

## **SUPPLEMENTAL MATERIALS**

### **Title:**

Deletion of the Syncytin A receptor Ly6e impairs syncytiotrophoblast fusion and placental morphogenesis causing embryonic lethality in mice.

### **Authors:**

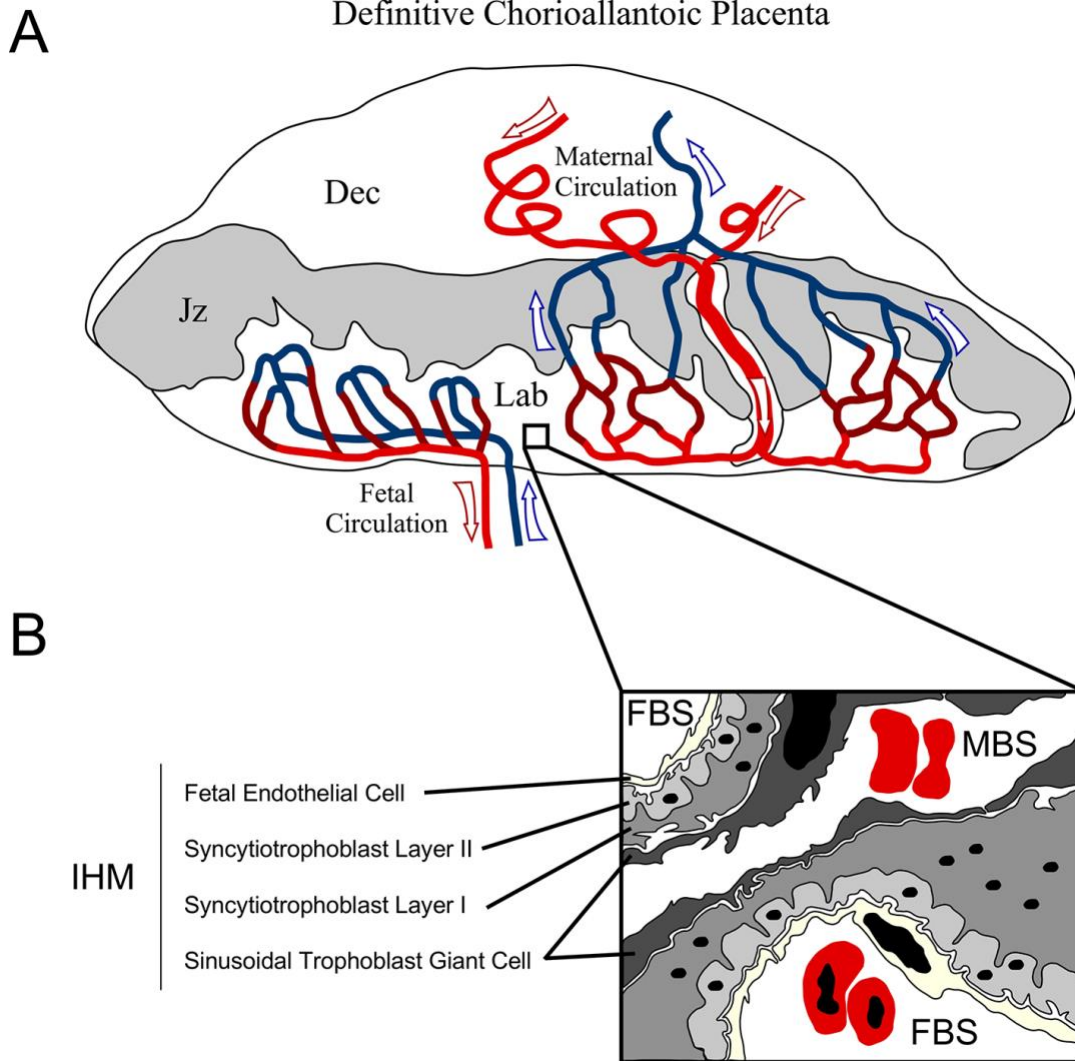
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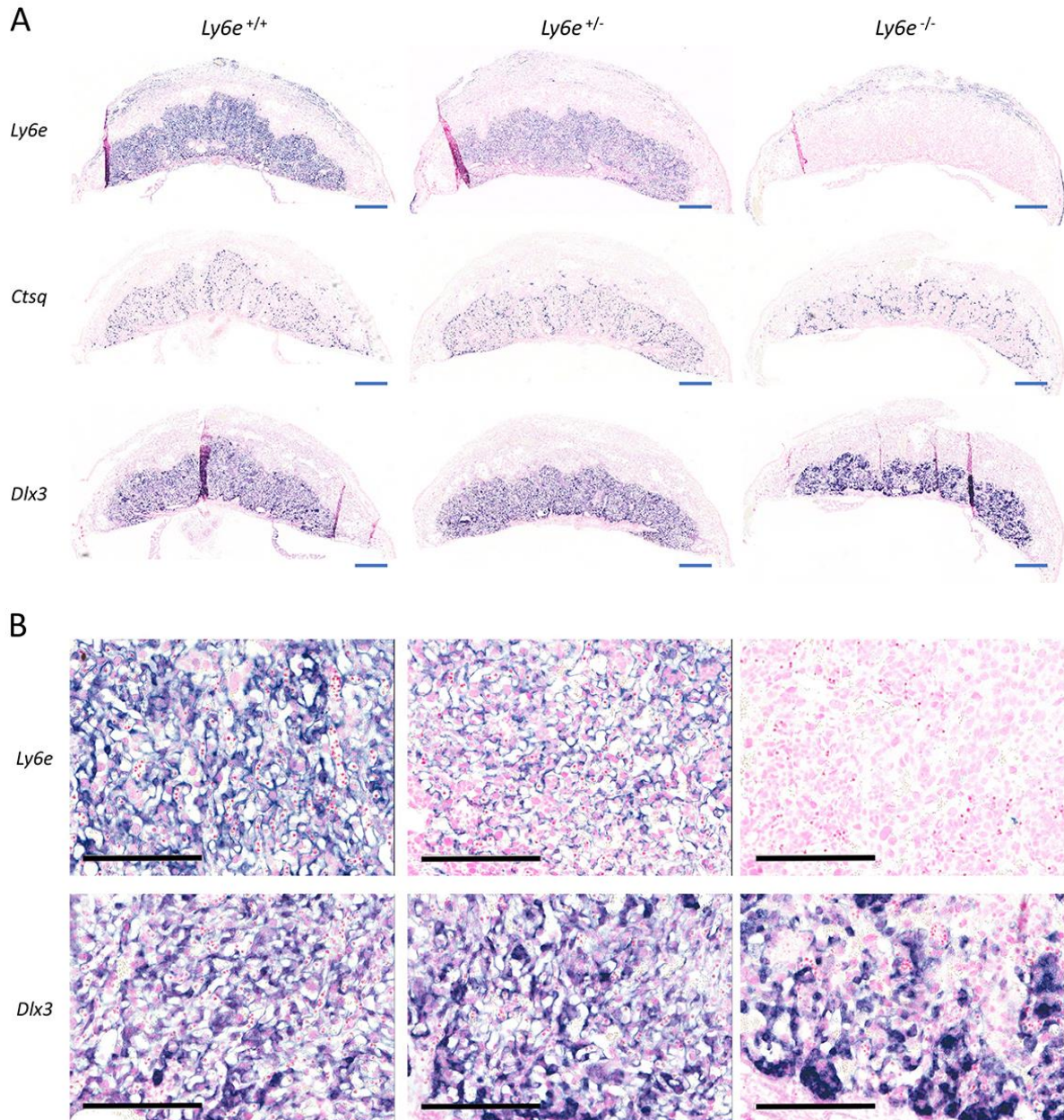
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Figure S1



**Supplemental Figure 1. Mouse Placental Architecture, Circulations, and Interhemal Membrane.** (A) The placenta is composed of three “zones” or “layers”: the maternal decidua (Dec), an inner “junctional zone” (Jz - made up of parietal trophoblast giant cells, glycogen trophoblast, and spongiotrophoblast), and an inner labyrinth layer (Lab), which is the site of maternal-fetal exchange. Placental circulations (~E12.5-E18.5) are also depicted. Maternal blood enters the placenta through the spiral arteries, which coalesce into ~2-3 central canals that traverse the middle Jz and bring blood to the base of the placental labyrinth layer. Blood then percolates up through a series of tortuous sinusoids until it empties into a venous return system at the Jz-Lab interface where it exits the placenta. Conversely, poorly oxygenated fetal blood enters the base of the labyrinth through larger vessels, flows to the top of the labyrinth layer through a capillary network and back out through the base of the placenta. (B) Maternal sinusoids and fetal capillaries are intertwined within the labyrinth, separated by a highly ordered cellular barrier called the interhemal membrane (IHM). The IHM is composed of 4 cell types: fetal endothelial cells that line the fetal vasculature, two intervening layers of syncytiotrophoblast cells, and a layer of sinusoidal trophoblast giant cells that line the maternal sinusoids.

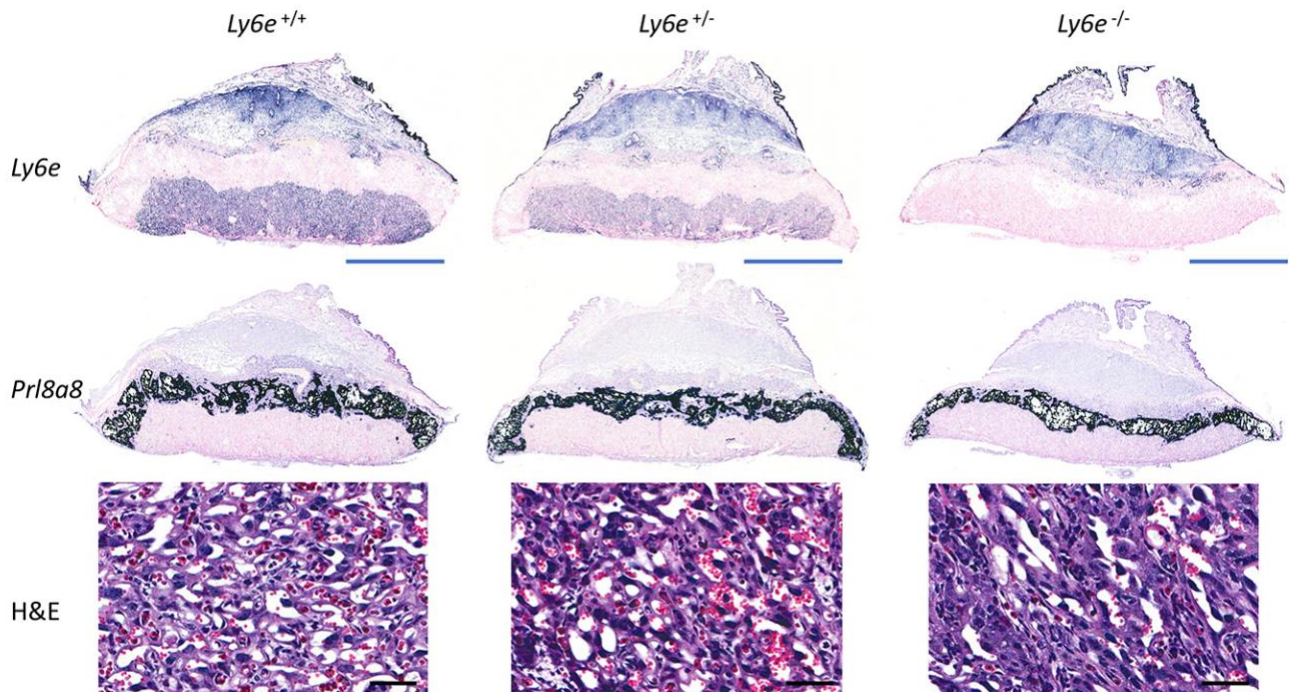
**Figure S2**



**Supplemental Figure S2. Comparison between E12.5 *Ly6e* wildtype, heterozygous and homozygous placentae by in situ hybridization.**

(A) Low magnification images of *Ly6e* wildtype, heterozygous, and mutant E12.5 placentae – in situ hybridization for *Ly6e*, *Ctsq* (marker of sinusoidal trophoblast giant cells, S-TGCs, lining maternal blood spaces), and *Dlx3* (a marker of labyrinth trophoblast cells). The distribution of S-TGCs throughout the labyrinth layer looks comparable between wildtype and heterozygous placentae, whereas *Ctsq*<sup>+</sup> S-TGCs appear abnormally distributed throughout the *Ly6e* mutant placenta. Blue bar = 500 μm. (B) High magnification images demonstrate normal distribution of maternal and fetal blood spaces throughout the labyrinth zone of *Ly6e*<sup>+/+</sup> and *Ly6e*<sup>+/-</sup> placentae, but abnormal organization of the blood spaces within the labyrinth layer of *Ly6e*<sup>-/-</sup> placentae. Black bar = 200 μm.

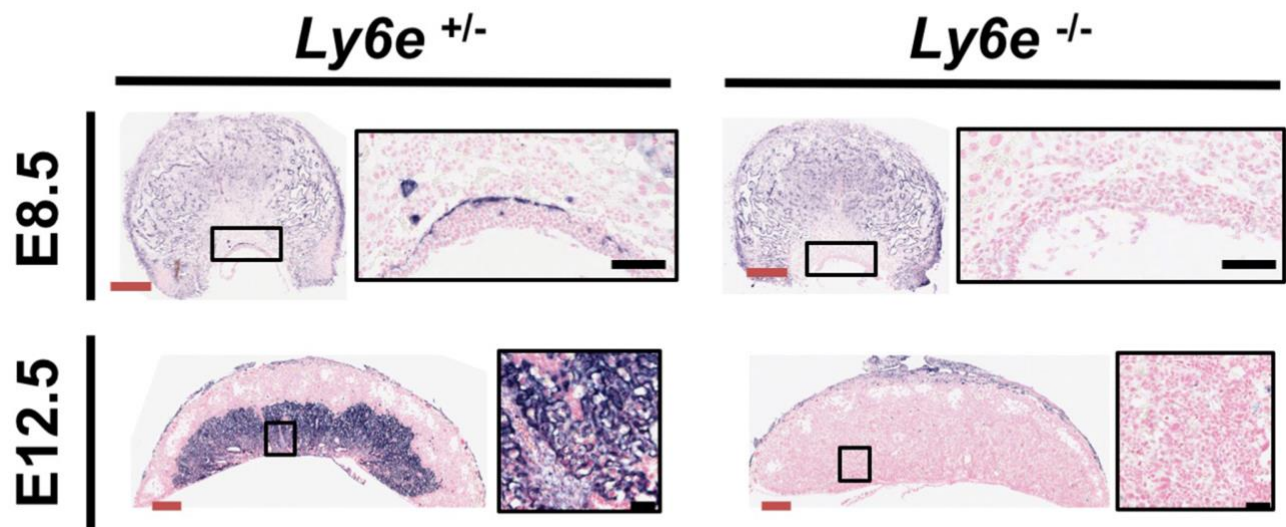
**Figure S3**



**Supplemental Figure S3. Comparison between E14.5 *Ly6e* wildtype, heterozygous and homozygous placentae by in situ hybridization.**

Low magnification images of *Ly6e* wildtype, heterozygous, and mutant E14.5 placentae – in situ hybridization for *Ly6e* and *Prl8a8* (marker of junctional zone spongiotrophoblast cells). Note: the labyrinth layer, beneath the *Prl8a8* positive junctional zone, is not different in size between the wildtype and heterozygous *Ly6e* placentae, but *Ly6e* homozygous mutant placentae have a reduced labyrinth thickness by E14.5. Blue bar = 2 mm. High magnification images of H&E sections demonstrate the normal distribution of maternal and fetal blood spaces throughout the labyrinth zones of *Ly6e* wildtype and heterozygous placentae, but abnormal organization of the blood spaces within the labyrinth layer of *Ly6e* mutant placentae. Black bar = 50  $\mu$ m.

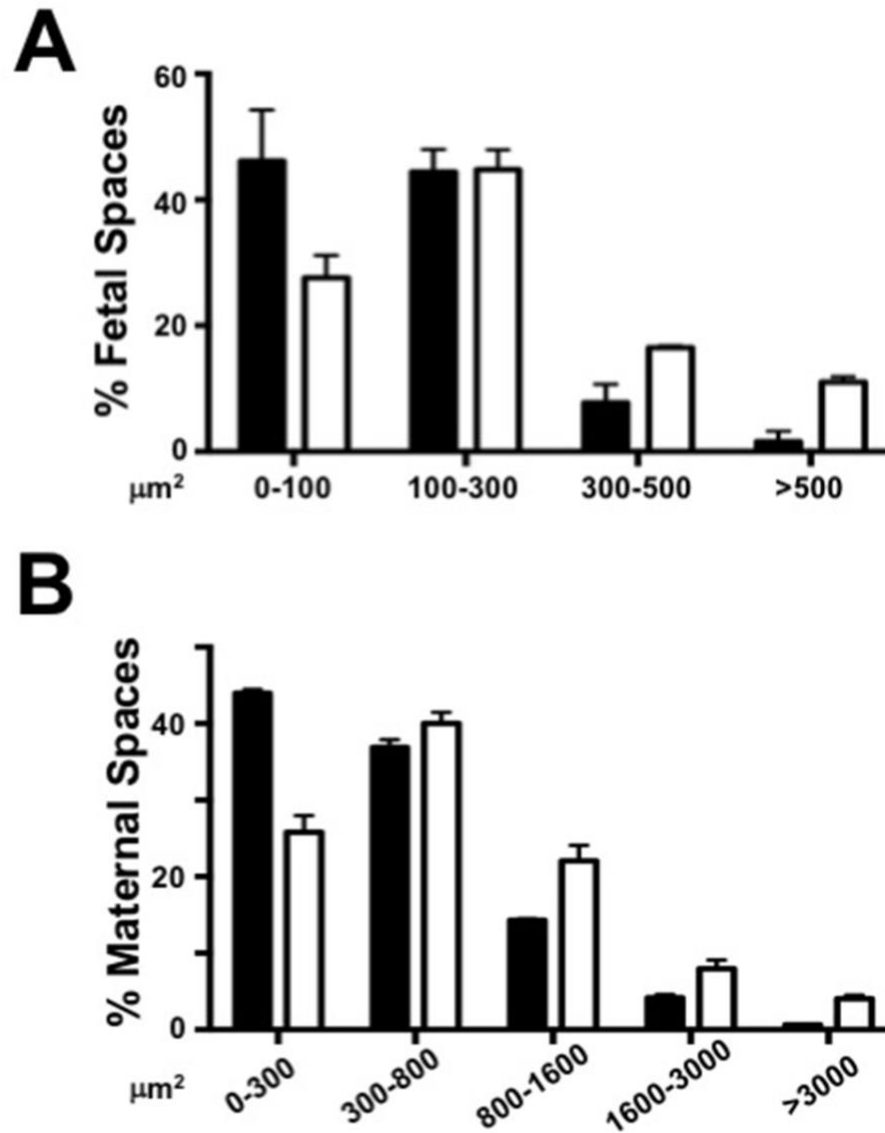
Figure S4.



**Supplemental Figure 4. In situ hybridization analysis of *Ly6e* in E8.5 and E12.5 *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae.**

In situ hybridization confirmed *Ly6e* expression to be absent in *Ly6e*<sup>-/-</sup> placentae, although positive staining could still be observed in the maternal decidua (which is genetically *Ly6e*<sup>+/-</sup>). Red bar, 500 μm; black bar, 100 μm.

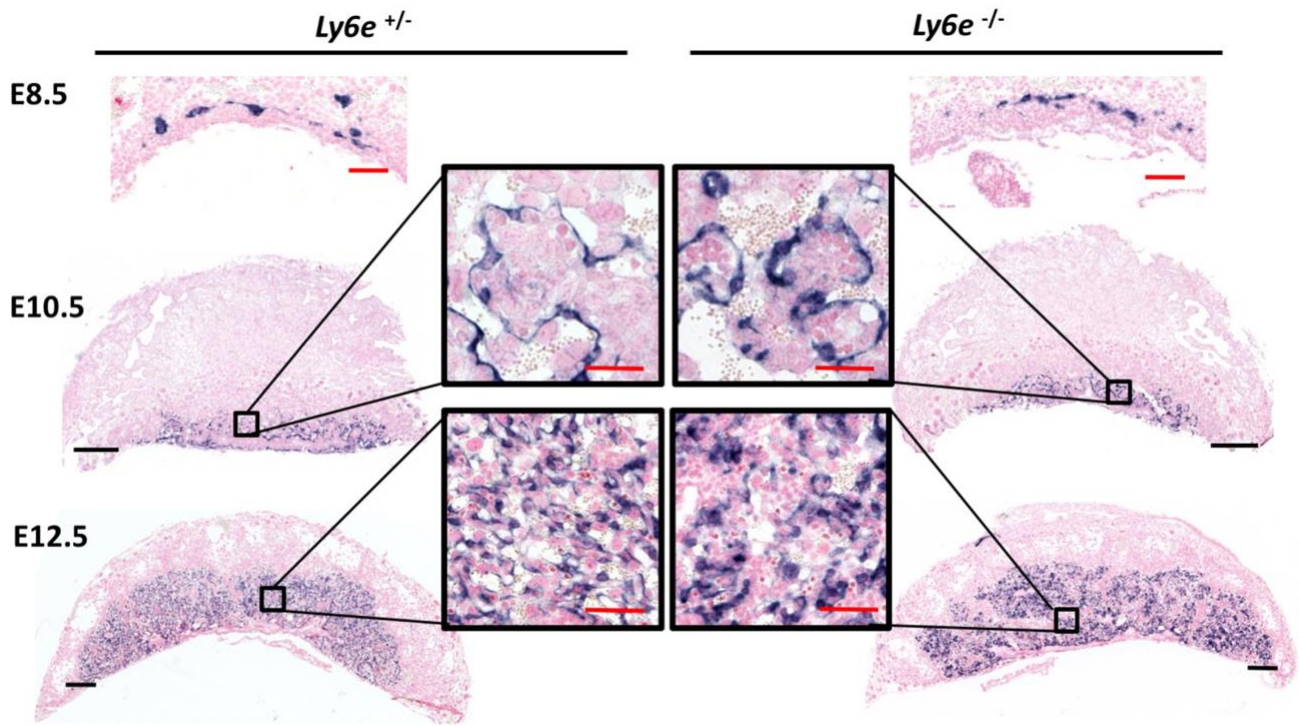
Figure S5



**Supplemental Figure S5. Distribution of maternal and foetal blood spaces in E12.5  $Ly6e^{+/+}$  and  $Ly6e^{-/-}$  placentae.**

(A) A reduction in the number of 100  $\mu\text{m}^2$  (or less) sized fetal blood spaces and an increase in larger blood spaces (>300  $\mu\text{m}^2$ ) was evident in H&E sections of  $Ly6e^{-/-}$  placentae. (B) A reduction in the number of 300  $\mu\text{m}^2$  (or less) sized maternal blood spaces and an increase in larger blood spaces (>1600  $\mu\text{m}^2$ ) was evident in H&E sections of  $Ly6e^{-/-}$  placentae. This shift in presence of larger spaces is consistent with impaired branching morphogenesis.

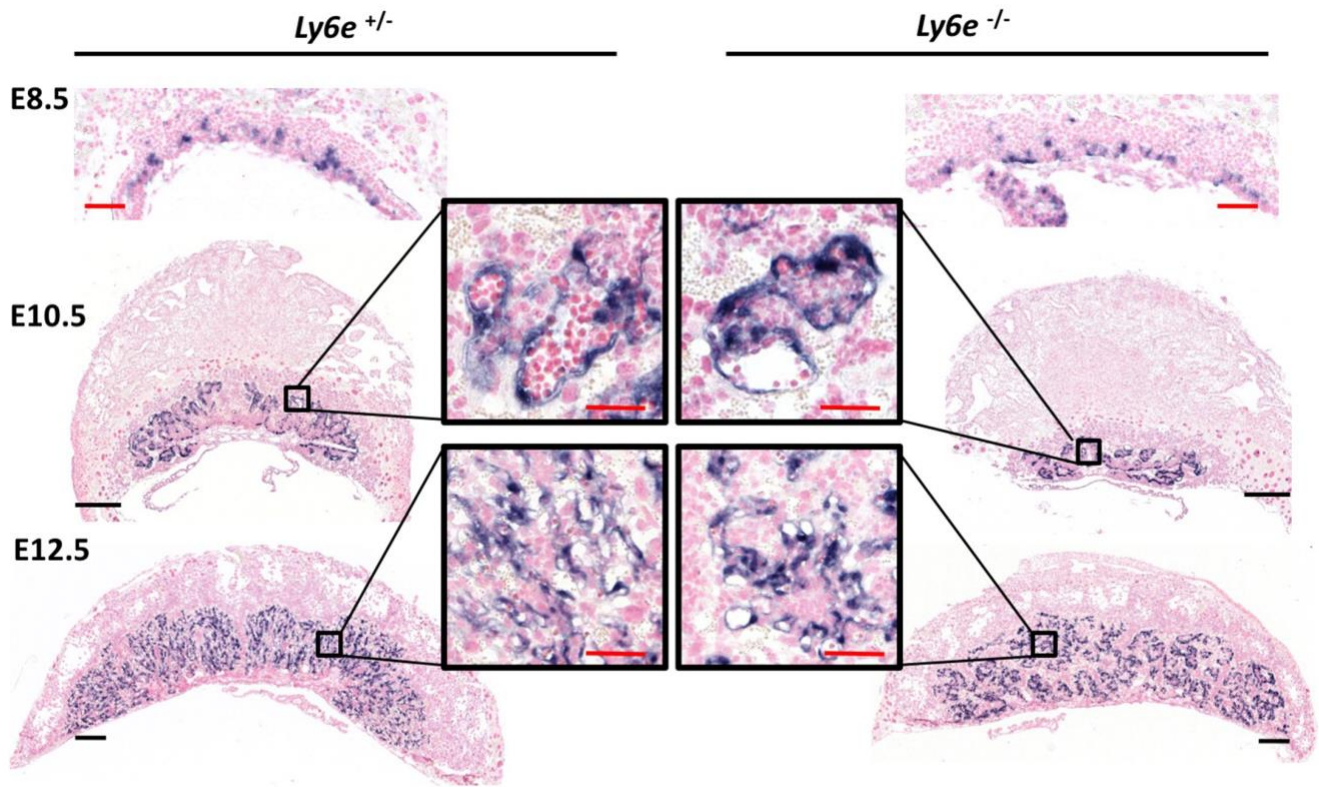
Figure S6



**Supplemental Fig 6. In situ hybridization analysis of *Syna* mRNA expression in *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at E8.5, E10.5, and E12.5.**

*Syna* mRNA expression is evident in the upper half of the chorion at E8.5, and later in the syncytiotrophoblast layer I cells of the E10.5-12.5 placenta in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae. Note: the morphology of *Syna*<sup>+</sup> cells in the *Ly6e*<sup>-/-</sup> placentae are more rounded, and less characteristically thinned and elongated than in the controls. Furthermore, the distribution of *Syna*<sup>+</sup> cells within the labyrinth of E10.5-E12.5 *Ly6e*<sup>-/-</sup> placentae appears disorganised. Black bar, 400 μm; red bar 100 μm.

Figure S7

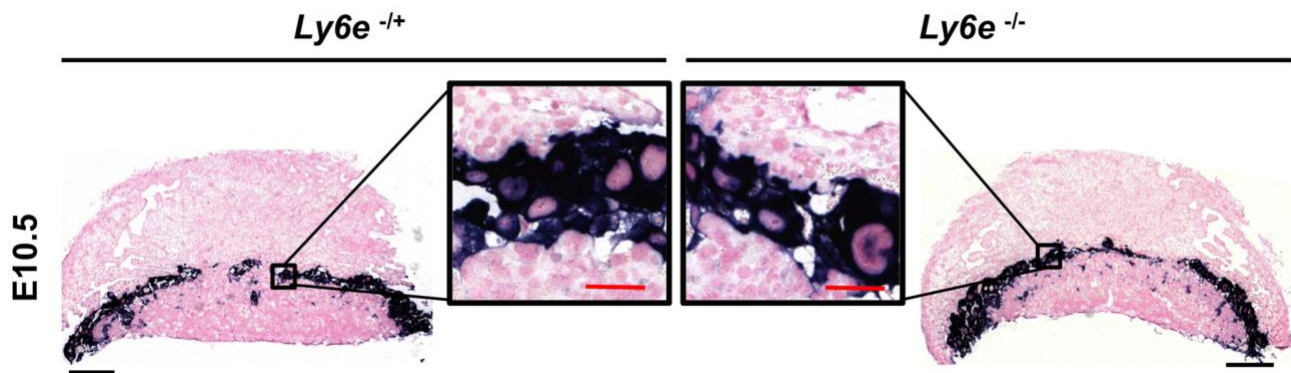


**Supplemental Fig 7. In situ hybridization analysis of *Gcm1* mRNA expression in *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at E8.5, E10.5, and E12.5.**

*Gcm1* mRNA expression is evident in clusters of cells along the leading edge of the chorion at E8.5, and later in the syncytiotrophoblast layer II cells of the E10.5-12.5 placenta in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae. Note: The distribution of *Gcm1*<sup>+</sup> cells within the labyrinth of E10.5-E12.5 *Ly6e*<sup>-/-</sup> placentae appears disorganised. Black bar, 400 μm; red bar 100 μm.



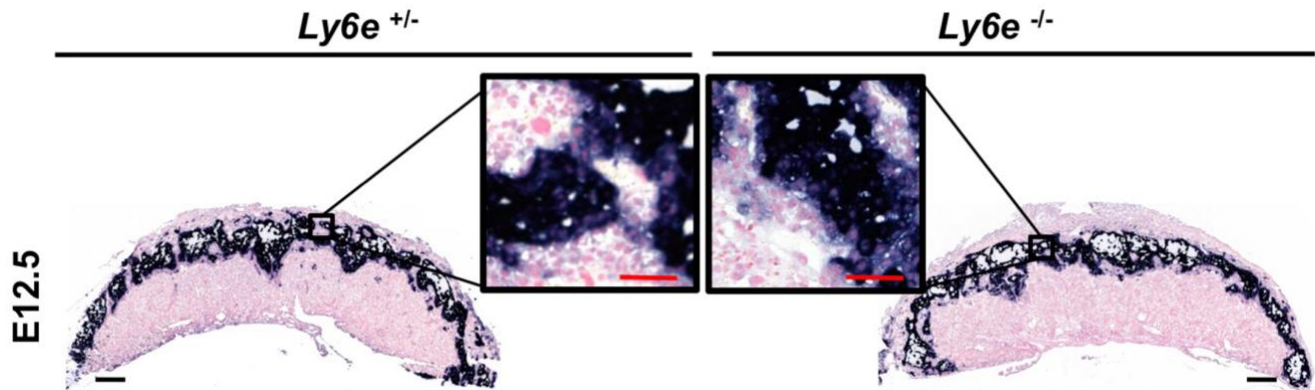
Figure S8



**Supplemental Fig 8. In situ hybridization analysis for *Prl3d* (*Pli*) mRNA expression in *Ly6e*<sup>+/+</sup> and *Ly6e*<sup>-/-</sup> placentae at E10.5.**

*Prl3d* positive parietal trophoblast giant cells (P-TGCs) can be clearly seen above the spongiotrophoblast layer in both *Ly6e*<sup>+/+</sup> and *Ly6e*<sup>-/-</sup> placentae. No obvious differences in P-TGCs were observed between *Ly6e*<sup>+/+</sup> and *Ly6e*<sup>-/-</sup> placentae at any developmental time point examined. Black bar, 400 μm; red bar 100 μm.

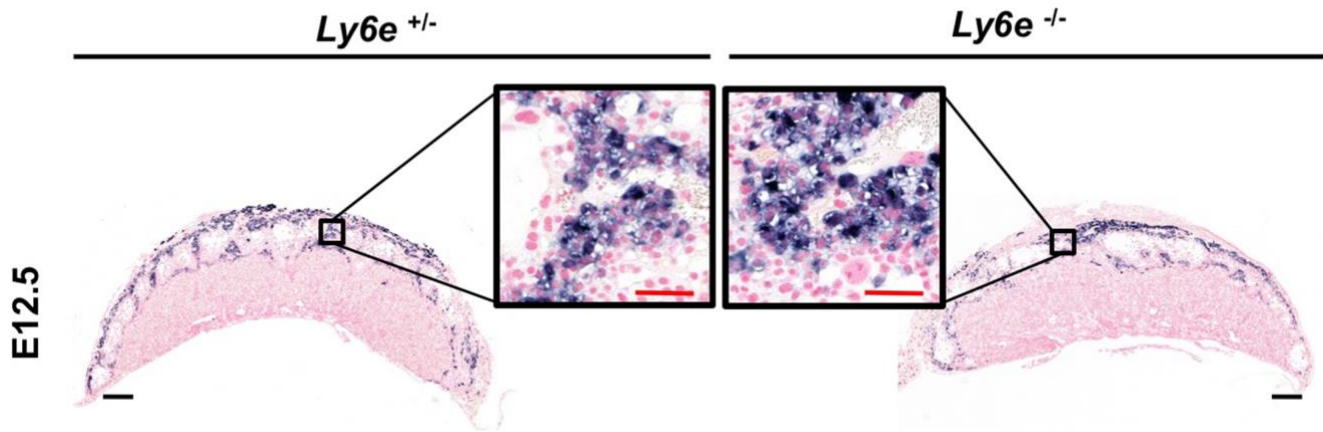
Figure S9



**Supplemental Fig 9. In situ hybridization analysis of *Prl8a8* mRNA expression in *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at E12.5.**

*Prl8a8* positive spongiotrophoblast cells can be clearly seen within the spongiotrophoblast layer in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae (E12.5). No obvious differences in *Prl8a8* staining were observed between *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at any developmental time point examined. Black bar, 400  $\mu$ m; red bar 100  $\mu$ m.

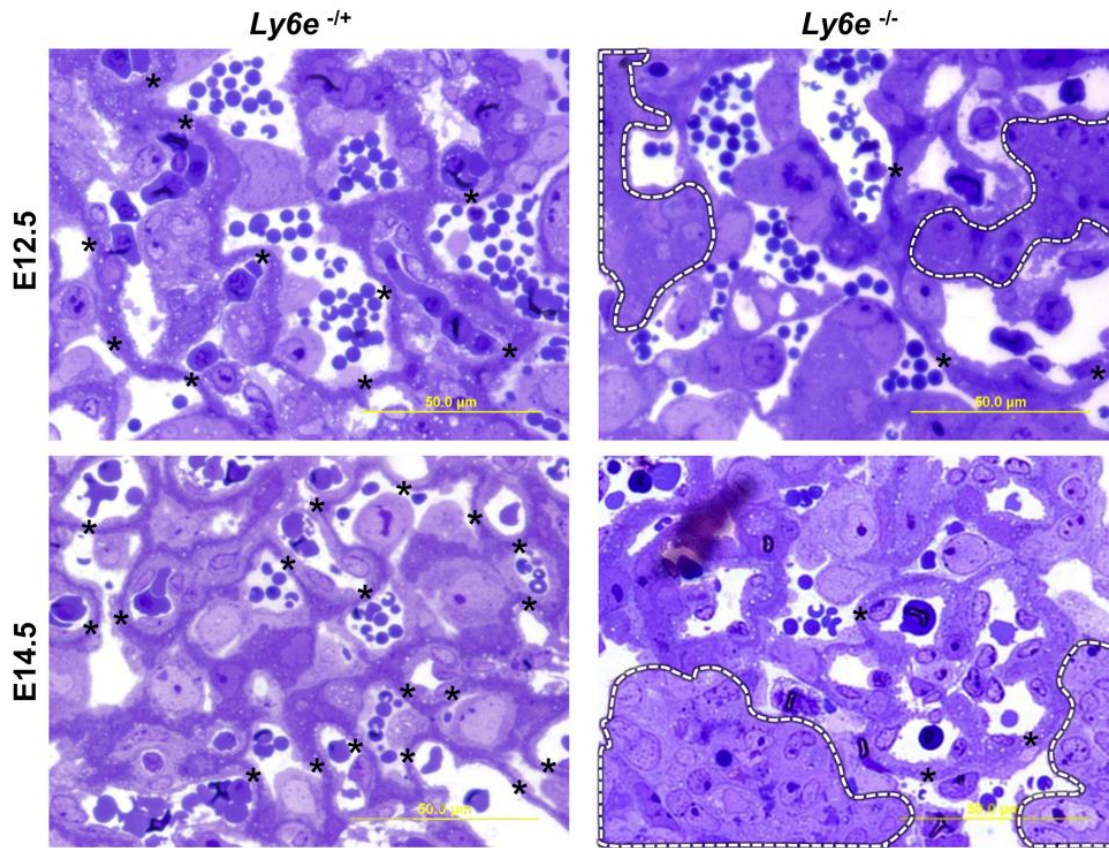
Figure S10



**Supplemental Fig 10. In situ hybridization analysis for *Prl7b1* mRNA expression in *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at E12.5.**

*Prl7b1* positive glycogen trophoblast cells can be clearly seen within the spongiotrophoblast layer and above the trophoblast giant cell layer within the decidua in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae (E12.5). In addition, *Prl7b1* positive spiral artery trophoblast giant cells can also be detected in placentae from both genotypes at E12.5. No obvious differences in *Prl7b1* staining were observed between *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae at any developmental time point examined. Black bar, 400 μm; red bar 100 μm.

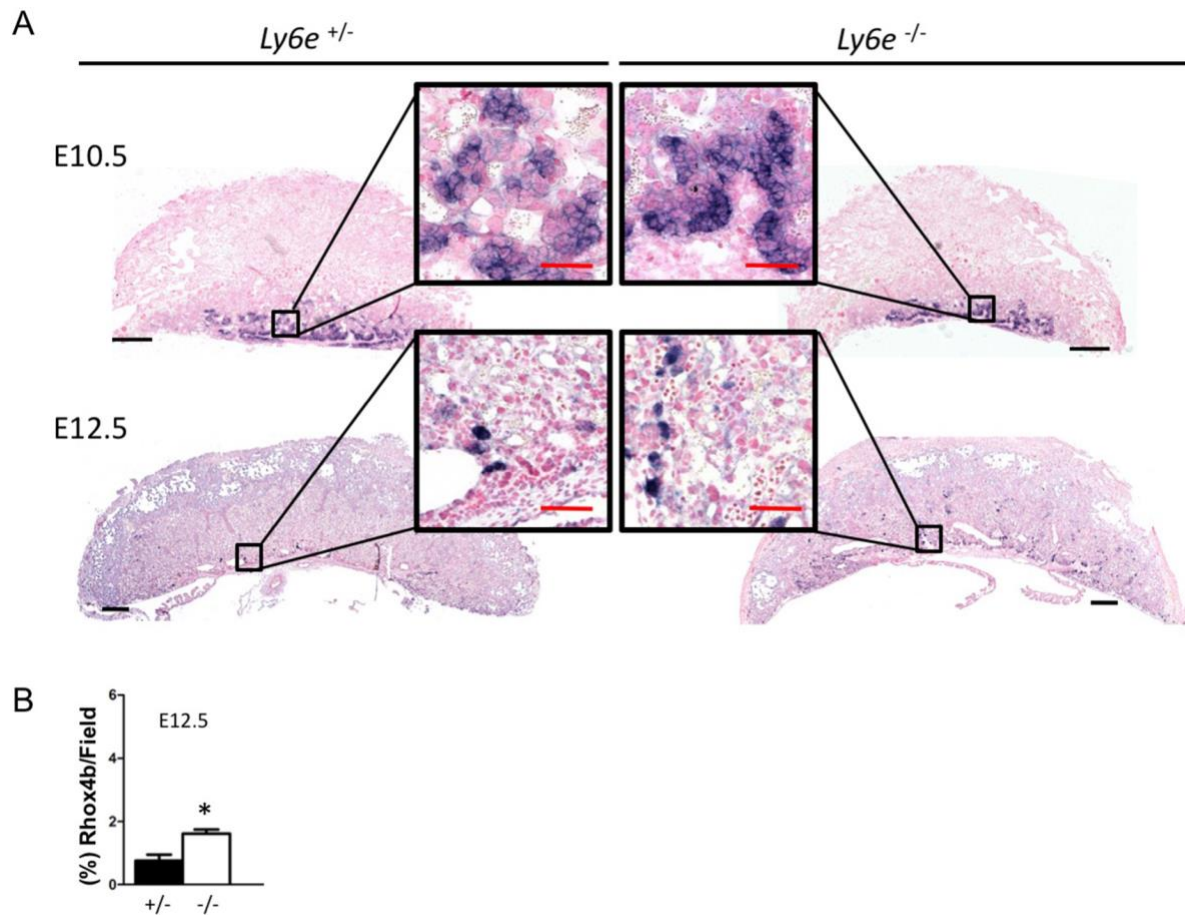
Figure S11



**Supplemental Fig 11. Increased number of densely packed clusters of trophoblast cells within the labyrinth layer of *Ly6e*<sup>-/-</sup> placentae.**

Toluidine blue stained resin sections (1 μm) of *Ly6e*<sup>-/-</sup> placentae at E12.5 (top right) and E14.5 (bottom right) show increased clusters of densely packed trophoblast cells (white hashed lines) within the labyrinth compared with *Ly6e*<sup>+/+</sup> placentae (top left and bottom left). While clusters of densely packed trophoblast can also be seen occasionally in the labyrinths of *Ly6e*<sup>+/+</sup> placentae, they are more common in *Ly6e*<sup>-/-</sup> mutants. \* - areas of characteristically thin interhaemal membrane separating endothelial-lined fetal vasculature from trophoblast giant cell-lined maternal sinusoids. Yellow bar, 50 μm.

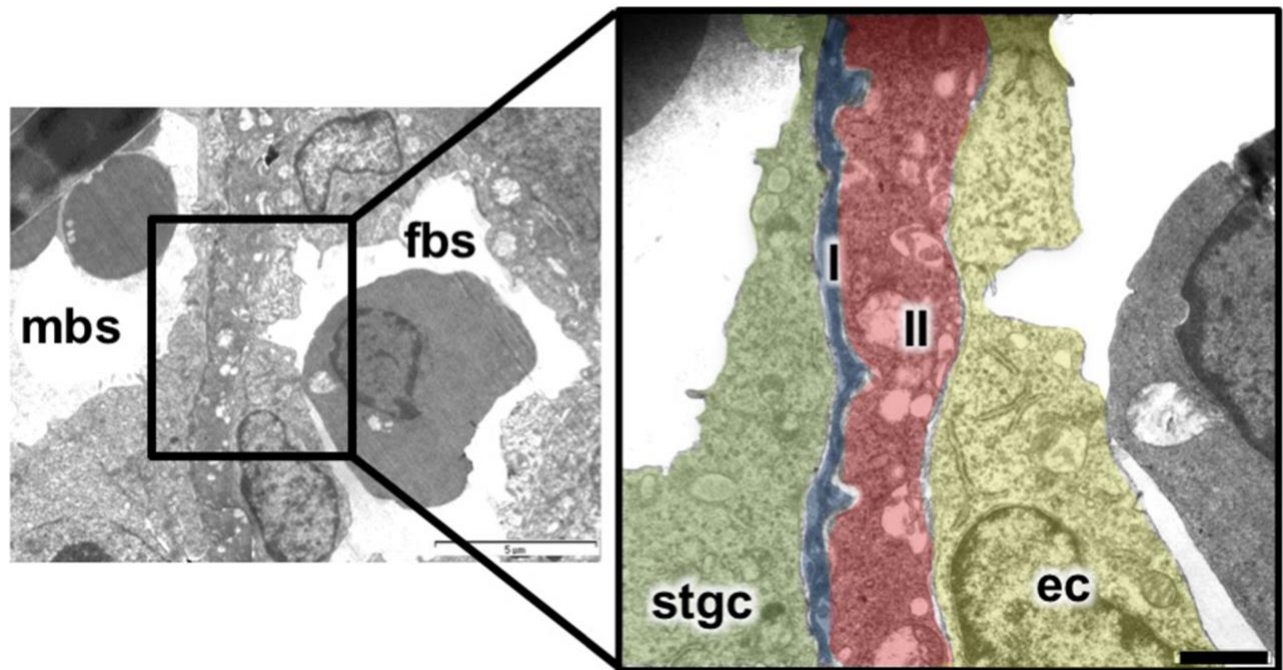
Figure S12



**Supplemental Fig 12. In situ hybridization analysis of *Rhox4b* mRNA expression in *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae E10.5, and E12.5.**

(A) *Rhox4b* mRNA expression is expressed in cuboidal trophoblast cells located predominantly at the base of the developing labyrinth in E10.5-12.5 placenta in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae. Note: although the number of *Rhox4b*<sup>+</sup> cells decreases with gestational age in both *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae, the number of *Rhox4b*<sup>+</sup> cells in the *Ly6e*<sup>-/-</sup> placentae is significantly increased compared to age-matched littermate controls. Black bar, 400 μm; red bar 100 μm. (B) In E12.5 *Ly6e*<sup>-/-</sup> placentae, clusters of *Rhox4b*<sup>+</sup> cells were distributed more broadly throughout the labyrinth layer, and covered a greater area than in heterozygous or WT littermate controls. Red bar, 100 μm. Error bars are +/- s.e.m.

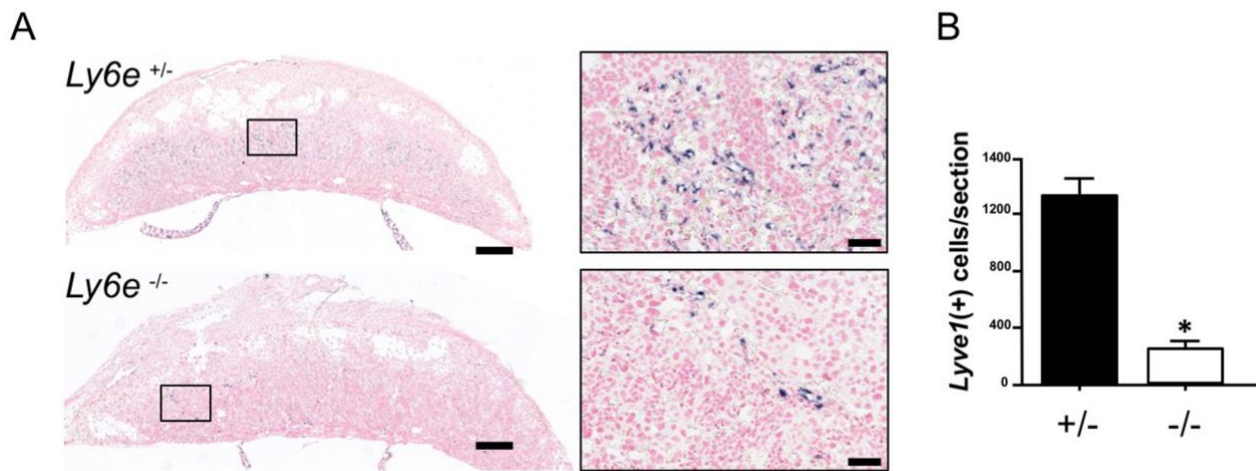
Figure S13



**Supplemental Fig 13. Ultrastructural defects in E12.5 *Ly6e*<sup>-/-</sup> placentae.**

Most interhaemal membranes observed in *Ly6e*<sup>-/-</sup> placentae displayed obvious ultrastructural defects. Some *Ly6e*<sup>-/-</sup> interhaemal membrane segments contained excessively thin and electron dense syncytiotrophoblast layer I, a phenotype never observed in heterozygous labyrinths. Black bar, 1 μm; white bar, 5 μm.

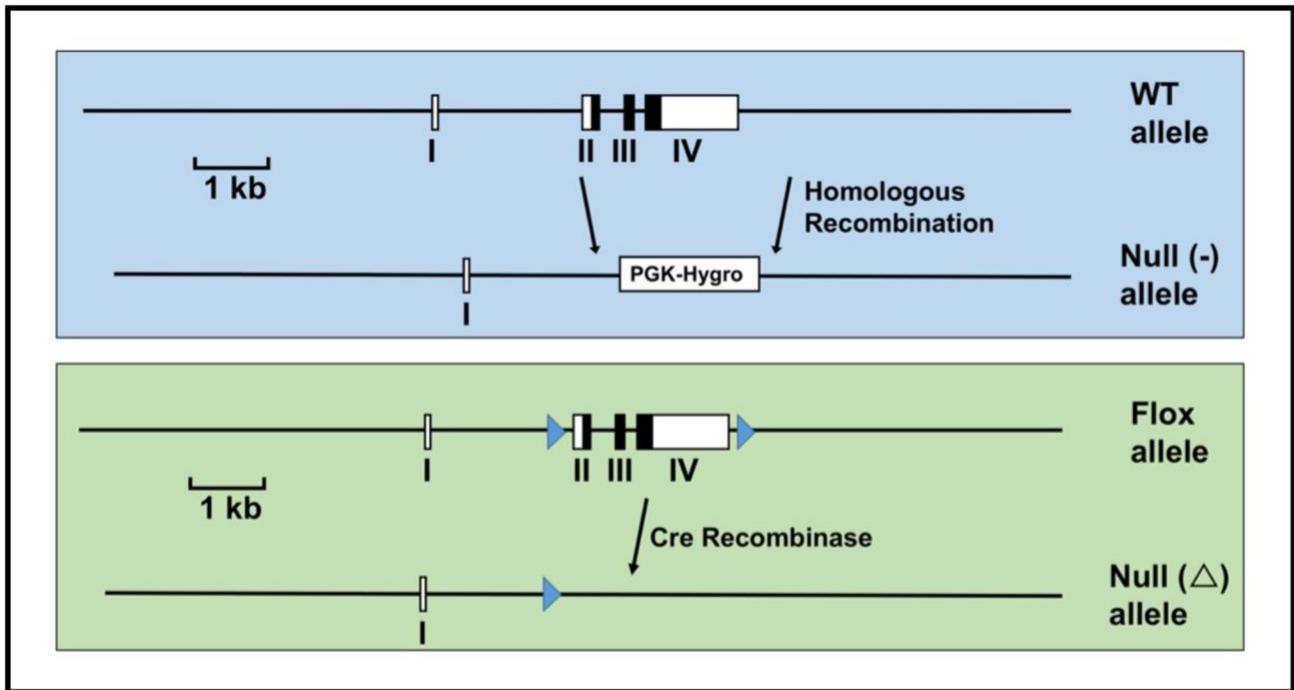
Figure S14



**Supplemental Fig 14. Reduced number of *Lyve1* positive cells in E12.5 *Ly6e*<sup>-/-</sup> placentae.**

(A) In situ hybridization for *Lyve1* in E12.5 *Ly6e*<sup>+/-</sup> and *Ly6e*<sup>-/-</sup> placentae. *Lyve1* expression is restricted to a subset of fetal endothelial cells, predominantly located in the “upper” portion of the labyrinth underlying the spongiotrophoblast layer. E12.5 *Ly6e*<sup>-/-</sup> placentae contained fewer *Lyve1* positive cells within the labyrinth. (B) A significant decrease (\*,  $p < 0.05$ ) in *Lyve1* cells was observed in *Ly6e*<sup>-/-</sup> placentae compared with *Ly6e*<sup>+/-</sup> controls. Black bar – *Ly6e*<sup>+/-</sup>; white bar – *Ly6e*<sup>-/-</sup>. Paired littermates (n=4 pairs) representing each of the two genotypes were used for all measurements; three sections from the midline per placenta were counted. Error bars are +/- s.e.m.

Figure S15

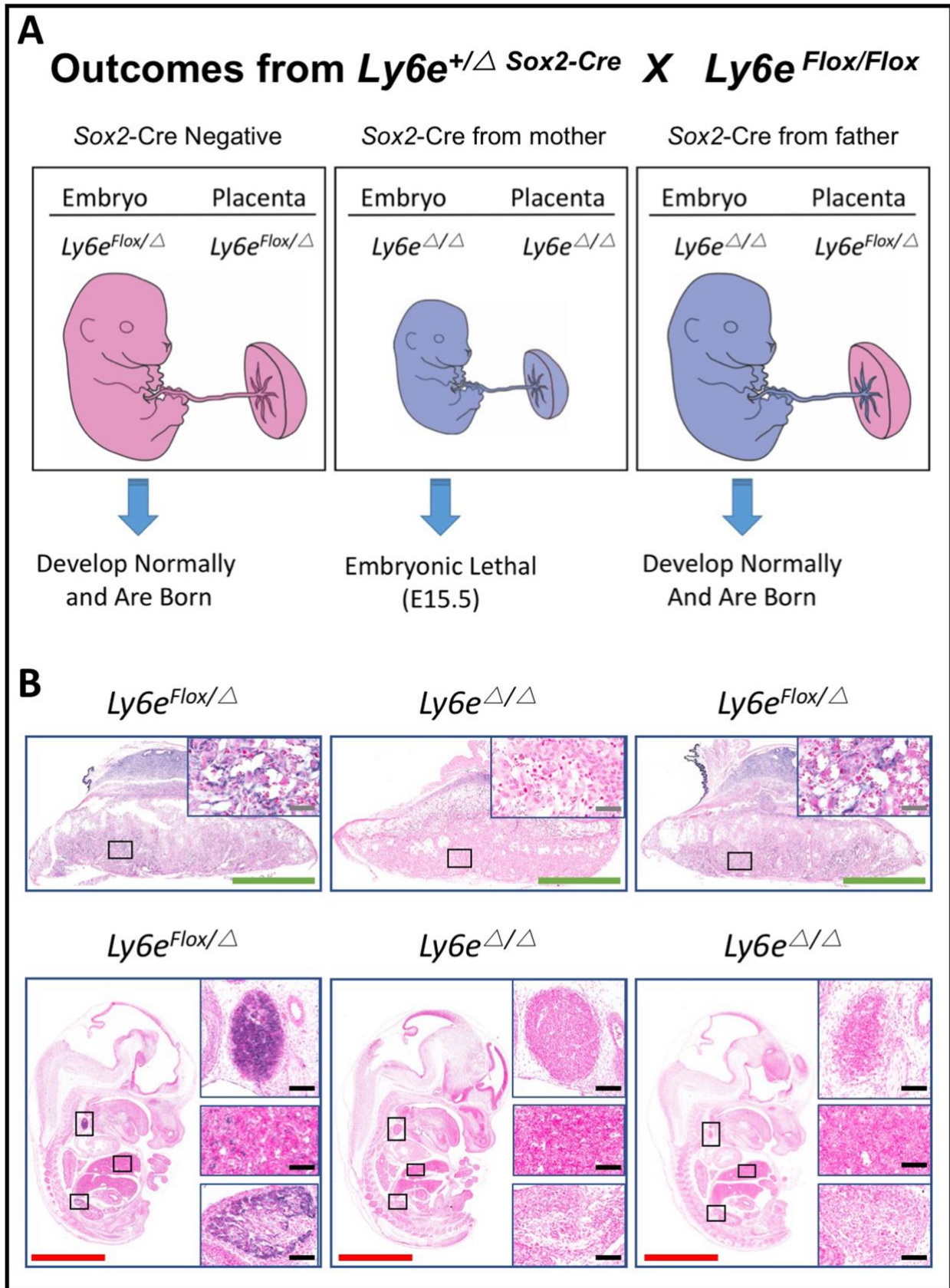


**Supplemental Figure 15. *Ly6e* Alleles and conditional crosses.**

Null *Ly6e* allele (-) published by Zammit et al.; exons 2-4 replaced with a PGK-Hygomycin cassette. Conditional allele (Flox) contains loxP sites flanking exons 2-4. Once deleted, the null allele ( $\Delta$ ) has the same genomic structure as the original (-) allele, without the PGK-Hyg cassette.



Figure S16



**Supplemental Figure 16. *Ly6e* conditional crosses.**

(A) Crosses with *Ly6e*<sup>+/ $\Delta$</sup> ; *TgSox2-Cre* males and *Ly6e*<sup>flox/flox</sup> females can produce *Ly6e* <sup>$\Delta$ / $\Delta$</sup> ; *TgSox2-Cre* embryos that retain a *Ly6e* <sup>$\Delta$ /Flox</sup>; *TgSox2-Cre* placenta, as *Sox2-Cre* is expressed in the epiblast and not expressed in trophoblast of the placenta. Note, crosses with *Ly6e*<sup>+/ $\Delta$</sup> ; *TgSox2-Cre* females and *Ly6e*<sup>flox/flox</sup> males result in both the embryo and placenta being *Ly6e* <sup>$\Delta$ / $\Delta$</sup>  due to *Sox2-Cre* expression in the ovary. Furthermore, crosses between *Ly6e*<sup>+/ $\Delta$</sup>  males and females produced *Ly6e* <sup>$\Delta$ / $\Delta$</sup>  embryos with the same phenotypes as *Ly6e*<sup>-/-</sup> embryos (see Supplemental Table 3).

(B) In situ hybridization for *Ly6e* in E12.5 placentae and embryos from crosses of *Ly6e*<sup>+/ $\Delta$</sup> ; *TgSox2-Cre* males with *Ly6e*<sup>flox/flox</sup> females. Genotypes of placenta/embryos align with those above in section A. Placentas are shown with a higher magnification of the labyrinth layer inset. Embryos are shown with 3 higher magnification insets of tissues that express *Ly6e* in the E12.5 embryo – thymus, liver, and adrenal gland. Note, embryos are missing tails due to sampling for genotyping. Green bar = 2 mm; grey bar = 50  $\mu$ m; Red bar = 3 mm; Black bar = 200  $\mu$ m.

**Supplemental Table 1**

<b>Gene Name</b>	<b>Sequence (5'-3')</b>	<b>RNA Polymerase site</b>
<i>Ly6e</i> forward	<b>AATTAACCCTCACTAAAGGG</b> TATCCCACTACT GGGCCTTG3	<b>T3</b>
<i>Ly6e</i> reverse	<b>TAATACGACTCACTATAGGG</b> TATCGGGGTTGG TCTTTCAG	<b>T7</b>
<i>Lyve1</i> forward	<b>AATTAACCCTCACTAAAGGG</b> CATCCCTCGGAT TTTCTCAA	<b>T3</b>
<i>Lyve1</i> reverse	<b>TAATACGACTCACTATAGGG</b> TCTGCAGGAACT GACAGTGG	<b>T7</b>
<i>Mest</i> forward	<b>AATTAACCCTCACTAAAGGG</b> GAGAGAGTGGT GGGTCCAAG	<b>T3</b>
<i>Mest</i> reverse	<b>TAATACGACTCACTATAGGG</b> CGATCACTCGAT GGAACCTC	<b>T7</b>

Supplemental Table 2

Comparisons Between <i>Ly6e</i> <sup>+/-</sup> and <i>Ly6e</i> <sup>-/-</sup> Placentae	
Gross Morphology (E12.5)	Phenotype in <i>Ly6e</i> <sup>-/-</sup> Placentae
Total Placental Volume	No Difference
Total Jz Volume	No Difference
Total Labyrinth Volume	No Difference
Maternal Blood Volume	<b>Decreased</b> in <i>Ly6e</i> <sup>-/-</sup> placentae
Fetal Blood Volume	<b>Decreased</b> in <i>Ly6e</i> <sup>-/-</sup> placentae
EM Morphology of Labyrinth (E12.5)	Phenotype in <i>Ly6e</i> <sup>-/-</sup> Placentae
S-TGC Layer	<b>Increased</b> vacuolization and abnormal apical morphology
SynT-I Layer	<b>Impaired fusion</b> , abnormal morphology elsewhere, including excessive thinning with increased electron density and increased vacuolization
SynT-II	<b>Increased</b> vacuolization
Fetal Endothelial Cells	No Difference
Diploid Trophoblast	<b>Increased</b> numbers of diploid trophoblast clusters within the labyrinth
In Situ Hybridization (E12.5)	Phenotype in <i>Ly6e</i> <sup>-/-</sup> Placentae
<i>Prl3d</i> ( <i>Pli</i> )	No Difference (Trophoblast Giant Cell marker)
<i>Prl8a8</i> ( <i>PlpCγ</i> )	No Difference (Spongiotrophoblast marker)
<i>Prl7b1</i> ( <i>PrlpN</i> )	No Difference (Glycogen Trophoblast marker)
<i>Ctsq</i>	Expressed – but <b>abnormal distribution throughout labyrinth.</b> (Sinusoidal Giant Cell marker)
<i>Syna</i>	Expressed (Syncytiotrophoblast Layer I marker)
<i>Gcm1</i>	Expressed (Syncytiotrophoblast Layer II marker)
<i>Mest</i>	Expressed – but <b>abnormal distribution throughout labyrinth.</b> (Fetal Endothelial marker)
<i>Rhox4b</i>	<b>Increased Expression</b> (Putative Labyrinth Progenitor marker)
<i>Lyve1</i>	<b>Severely Reduced Expression</b> (Marker of a Subset of Fetal Endothelial Cells)
Immunohistochemistry (E12.5)	Phenotype in <i>Ly6e</i> <sup>-/-</sup> Placentae
Phosphohistone H3	<b>Increased</b> staining within labyrinth layer

Supplemental Table 3

<b>Viability Data</b>					
<b><i>Ly6e</i><sup>+/-</sup> ♂ x <i>Ly6e</i><sup>+/-</sup> ♀</b>			<b>Embryo/Pup Genotype</b>		
<b>Age</b>	<b>Embryos/Pups (No. Litters)</b>	<b><i>Ly6e</i><sup>+/+</sup></b>	<b><i>Ly6e</i><sup>+/-</sup></b>	<b><i>Ly6e</i><sup>-/-</sup></b>	
E10.5	20 (3)	8	8	4	
E12.5	53 (7)	5	37	11	
E14.5	90 (12)	34	52	4	
E15.5	18 (3)	6	11	1	
P14	92 (14)	30	62	0	
<b><i>Ly6e</i><sup>+/<math>\Delta</math></sup> ♂ x <i>Ly6e</i><sup>+/<math>\Delta</math></sup> ♀</b>			<b>Pup Genotype</b>		
<b>Age</b>	<b>Embryos/Pups (No. Litters)</b>	<b><i>Ly6e</i><sup>+/+</sup></b>	<b><i>Ly6e</i><sup>+/<math>\Delta</math></sup></b>	<b><i>Ly6e</i><sup><math>\Delta</math>/<math>\Delta</math></sup></b>	
P14	28 (4)	10	18	0	
<b><i>Ly6e</i><sup>Flox/Flox</sup> ♂ x <i>Ly6e</i><sup>+/<math>\Delta</math> Sox2-Cre</sup> ♀</b>			<b>Pup Genotype</b>		
<b>Age</b>	<b>Embryos/Pups (No. Litters)</b>	<b><i>Ly6e</i><sup>+/<math>\Delta</math>/Flox</sup></b>	<b><i>Ly6e</i><sup>+/<math>\Delta</math> Sox2-Cre</sup></b>	<b><i>Ly6e</i><sup><math>\Delta</math>/Flox</sup></b>	<b><i>Ly6e</i><sup><math>\Delta</math>/<math>\Delta</math> Sox2-Cre</sup></b>
P0	12 (3)	5	3	4	0
<b><i>Ly6e</i><sup>+/<math>\Delta</math> Sox2-Cre</sup> ♂ x <i>Ly6e</i><sup>Flox/Flox</sup> ♀</b>			<b>Pup Genotype</b>		
<b>Age</b>	<b>Embryos/Pups (No. Litters)</b>	<b><i>Ly6e</i><sup>+/<math>\Delta</math>/Flox</sup></b>	<b><i>Ly6e</i><sup>+/<math>\Delta</math> Sox2-Cre</sup></b>	<b><i>Ly6e</i><sup><math>\Delta</math>/Flox</sup></b>	<b><i>Ly6e</i><sup><math>\Delta</math>/<math>\Delta</math> Sox2-Cre</sup></b>
P0	63 (7)	17	12	23	11