

Supporting Information

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Temperature Data

To obtain climate data for the years of historical surveys, we downsampled the 0.5° gridded time series of mean monthly temperatures from the University of East Anglia (CRU TS 3.10, ref. 1) for the whole Alpine arc to a resolution of 100 × 100 m for the period 1900–2014. Therefore, we statistically downsampled the 1-km WorldClim data (average values for the period 1950–2000, available online at www.worldclim.org) to 100 × 100 m using the methods applied in Dullinger et al. (2). Using the CRU dataset, we subsequently calculated anomalies between the yearly values and the average values for the reference period 1950–2000 used by WorldClim at the original 0.5° resolution. These anomalies were then spatially interpolated to 100 × 100 m and added to the downsampled WorldClim climatic maps to create a time series of absolute values at 100 × 100 m resolution.

We then assigned each plot a mean decadal temperature, i.e., the mean annual temperature of the 10 y preceding the historical survey. Subsequently, historical air temperature positions of rear edges, optima, and leading edges were calculated in the same manner as historical elevational positions (see *Statistical Analyses*). The relations between elevational shifts of rear edges, optima, and leading edges and their respective historical temperature positions as well as between proportional changes of elevational range size, abundances, their sum, and the historical temperature optima of the species were determined with linear regression models using each range attribute separately as response variable and using temperature as predictor.

For characterizing air temperature change between the historical and recent survey by one parameter, we calculated the difference between historical and mean decadal temperature assigning all historical plots the average sampling year (i.e., 1952).

Correlations and Skewness of Elevational Shifts

We calculated Pearson's correlation coefficients of shifts of rear edges, optima, and leading edges. The dynamics of rear and leading edges were significantly correlated with optimum shifts (rear–optimum: paired Pearson's $r = 0.35$, $df = 181$, $t = 5.06$, $P < 0.001$; leading–optimum: paired Pearson's $r = 0.34$, $df = 181$, $t = 4.92$, $P < 0.001$, Fig. S6) but rear and leading edge shifts were uncorrelated ($df = 181$, $t = 1.54$, $P = 0.127$).

Skewness of species' historical and recent distributions was calculated using the function *skewness* as implemented in the R-package *moments* (3). Changes were then derived by subtracting the historical from the recent value for each species separately. The average skewness of elevational distributions did not change (Fig. S6), and there is hence no indication that a “lean” type of range dynamics, with optima shifting at a different pace than one or both of the limits (4, 5), is prevalent among these species.

Species-Specific Traits

We compiled, in total, 10,046 records of species-specific traits from our own measurements, the online database TRY (6–40), the Tundra Trait Team, and literature (41–55). In a first step, we assessed averages for each trait and species from all available records. Then we calculated a compound index based on these traits, for both persistence- and dispersal-related traits (herein-

after named persistence and dispersal, respectively). Persistence was calculated as the first ordination axis using the function *dudi.mix* as implemented in the package *ade4* (56) on the traits dominance (five levels from scattered to dominant), life span (years), life strategy (competition, intermediate, stress), and number of vegetative offsprings per year (count). High values represent long-living, dominant, and competitive species that produce high numbers of vegetative offsprings. For dispersal, we first calculated seed retention times in several types of animal fur and seed survival in guts based on the traits seed surface structure and seed mass following Römermann et al. (52) and Mouissie et al. (57). Analogous to persistence, we then calculated dispersal as the first ordination axis using the function *dudi.pca* as implemented in the package *ade4* (56) on the traits terminal velocity (m/s), retention in fur of cattle, deer, rabbit, horse, sheep, and bear (% of seeds remaining in fur after 1 h), gut survival (reduction in germination rate), and seed release height (centimeters). Low values represent species that can disperse over longer distances more easily. We then tested for significant relationships between these two indicators and rear edge, optimum, and leading edge shifts, respectively, using linear regression models.

Pasture Data

We derived proportional areas used as pastures in decadal time steps from “The History Database of the Global Environment” (HYDE v. 3.1, refs. 58 and 59) provided by the Netherlands Environmental Assessment Agency with a spatial resolution of 5'. For our analyses, we used the decadal value of 1950 for all historical plots (see *Temperature Data*) and calculated proportional changes by subtracting the historical from the most recent value (2005) and dividing it by the historical. To analyze whether proportional changes of the area used as pasture were significantly different from zero, we applied an intercept-only linear regression model ($df = 1,548$, $t = -302.80$, $P < 0.001$).

Community Density, Community Richness, and Turnover of Cooccurring Species

Changes of community density were calculated as the difference between the recent and the historical total cover (in percent) of vascular plant species per plot. Changes in community richness were computed as the difference in the total number of vascular plant species per plot. For these analyses, we used all 1,070 recorded species. Changes of community density could only be calculated for 860 plots, since this information was not reported for all of the historical plots. Turnover of cooccurring species between the historical and the recent surveys was calculated for those 183 plant species for which whole range dynamics were available (but all species are considered to be potential cooccurring species of the 183 target species). Based on presence/absence data of species cooccurring with a focal species in any plot, we calculated Bray–Curtis dissimilarities using the function *vegdist* as implemented in the package *vegan* (60). Whether relations between changes of density and richness per plot and plot elevation as well as between turnover of cooccurring species per species and the historical optimum of the species were significant was tested with linear regression models.

1. Harris I, Jones PD, Osborn TJ, Lister DH (2014) Updated high-resolution grids of monthly climatic observations—The CRU TS3.10 Dataset. *Int J Climatol* 34: 623–642.
2. Dullinger S, et al. (2012) Extinction debt of high-mountain plants under twenty-first century climate change. *Nat Clim Chang* 2:619–622.
3. Komsta L, Novomestky F (2015) Moments: Moments, Cumulants, Skewness, Kurtosis and Related Tests, Version 0.14. Available at <http://CRAN.R-project.org/package=moments>. Accessed July 27, 2017.
4. Kelly AE, Goulden ML (2008) Rapid shifts in plant distribution with recent climate change. *Proc Natl Acad Sci USA* 105:11823–11826.

5. Lenoir J, Svenning JC (2015) Climate-related range shifts—A global multidimensional synthesis and new research directions. *Ecography* 38:15–28.
6. Kattge J, et al. (2011) TRY – A global database of plant traits. *Global Change Biology* 17:2905–2935.
7. Wohlfahrt G, et al. (1999) Inter-specific variation of the biochemical limitation to photosynthesis and related leaf traits of 30 species from mountain grassland ecosystems under different land use. *Plant Cell Environ* 22:1281–1296.
8. Chatz B, et al. (2012) Global convergence in the vulnerability of forests to drought. *Nature* 491:752–755.
9. Campetella G, et al. (2011) Patterns of plant trait-environment relationships along a forest succession chronosequence. *Agric Ecosyst Environ* 145:38–48.
10. Cioarlan V (2009) *The Illustrated Flora of Romania: Pteridophyta et Spermatoptya* (Editura Ceres, Bucharest, Romania).
11. Cornelissen JHC (1996) An experimental comparison of leaf decomposition rates in a wide range of temperate plant species and types. *J Ecol* 84:573–582.
12. Cornelissen JHC, et al. (2003) Functional traits of woody plants: Correspondence of species rankings between field adults and laboratory-grown seedlings? *J Veg Sci* 14:311–322.
13. Cornelissen JHC, et al. (2004) Leaf digestibility and litter decomposability are related in a wide range of subarctic plant species and types. *Funct Ecol* 18:779–786.
14. Cornelissen JHC, Diez PC, Hunt R (1996) Seedling growth, allocation and leaf attributes in a wide range of woody plant species and types. *J Ecol* 84:755–765.
15. Dainese M, Bragazza L (2012) Plant traits across different habitats of the Italian Alps: A comparative analysis between native and alien species. *Alp Bot* 122:11–21.
16. Diaz S, et al. (2004) The plant traits that drive ecosystems: Evidence from three continents. *J Veg Sci* 15:295–304.
17. Everwand G, Fry EL, Eggers T, Manning P (2014) Seasonal variation in the capacity for plant trait measures to predict grassland carbon and water fluxes. *Ecosystems (NY)* 17:1095–1108.
18. Fitter AH, Peat HJ (1994) The ecological flora database. *J Ecol* 82:415–425.
19. Fry EL, Power SA, Manning P (2014) Trait-based classification and manipulation of plant functional groups for biodiversity–ecosystem function experiments. *J Veg Sci* 25:248–261.
20. Garnier E, et al. (2007) Assessing the effects of land-use change on plant traits, communities and ecosystem functioning in grasslands: A standardized methodology and lessons from an application to 11 European sites. *Ann Bot* 99:967–985.
21. Green W (2009) USDA PLANTS compilation, version 1. *NRCS: The PLANTS Database* (Natl Plant Data Cent, Baton Rouge, LA).
22. Hickler T (1999) Plant functional types and community characteristics along environmental gradients on Öland's Great Alvar (Sweden). Master of Science (Univ Lund, Lund, Sweden).
23. Hill MO, Preston CD, Roy DB (2004) *PLANTATT–Attributes of British and Irish Plants: Status, Size, Life History, Geography and Habitats* (Cent Ecol Hydrol, Huntingdon, UK).
24. Kleyer M, et al. (2008) The LEDA Traitbase: A database of life-history traits of the Northwest European flora. *J Ecol* 96:1266–1274.
25. Klimešová J, de Bello F (2009) CLO-PLA: The database of clonal and bud bank traits of central European flora. *J Veg Sci* 20:511–516.
26. Kühn I, Durka W, Klotz S (2004) BiolFlor—A new plant-trait database as a tool for plant invasion ecology. *Divers Distrib* 10:363–365.
27. Moretti M, Legg C (2009) Combining plant and animal traits to assess community functional responses to disturbance. *Ecography* 32:299–309.
28. Medlyn BE, et al. (2001) Stomatal conductance of forest species after long-term exposure to elevated CO₂ concentration: A synthesis. *New Phytol* 149:247–264.
29. Milla R, Reich PB (2011) Multi-trait interactions, not phylogeny, fine-tune leaf size reduction with increasing altitude. *Ann Bot* 107:455–465.
30. Ordoñez JC, et al. (2010) Plant strategies in relation to resource supply in mesic to wet environments: Does theory mirror nature? *Am Nat* 175:225–239.
31. Paula S, et al. (2009) Fire-related traits for plant species of the Mediterranean Basin. *Ecology* 90:1420.
32. Prentice IC, et al. (2011) Evidence of a universal scaling relationship for leaf CO₂ drawdown along an aridity gradient. *New Phytol* 190:169–180.
33. Quested HM, et al. (2003) Decomposition of sub-Arctic plants with differing nitrogen economies: A functional role for hemiparasites. *Ecology* 84:3209–3221.
34. Royal Botanic Gardens KEW (2008) Seed Information Database (SID), version 7.1 (Roy Bot Gardens Kew, Wakehurst, UK).
35. Schweingruber FH, Landolt W (2005) *The Xylem Database* (Swiss Federal Institute for Forest, Snow and Landscape Research, Birmensdorf, Switzerland).
36. Gachet S, Véla E, Tatoni T (2005) BASECO: A floristic and ecological database of Mediterranean French flora. *Biodivers Conserv* 14:1023–1034.
37. Spasojevic MJ, Suding KN (2012) Inferring community assembly mechanisms from functional diversity patterns: The importance of multiple assembly processes. *J Ecol* 100:652–661.
38. Minden V, Andratschke S, Spalke J, Timmermann H, Kleyer M (2012) Plant trait-environment relationships in salt marshes: Deviations from predictions by ecological concepts. *Perspect Plant Ecol Evol Syst* 14:183–192.
39. Wirth C, Lichstein JW (2009) The imprint of species turnover on old-growth forest carbon balances—Insights from a trait-based model of forest dynamics. *Old-Growth Forests: Function, Fate and Value*, eds Wirth C, Gleixner G, Heimann M (Springer, Berlin), pp 81–113.
40. Wright IJ, et al. (2004) The worldwide leaf economics spectrum. *Nature* 428:821–827.
41. Hintze C, et al. (2013) D3: The dispersal and diaspore database—Baseline data and statistics on seed dispersal. *Perspect Plant Ecol Evol Syst* 15:180–192.
42. Landolt E, et al. (2010) *Flora Indicativa: Ecological Indicator Values and Biological Attributes of the Flora of Switzerland and the Alps* (Haupt, Bern, Switzerland), 2nd Ed.
43. Fischer MA, Oswald K, Adler W (2008) *Exkursionsflora für Österreich, Liechtenstein und Südtirol* (Biologiezentrum Oberösterreichischen Landesmuseen, Linz, Austria), 3rd Ed.
44. Kutschera L, Lichtenegger E (1982) *Wurzelatlas Mitteleuropäischer Grünlandpflanzen* (Gustav Fischer, Stuttgart, NY).
45. Müller-Schneider P (1986) Verbreitungsbioökologie der Blütenpflanzen Graubündens. *Veröffentlichungen Geobotanischen Instituts ETH Stiftung Rubel* 85:1–263.
46. Schroeter C (1926) *Das Pflanzenleben der Alpen* (Albert Raastein, Zurich).
47. Thompson K, Band SR, Hodgson JG (1993) Seed size and shape predict persistence in soil. *Funct Ecol* 7:236–241.
48. Rothmaler W (2002) *Exkursionsflora von Deutschland* (Spektrum Akademischer, Heidelberg), 9th Ed.
49. Hegi G (1966–2008) *Illustrierte Flora von Mitteleuropa* (Weissdorn, Jena, Germany).
50. Cerabolini B, Ceriani RM, Caccianiga M, Andreis RD, Raimondi B (2003) Seed size, shape and persistence in soil: A test on Italian flora from Alps to Mediterranean coasts. *Seed Sci Res* 13:75–85.
51. Tackenberg O (2001) *Methoden zur Bewertung gradueller Unterschiede des Ausbreitungspotentials von Pflanzenarten: Modellierung des Windausbreitungspotentials und regelbasierte Ableitung des Fernausbreitungspotentials* (J. Cramer, Berlin).
52. Römermann C, Tackenberg O, Poschlod P (2005) How to predict attachment potential of seeds to sheep and cattle coat from simple morphological seed traits. *Oikos* 110: 219–230.
53. Bruun HH, Österdahl S, Moen J, Angerbjörn A (2005) Distinct patterns in alpine vegetation around dens of the Arctic fox. *Ecography* 28:81–87.
54. Pluess AR, Schütz W, Stöcklin J (2005) Seed weight increases with altitude in the Swiss Alps between related species but not among populations of individual species. *Oecologia* 144:55–61.
55. Tamme R, et al. (2014) Predicting species' maximum dispersal distances from simple plant traits. *Ecology* 95:505–513.
56. Dray S, Dufour A-B (2007) The ade4 package: Implementing the duality diagram for ecologists. *J Stat Softw* 22:1–20.
57. Mouissie AM, Van der Veen CE, Veen GF, Van Diggelen R (2005) Ecological correlates of seed survival after ingestion by fallow deer. *Funct Ecol* 19:284–290.
58. Goldewijk KK, Beusen A, Janssen P (2010) Long-term dynamic modeling of global population and built-up area in a spatially explicit way: HYDE 3.1. *Holocene* 20: 565–573.
59. Goldewijk KK, Beusen A, van Drecht G, de Vos M (2011) The HYDE 3.1 spatially explicit database of human-induced global land-use change over the past 12,000 years. *Glob Ecol Biogeogr* 20:73–86.
60. Oksanen J, et al. (2016) vegan: Community Ecology Package, Version 2.3-5. Available at <http://CRAN.R-project.org/package=vegan>. Accessed August 10, 2016.

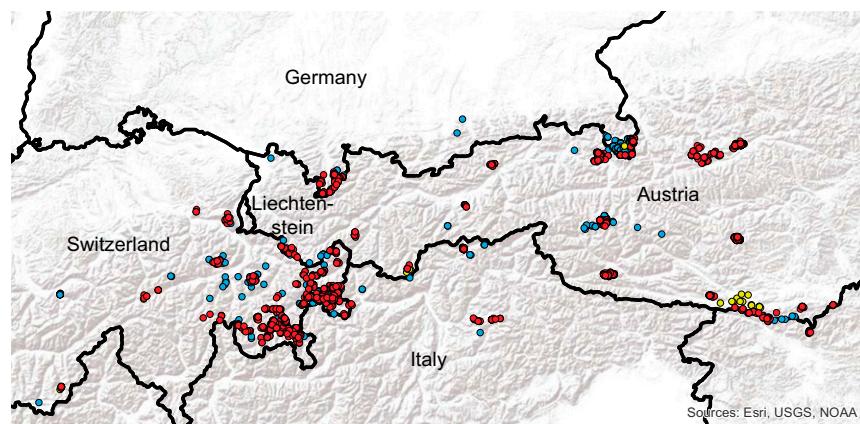


Fig. S1. Geographical distribution of plots; 1,549 resurveyed plots were used for plot-related analyses (red), 27 plots were resurveyed but not included in plot-related analyses (yellow, i.e., in total 1,576 resurveyed plots), and 435 plots were relocated but not resurveyed (blue), totaling 2011 relocated plots from 26 original publications (1–26).

1. Aichinger E (1933) *Vegetationskunde der Karawanken* (Verlag von Gustav Fischer, Jena, Germany).
2. Aichinger E (1958) Pflanzensoziologische Studien am Südfuß der Hochalmspitze. *Carinthia II* 68:120–139.
3. Braun-Blanquet G, Braun-Blanquet J (1931) Recherches phytogéographiques sur le massif du Gross Glockner (Hohe Tauern). *Rev Geogr Alp* 19:675–735.
4. Gumpelmayer F (1967) *Die Vegetation und ihre Gliederung in den Leoganger Steinbergen* (Univ Innsbruck, Innsbruck, Austria).
5. Hartl H (1963) Die Vegetation des Eisenhuts im Kärntner Nockgebiet. *Carinthia II* 73:293–336.
6. Höpflinger F (1957) Die Pflanzengesellschaften des Grimminggebietes. *Mitteilungen Naturwissenschaftliche Vereins Steiermark* 87:74–113.
7. Oberdorfer E (1959) Borstgras- und Krummseggenrasen in den Alpen. *Beiträge Naturkundlichen Forschung Südwestdeutschland* 18:117–143.
8. Pignatti-Wikus E (1960) Pflanzensoziologische Studien im Dachsteingebiet. *Beiträge Alpinen Karstforschung* 13:87–168.
9. Thimm I (1953) Die Vegetation des Sonnwendgebirges (Rofan) in Tirol (subalpine und alpine Stufe). *Berichte Naturwissenschaftlich Medizinischen Vereins Innsbruck* 50:5–166.
10. Wendelberger G (1953) Über einige hochalpine Pioniergesellschaften aus der Glockner- und Muntanitzgruppe in den Hohen Tauern. *Verhandlungen Zoologisch Botanischen Gesellschaft Österreichs* 93:100–109.
11. Wikus E (1952) Die Pflanzengesellschaften der Lienzer Dolomiten oberhalb der Baumgrenze. Doctoral thesis (Univ Vienna, Vienna).
12. Braun-Blanquet J, Braun-Blanquet G, Trepp W, Bach R, Richard F (1964) Pflanzensoziologische und bodenkundliche Beobachtungen im Samnaun. *Jahresbericht Naturforschenden Gesellschaft Graubündens* 90:3–48.
13. Braun-Blanquet J, Pallmann H, Bach R (1954) *Pflanzensoziologische und bodenkundliche Untersuchungen im Schweizerischen Nationalpark und seinen Nachbargebieten: Vegetation und Böden der Wald- und Zwergrauwaldgesellschaften (Vaccinio-Piceetalia)* (Nationalpark-Museum, Chur, Switzerland).
14. Flütsch P (1930) Über die Pflanzengesellschaften der alpinen Stufe des Berninagebietes. *Jahresbericht Naturforschenden Gesellschaft Graubündens* 68:5–59.
15. Lüdi W (1948) *Die Pflanzengesellschaften der Schinigeplatte bei Interlaken und ihre Beziehungen zur Umwelt: eine vergleichend ökologische Untersuchung* (Geobotanisches Forschungsinstitut Rübel, Zürich).
16. Wagner H (1944) Pflanzensoziologische Beobachtungen in der Ramsau bei Schladming. *Rundbrief der Zentralstelle für Vegetationskartierung des Reiches*, 14, Beilage.
17. Schnyder A (1930) Floristische und Vegetationsstudien im Alviergebiet. *Vierteljahrsschrift der Naturforschenden Gesellschaft in Zürich* 17(Jahrgang 75) (Beiblatt, Zürich).
18. Pallmann H, Haffter P (1933) Pflanzensoziologische und bodenkundliche Untersuchungen im Oberengadin: mit besonderer Berücksichtigung der Zwergrauwaldgesellschaften der Ordnung Rhodoreto-Vaccinietalia. *Berichte Schweizerischen Botanischen Gesellschaft* 42:357–466.
19. Zollitsch B (1968) Soziologische und ökologische Untersuchungen auf Kalkschiefern in hochalpinen Gebieten. Teil I. *Berichte Bayerischen Botanischen Gesellschaft* 40:67–100.
20. Albrecht J (1969) *Soziologische und ökologische Untersuchungen alpiner Rasengesellschaften insbesondere auf Kalk-Silikat-Gesteinen*. Dissertationes Botanicae (Cramer, Berlin).
21. Braun J (1913) Die Vegetationsverhältnisse der Schneestufe in den Rätisch-Leontischen Alpen: Ein Bild des Pflanzenlebens an seinen äußersten Grenzen. *Neue Denkschrift Schweizerischen Naturforschenden Gesellschaft* 48:1–339.
22. Braun-Blanquet J (1969) *Die Pflanzengesellschaften der rätischen Alpen im Rahmen ihrer Gesamtverbreitung. Teil I* (Bischofberger, Chur, Switzerland).
23. Braun-Blanquet J, Jenny J (1926) Vegetationsentwicklung und Bodenbildung in der alpinen Stufe der Zentralalpen (Klimaxgebiet des *Caricion curvulae*): mit besonderer Berücksichtigung der Verhältnisse im schweizerischen Nationalparkgebiet. *Denkschriften Schweizerischen Naturforschenden Gesellschaft* 63:183–349.
24. Wagner H (1965) Die Pflanzendecke der Komperdellalm in Tirol. *Documents Carte Vegetation Alpes* 3:7–59.
25. Lippert W (1966) Die Pflanzengesellschaften des Naturschutzgebietes Berchtesgaden. *Berichte Bayrischen Botanischen Gesellschaft* 39:67–122.
26. Oberdorfer E (1950) Beitrag zur Vegetationskunde des Allgäu. *Beiträge Naturkundlichen Forschung Südwestdeutschland* 9:29–98.

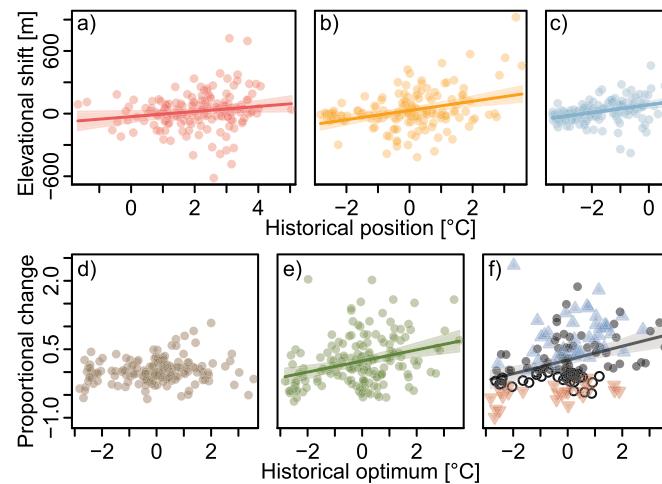


Fig. S2. Relationships between changes of range attributes and the historical ambient air temperature of these attributes for 162 mountain plant species of the European Alps. (A) Rear edges, (B) optima, (C) leading edges, (D) elevational range size, (E) abundance, and (F) sum of proportional elevational range size and abundance changes. Lines and their shades represent significant linear regression models (Table S2) with their confidence intervals. In F, blue pyramids depict winners (i.e., species with increased elevational ranges and increased abundances), red inverted pyramids depict losers (i.e., those with decreases in both of these range attributes), and dots symbolize species which combine loss in one attribute with gain in the other one. Closed darker dots can be considered as net winners (gain in one attribute > loss in the other one), and open dots can be considered as net losers (gain in one attribute < loss in the other one). Panels differ in size due to differing temperature ranges of the respective historical positions.

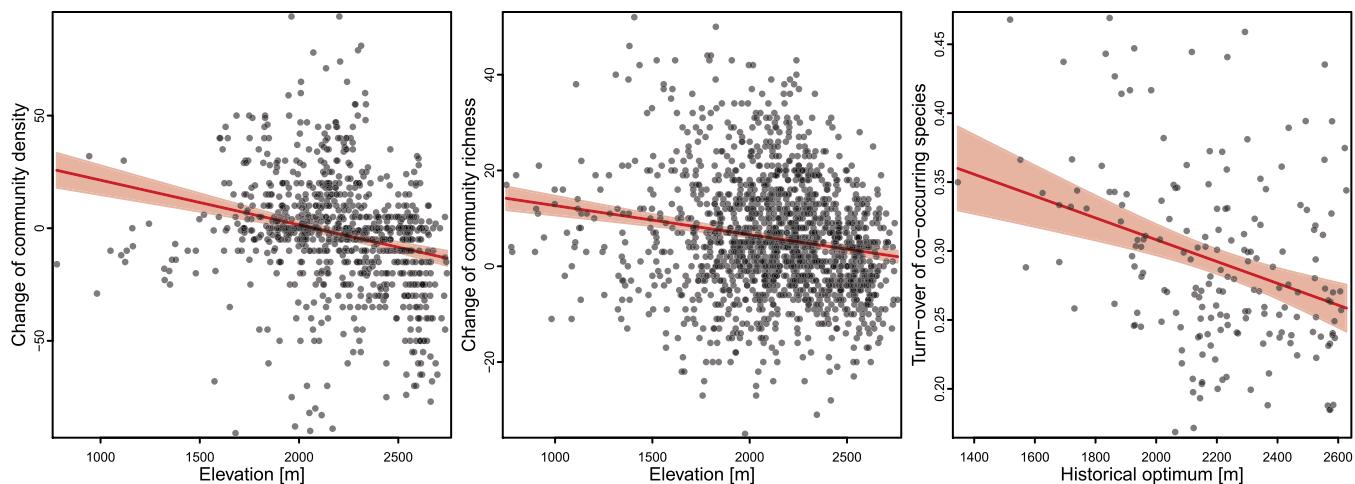


Fig. S3. Relationships between community changes and elevation. Lines and their shades represent significant linear regression models (Table S3) with their confidence intervals. Change of community density is based on the total vascular plant cover (in percent) of 860 resurveyed plots; change of community richness on the total number of vascular plant species of 1,549 resurveyed plots; turnover of cooccurring species was computed for those 183 mountain plant species for which elevational range dynamics could be calculated. Individual records are depicted as grey dots and are darker with more records at the same position.

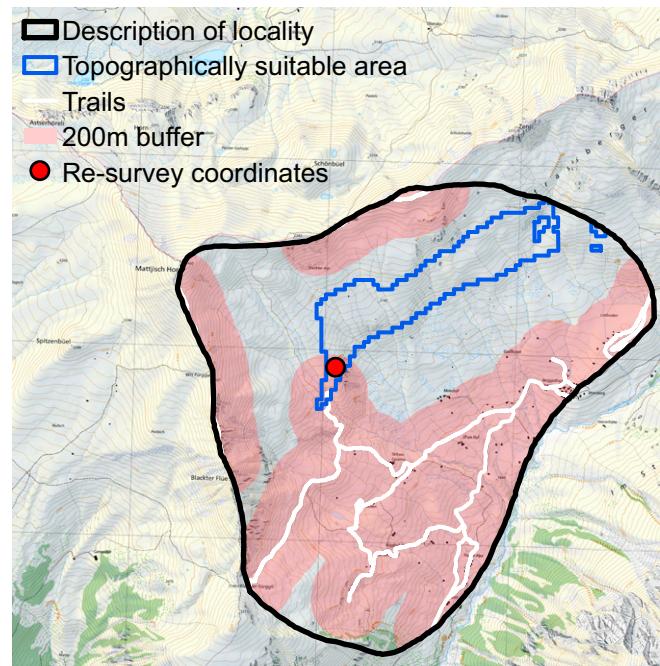


Fig. S4. Relocation design exemplified for one plot. Topographic background map is derived from the Federal Office of Topography swisstopo, available online at www.swisstopo.ch.

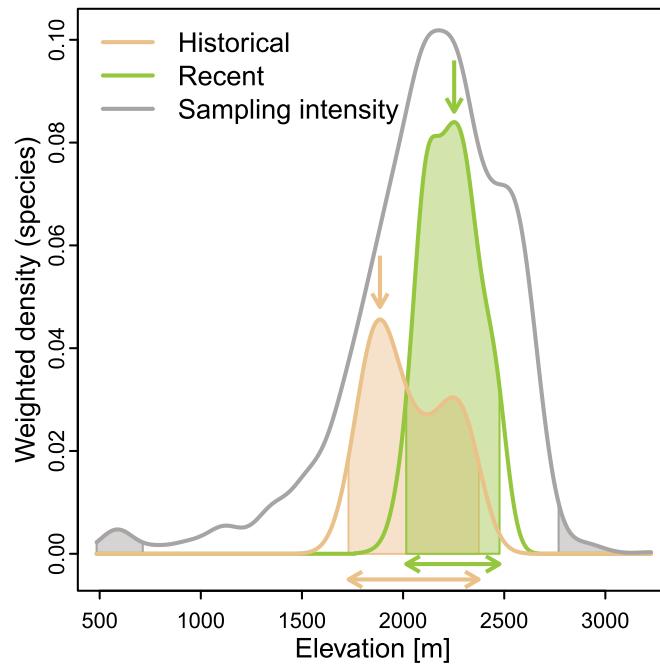


Fig. S5. Schematic illustration of the calculation of range attributes. Colored lines represent calculated density distributions of the species; the gray line represents the density distribution of the sampling intensity with reduced density values to fit the plot. Species abundance is defined as the sum of all density values below the respective curve, i.e., as the “integral” of the density distribution. Areas between the 5% and 95% quantiles of the species abundance are shaded in color, with their lower vertical boundary representing the species’ rear edge and the upper vertical boundary representing its leading edge. Vertical arrows show the position of the species’ optimum, i.e., the adapted density distributions’ peak. Horizontal arrows depict the species’ elevational range size. Gray shaded areas depict the 1% and 99% quantile of the sampling intensities distribution.

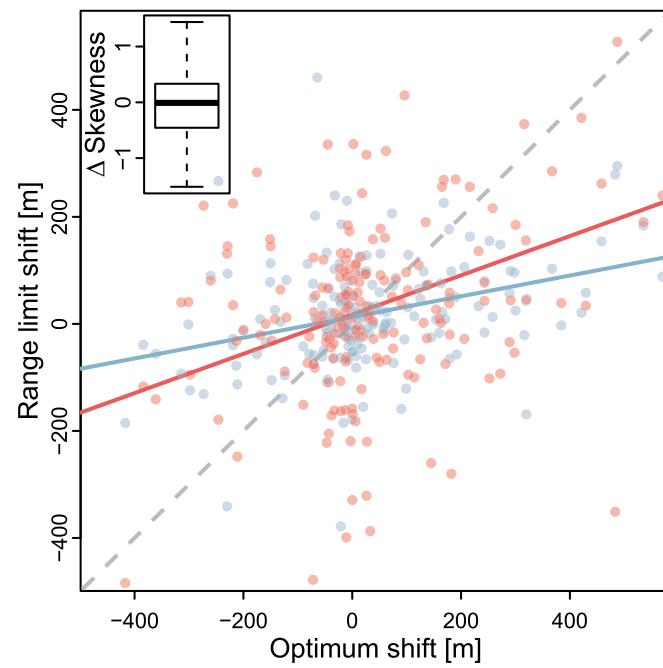


Fig. S6. Relationships between elevational shifts of range limits and optima for 183 mountain plant species of the European Alps. Rear edge shifts are depicted as red dots, and leading edge shifts are depicted as blue dots. Lines in the respective colors represent linear regression models. Dots above the diagonal dashed line represent species with range limits shifting faster than their optimum and below those with faster optimum shifts. (*Inset*) The boxplot depicts changes of optimum skewness.

Table S1. Changes of range attributes for 183 mountain plant species of the European Alps

Species	Rear edge shift, m	Optimum shift, m	Leading edge shift, m	Proportional elevational range size change	Proportional abundance change
<i>Achillea atrata</i>	373	316	-21	-0.31	0.26
<i>Achillea clavennae</i>	59	10	-38	-0.09	0.14
<i>Acinos alpinus</i>	-44	-39	21	0.07	0.77
<i>Adenostyles alliariae</i>	-122	17	-46	0.12	1.10
<i>Agrostis alpina</i>	-11	7	-60	-0.07	0.60
<i>Ajuga pyramidalis</i>	-141	-361	-56	0.10	0.65
<i>Androsace chamaejasme</i>	-24	6	-21	0.00	0.34
<i>Androsace obtusifolia</i>	-32	-213	-78	-0.08	0.08
<i>Anemonastrum narcissiflorum</i>	262	458	154	-0.15	0.70
<i>Arabis bellidifolia s.lat.</i>	-280	182	98	0.43	0.21
<i>Arabis caerulea</i>	-28	39	3	0.07	0.54
<i>Arctostaphylos alpinus</i>	158	38	-1	-0.28	-0.45
<i>Arctostaphylos uva-ursi</i>	285	367	103	-0.28	0.48
<i>Arenaria ciliata s.str.</i>	-70	-12	-8	0.10	-0.37
<i>Aster alpinus</i>	-3	-18	-15	-0.01	-0.44
<i>Avenella flexuosa</i>	-28	206	106	0.22	0.10
<i>Avenula versicolor</i>	-67	63	22	0.11	0.14
<i>Bartsia alpina</i>	39	384	-3	-0.05	0.35
<i>Betonica alopecuros</i>	132	-21	-378	-0.49	0.40
<i>Botrychium lunaria</i>	63	102	31	-0.03	0.08
<i>Calamagrostis villosa</i>	-62	-11	-4	0.13	0.00
<i>Campanula barbata</i>	125	16	-10	-0.16	0.05
<i>Carduus defloratus s.lat.</i>	0	-159	-34	-0.03	0.83
<i>Carex atrata agg.</i>	-109	-43	6	0.16	0.66
<i>Carex capillaris</i>	-95	-298	-124	-0.03	0.42
<i>Carex ericetorum</i>	-75	-71	-58	0.02	-0.34
<i>Carex ferruginea</i>	527	487	295	-0.19	0.72
<i>Carex firma</i>	16	31	-18	-0.05	0.15
<i>Carex mucronata</i>	-399	-11	55	0.59	-0.50
<i>Carex ornithopoda agg.</i>	-151	-90	-42	0.11	1.29
<i>Carex rupestris</i>	22	-71	-75	-0.20	-0.64
<i>Carex sempervirens</i>	-24	-79	-56	-0.04	0.57
<i>Cerastium arvense</i>	85	80	81	0.00	0.12
<i>Cerastium carinthiacum</i>	-387	33	24	0.58	-0.04
<i>Cerastium uniflorum</i>	156	319	43	-0.24	0.11
<i>Chaerophyllum villarsii</i>	283	-175	-57	-0.41	0.95
<i>Chamorchis alpina</i>	-73	-82	112	0.34	-0.47
<i>Cirsium acaule</i>	78	13	126	0.08	0.94
<i>Cirsium spinosissimum</i>	-163	-21	38	0.31	2.03
<i>Coeloglossum viride</i>	-171	-38	-1	0.27	1.11
<i>Crepis jacquinii s.lat.</i>	-260	145	-25	0.35	-0.48
<i>Cystopteris fragilis</i>	12	-121	82	0.09	-0.05
<i>Daphne mezereum</i>	44	291	99	0.07	0.43
<i>Daphne striata</i>	35	-219	-28	-0.10	0.05
<i>Dianthus sylvestris</i>	12	-23	44	0.04	-0.46
<i>Draba aizoides agg.</i>	87	-22	-63	-0.20	-0.11
<i>Draba hoppeana</i>	89	-10	-2	-0.36	-0.20
<i>Dryas octopetala</i>	93	24	22	-0.10	0.25
<i>Empetrum nigrum agg.</i>	81	1	53	-0.06	-0.10
<i>Erigeron alpinus agg.</i> (incl. <i>uniflorus</i>)	111	6	-11	-0.13	-0.29
<i>Euphrasia minima</i>	225	-219	-40	-0.26	0.32
<i>Euphrasia officinalis</i>	50	-20	186	0.21	1.59
<i>Euphrasia salisburgensis</i>	-162	-33	51	0.26	0.82
<i>Festuca alpina</i>	-46	75	71	0.16	0.07
<i>Festuca halleri agg.</i>	-24	-47	133	0.27	0.50
<i>Festuca pumila</i>	16	-55	-61	-0.11	0.49
<i>Festuca varia agg.</i>	96	99	-121	-0.22	0.56
<i>Festuca violacea agg.</i>	-219	-3	-31	0.26	0.36
<i>Galium pusillum agg.</i>	21	-15	-17	-0.04	0.46
<i>Gentiana acaulis</i>	68	-55	34	-0.04	0.08
<i>Gentiana brachyphylla</i>	-82	-69	11	0.23	-0.19

Table S1. Cont.

Species	Rear edge shift, m	Optimum shift, m	Leading edge shift, m	Proportional elevational range size change	Proportional abundance change
<i>Gentiana nivalis</i>	161	60	-2	-0.17	-0.31
<i>Gentiana orbicularis</i>	-158	0	-52	0.27	-0.46
<i>Gentiana punctata</i>	-7	-26	7	0.02	0.02
<i>Gentianella anisodonta</i>	269	168	226	-0.07	0.64
<i>Gentianella aspera</i>	54	-10	230	0.31	-0.17
<i>Gentianella campestris</i>	158	-151	78	-0.09	-0.19
<i>Geranium sylvaticum</i>	-27	137	71	0.11	0.42
<i>Geum montanum</i>	97	-14	69	-0.03	0.25
<i>Globularia nudicaulis</i>	-484	-417	-185	0.33	0.39
<i>Gnaphalium hoppeanum/supinum</i>	216	258	13	-0.24	-0.05
<i>Hedysarum hedsyaroides</i>	-37	-34	-65	-0.04	0.10
<i>Helianthemum alpestre</i>	81	59	-13	-0.13	0.10
<i>Heracleum austriacum</i>	40	-314	-93	-0.17	1.35
<i>Hieracium alpinum</i>	77	243	80	0.00	-0.12
<i>Hieracium glanduliferum</i>	-182	6	31	0.39	-0.30
<i>Hieracium lactucella</i>	2	163	76	0.10	-0.54
<i>Hieracium pilosum/villosum</i>	53	-68	202	0.19	-0.24
<i>Homogyne alpina</i>	-20	179	49	0.09	0.43
<i>Homogyne discolor</i>	256	216	163	-0.15	2.02
<i>Huperzia selago</i>	316	26	128	-0.19	-0.33
<i>Hypochaeris uniflora</i>	34	429	58	0.04	-0.28
<i>Juncus jacquinii</i>	35	-7	-26	-0.12	0.18
<i>Juncus monanthos</i>	41	-301	-1	-0.05	1.54
<i>Juncus trifidus</i>	120	-12	52	-0.10	0.00
<i>Juniperus communis</i>	110	122	65	-0.07	0.24
<i>Kobresia myosuroides</i>	-64	-35	-17	0.09	-0.24
<i>Laserpitium halleri</i>	-89	-124	-96	-0.01	-0.41
<i>Leontodon helveticus</i>	30	-10	37	0.01	0.43
<i>Leontodon montanus</i>	2	131	98	0.13	-0.07
<i>Leontopodium alpinum</i>	60	-7	-42	-0.12	-0.40
<i>Ligusticum mutellina</i>	-31	-148	-105	-0.09	0.79
<i>Ligusticum mutellinoides</i>	66	-45	-104	-0.25	-0.03
<i>Loiseleuria procumbens</i>	104	-23	113	0.02	-0.22
<i>Lonicera caerulea</i>	-3	-16	-185	-0.34	-0.54
<i>Luzula alpinopilosa</i>	51	73	206	0.31	0.82
<i>Luzula lutea</i>	256	165	22	-0.31	0.08
<i>Luzula sylvatica</i>	-222	-47	-109	0.19	0.25
<i>Minuartia verna</i> agg.	-5	10	11	0.02	-0.20
<i>Myosotis alpestris</i>	-29	-42	5	0.03	0.10
<i>Myosotis sylvatica</i>	124	-71	48	-0.12	-0.31
<i>Nigritella nigra</i> agg.	-19	-33	102	0.20	-0.20
<i>Oxalis acetosella</i>	-329	0	-179	0.24	-0.03
<i>Oxytropis campestris</i>	-161	-13	-66	0.16	-0.42
<i>Oxytropis montana</i> agg.	-24	-12	54	0.12	0.28
<i>Pedicularis asplenifolia</i>	111	7	-82	-0.57	-0.42
<i>Pedicularis rostratocapitata</i>	42	14	-24	-0.08	-0.19
<i>Pedicularis tuberosa</i>	178	52	131	-0.08	-0.36
<i>Pedicularis verticillata</i>	185	300	25	-0.19	-0.26
<i>Persicaria vivipara</i>	9	160	-17	-0.03	0.12
<i>Peucedanum ostruthium</i>	-24	20	-155	-0.17	0.93
<i>Phleum alpinum</i> agg.	-54	298	96	0.21	0.74
<i>Phleum hirsutum</i>	131	-230	-341	-0.53	0.30
<i>Phyteuma betonicifolium</i>	58	179	-65	-0.17	-0.11
<i>Phyteuma hemisphaericum</i>	9	-142	26	0.03	-0.03
<i>Phyteuma sieberi</i>	-71	50	59	0.26	-0.34
<i>Pinus mugo</i> agg.	-617	-128	-139	0.72	0.73
<i>Plantago alpina</i>	145	-150	113	-0.03	0.81
<i>Plantago atrata</i>	46	320	-169	-0.27	0.69

Table S1. Cont.

Species	Rear edge shift, m	Optimum shift, m	Leading edge shift, m	Proportional elevational range size change	Proportional abundance change
<i>Plantago strictissima</i>	20	-32	240	0.40	0.53
<i>Poa nemoralis</i>	5	-64	460	1.08	0.15
<i>Polygala alpestris</i>	-21	-169	9	0.04	0.31
<i>Potentilla aurea</i>	64	-132	4	-0.07	0.58
<i>Potentilla brauneana</i>	695	923	121	-0.50	1.59
<i>Potentilla crantzii</i>	-117	-384	-39	0.08	-0.09
<i>Potentilla grandiflora</i>	43	243	-16	-0.07	-0.33
<i>Primula auricula</i>	-75	171	9	0.09	-0.56
<i>Primula hirsuta</i>	-168	1	-52	0.21	0.00
<i>Primula integrifolia</i>	93	25	-66	-0.27	-0.15
<i>Primula minima</i>	-34	-48	-10	0.04	0.03
<i>Primula wulfeniana</i>	-11	-200	-13	0.00	0.05
<i>Pseudorchis albida</i>	102	-7	34	-0.10	-0.28
<i>Pulsatilla alpina s.lat.</i>	336	2	-12	-0.43	-0.17
<i>Pulsatilla vernalis</i>	270	190	50	-0.26	0.09
<i>Ranunculus alpestris</i>	385	421	21	-0.30	0.21
<i>Ranunculus hybridus</i>	-63	45	33	0.15	0.22
<i>Rhinanthus glacialis</i>	-351	483	279	0.79	0.24
<i>Rhododendron ferrugineum</i>	6	37	73	0.14	0.29
<i>Rhododendron hirsutum</i>	73	139	73	0.00	1.05
<i>Rumex alpestris</i>	3	-54	17	0.01	0.78
<i>Rumex scutatus</i>	38	178	177	0.13	-0.05
<i>Salix alpina</i>	-95	103	129	0.48	-0.55
<i>Salix reticulata</i>	721	192	-57	-0.59	1.11
<i>Salix retusa agg.</i>	335	-45	-16	-0.34	0.37
<i>Saponaria pumila</i>	136	232	33	-0.21	0.00
<i>Saussurea alpina</i>	-220	27	-8	0.44	-0.57
<i>Saxifraga aizoides</i>	-102	252	155	0.44	1.35
<i>Saxifraga androsacea</i>	768	71	2	-0.52	-0.31
<i>Saxifraga caesia</i>	94	6	21	-0.09	-0.04
<i>Saxifraga paniculata</i>	427	96	-29	-0.36	-0.20
<i>Saxifraga rudolphiana</i>	323	62	2	-0.69	0.15
<i>Selaginella selaginoides</i>	-16	76	-73	-0.06	0.83
<i>Sempervivum montanum</i>	27	154	8	-0.03	0.33
<i>Senecio abrotanifolius</i>	-205	-43	67	0.40	0.30
<i>Senecio carniolicus</i>	132	-8	-68	-0.33	0.27
<i>Sesleria ovata</i>	184	-8	-26	-0.40	-0.41
<i>Sesleria sphaerocephala</i>	-93	272	148	0.49	-0.42
<i>Sibbaldia procumbens</i>	145	-229	94	-0.07	-0.08
<i>Silene acaulis s.lat.</i>	9	-2	17	0.01	0.17
<i>Silene alpestris</i>	190	535	184	-0.01	0.90
<i>Silene rupestris</i>	91	52	25	-0.08	-0.34
<i>Soldanella alpina</i>	39	90	-159	-0.20	1.03
<i>Soldanella minima agg.</i>	240	570	88	-0.20	1.52
<i>Soldanella pusilla</i>	120	-53	-50	-0.22	0.25
<i>Thesium alpinum</i>	-107	-22	5	0.12	0.11
<i>Thlaspi rotundifolium</i>	11	30	33	0.03	-0.21
<i>Trifolium alpinum</i>	190	135	72	-0.16	-0.06
<i>Trifolium badium</i>	-248	-211	-113	0.23	0.63
<i>Trifolium thalii</i>	-100	114	14	0.14	0.21
<i>Trisetum alpestre</i>	-14	15	96	0.30	1.00
<i>Trollius europaeus</i>	-66	125	47	0.11	0.39
<i>Vaccinium myrtillus</i>	-34	289	57	0.16	0.29
<i>Vaccinium uliginosum agg.</i>	173	-5	81	-0.14	0.02
<i>Valeriana celtica</i>	52	2	12	-0.08	0.32
<i>Valeriana montana</i>	-478	-72	75	0.59	0.60
<i>Valeriana tripteris</i>	-179	-246	267	0.90	0.13
<i>Veronica alpina</i>	-19	57	-8	0.01	0.00
<i>Veronica aphylla</i>	221	-273	-131	-0.29	1.39

Table S1. Cont.

Species	Rear edge shift, m	Optimum shift, m	Leading edge shift, m	Proportional elevational range size change	Proportional abundance change
<i>Veronica bellidoides</i>	244	18	11	-0.28	-0.34
<i>Veronica fruticans</i>	81	-260	90	0.01	0.31
<i>Veronica serpyllifolia</i>	-321	26	99	0.64	0.33
<i>Viola biflora</i>	54	-13	42	-0.01	0.60
<i>Viola calcarata</i>	33	25	24	-0.01	0.25

Nomenclature follows Fischer et al. (1), supplemented by Lauber et al. (2).

1. Fischer MA, Oswald K, Adler W (2008) *Exkursionsflora für Österreich, Liechtenstein und Südtirol* (Biologiezentrum Oberösterreichischen Landesmuseen, Linz, Austria), 3rd Ed.
2. Lauber K, Wagner G, Gygax A (2012) *Flora Helvetica* (Haupt, Bern, Switzerland), 5th Ed.

Table S2. Relationships between changes of range attributes and different predictor variables for mountain plant species of the European Alps

Range attribute	Predictor variable	Effect \pm SE	df	t value	P value	R ²
Elevation						
Rear edge shift, m	(Intercept)	320.8 \pm 94.3				
	Historical rear edge, m	-0.2 \pm 0.1	181	-3.12	0.002	0.05
Optimum shift, m	(Intercept)	642.1 \pm 103.0				
	Historical optimum, m	-0.3 \pm 0.0	181	-5.94	<0.001	0.16
Leading edge shift, m	(Intercept)	550.7 \pm 93.8				
	Historical leading edge, m	-0.2 \pm 0.0	181	-5.68	<0.001	0.15
Proportional elevational range size change	(Intercept)	0.3 \pm 0.2				
	Historical optimum, m	-1.3 ^{e-4} \pm 0.8 ^{e-4}	181	-1.78	0.076	0.01
Proportional abundance change	(Intercept)	1.5 \pm 0.3				
	Historical optimum, m	-6.0 ^{e-4} \pm 1.4 ^{e-4}	181	-4.26	<0.001	0.09
Sum of proportional elevational range size and proportional abundance changes	(Intercept)	1.9 \pm 0.3				
	Historical optimum, m	-7.4 ^{e-4} \pm 1.6 ^{e-4}	181	-4.70	<0.001	0.10
Air temperature						
Rear edge shift, m	(Intercept)	-28.7 \pm 27.3				
	Historical rear edge, °C	24.5 \pm 11.5	160	2.13	0.035	0.02
Optimum shift, m	(Intercept)	30.0 \pm 13.3				
	Historical optimum, °C	44.1 \pm 9.8	160	4.51	<0.001	0.11
Leading edge shift, m	(Intercept)	84.4 \pm 14.9				
	Historical leading edge, °C	37.4 \pm 7.3	160	5.14	<0.001	0.14
Proportional elevational range size change	(Intercept)	2.2 ^{e-2} \pm 2.1 ^{e-2}				
	Historical optimum, °C	2.9 ^{e-2} \pm 1.5 ^{e-2}	160	1.92	0.056	0.02
Proportional abundance change	(Intercept)	24.3 ^{e-2} \pm 4.0 ^{e-2}				
	Historical optimum, °C	12.2 ^{e-2} \pm 3.0 ^{e-2}	160	4.12	<0.001	0.09
Sum of proportional elevational range size and proportional abundance changes	(Intercept)	26.5 ^{e-2} \pm 4.3 ^{e-2}				
	Historical optimum, °C	15.1 ^{e-2} \pm 3.1 ^{e-2}	160	4.82	<0.001	0.12
Temperature indicator value						
Rear edge shift, m	(Intercept)	150.5 \pm 45.2				
	Temperature indicator	-67.9 \pm 24.1	176	-2.82	0.005	0.04
Optimum shift, m	(Intercept)	30.5 \pm 44.4				
	Temperature indicator	2.9 \pm 23.7	176	0.12	0.903	0.00
Leading edge shift, m	(Intercept)	-35.5 \pm 24.6				
	Temperature indicator	31.2 \pm 13.1	176	2.37	0.019	0.03
Proportional elevational range size change	(Intercept)	-24.5 ^{e-2} \pm 6.3 ^{e-2}				
	Temperature indicator	14.4 ^{e-2} \pm 3.4 ^{e-2}	176	4.25	<0.001	0.09
Proportional abundance change	(Intercept)	9.2 ^{e-2} \pm 12.7 ^{e-2}				
	Temperature indicator	7.2 ^{e-2} \pm 6.8 ^{e-2}	176	1.06	0.289	0.00
Sum of proportional elevational range size and proportional abundance changes	(Intercept)	-15.3 ^{e-2} \pm 14.0 ^{e-2}				
	Temperature indicator	21.6 ^{e-2} \pm 7.5 ^{e-2}	176	2.90	0.004	0.04
Nutrient indicator value						
Rear edge shift, m	(Intercept)	49.5 \pm 51.6				
	Nutrient indicator	-8.6 \pm 23.1	179	-0.37	0.710	0.00
Optimum shift, m	(Intercept)	-13.4 \pm 49.2				
	Nutrient indicator	21.6 \pm 22.0	179	0.98	0.328	0.00
Leading edge shift, m	(Intercept)	42.0 \pm 27.9				
	Nutrient indicator	-10.6 \pm 12.5	179	-0.85	0.396	0.00
Proportional elevational range size change	(Intercept)	-1.3 ^{e-2} \pm 7.4 ^{e-2}				
	Nutrient indicator	1.0 ^{e-2} \pm 3.3 ^{e-2}	179	0.29	0.775	0.01
Proportional abundance change	(Intercept)	-46.8 ^{e-2} \pm 13.4 ^{e-2}				
	Nutrient indicator	31.9 ^{e-2} \pm 6.0 ^{e-2}	179	5.34	<0.001	0.13
Sum of proportional elevational range size and proportional abundance changes	(Intercept)	-48.0 ^{e-2} \pm 15.1 ^{e-2}				
	Nutrient indicator	32.9 ^{e-2} \pm 6.8 ^{e-2}	179	4.86	<0.001	0.11
Persistence- and dispersal-related traits						
Rear edge shift, m	(Intercept)	29.8 \pm 13.9				
	Persistence	-7.2 \pm 10.3	181	-0.69	0.489	0.00
Optimum shift, m	(Intercept)	33.9 \pm 13.4				
	Persistence	1.0 \pm 9.9	181	0.10	0.923	0.01

Table S2. Cont.

Range attribute	Predictor variable	Effect \pm SE	df	t value	P value	R^2
Optimum shift, m	(Intercept)	33.9 \pm 13.4	181	0.03	0.973	0.01
	Dispersal	0.3 \pm 8.5				
Leading edge shift, m	(Intercept)	19.5 \pm 7.5	181	-1.54	0.126	0.01
	Dispersal	-7.3 \pm 4.7				

Relationships were calculated using linear regression models. The number of observations differs between predictor variables due to data availability. Temperature indicator values: 1, alpine to nival; 1.5, lower alpine to upper subalpine; 2, subalpine; 2.5, lower subalpine to upper montane; 3, montane; 3.5, lower montane to upper colline; 4, colline. Nutrient indicator values: 1, very nutrient-poor; 2, nutrient-poor; 3, moderately nutrient-poor to moderately nutrient-rich; 4, nutrient-rich.

Table S3. Relationships between community changes and elevation

Type of community change	Elevation	Effect \pm SE	df	t value	P value	R^2
Change of community density	(Intercept)	41.2 \pm 6.0	858	-7.36	<0.001	0.06
	Plot elevation, m	-2.0 ^{e-2} \pm 0.3 ^{e-2}				
Change of community richness	(Intercept)	18.7 \pm 1.9	1,547	-6.88	<0.001	0.03
	Plot elevation, m	-6.1 ^{e-3} \pm 0.9 ^{e-3}				
Turnover of cooccurring species	(Intercept)	4.7 ^{e-1} \pm 0.4 ^{e-1}	181	-4.60	<0.001	0.10
	Historical optimum, m	-7.9 ^{e-5} \pm 1.7 ^{e-5}				

Change of community density is based on the total vascular plant cover (in percent) of 860 resurveyed plots; change of community richness is based on the total number of vascular plant species of 1,549 resurveyed plots; turnover of cooccurring species was computed for those 183 mountain plant species for which elevational range dynamics could be calculated.