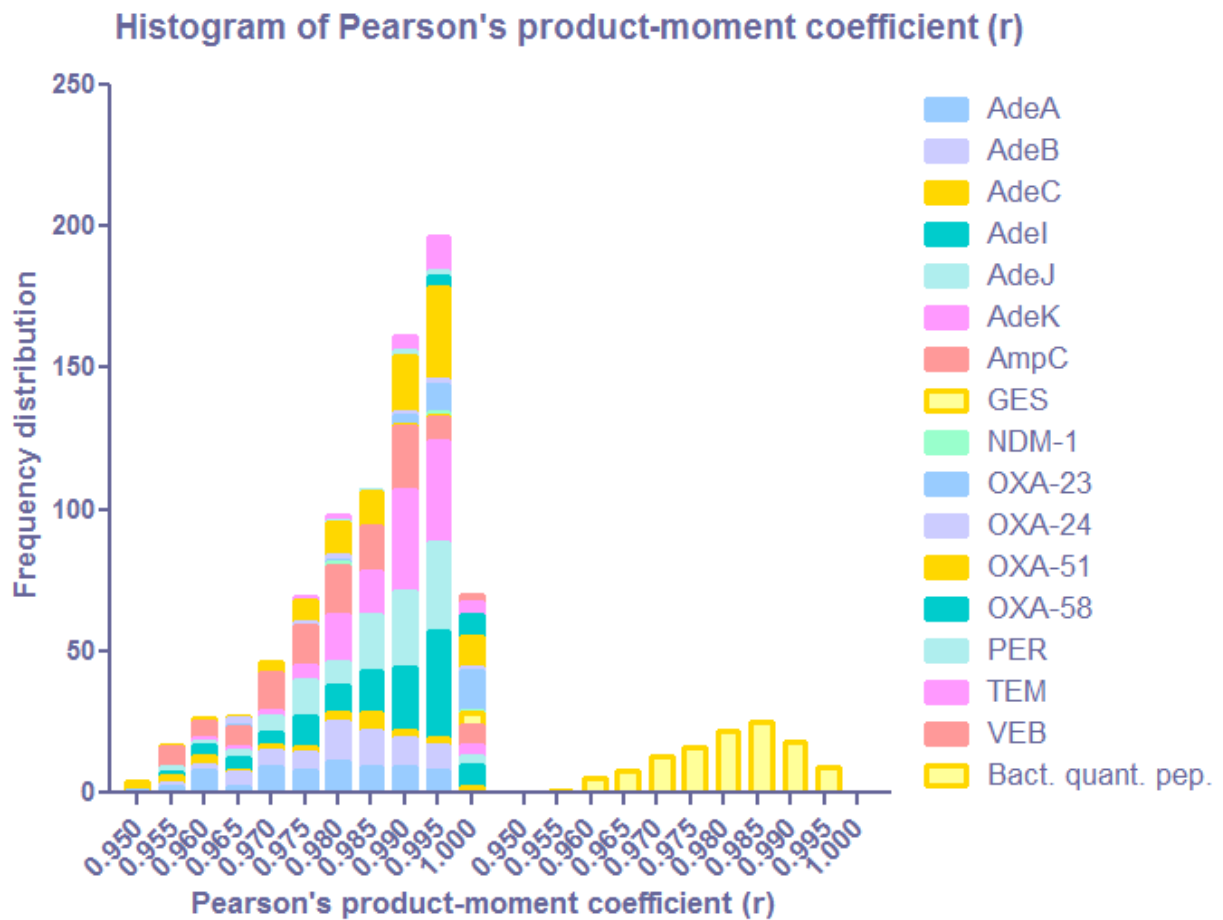


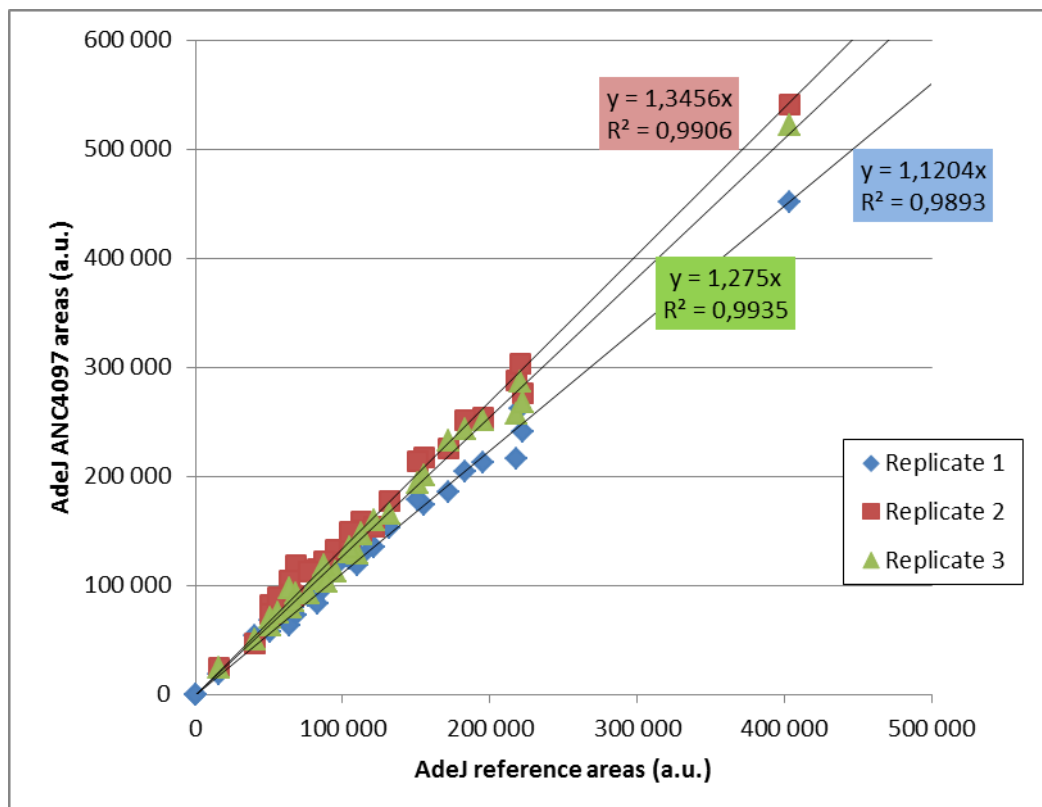
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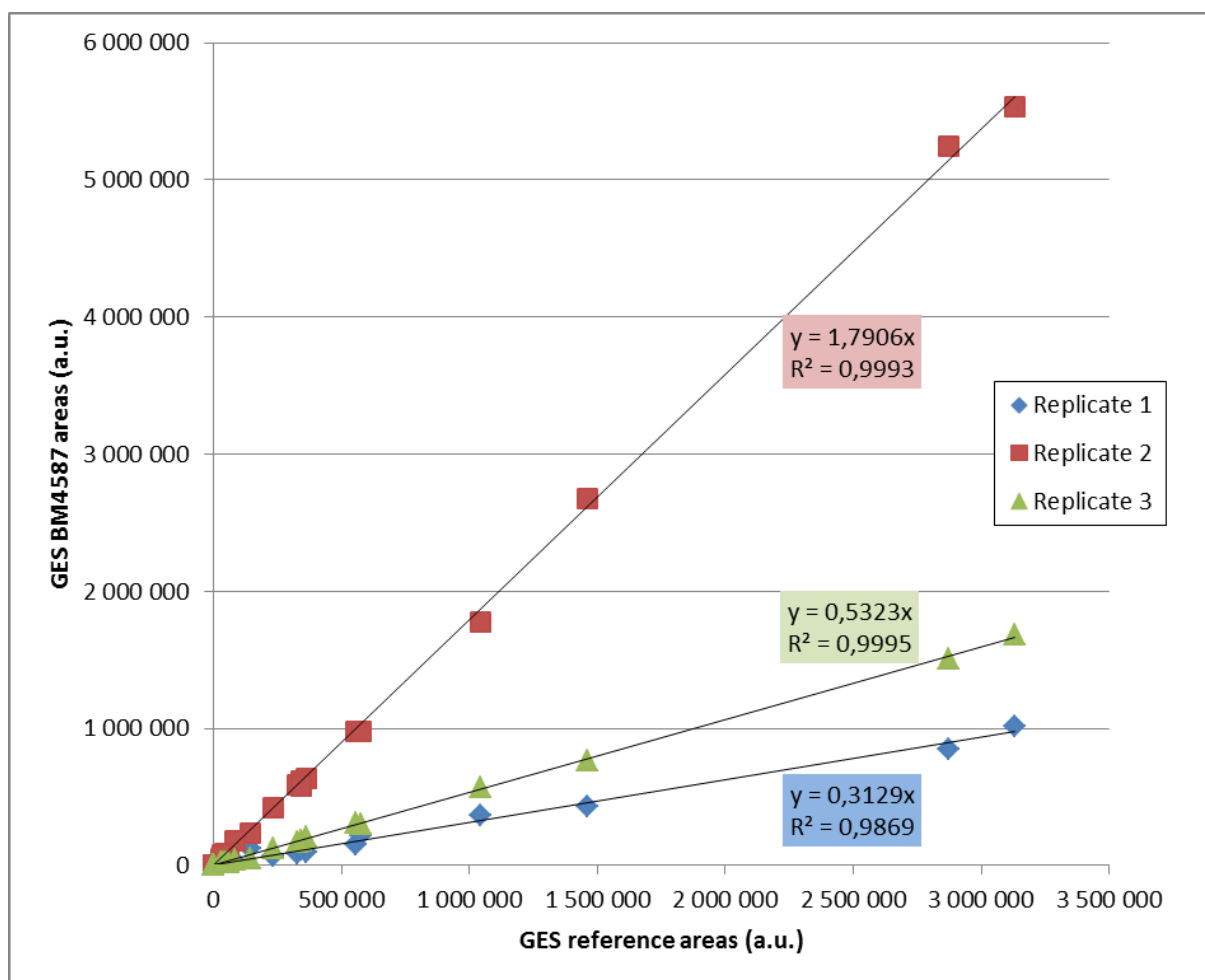


**Supplemental Fig. S1: Histogram of Pearson's product-moment coefficients (r)** calculated between the non-zero transition areas of each strain and reference transition areas estimated by averaging all the non-zero transition areas from the whole data set. The distribution of bacterial quantotypic peptides are on the right and the cumulative distribution of other proteins on the left.



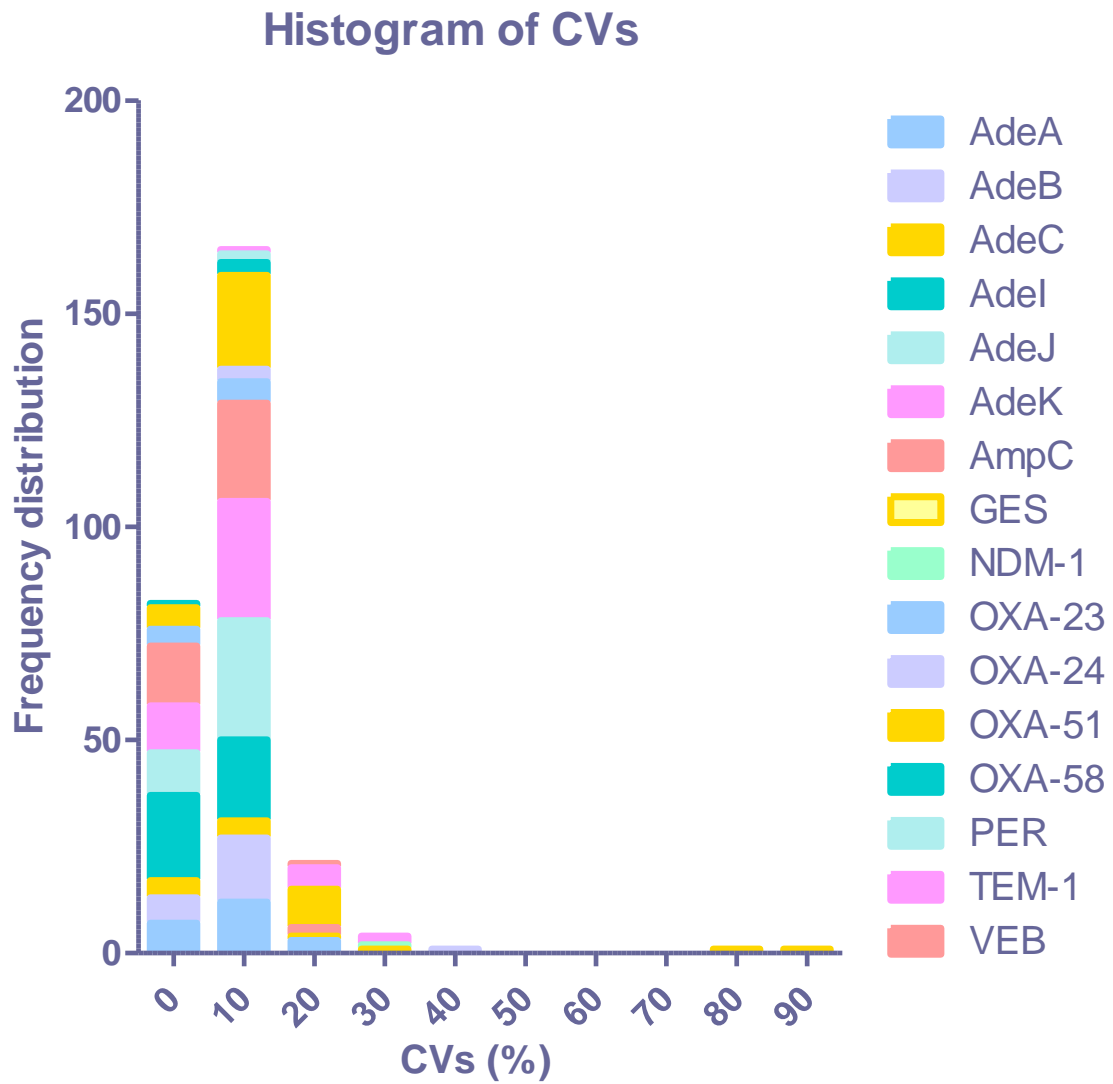
**Supplemental Fig. S2a: correlation curves for AdeJ measurement via SRM in ANC4097 sample, relative protein quantitation 2.4/4.9%.**

	Protein class	Protein Name	Peptide	Transitions	Retention Time(min)	Quantotypic and Proteotypic	Used for quantitation
247	Efflux pump	AdeJ	GEDLLDATLHAAK	AdeJ.GEDLLDATLHAAK.+2y8	15,44	Used	
248	Efflux pump	AdeJ	GEDLLDATLHAAK	AdeJ.GEDLLDATLHAAK.+3y7	15,44	Used	
249	Efflux pump	AdeJ	GEDLLDATLHAAK	AdeJ.GEDLLDATLHAAK.+3y8	15,44	Used	
250	Efflux pump	AdeJ	GMALNMIK	AdeJ.GMALNMIK.+2y5	16,8	Used	
251	Efflux pump	AdeJ	GMALNMIK	AdeJ.GMALNMIK.+2y6	16,8	Used	
252	Efflux pump	AdeJ	GMALNMIK	AdeJ.GMALNMIK.+2y7	16,8	Used	
253	Efflux pump	AdeJ	IATGANALDTAEAVEQR	AdeJ.IATGANALDTAEAVEQR.+3y4	13	Used	
254	Efflux pump	AdeJ	IATGANALDTAEAVEQR	AdeJ.IATGANALDTAEAVEQR.+3y5	13	Used	
255	Efflux pump	AdeJ	IATGANALDTAEAVEQR	AdeJ.IATGANALDTAEAVEQR.+3y6	13	Used	
256	Efflux pump	AdeJ	IAWGGSYINDFVDR	AdeJ.IAWGGSYINDFVDR.+2y6	18,5	Used	
257	Efflux pump	AdeJ	IAWGGSYINDFVDR	AdeJ.IAWGGSYINDFVDR.+2y7	18,5	Used	
258	Efflux pump	AdeJ	IAWGGSYINDFVDR	AdeJ.IAWGGSYINDFVDR.+3y6	18,5	Used	
259	Efflux pump	AdeJ	IWLDPK	AdeJ.IWLDPK.+2y3	13,56	Used	TopTra2TopPep3
260	Efflux pump	AdeJ	IWLDPK	AdeJ.IWLDPK.+2y5	13,56	Used	TopTra2TopPep3
261	Efflux pump	AdeJ	IWLDPK	AdeJ.IWLDPK.+2y6	13,56	Used	
265	Efflux pump	AdeJ	NYPTGLADK	AdeJ.NYPTGLADK.+2y5	9	Used	
266	Efflux pump	AdeJ	NYPTGLADK	AdeJ.NYPTGLADK.+2y6	9	Used	TopTra2TopPep3
267	Efflux pump	AdeJ	NYPTGLADK	AdeJ.NYPTGLADK.+2y7	9	Used	TopTra2TopPep3
268	Efflux pump	AdeJ	SFNNGFDR	AdeJ.SFNNGFDR.+2y4	9,4	Used	
269	Efflux pump	AdeJ	SFNNGFDR	AdeJ.SFNNGFDR.+2y5	9,4	Used	
270	Efflux pump	AdeJ	SFNNGFDR	AdeJ.SFNNGFDR.+2y6	9,4	Used	
271	Efflux pump	AdeJ	TPPETQIGSLIQR	AdeJ.TPPETQIGSLIQR.+2y6	14,9	Used	
272	Efflux pump	AdeJ	TPPETQIGSLIQR	AdeJ.TPPETQIGSLIQR.+2y7	14,9	Used	
273	Efflux pump	AdeJ	TPPETQIGSLIQR	AdeJ.TPPETQIGSLIQR.+3y6	14,9	Used	
274	Efflux pump	AdeJ	VELGSDNYQFDSK	AdeJ.VELGSDNYQFDSK.+2y3	12,14	Used	
275	Efflux pump	AdeJ	VELGSDNYQFDSK	AdeJ.VELGSDNYQFDSK.+2y4	12,14	Used	
276	Efflux pump	AdeJ	VELGSDNYQFDSK	AdeJ.VELGSDNYQFDSK.+2y7	12,14	Used	
277	Efflux pump	AdeJ	VMSEDHTDPVTATSR	AdeJ.VMSEDHTDPVTATSR.+3y4	8	Used	
278	Efflux pump	AdeJ	VMSEDHTDPVTATSR	AdeJ.VMSEDHTDPVTATSR.+3y5	8	Used	
279	Efflux pump	AdeJ	VMSEDHTDPVTATSR	AdeJ.VMSEDHTDPVTATSR.+3y7	8	Used	
280	Efflux pump	AdeJ	VYVQGDAGSR	AdeJ.VYVQGDAGSR.+2y6	5,6	Used	TopTra2TopPep3
281	Efflux pump	AdeJ	VYVQGDAGSR	AdeJ.VYVQGDAGSR.+2y7	5,6	Used	TopTra2TopPep3
282	Efflux pump	AdeJ	VYVQGDAGSR	AdeJ.VYVQGDAGSR.+2y8	5,6	Used	

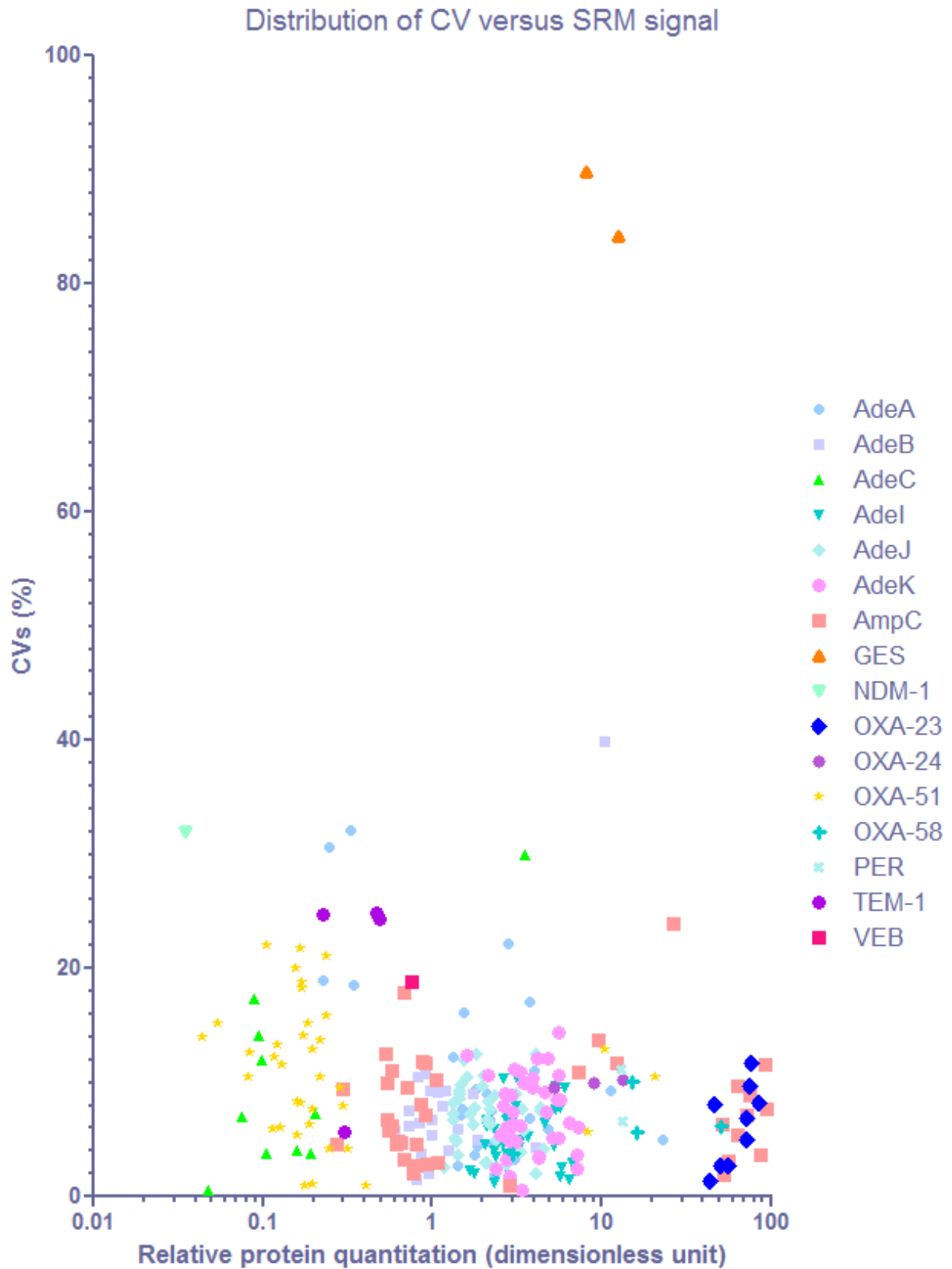


**Supplemental Fig. S2b: correlation curves for GES measurement via SRM in BM4701 sample, relative protein quantitation 8.1/89.8%.**

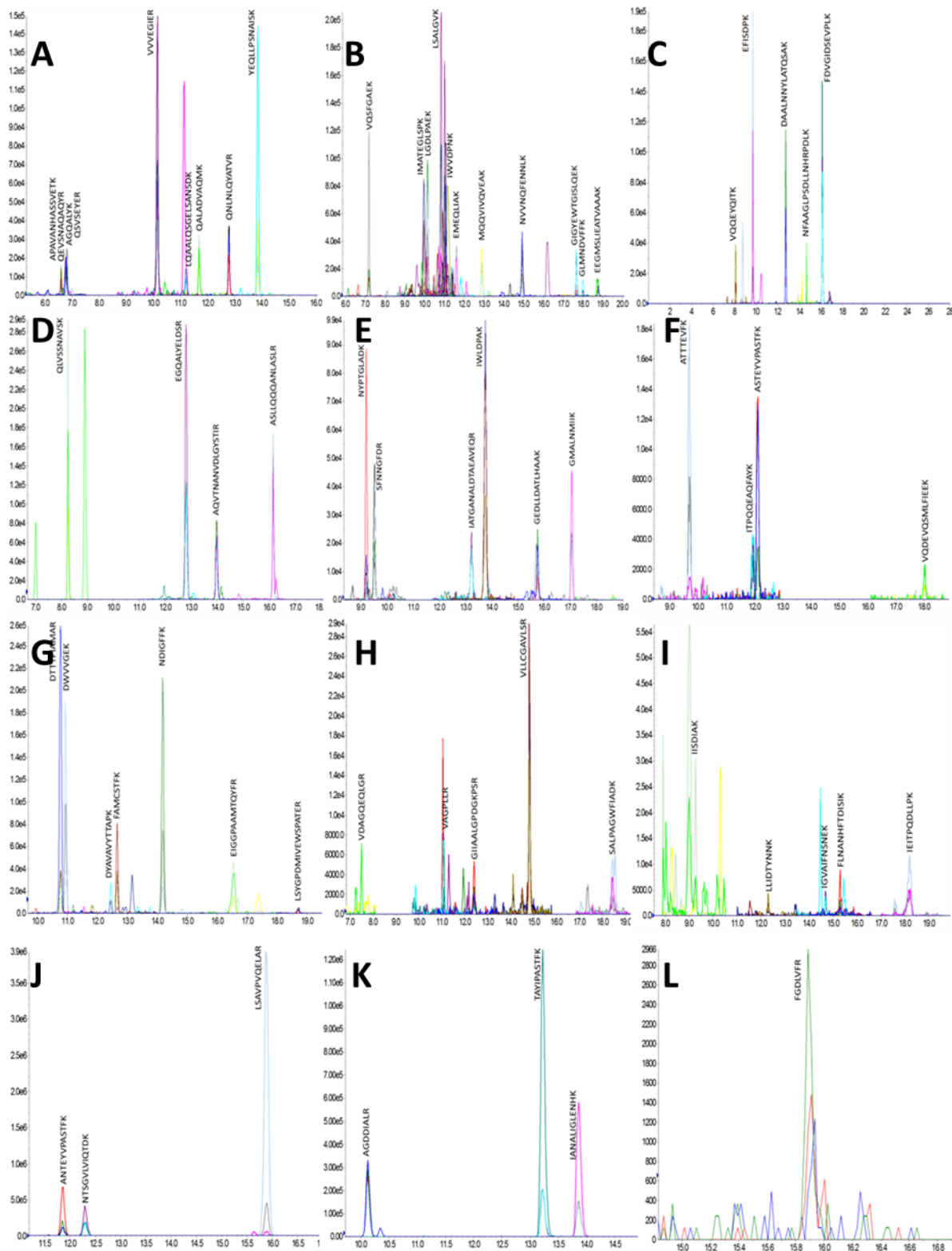
	Protein class	Protein Name	Peptide	Transitions	Retention Time(min)	Quantotypic and Proteotypic	Used for quantitation
343	Wide-Spectrum $\beta$ -lactamase	GES	AAEIGVAIVDPQGEIVAGHR	GES.AAEIGVAIVDPQGEIVAGHR.+3y13+2	15,5	Used	
344	Wide-Spectrum $\beta$ -lactamase	GES	AAEIGVAIVDPQGEIVAGHR	GES.AAEIGVAIVDPQGEIVAGHR.+3y14+2	15,5	Used	
345	Wide-Spectrum $\beta$ -lactamase	GES	AAEIGVAIVDPQGEIVAGHR	GES.AAEIGVAIVDPQGEIVAGHR.+3y16+2	15,5	Used	
346	Wide-Spectrum $\beta$ -lactamase	GES	AAQIGVAIVDPQGEIVAGHR	GES.AAQIGVAIVDPQGEIVAGHR.+3y13+2	15,5	Used	
347	Wide-Spectrum $\beta$ -lactamase	GES	AAQIGVAIVDPQGEIVAGHR	GES.AAQIGVAIVDPQGEIVAGHR.+3y14+2	15,5	Used	TopTra2TopPep3
348	Wide-Spectrum $\beta$ -lactamase	GES	AAQIGVAIVDPQGEIVAGHR	GES.AAQIGVAIVDPQGEIVAGHR.+3y16+2	15,5	Used	TopTra2TopPep3
352	Wide-Spectrum $\beta$ -lactamase	GES	DWVVGEK	GES.DWVVGEK.+2y4	11,2	Used	TopTra2TopPep3
353	Wide-Spectrum $\beta$ -lactamase	GES	DWVVGEK	GES.DWVVGEK.+2y5	11,2	Used	TopTra2TopPep3
354	Wide-Spectrum $\beta$ -lactamase	GES	DWVVGEK	GES.DWVVGEK.+2y6	11,2	Used	
355	Wide-Spectrum $\beta$ -lactamase	GES	DYAVAVYTTAPK	GES.DYAVAVYTTAPK.+2y6	12,5	Used	
356	Wide-Spectrum $\beta$ -lactamase	GES	DYAVAVYTTAPK	GES.DYAVAVYTTAPK.+2y7	12,5	Used	
357	Wide-Spectrum $\beta$ -lactamase	GES	DYAVAVYTTAPK	GES.DYAVAVYTTAPK.+2y8	12,5	Used	
370	Wide-Spectrum $\beta$ -lactamase	GES	LSYGPDMIVWSPATER	GES.LSYGPDMIVWSPATER.+3y5	19,1	Used	
371	Wide-Spectrum $\beta$ -lactamase	GES	LSYGPDMIVWSPATER	GES.LSYGPDMIVWSPATER.+3y6	19,1	Used	
372	Wide-Spectrum $\beta$ -lactamase	GES	LSYGPDMIVWSPATER	GES.LSYGPDMIVWSPATER.+3y7	19,1	Used	
373	Wide-Spectrum $\beta$ -lactamase	GES	NDIGFFK	GES.NDIGFFK.+2y4	14,4	Used	TopTra2TopPep3
374	Wide-Spectrum $\beta$ -lactamase	GES	NDIGFFK	GES.NDIGFFK.+2y5	14,4	Used	TopTra2TopPep3
375	Wide-Spectrum $\beta$ -lactamase	GES	NDIGFFK	GES.NDIGFFK.+2y6	14,4	Used	



**Supplemental Fig. S3a : histogram of CV distributions.** CVs were calculated using SRM measurements from three biological replicates.



**Supplemental Fig. S3b : CV versus average of relative protein quantitation.**  
 CVs and averages were calculated using SRM measurements from three biological replicates.

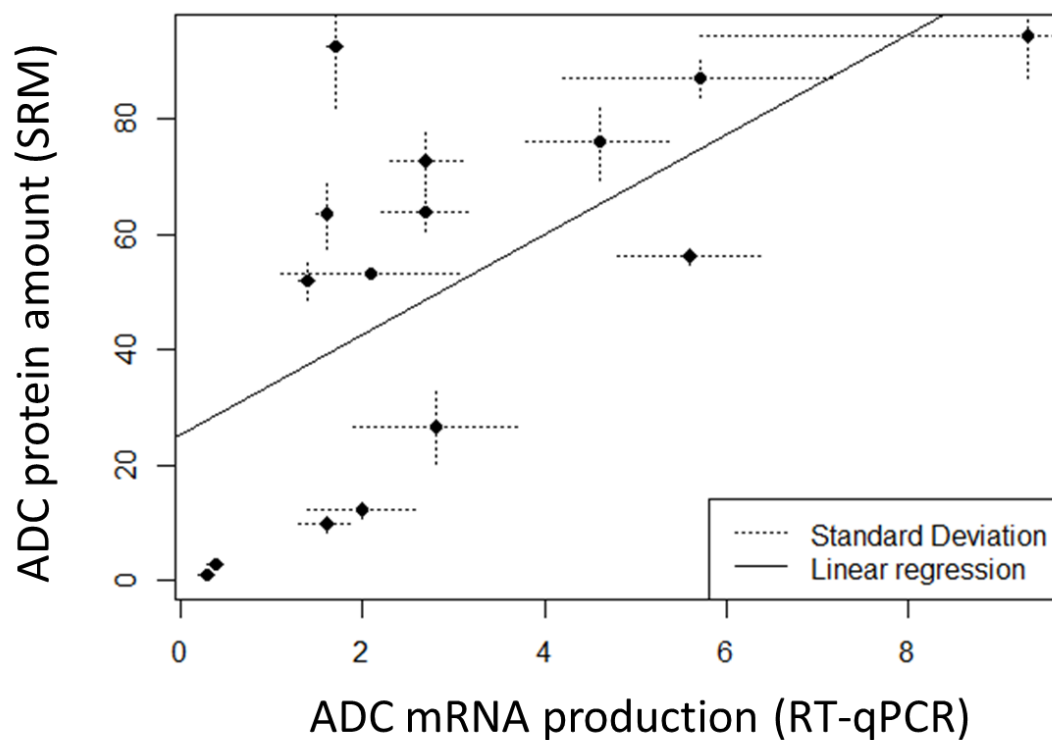


**Supplemental Fig. S4 : characteristic peptide peaks by SRM** (in replicate 1), from top to bottom, left to right. **A:** AdeA in BM4704 (4.8/5.9%); **B:** AdeB in CIP 70.34<sup>T</sup> (4.1/4.3%); **C:** AdeC in NIPH 290 (3.5/3.0); **D:** AdeI in NIPH 335 (5.7 /1.8%); **E:** AdeJ in NIPH 527 (1.8/12.5%); **F:** OXA-51 like in NIPH 190 (0.3/7.9%); **G:** GES in BM4701 (8.1/89.8%); **H:** TEM-1 in ANC 4097 (0.49/24.3%); **I:** VEB in AYE (0.76/18.8%); **J:** OXA-23 in ANC 4097 (46.1/2.7%); **K:** OXA-58 in NIPH 1362: 15.1/10%; **L:** NDM-1 in ANC 4097 (0.04/31.9%). Peaks were detected using MultiQuant<sup>TM</sup> 2.1 and areas under 3,000 Sciex arbitrary unit (a.u.) were set to null.

```
>acin_baum_NIPH_1362_redo
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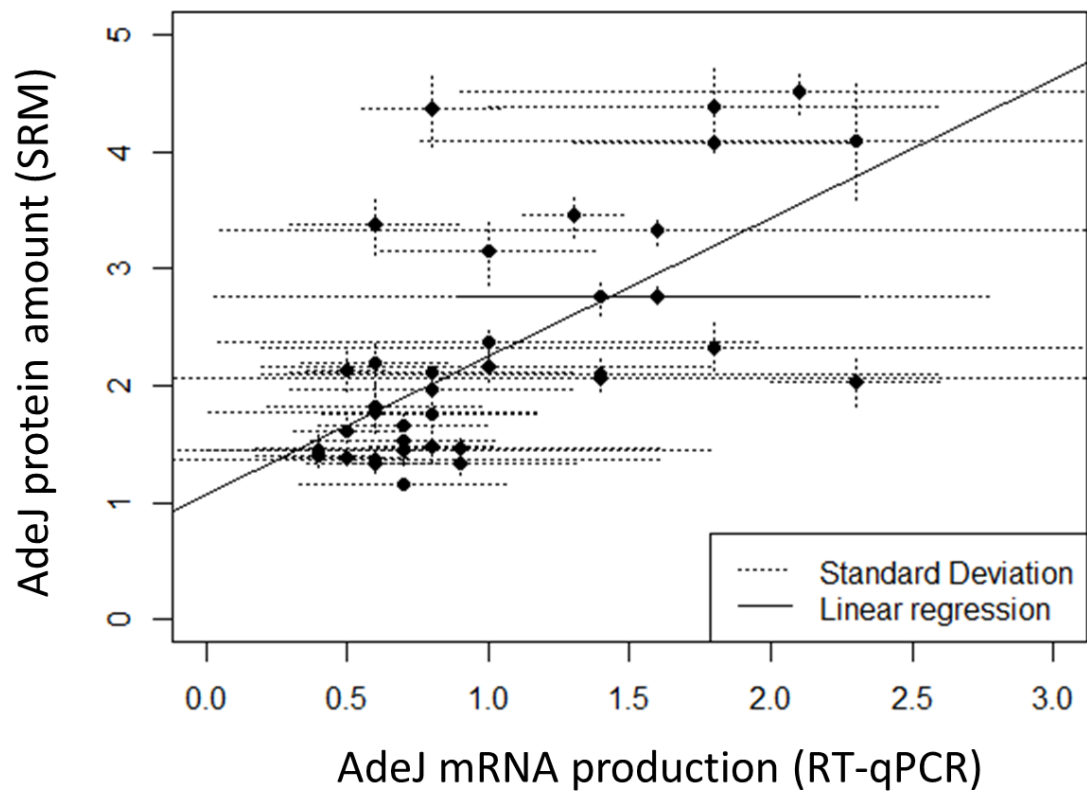
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NMQIGTEVDQFWLKGPLTITPIQEVKFVYDLAQGQLPFKPEVQQQVKEMLYVERRGENRLYAKSGWGMVAVDPO  
VGWYVGFVEKADGQVVAFALNMQMKAAGDDIALRKQLSLDVLDKLGVFHYL
```

**Supplemental Fig. S5** : OXA-58 gene identified in NIPH 1362 by new WGS.



**Supplemental Fig. S6a : ADC protein versus mRNA production.** Correlation between mRNA synthesis (RT-qPCR) expression and protein quantitation (SRM). For SRM, 3 samples were analyzed from 3 different biological cultures. For RT-qPCR, at least 2 samples were analyzed from the same biological culture. Pearson's product-moment correlation p-values was 0.01057.





**Supplemental Fig. S6b : AdeJ protein *versus* mRNA production.** Correlation between AdeJ mRNA synthesis (RT-qPCR) expression and protein quantitation (SRM). For SRM, 3 samples were analyzed from 3 different biological cultures. For RT-qPCR, at least 2 samples were analyzed from the same biological culture. Pearson's product-moment correlation p-values was  $1.379 \times 10^{-5}$ .

## Supplemental Tables

Sample Name	Reference (PMID numbers)	DNA Purity from NanoDrop A(260)/A(280)	DNA quantity from QuBit (ng/uL)
<b>NIPH1362</b>	Nemec 2004, Diancourt 2010	1.83	83.6
<b>BM4701</b>	Jeannot 2014	1.89	16.8
<b>BM4702</b>	Jeannot 2014	1.85	46.3
<b>BM4703</b>	Jeannot 2014	1.93	58.1
<b>BM4704</b>	Jeannot 2014	1.94	48.5
<b>BM4705</b>	Jeannot 2014	1.93	70.1
<b>BM4706</b>	Jeannot 2014	1.84	37.3
<b>BM4707</b>	Jeannot 2014	1.86	77.2
<b>BM4708</b>	Jeannot 2014	1.86	53.6
<b>BM4709</b>	Jeannot 2014	1.88	34.6
<b>BM4710</b>	Jeannot 2014	1.85	85.4
<b>BM4711</b>	Jeannot 2014	1.87	24.7
<b>BM4712</b>	Jeannot 2014	1.89	105.0
<b>BM4713</b>	Jeannot 2014	1.83	58.3

**Supplemental Table S1 : strains sequenced in this study.**

## Supplemental Table S2 to S4 : see specific files.

### OXA-23

>tr|A0A0E1PW85|A0A0E1PW85\_ACIBA OXA-23 protein OS=Acinetobacter baumannii NCGM 237  
GN=blaOXA-23 PE=4 SV=1  
MNKYFTCYVVASLFLSGCTVQHNLINETPSQIVQGHNQVIHQYFDEKNTSGVLVIQTDKKINLYGNALS  
RANTEYVPASTFKMLNALIGLENQKTDINEIFKWKGEKRSFTAWEKDMTLGEAMKLSAVPVYQELARR  
IGLDLMQKEVKRIGFGNAEIGQQVDNFWLVGPLKVTPIQEVFVSQLAHTQLPFSEKVVQANVKNMLLL  
EESNGYKIFGKTGWAMDIKPQVGWLTGWVEQPDGK  
IVAFALNMEMRSEMPASIRNELLMKSLKQLNII

### OXA-24

>tr|Q8RLA6|Q8RLA6\_ACIBA Beta-lactamase OS=Acinetobacter baumannii GN=blaOXA-33 PE=1 SV=1  
MKKFILPIFSISILVSLACSSIKTKSEDNFHISSQQHEKAIKSYFDEAQTQGVIIEGKLNSTYGNALARA  
NKEYVPASTFKMLNALIGLENHKATTNEIFKWDGKKRTYPMWEKDMTLGEAMALSAPVYQELARR  
TGLELMQKEVKRVNFGNTNIGTQVDNFWLVGPLKITPVQEVNFADDLAHNRLPFKLETQEEVKKMLLI  
KEVNGSKIYAKSGWGMGVTPQVGWLTGWVEQANGKKIPFSLNLEMKEGMSGIRNEITYKSLENLGII

### OXA-51

>tr|Q5QT35|Q5QT35\_ACIBA Beta-lactamase OS=Acinetobacter baumannii GN=oxa-51 PE=1 SV=1  
MNIKTLLLITSAIFISACSPYIVTANPNHSASKSDEKAEEKIKNLFNEVHTTGVLVIQQGQTQOSYGNDLAR  
ASTEYVPASTFKMLNALIGLEHHKATTTEVFKWDGQKRLFPWEKDMTLGDAMKASAIPVYQDLARRI  
GLELMSKEVKRVGYGNADIGTQVDNFWLVGPLKITPQQAQFAFKLANKTLPFSPKVQDEVQSMLFIE  
EKNGNKIYAKSGWGDVDPQVGWLTGWVVPQGNIVAFSLNLEMKKGIPSSVRKEITYKSLEQLGIL

### OXA-58

>tr|Q2TR58|Q2TR58\_ACIBA Beta-lactamase OS=Acinetobacter baumannii GN=blaOXA-58 PE=1 SV=1  
MKLLKILSLVCLISIGACAEHSMSRAKTSTIPQVNNIIDQNVQALFNEISADAVFVTYDGGQNIKKYGTH  
LDRAKTAYIPASTFKIANALIGLENHKATSTEIFKWDGKPRFFKAWDKDFTLGEAMQASTVPVYQELAR  
RIGPSLMQSELQRIGYGNMQIGTEVDQFWLKGPLTITPIQEVKVVYDLAQQQLPFKPEVQQQVKEMLYV  
ERRGENRLYAKSGWGMAVDPQVGWYVGFVEKADGQVVAFALNMQMKGDDIALRKQLSLDVLDKL  
GVFHYL

## Supplemental Table S5 : specificity of peptides used for OXA quantitation.

Peptides in green and yellow are unique and therefore specific. Sequence LSAVPVYQELAR (in blue) is shared by OXA-23 and OXA-24 but cannot be detected in OXA-24 digested using trypsin, because of upstream sequence alteration (DMTLGEAMK *versus* DMTLGEAMAL).