

Figure S1. LAP2*a* is downregulated in HGPS fibroblasts. (A) Primary human fibroblasts were fixed with 4% PFA and processed for immunofluorescence using anti LAP2*a* antibody (green) and progerin antibody (red). DNA was stained with DAPI (blue). Scale bar: 20μ m. (B) Average mean LAP2*a* fluorescence intensities of one wild-type (WT 1) and three different HGPS primary human fibroblast cell lines (HGPS 1, 2 and 3) were measured in 250 nuclei each and plotted in a histogram (n=3). (C) One wild-type (WT 1) and three different HGPS primary human fibroblast cell lines (HGPS 1, 2 and 3) were stained with anti-progerin antibody. Average mean progerin fluorescence intensities were measured in 250 nuclei each and plotted in a histogram (n=3). (D) Growth curves of a three different WT cell lines (p17) over 6 days (n=3). (E) Growth curves of HGPS 2 cell line at middle (p15) and late (p21) passage (n=3).

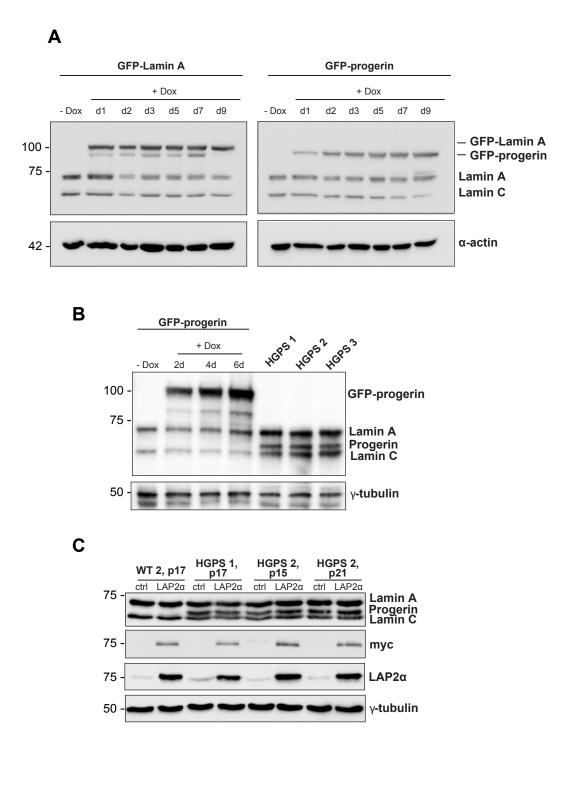


Figure S2. Immunoblot analysis of hTERT TetOn cells and primary human fibroblasts. (A) Immunoblot analysis of total cell lysates of GFP-Lamin A and GFP-progerin expressing cells at different time-points after induction with 0.5 μ g ml⁻¹ doxycycline using anti-lamin A/C antibody. α -ctin levels served as a loading control. (B) Immunoblot analysis of GFP-progerin cells at different time-points of induction with 1.0 μ g ml⁻¹ doxycycline and primary HGPS-derived fibroblasts using anti-lamin A/C antibody. γ -tubulin served as a loading control. (C) Immunoblot analysis of one wild-type and three different HGPS cell lines using anti-lamin A/C, anti-myc and anti-LAP2 α antibodies following lentiviral transfections with myc-tagged human LAP2 α or GFP (ctrl) plasmids. γ -tubulin levels served as a loading control.

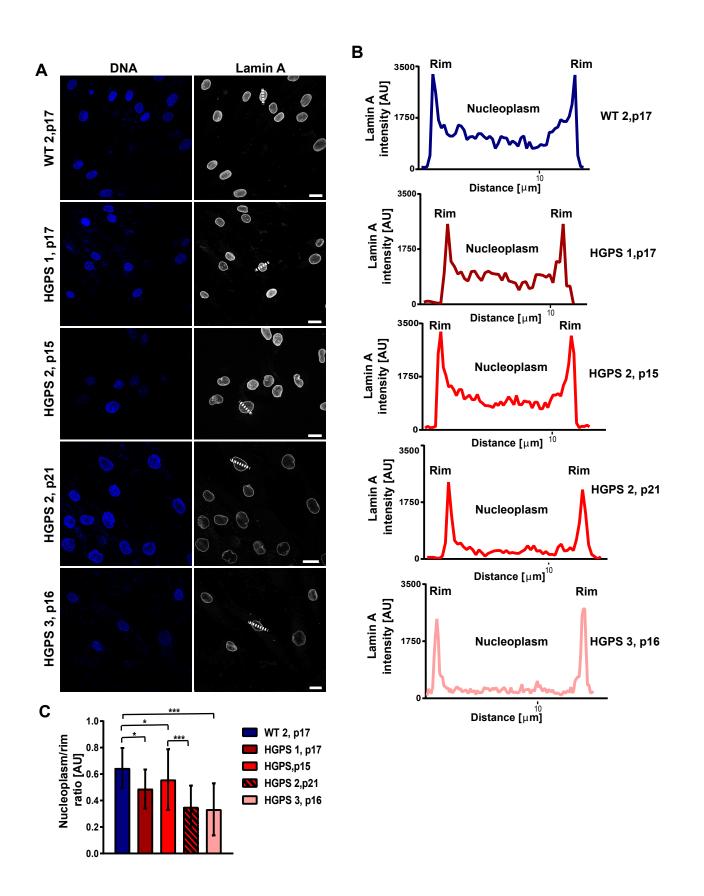
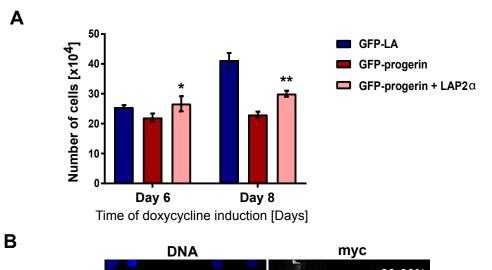
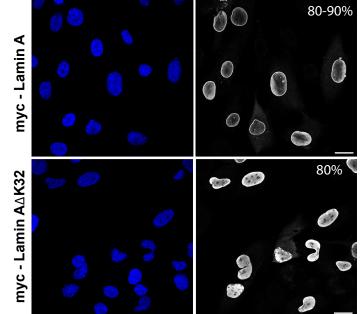
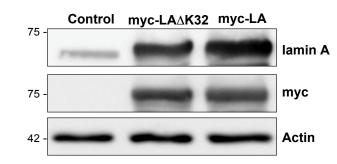


Figure S3. Nucleoplasmic pool of A-type lamins is not affected in low progerin-expressing cells. (A) Primary human fibroblasts were fixed with 4% PFA and processed for immunofluorescence using antibody specifically recognizing lamin A, not lamin C or progerin. DNA was stained with DAPI (blue). Scale bar: 20 μ m. (B) Fluorescence intensity of the lamin A signal was measured across nuclei (dotted line) and plotted. (C) Nucleoplasmic over rim ratio of the lamin A staining was measured in 100 nuclei each of one wild-type and three different HGPS cell lines and plotted in a histogram.









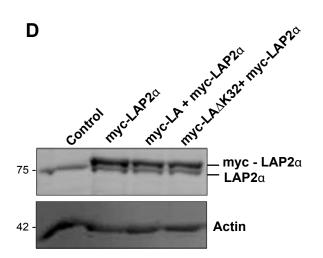


Figure S4. Ectopic expression of human myc-Lamin A, human myc-LaminΔK32 and human myc-LAP2*a* in hTERT-immortalized skin fibroblasts. (A) hTERT–immortalized fibroblasts inducibly expressing GFP-progerin were transfected either with a GFP-expressing (ctrl) or myc-LAP2*a* expressing construct on two consecutive days prior to the induction of progerin expression and the cell number was determined on days 6 and 8 post-induction (1.0 µg ml⁻¹ doxycycline, n=3). GFP-Lamin A expressing cells were used as control. (B) Immunofluorescence analysis of myc-Lamin A- (upper panel) and myc-LaminAΔK32- (lower panel) expression using α-myc antibody (grey) and DAPI (blue). Scale bar: 20 µm. The percentage indicates transfection efficiency. (C) Immunoblot analysis of total cell lysates of myc-Lamin Aand myc-LaminΔK32-expressing cells using anti-myc and anti-lamin A/C antibodies. *γ*-tubulin levels served as a loading control. (D) Uninduced cells were transfected either with a control GFP-expressing construct (GFP ctrl), myc-LAP2α- expressing construct alone or in combination with the myc-Lamin A and myc-LaminAΔK32 construct. The level of LAP2α expression was assessed by immunoblot analysis using anti-LAP2α specific antibody.