 dimensional rendering of cleared P2 hearts from β actin^{CreER} ; R26^{VT2/GK}, $Nkx2.5$ ^{CreER} ; R26^{VT2/GK}, and α MHC^{CreER} ; R26^{VT2/GK}. Examples of sagittal, coronal, and axial planes captured using Imaris 7.7.2 software (Bitplane AG, Zurich).



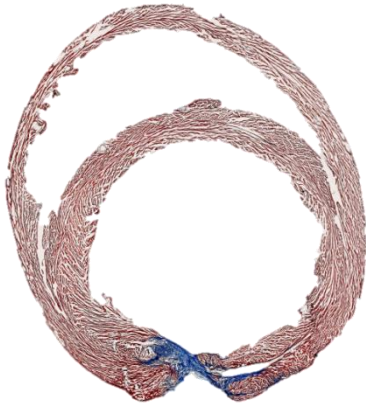
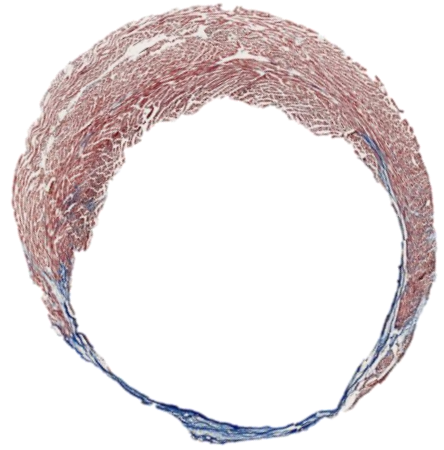
Supplementary Figure 10: NRG1 treatment induces postnatal CM proliferation. **a**, Experimental design of NRG1 treatment. Arrow indicates tamoxifen administration at P15. NRG1 (n=4) or BSA control (n=3) were given daily to *αMHC^{CreER} ; R26^{VT2/GK}* mice from P20 to P28. **b**, Quantification reveals increased formation of CM clusters following NRG1 treatment. **c**, Representative fluorescent microscope images of control and NRG1-treated hearts. Stars indicate clusters of CMs. § NRG1 vs control, $p < 0.05$. Scale bar 50 μm . All measurements shown are depicted as mean \pm s.em.



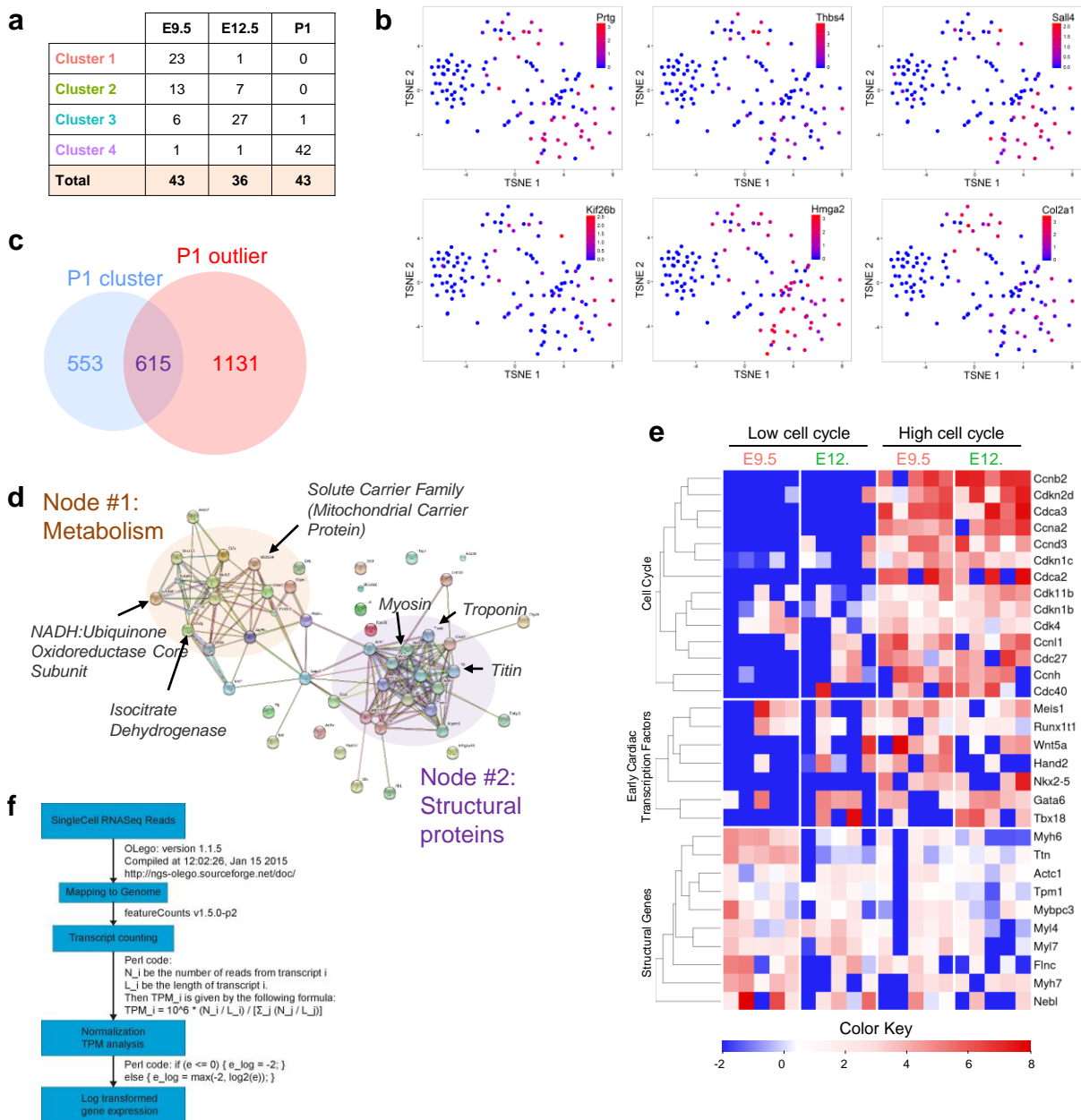
Supplementary Figure 11: Smooth muscle cell and fibroblast clones of WT1 origin. **a**, Experimental design. *Wt1^{CreER}* ; *R26^{VT2/GK}* mice received tamoxifen at E9.5 and were analyzed at P0. **b-c**, Representative confocal microscope images of **b**, a clone (blue) consisting of smooth muscle cells (α -SMA in red) and **c**, a clone of fibroblasts (DDR2 in red). Insets show the individual pseudocolors in GFP background. Scale bars 20 μ m.



Supplementary Figure 12: BrdU incorporation in CMs during development. **a**, Representative flow cytometric analysis of αMHC-GFP cells sorted for qPCR or single cell analysis. Gated areas show cells selected for sorting. **b-d**, Representative flow cytometric analysis plots of BrdU incorporation in *Nkx2.5^{+Cre}; R26R-Tdt^{+/fl}; αMHC^{+GFP}* hearts at **(b)** E9.5, **(c)** E12.5, and **(d)** P1.

a**b**

Supplementary Figure 13: Mason's trichrome staining of infarcted hearts. Representative images of a heart section 21 days following LAD ligation at **(a)** P1 and **(b)** 2 months of age. A small area of fibrosis (blue) is still apparent **(a)** after neonatal injury while injury in adult results in large scar formation **(b)**.



Supplementary Figure 14: Transcriptional profiling of CMs during early embryonic and postnatal development. **a**, Table depicting numbers of single cells found within each km cluster. **b**, Expression of genes found only in E9.5 cells (*Thbs4*, *Kif26b*, *Col2a1*, *Prtg*) or were highly enriched (*Sall4*, *Hmga2*) within cells from this timepoint. **c**, Venn diagram depicting the number of differentially expressed genes between P1 cluster and P1 outlier. **d**, Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) analysis comparing the top 50 genes more highly expressed in the P1 cluster compared to P1 outlier. Two nodes were identified encoding for genes of metabolism (bronze circle) and structural proteins (purple circle). **e**, Single cells at E9.5 and E12.5 were selected based on low or high cell cycle activity, as determined by expression levels of common cell cycle genes. Genes encoding for cardiac structural proteins and transcription factors were examined within the selected cells showing a possible link between cell cycle activity and CM maturity. **f**, RNA Sequencing Analysis Workflow. RNASeq data was mapped with OLego version 1.1.5 and normalized using Transcripts per Millions analysis. featureCounts version v1.5.0-p2 was used to estimate total number of reads. Expression levels were then log transformed using algorithm depicted. ** $p < 0.01$. Mean \pm SEM.

Supplementary Table 1: List of genes for used for heatmap in Figure 5b.

Gene Group	List of Genes			
Muscle / Cardiac Cell Differentiation	Myh9 Myl6 Myh10 Tgfb2 Tbx5	Tiparp Myh6 Actg1 Foxp1 Tagln2	Lgals1 Tbx18 Dlg1 Sort1 Rhoa	Mbnl1 Ttn Dicer1 Ppp3ca Tmod1
Gene Silencing / Histone Modification	Tyms Hdac1 Ilf3 Smarca5 Ehmt1 Prmt7	Hells Sirt6 Gspt1 Gamt Dhfr Ezh2	Coq5 Trmt112 Ncbp2 Kdm6b Wbscr22 Tgs1	Rrp8 Brcc3 Mat2a N6amt1 Tnrc6b
Cell Cycle	Cks1b Myh9.1 Cdk4 Trp53 Cdca8 Ranbp1 Mcm7 Myh10.1 Gmnn Sik1 Cfl1 Mcm2 Nudc Mcm5 Tpx2 Hcfc1	Ccnd1 Haus7 Tgfb2.1 Spin1 Pmf1 Tipin Hells.1 Cenpe Tacc3 Myh6.1 Actg1.1 Anapc5 Ccnd3 Haus8 Cdk7 Cdk2	Smad3 Nde1 Fam33a Dhfr.1 Calm2 Erh Cdk11b Ak1 Actb Mfn2 Ccar1 Ccnh Npm1 4933424B01Rik Rpl24 Tcf3	Anapc11 Rbbp4 Rcc2 Rbm7 Bcl2 E2f3 Haus1 Msh2 Nedd1 Gak Rac1 Rhoa.1 Mapre1 Lrrcc1
Metabolism	Gbas Uqcrfs1 Uqcrb Myh7	Uqcrq Atp5h 2410091C18Rik Itpr1	Cycs Atp5l Atp5o Cox6b1	Atp5j Cox7c Cox6c
Cell Growth / Migration	Cdk4.1 Trp53.1 Gtpbp4	Tgfb2.2 Tbx5.1 Celf1	Hif1a Dlg1.1 Mkks	Bcl2.1 Rac1.1
Ion Channels / Transporters	Impdh2 Tnfaip1	Pkm2 Cdk2.1	Atp1b2 Abcc9	Mat2a.1 Gmpr2
Muscle / Cardiac Cell Structure & Contraction	Ankrd1 Myh9.2 Actn1 Tpm2 Myl6.1 Cnn3 Myh10.2 Cald1	Tbpl1 Polr1e Haus7.1 Zdbf2 Myh7.1 Cenpe.1 Myh6.2 Actg1.2	Hmgn2 Itpr1.1 Abcc9.1 Cycs.1 Gnpat Rpl24.1 Dlg1.2 Hist1h2ab	Ank2 Ldb3 Ttn.1 Hist1h2an Dicer1.1 Rpl9 Cacna1c Ppp3ca.1