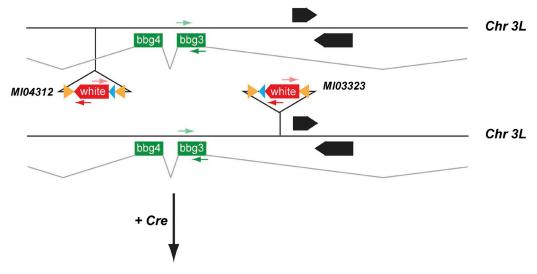
Supplemental material



Forest et al., https://doi.org/10.1083/jcb.201705107

A bbg locus after RMCE insertion of LoxP sites and White genes in the MI04312 and MI03323 elements here shown in trans



B bbg locus after Cre-mediated excision of the LoxP sites

C PCR validation of the deletion for the bbg¹⁰ and bbg³⁴ alleles

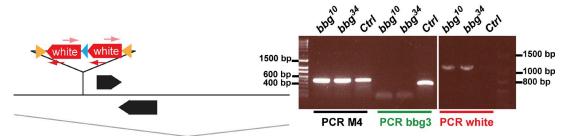


Figure S1. Generation of new bbg alleles by recombinase-mediated cassette exchange. (A) Schematic representation of the bbg locus focusing on the third and fourth exons (bbg3 and bbg4 in green). The two MiMIC insertions flanking these exons have been engineered by recombination-mediated cassette exchange (RMCE) to introduce a w+ marker and a LoxP site (in blue). Green and red arrows represent specific primer sets used for mapping. (B) bbg locus after a boost of Cre recombinase. (C) PCR validation of these events on adult fly homozygous genomic DNA. PCR M4 is a PCR with primers for the E(spl)-m4 gene and was unaffected in this procedure. This approach led to specific deletions (shown in this image are the bbg10 and bbg34 alleles) at high frequency.

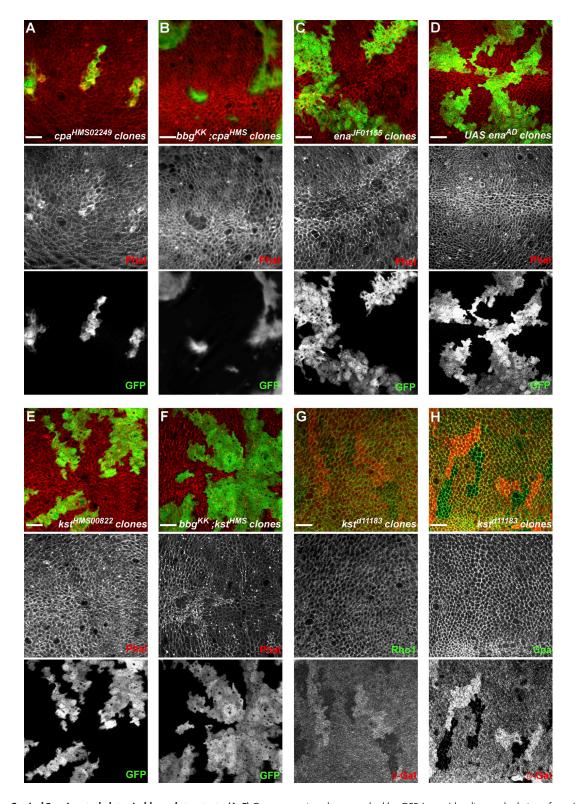


Figure S2. Cortical F-actin cytoskeleton in *bbg* or *kst* mutants. (A–F) Overexpression clones marked by GFP (green) leading to depletion of cpa (A), double depletion of cpa and bbg (B), depletion of ena (C), overexpression of ena (D), depletion of kst (E), or double depletion of bbg and kst (F), and showing the organization of the cortical apical F-actin (red). No changes in F-actin could be observed after ena level modulation or kst depletion. A strong decrease of apical F-actin was seen only in bbg-depleted cells (B and F), imposing even on the cpa depletion—induced F-actin enrichment (A). (**G and H**) kst^{d+1183} mutant clones in the larval wing disc marked by the absence of β -galactosidase (red) and marked for the actin regulators Rho1 (G, green) and Cpa (H, green). No significant difference was observed between WT and kst mutant cells. Bars, 10 μ m.

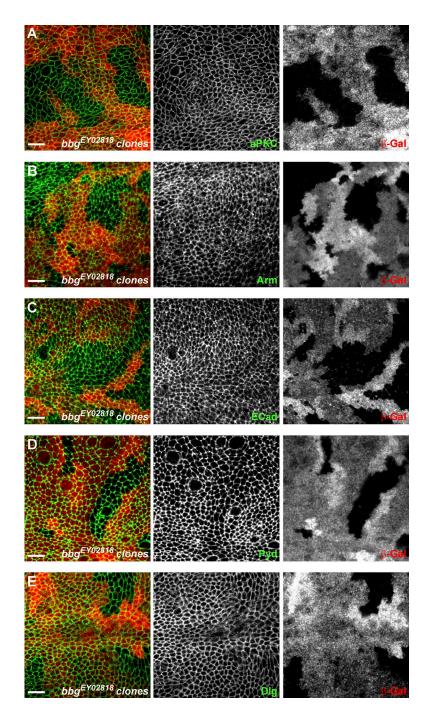


Figure S3. **Epithelial apical–basal polarity is unaffected in** bbg **mutants. (A–E)** $bbg^{EYO2818}$ mutant clones in the larval wing disc marked by the absence of β -Galactosidase (red) and marked for the apical–basal markers aPKC (A, green), armadillo (B, green), E-Cad (C, green), polychaetoid (D, green), and Dlg (E, green). No significant difference was observed between WT and bbg mutant cells. Bars, $10 \mu m$.

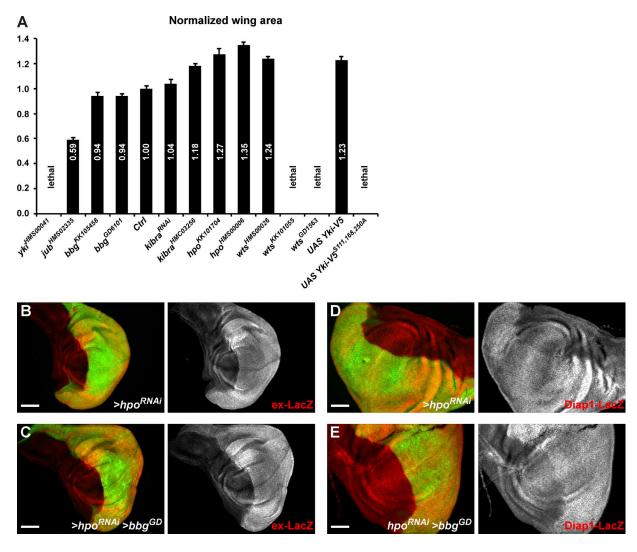


Figure S4. **Bbg and the Hippo pathway.** (A) Adult female *Drosophila* wing size after nubG4-driven RNAi knockdown at 25°C of the indicated genotypes. The wing area for each genotype is expressed as a ratio compared with the mean wing area of the nubG4>UAS GFP controls (Ctrl). SD is shown; n=17-31 independent female wings. Some combinations did not produce any viable adults and are labeled "lethal." (B–E) Pouch region of third instar larva wing imaginal discs. Effect of hh-Gal4-driven RNAi knockdown in the posterior compartment marked by GFP (green) on the expression of the Yki activity reporters ex-LacZ (B and C, red) and Diap1-LacZ (D and E, red). The up-regulations of ex-LacZ and Diap1-LacZ caused by dx-knockdown (B and D) were not affected by the further knockdown of dx-by dx-caused by dx-caused dx-caus