

Supporting Information

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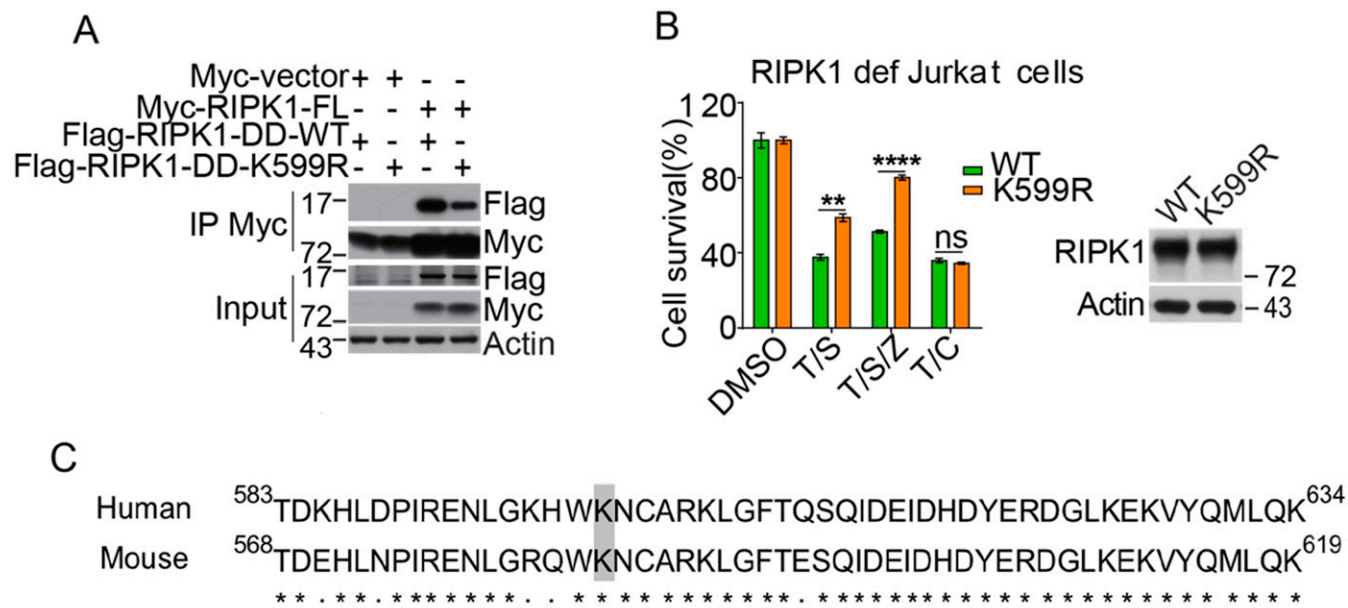


Fig. S1. Effect of hRIPK1 K599R mutation on RIPK1 dimerization and cell death. (A) HEK293T cells were cotransfected with expression vectors of Myc-tagged full-length RIPK1, Flag-RIPK1-DD, and Flag-RIPK1-DD-K599R as indicated for 24 h, the cells were then lysed with 0.5% Nonidet P-40 buffer, and cell lysates were immunoprecipitated with anti-Myc antibody-conjugated agarose. (B) RIPK1-deficient Jurkat cells were infected with retrovirus encoding Flag-tagged hRIPK1 WT and Flag-tagged hRIPK1 K599R by the Tet-On Advanced Inducible Expression System. Different RIPK1-reconstituted Jurkat cells were treated with 1 μ g/mL doxycycline for 48 h to induce the expression of RIPK1. The cells were then pretreated with 100 nM SM-164, 100 nM 5Z-7, or CHX 1 μ g/mL for 2 h and treated with 100 ng/mL TNF α and 50 μ M zVAD.fmk for 10 h. The cell survival was measured by CellTiterGlo. Results shown are averages of triplicates \pm SEM (** P < 0.01; **** P < 0.0001). (Right) Western blot analysis of RIPK1 expression levels in RIPK1-deficient Jurkat cells infected with retrovirus encoding Flag-tagged hRIPK1 WT and K599R. (C) Sequence alignment of the death domains of human and murine RIPK1 around the K599/K584 region. K599/K584 residues conserved in humans and mice are highlighted in gray.

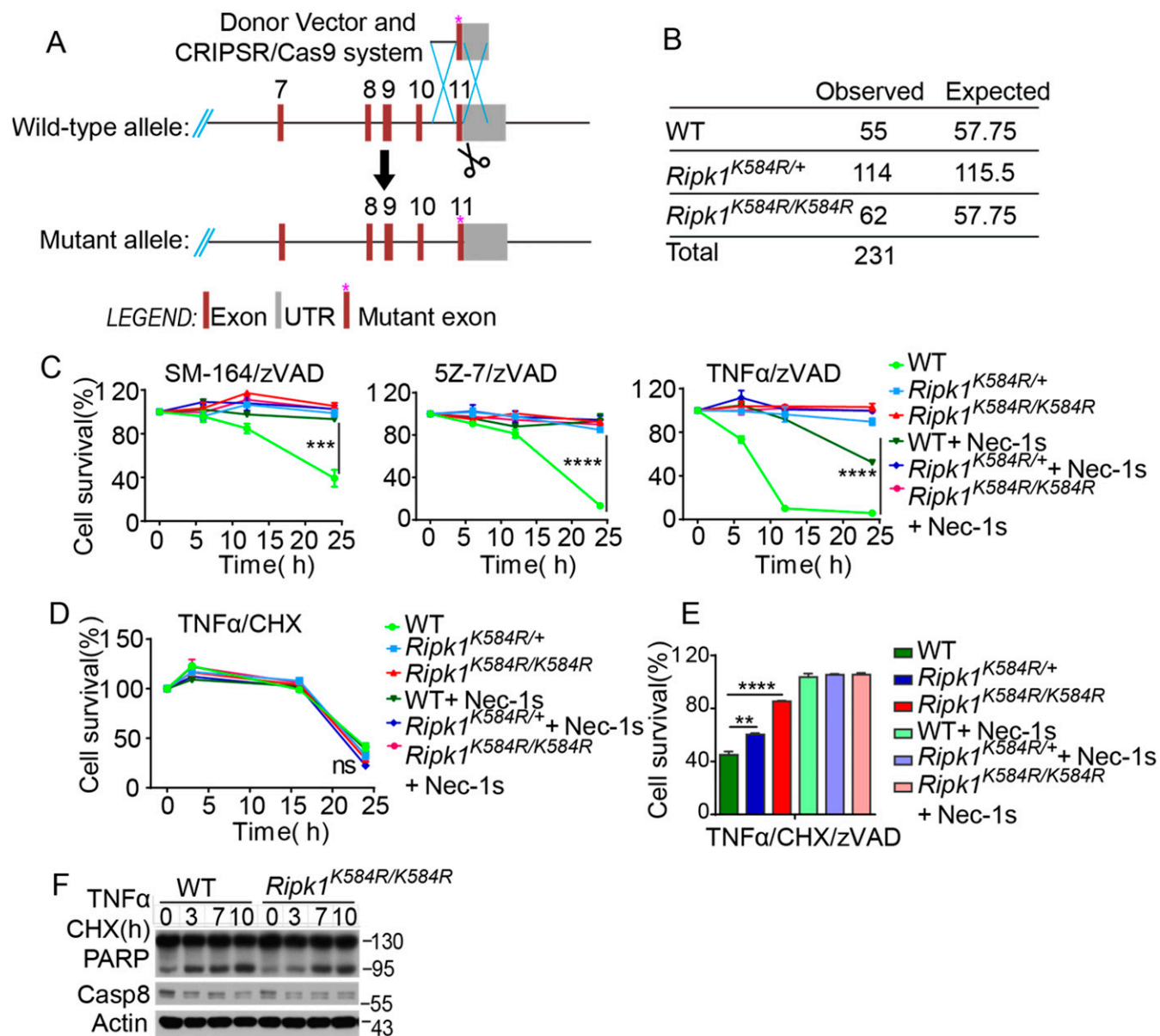


Fig. S2. *Ripk1*^{K584R} cells are resistant to RIPK1-dependent cell death but not RIPK1-independent apoptosis. (A) A schematic diagram of the murine *Ripk1* locus and the *Ripk1* K584R allele. (B) Numbers of surviving offspring from mating of *Ripk1*^{K584R/+} parents. (C) WT, *Ripk1*^{K584R/+}, and *Ripk1*^{K584R/K584R} immortalized MEFs were pretreated with 25 nM SM-164 (Left) or 100 nM 5Z-7 (Middle) for 2 h and then treated with 50 μ M zVAD.fmk or treated with 20 ng/mL TNF α and 50 μ M zVAD.fmk (Right) as indicated for different time points. (D and E) WT, *Ripk1*^{K584R/+}, and *Ripk1*^{K584R/K584R} immortalized MEFs were treated with TNF α +CHX to induce apoptosis (D) or treated with TNF α +CHX+zVAD.fmk to induce necroptosis (E). The cell survival was measured by CellTiterGlo (A–C). (* P < 0.05; ** P < 0.01; *** P < 0.001; **** P < 0.0001.) (F) WT and *Ripk1*^{K584R/K584R} immortalized MEFs were treated with TNF α +CHX for different time points, and cells were lysed with 0.5% Nonidet P-40 buffer and analyzed by Western blotting with caspase and PARP cleavage to indicate apoptosis.

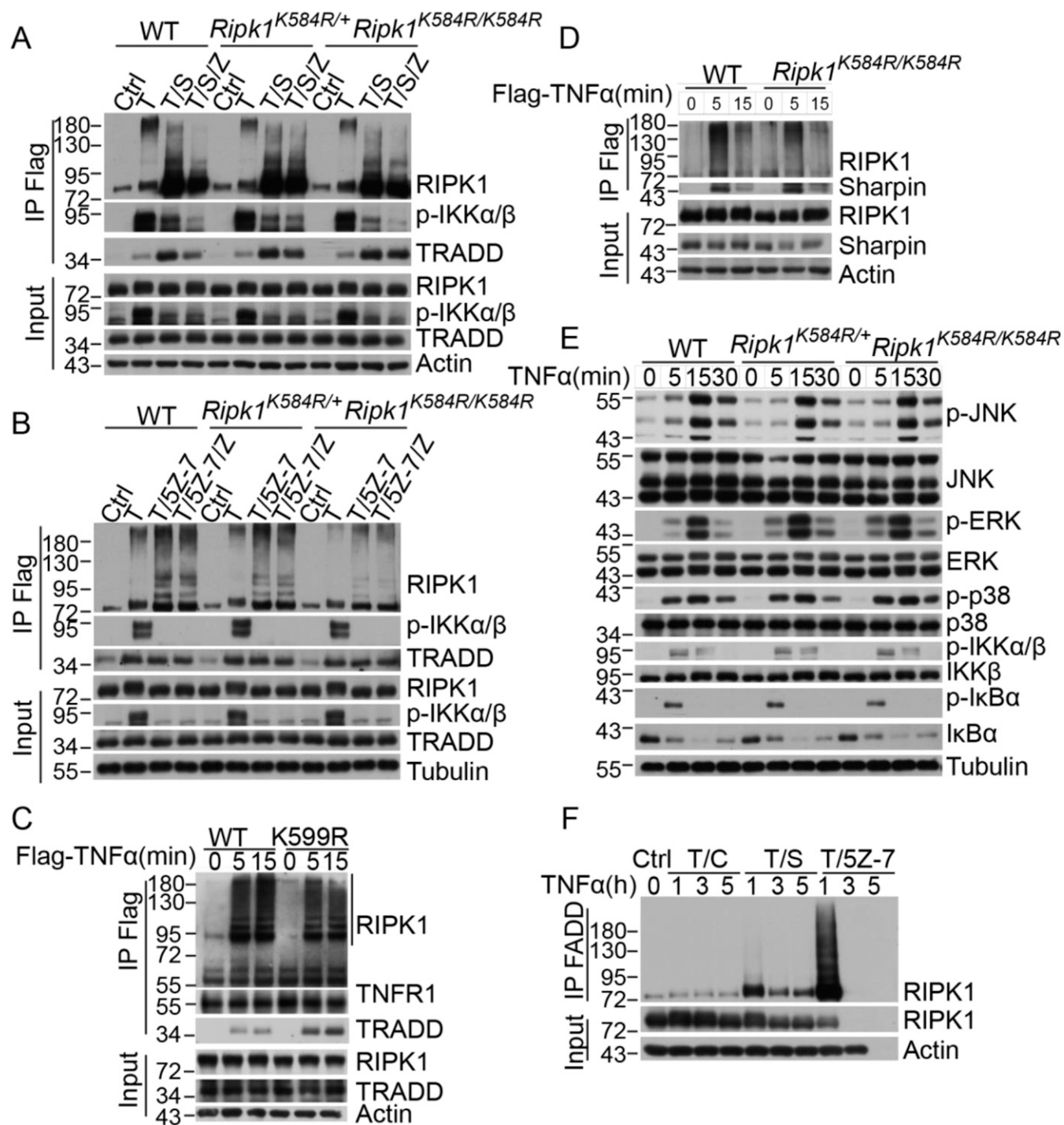


Fig. S3. The *Ripk1*^{K584R} mutation disrupts the formation of complex II. (A and B) Primary WT, *Ripk1*^{K584R/+}, and *Ripk1*^{K584R/K584R} MEFs were pretreated with SM-164 ± zVAD (A) or 5Z-7 ± zVAD (B) for 1 h as indicated, and then Flag-TNFα was added for 5 min, cells were then lysed with 0.5% Nonidet P-40 buffer, and cell lysates were immunoprecipitated with anti-Flag antibody-conjugated agarose. All immunoprecipitated proteins and whole-cell lysates were analyzed by Western blotting with indicated antibodies. (C) RIPK1-deficient Jurkat cells infected with retrovirus encoding HA-tagged WT or K599R hRIPK1 were treated with Flag-TNFα for the indicated time. The cells were then lysed with 0.5% Nonidet P-40 buffer and immunoprecipitated with anti-Flag antibody-conjugated agarose. (D) Primary WT and *Ripk1*^{K584R/K584R} MEFs were treated with TNFα and then lysed with 0.5% Nonidet P-40 buffer and immunoprecipitated with anti-Flag antibody-conjugated agarose. (E) WT, *Ripk1*^{K584R/+}, and *Ripk1*^{K584R/K584R} primary MEFs were treated with 50 ng/mL TNFα at various time points. The cell lysates were analyzed by phosphorylated and total JNK, ERK, p38, IKKα/β, IκBα, and actin as indicated. (F) Immortalized MEFs were pretreated with CHX, SM-164, or 5Z-7 for 2 h and then treated with TNFα as indicated at different time points. The cells were lysed with 0.5% Nonidet P-40 buffer and immunoprecipitated with anti-FADD antibody. The immunoprecipitated complexes and whole-cell lysates were analyzed by Western blotting with RIPK1.

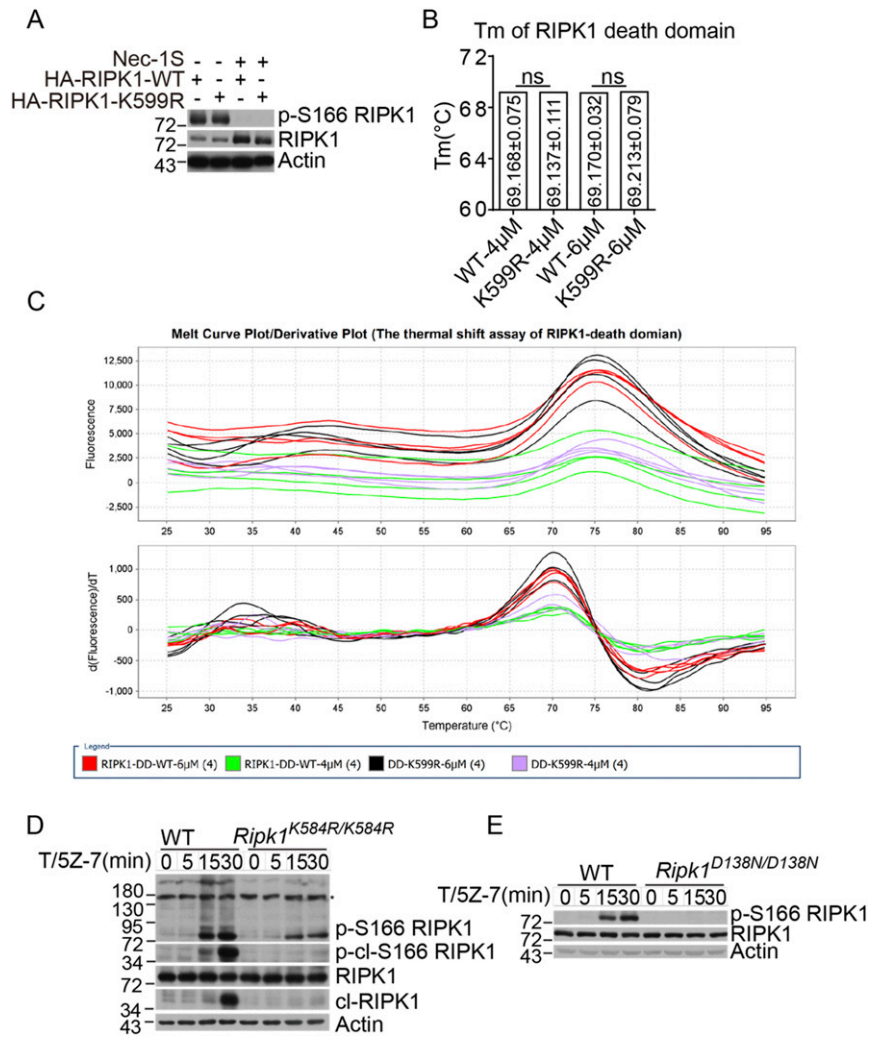


Fig. 54. The RIPK1 K584R mutation does not directly inhibit RIPK1 kinase activity or affect protein stability. (A) The RIPK1 KO 293T cells were transfected with HA-tagged RIPK1 WT or K599R expression plasmids in the presence or absence of 10 μ M Nec-1s. The cell lysates were collected 24 h after transfection and analyzed by Western blotting with the indicated antibodies. (B and C) The melting temperature (T_m) (B) and thermal stability profiles (C) of WT and K599R hRIPK1-DD. hRIPK1 (583-671AA) was purified from the *E. coli* expression system following overnight induction at 4 °C. hRIPK1-DD WT and K599R (4 μ M, 6 μ M) protein thermal stability were analyzed through the use of differential scanning calorimetry by real-time PCR and the melting temperatures were calculated by Protein Thermal Shift Software. Four replicates for each reaction were performed. (D and E) WT and *Ripk1*^{K584R/K584R} (D) and WT and *Ripk1*^{D138N/D138N} (E) immortalized MEFs were pretreated with 5Z-7 for 2 h and then treated with TNF α for the indicated time. The cells were lysed with 0.5% Nonidet P-40 buffer and analyzed by Western blotting analysis using p-S166 RIPK1 and RIPK1 antibodies.