

Figure S2 (relates to Figure 2). Tor signaling is activated in tracheal epithelial cells upon regeneration.

A. Complete set of panels for data shown in Fig.2B. Confocal images of a representative area of the tracheal epithelium of either control or SO₂ treated mice. IHC to detect Trp63 (BCs), Krt8 (differentiated cells) and p-S6 (Tor signaling activity). For each time-point pictures show one representative epithelium (from n=5 for SO₂ treated and n=3 for controls).

B. Complete set of panels for data shown in Fig.2D and Fig.2E. Confocal images of a representative area of the tracheal epithelium of either control or SO₂ treated mice. IHC to detect Krt5 (BCs) and Krt8 (differentiated cells). Arrowheads point to examples of Krt5+ cells (green), Krt8+ cells (red) and Krt5+/Krt8+ cells (yellow). For each time-point pictures show one representative epithelial section (from n=5 for SO₂ treated and n=3 for controls).

C. Confocal images of an area of the tracheal epithelium of wild type mice, either mock treated or one day after SO₂ exposure. IHC detecting Ki67 (green and white) and p-S6 (red and white). Images show representative epithelia (n>5).

D. Western blot to detect phosphorylated ribosomal protein S6 (p-S6) in whole trachea at indicated time points after SO₂ exposure. Total S6 and alpha-tubulin were detected as loading controls. Representative blot is shown; quantification is shown in Fig. 2C.

E & F Confocal images of tracheospheres treated with Rapamycin (200nM) for 6 days. In E, Spheres were stained for a mitotic marker (pH3, green and white), a BC marker (Trp63, red) and a differentiated cell marker (Krt8, white). Quantification of pH3+ cell numbers is shown in the chart (T test). In F, Spheres were stained for an apoptotic marker (Cleaved Caspase 3, green and white), a BC marker (Trp63, red) and a differentiated cell marker (Krt8, white). For both DAPI is shown in blue. Panels show one representative out of n≥20 spheres.

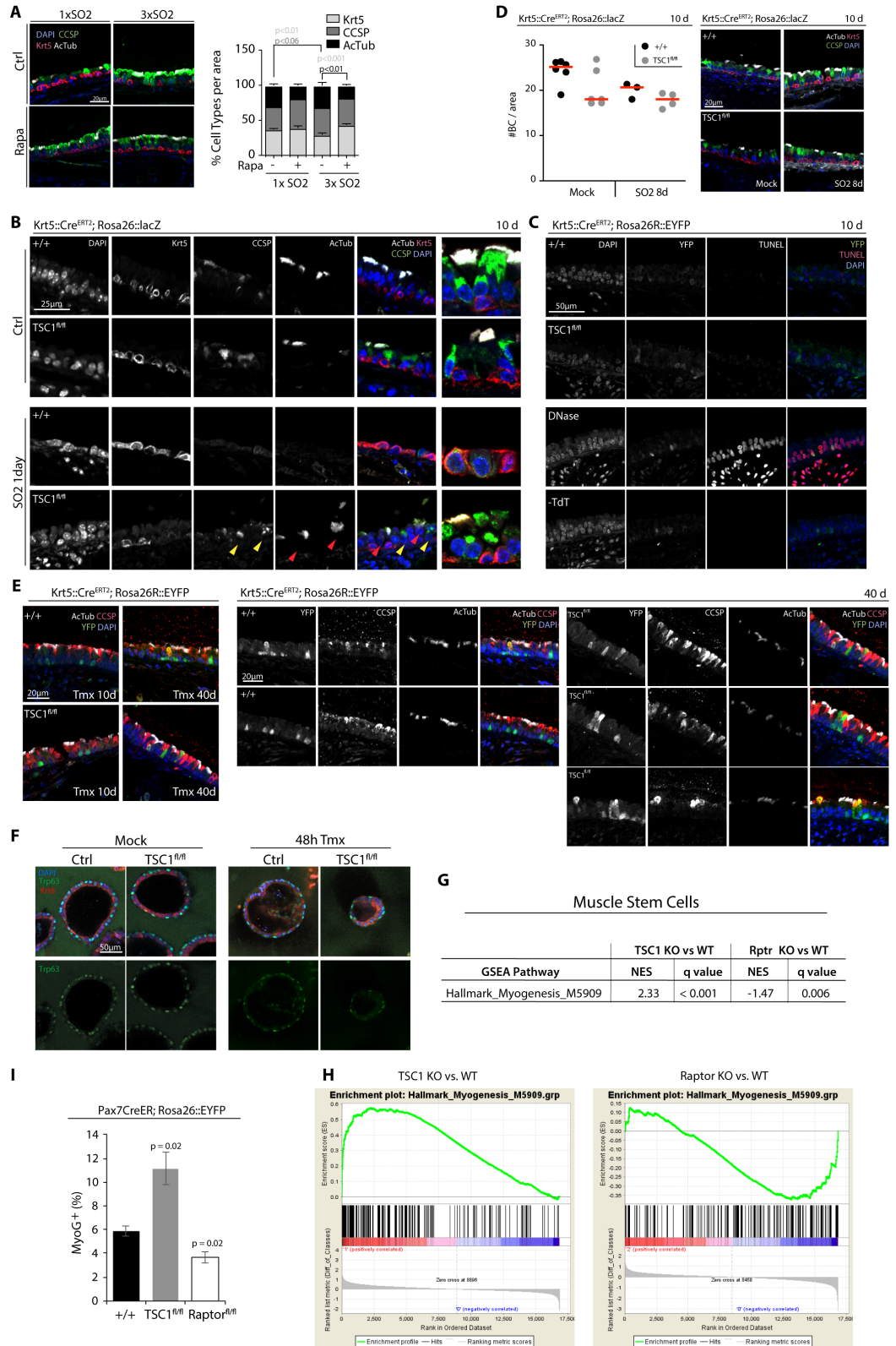


Figure S3 (Related to Figure 3). SC loss and differentiation in response to TOR activation in the mouse tracheal epithelium and muscle

A. Confocal images of an area of the tracheal epithelium and quantification of BCs (Krt5), secretory cells (CCSP) and ciliated cells (AcTub) in young animal two weeks after the first or third SO₂ exposure (SO₂ exposure was performed every other week). IHC detecting BCs (red), secretory cells (CCSP, green) and ciliated cells (AcTub, white). Images show representative epithelia (n=5). Bars representing the mean +/- SEM. ANOVA U-test (n>4).

B. Confocal images of an area of the tracheal epithelium of controls (Krt5::Cre^{ERT2}, Rosa26::lacZ / +) or TSC1 deleter mice (Krt5::Cre^{ERT2}, Rosa26::lacZ / TSC1^{fl/fl}) of SO₂ or Mock treated mice at one day after exposure. The SO₂ exposure was performed 10 days after the last Tamoxifen injection (4 IP injections every other day). IHC detecting BCs (Krt5, red and white), Secretory Cells (CCSP, green and white) and Ciliated Cells (AcTub, white). Images show representative epithelia (n≥4).

C. Confocal images of an area of the tracheal epithelium of controls (Krt5::Cre^{ERT2}, Rosa26::eYFP / +) or TSC1 deleter mice (Krt5::Cre^{ERT2}, Rosa26::eYFP / TSC1^{fl/fl}) at 10 days after the last Tamoxifen injection (3 injections, every other day). TUNEL staining detecting apoptotic cells (red and white) was performed in addition to a positive control (DNase treated section) and a negative control (without the TdT enzyme). Images show representative epithelia (n≥4).

D. On the left, quantification of BCs (Trp63 positive cells) in the tracheal epithelium of controls (Krt5::Cre^{ERT2}, Rosa26::lacZ / +) or TSC1 deleter mice (Krt5::Cre^{ERT2}, Rosa26::lacZ / TSC1^{fl/fl}), mock or SO₂ treated. The SO₂ exposure was performed 10 days after the last Tamoxifen injection (4 IP injections every other day). Each data point represents the mean for one trachea. Red lines represent the median. ANOVA U-test. On the right, confocal images of representative tracheal epithelia at 8 days after SO₂ stained with different cell type markers (Krt5 in red for BCs, CCSP in green for secretory cells and AcTub in white for ciliated cells). n≥3.

E. Confocal images of an area of the tracheal epithelium of controls (Krt5::Cre^{ERT2}, Rosa26::eYFP / +) or TSC1 deleter mice (Krt5::Cre^{ERT2}, Rosa26::eYFP / TSC1^{fl/fl}) at 10 or 40 days after the last Tamoxifen injection (3 injections, every other day). IHC detecting eYFP clones (green), Secretory Cells (CCSP, red) and Ciliated Cells (AcTub, white). Images show representative epithelia (n≥4).

F. Confocal images of tracheospheres derived from control (Krt5::Cre^{ERT2}, Rosa26::lacZ / +) or TSC1 deleter mice (Krt5::Cre^{ERT2}, Rosa26::lacZ / TSC1^{fl/fl}). Spheres were grown for 9 days, then exposed to Tamoxifen for 2 days. Trp63+ cells (BCs, green) and differentiated cells (Krt8+, red) are detected by IHC (DAPI blue).

G., H. mTORC1 signaling promotes expression of genes involved in skeletal muscle myogenesis in MuSCs. Data are displayed as normalized enrichment scores (NES) and FDR-adjusted p-values (q value) from gene-set enrichment analysis (GSEA) of gene expression profiles of total RNA purified from YFP+ MuSCs immediately after isolation

from adult WT, TSC1 KO, or Raptor KO mice. Gene expression profiling experiments were described in Rodgers et al., Nature 2014 and the data are deposited as GSE55490.

I. mTORC1 signaling promotes expression of MyoG in MuSCs. Data are presented as percent MyoG MuSCs, 12 hours after FACS isolation of YFP+ cells from adult WT, TSC1 KO, Raptor KO mice, as measured by IF staining. Mean and sem, P values were calculated using two tailed students tTest (WT; n=3, TSC1 KO; n = 3, Raptor KO; n = 4).

WT: *Pax7CreER* /+;*rosa26EYFP* /+; TSC1 KO: *TSC1*^{fl/fl}; *Pax7CreER* /+;*rosa26EYFP* /+;
Raptor KO: *RPTR*^{fl/fl}; *Pax7CreER* /+;*rosa26EYFP* /+.

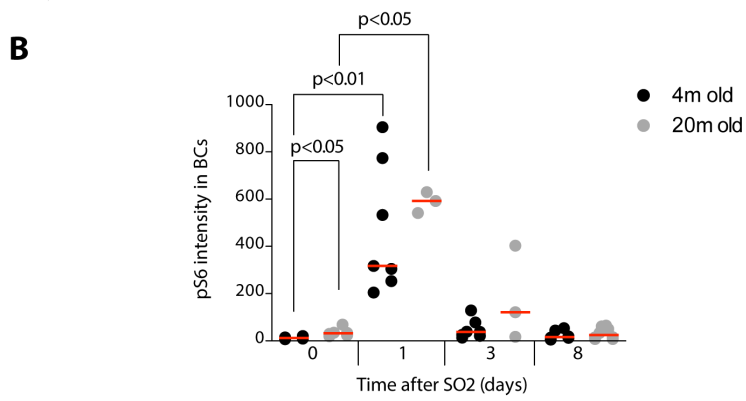
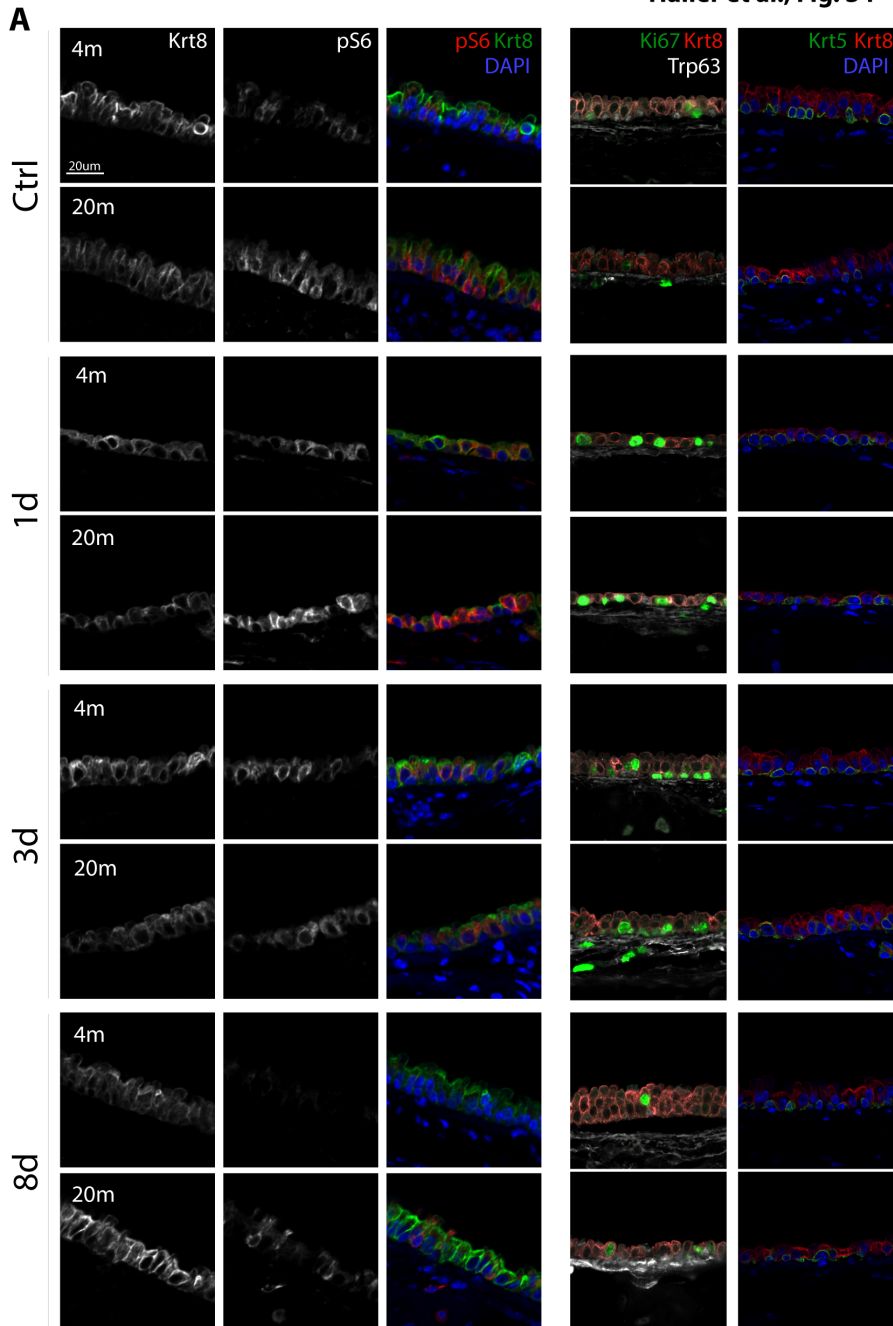


Figure S4 (related to Figure 4). Delayed regeneration of the airway epithelium of old mice.

A: Images corresponding to the quantifications in Fig.4A. Confocal images of an area of the tracheal epithelium of either 4 month (4m) or 20 month (20m) old wild-type (C57BL/6J) mice at the indicated timepoints after SO₂ exposure. IHC detecting Trp63 (white), pS6 (red), Krt5 (green), Krt8 (red or green), and Ki67 (green). Images show representative epithelia (n≥3 mice).

B: Quantification of p-S6 levels in the BCs (fluorescence intensity normalized to background) in the conditions shown in A. Mean and SEM are shown, ANOVA U-Test.

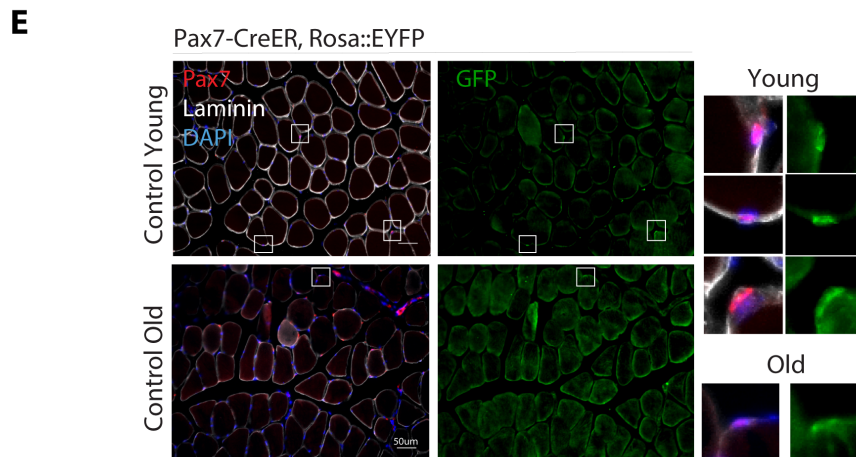
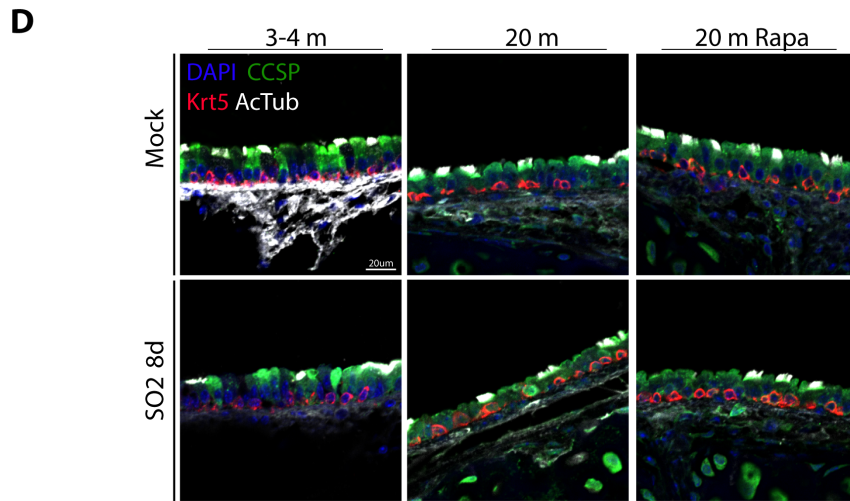
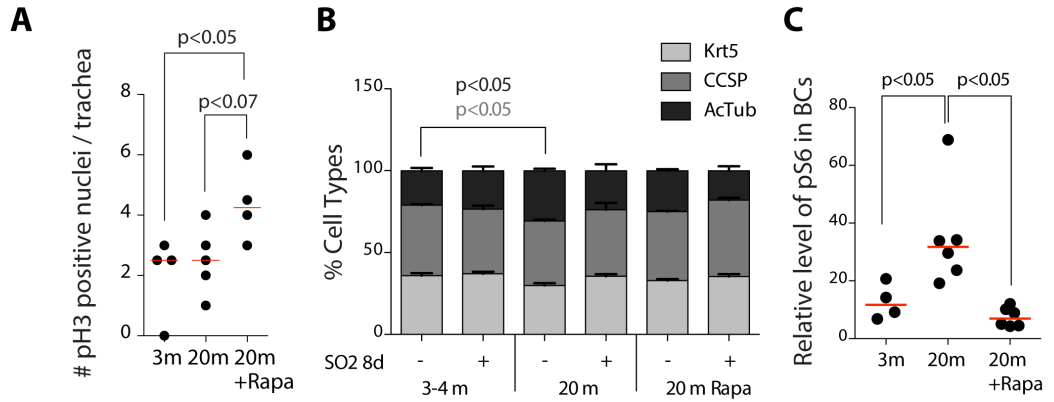


Figure S5 (related to Figure 5). Age-related stem cell phenotypes in trachea and muscle

A. Quantification of mitotic figures (pH3 staining) in young (3 months old), old (20 months old) and old Rapamycin treated (3 month of Rapamycin 42ppm) epithelia (no injury). Each data point represents the mean for one trachea. Red lines represent the median. ANOVA U-test.

B. Quantification of BCs (Krt5), Secretory cells (CCSP) and Ciliated Cells (AcTub) in an area of the tracheal epithelium of young, old and old Rapamycin treated mice after SO₂ exposure. Bars representing the mean +/- SEM. ANOVA U-test (n>4).

C. Quantification of p-S6 levels (fluorescence, normalized to the background) in BCs of young (3m old), old (20m old) and old Rapamycin treated mice (3 month of Rapamycin starting at 17 months of age). Each data point represents the mean for one trachea. Red lines represent the median.

D. Confocal images of an area of the tracheal epithelial of young (3 month), old (20 month) and old Rapamycin treated for 3 month (20 month) at 8 days after SO₂ exposure or mock exposed. IHC detecting BCs (red), secretory cells (CCSP, green) and ciliated cells (AcTub, white). Images show representative epithelia (n≥4).

E. Representative images of IHC staining of TA (tibial anterior) muscles from young and aged control mice. White squares surround GFP⁺-Pax7⁺ MuSCs. Higher resolution images of these cells are displayed on the right. The scale bar represents 50µm.