

*Supporting Information for*

**Nanowire Arrays Restore Vision in Blind Mice**

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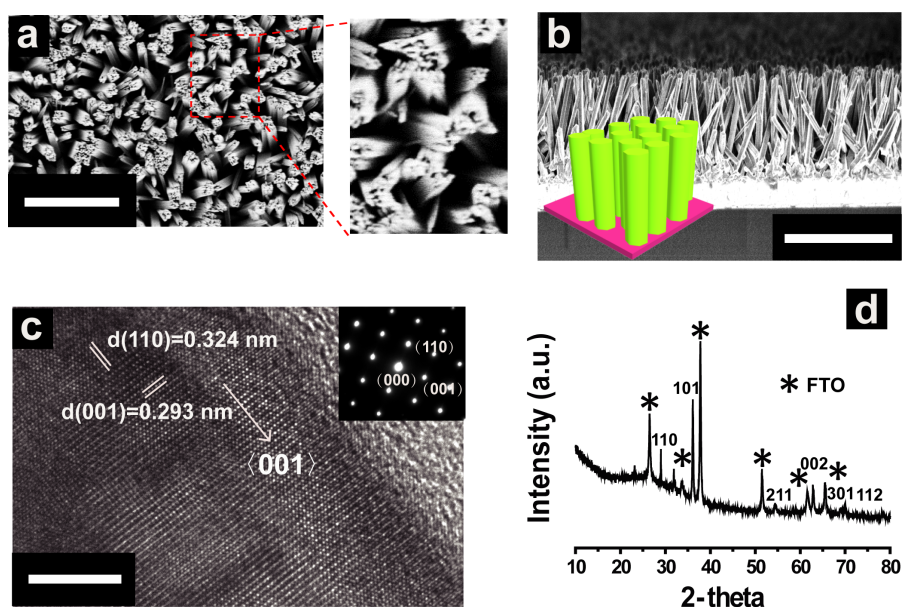
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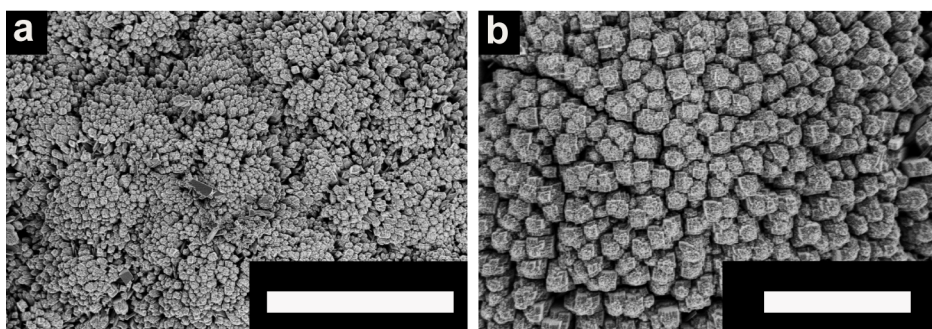
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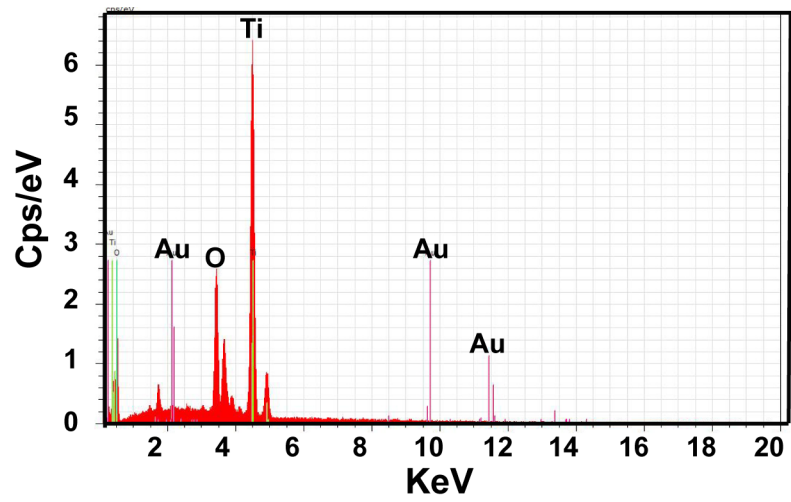
## Supporting Figures



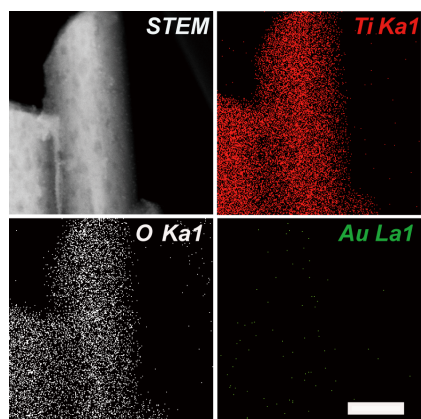
**Supplementary Figure 1** | (a) Top-view and (b) side-view SEM images of TiO<sub>2</sub> NW arrays on an FTO substrate. (c) HRTEM image and (inset) selected area electron diffraction pattern of a representative single-crystalline TiO<sub>2</sub> NW. (d) XRD pattern of TiO<sub>2</sub> NWs on an FTO substrate. The peaks of FTO are indicated by asterisks. Scale bars: 1  $\mu$ m (a), 2  $\mu$ m (b), 5 nm (c).



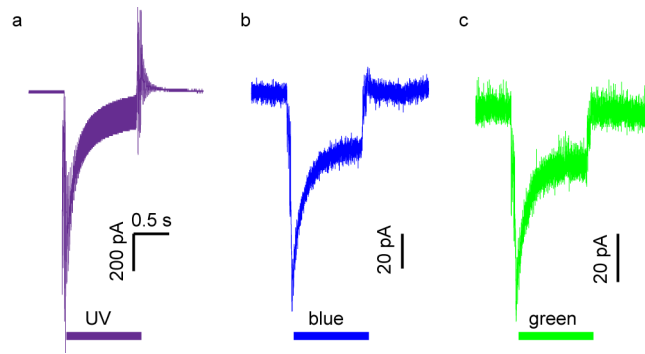
**Supplementary Figure 2 | (a, b)** Top-view SEM images of Au nanoparticle-decorated TiO<sub>2</sub> nanowire arrays on the PDMS. Scale bars: 10 μm (a), 2 μm (b).



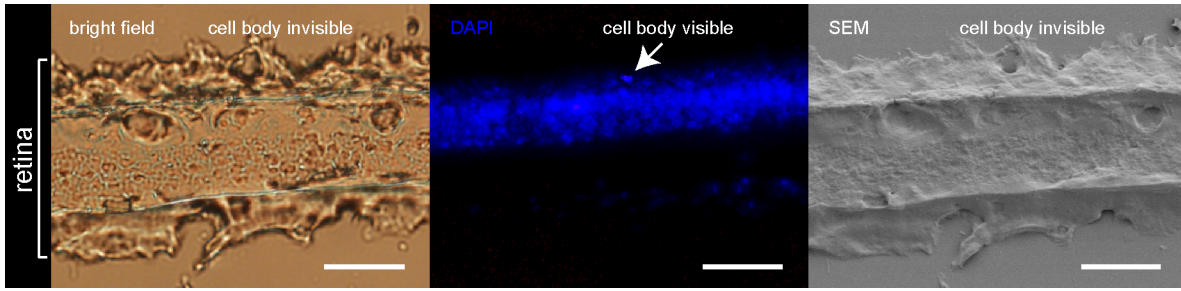
**Supplementary Figure 3** | EDX spectrum of the Au-TiO<sub>2</sub> NWs. The molar percentages of Au and Ti were calculated as ~ 0.11% and 71.77%, respectively.



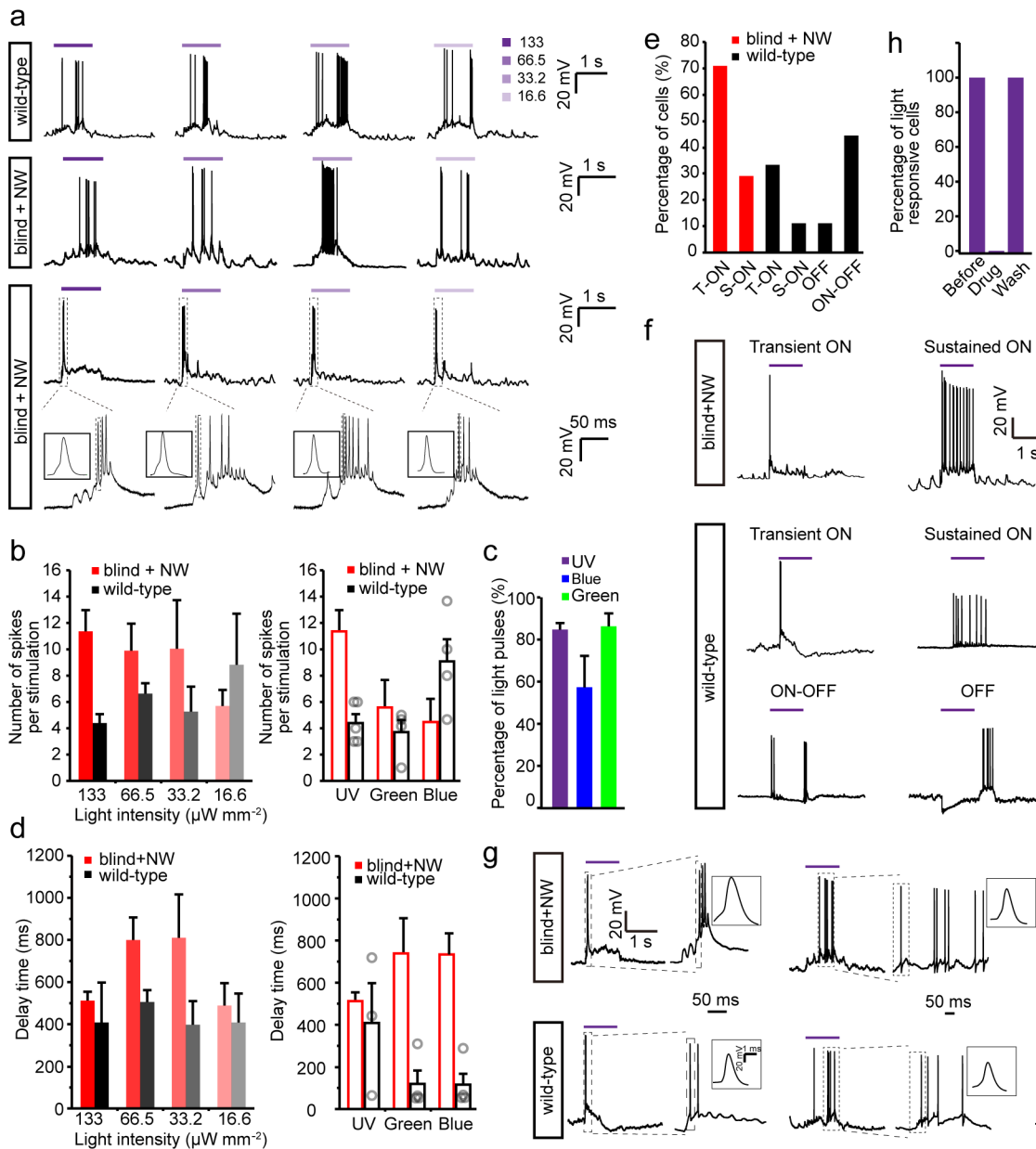
**Supplementary Figure 4** | Scanning TEM image and corresponding EDX elemental mapping data of Au-TiO<sub>2</sub> NWs. Scale bar: 100 nm.



**Supplementary Figure 5** | Photocurrent versus time from an Au-TiO<sub>2</sub> NW array generated by illumination of UV (a) , blue (b) and green (c) light, respectively. Different filters were applied to obtain UV (375/28 nm, 133  $\mu\text{W mm}^{-2}$ ), blue (470/20 nm, 691  $\mu\text{W mm}^{-2}$ ) and green light (546/12 nm, 470  $\mu\text{W mm}^{-2}$ ).



**Supplementary Figure 6** | Bright light, DAPI fluorescence and Scanning Electron Microscope images of the same 30  $\mu\text{m}$  retinal slice. Scale bars: 50  $\mu\text{m}$ .

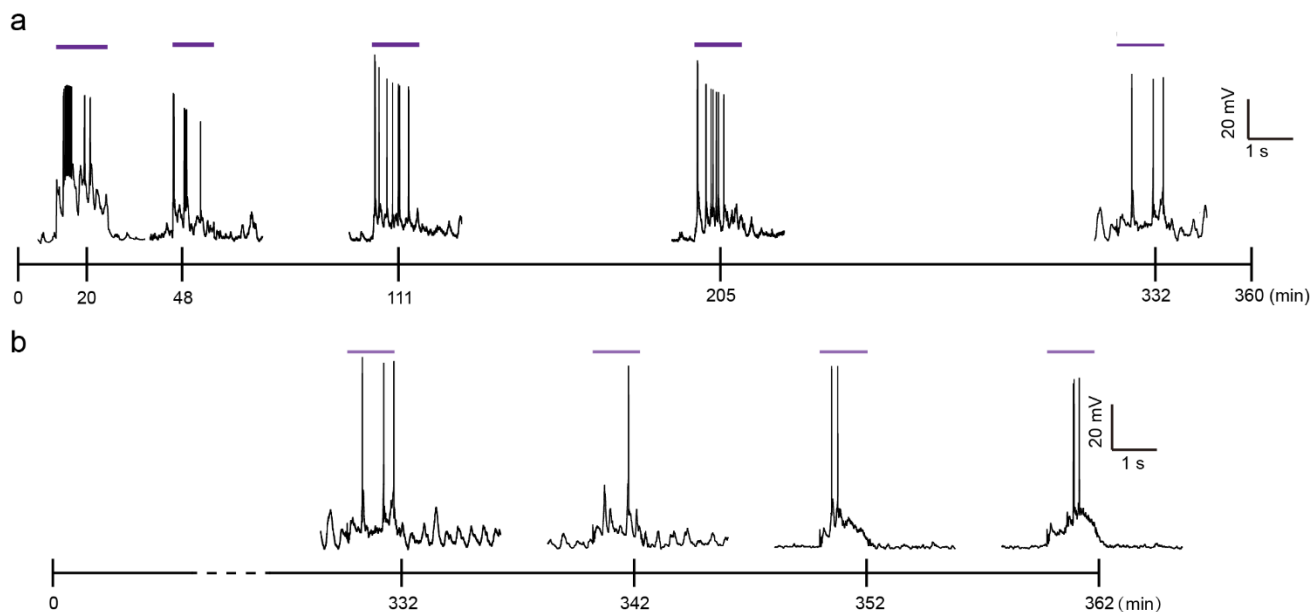


**Supplementary Figure 7 | Light responses in NW array-interfaced blind mouse retinas. (a)**

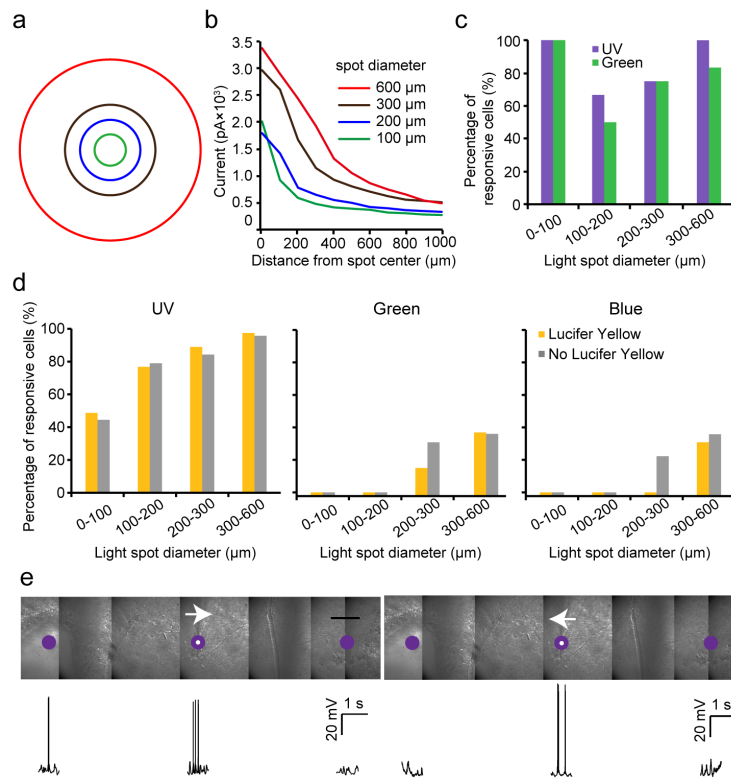
RGC responses to UV light with different light intensities. The light intensity values are shown with units of  $\mu\text{W mm}^{-2}$ . Purple lines represent the presence of UV light. The bottom panel shows zoom-in spike activities. **(b)** Number of spikes per stimulation. Blind + NW:  $n = 43, 23, 13, 17$  RGCs in 23, 17, 11, 10 retinas for 133, 66.5, 33.2, 16.6  $\mu\text{W mm}^{-2}$ , respectively; Green,  $n = 8$  RGCs, 5 retinas; Blue,  $n = 5$  RGCs, 4 retinas. Wild-type:  $n = 5, 5, 7, 6$  RGCs in 4, 4, 6, 4 retinas for 133, 66.5, 33.2, 16.6  $\mu\text{W mm}^{-2}$ , respectively; Green,  $n = 4$  RGCs, 2 retinas; Blue,  $n = 4$  RGCs, 3 retinas. **(c)** Percentage of light pulses triggering spiking activities in all light-responsive RGCs for UV ( $n = 65$



RGCs, 25 retinas), blue (n = 5 RGCs, 4 retinas) and green (n = 9 RGCs, 6 retinas) light in NW array-interfaced blind retinas. **(d)** Latency to the onset of near UV, green, blue light. Blind + NW: n = 45, 22, 13, 18 RGCs in 24, 17, 11, 10 retinas for 133, 66.5, 33.2, 16.6  $\mu\text{W mm}^{-2}$ , respectively; Green, n = 11 RGCs, 5 retinas; Blue, n = 5 RGCs, 4 retinas. Wild-type: n = 3, 3, 6, 6 RGCs in 3, 3, 3, 4 retinas for 133, 66.5, 33.2, 16.6  $\mu\text{W mm}^{-2}$ , respectively; Green, n = 4 RGCs, 2 retinas; Blue, n = 4 RGCs, 3 retinas. **(e)** Percentages of RGCs with different light-response patterns and examples shown in **(f)**. Blind + NW: n = 86 RGCs, 25 retinas; Wild-type: n = 9 RGCs, 9 retinas. **(g)** Spike details in wild-type and NW-interfaced blind retinas. **(h)** Percentage of responsive RGCs before, during and after application of glutamate receptor antagonist in light-responsive RGCs (n = 6 RGCs, 6 retinas). Data are presented as mean and S.E.M.



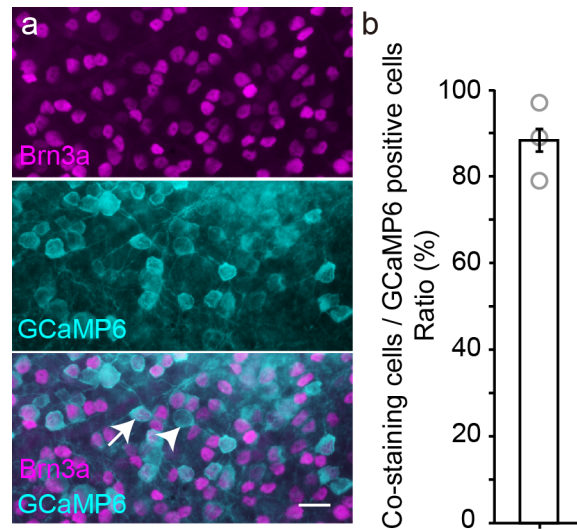
**Supplementary Figure 8 | Time lapse responses in NW array interfaced retinas. (a)** Light responses of RGCs *in vitro*. Time 0 represents the moment of putting the retina on top of the NW array. The first RGC was successfully patched 20 min later. **(b)** Light responses of one RGC *in vitro*. This RGC was successfully patched 332 min after putting the retina on top of the NW array.



**Supplementary Figure 9** | (a) Schematic for the light spot with different diameter. (b) The photocurrent of NW array triggered by different-size light spot measured in different distance from center of the spot. (c) Percentage of light-responsive cells with different light spot diameter in wild-type mice. UV: n = 9, 9, 8, 6 RGCs in 6, 6, 6, 5 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. Green: n = 9, 8, 8, 6 RGCs in 6, 5, 6, 5 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. (d) Percentage of light-responsive cells in response to different-size light spot with or without Lucifer Yellow (LY) in the internal solution. UV: LY, n = 37, 43, 27, 38 RGCs in 20, 21, 15, 20 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. No-LY, n = 18, 19, 19, 23 RGCs in 13, 13, 13, 16 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. Green: LY, n = 24, 19, 20, 38 RGCs in 14, 9, 11, 17 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. No-LY, n = 13, 12, 13, 25 RGCs in 9, 9, 9, 14 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. Blue: LY, n = 13, 11, 11, 13 RGCs in 6, 5, 6, 6 retinas for 0 – 100, 100 – 200, 200 – 300 and 300 – 600  $\mu\text{m}$ , respectively. No-LY, n = 8, 9, 9,

14 RGCs in 5, 6, 6, 8 retinas for 0 – 100, 100 – 200, 200 -300 and 300 – 600  $\mu\text{m}$ , respectively. **(e)**

The responses to moving UV light spots (purple circles, 55  $\mu\text{m}$ ) with two opposite directions. Arrows represent the moving direction of spots. White dots represent the location of the neuron. Top panel: DIC images of retina and three example locations of the light spot. Bottom panel: The corresponding light responses of different light spot locations. Average moving velocity: 0.0936  $\text{mm s}^{-1}$  (left), 0.1674  $\text{mm s}^{-1}$  (right). Scale bar: 100  $\mu\text{m}$  **(e)**.



**Supplementary Figure 10 | Most of the GCaMP6 positive cells are RGCs. (a)**

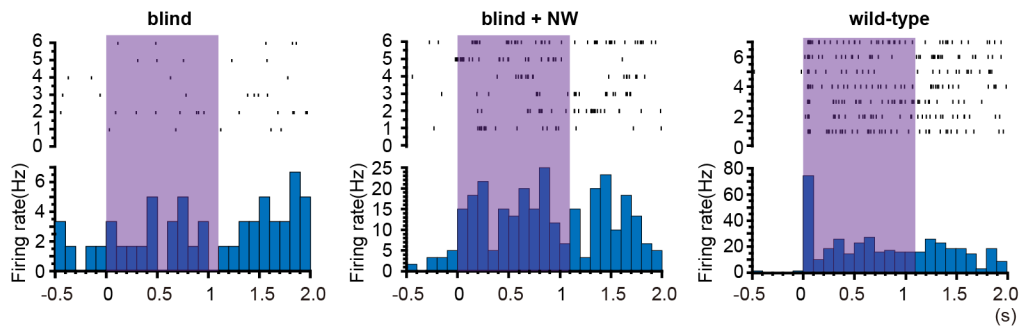
Immunohistochemistry for Brn3a (up), GCaMP6 (middle), and merge of Brn3a and GCaMP6

(bottom) in the retina of blind mice. Arrowhead with long tail: co-staining of Brn3a and GCaMP6.

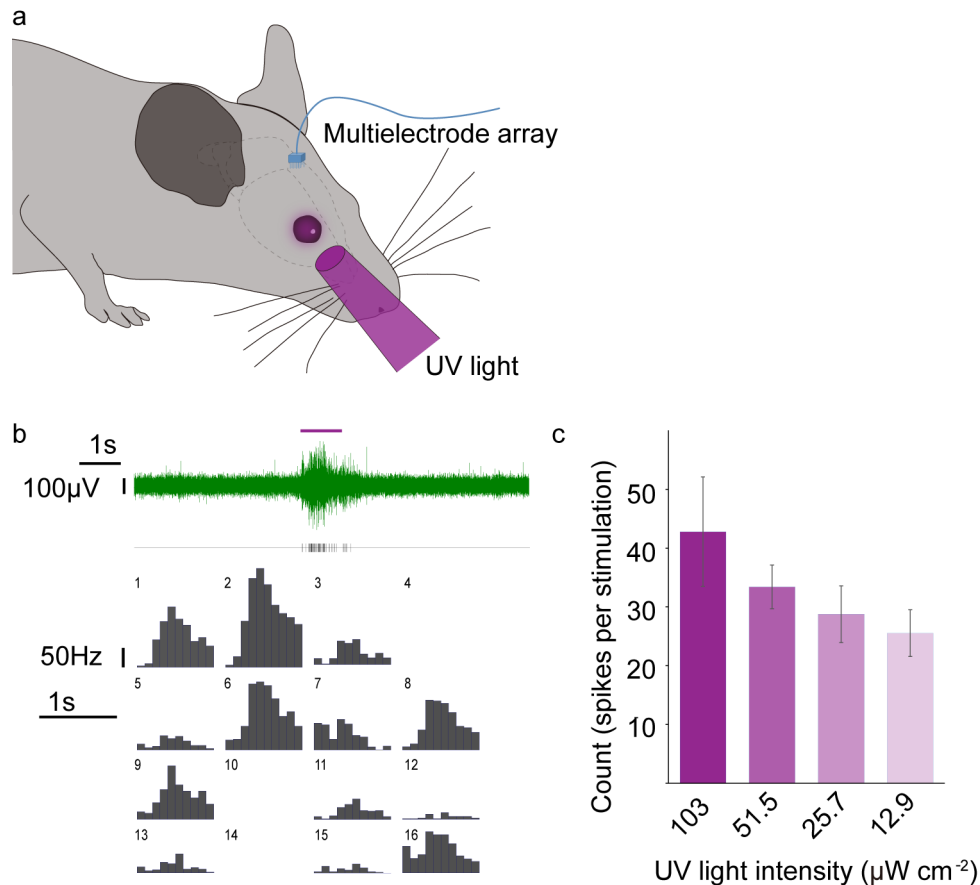
Arrowhead with short tail: GCaMP6. **(b)** The ratio of Brn3a and GCaMP6 co-staining cells to

GCaMP6 cells in the retina of blind mice (n = 3 areas from 1 retina). Data are presented as mean and

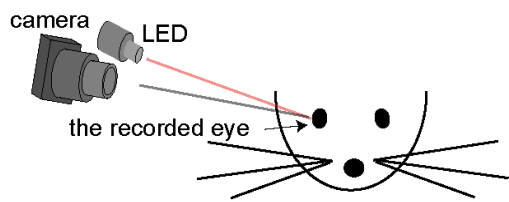
S.E.M. Scale bar: 20  $\mu$ m **(a)**.



**Supplementary Figure 11 | Raster plots and post stimulus time histogram (PSTH) of spikes from SC neurons.** Left: blind mice, Middle: NW array-implanted blind mice 2 days after implant, Right: wild-type mice. The purple shade area indicates the presentation of light.



**Supplementary Figure 12 | Response to UV light in primary visual cortex *in vivo* in anaesthetized wild-type mice** (a) Schematic of the extracellular recording setup *in vivo*. UV light was presented in front of one eye through a light guide. Spiking activities in the contralateral primary visual cortex were recorded using a 16 channel multi-electrode array. (b) Top: Green trace showed the high-pass filtered (>300 Hz) raw data recorded from one channel. Middle: Raster plot of spikes in the top trace. Each vertical black line represented one spike. Bottom: PSTH (post-stimulation time histogram) firing rate of neurons recorded from each channel. The numbers represented the channel. Each column represented average firing rate of the neuron within 100 ms. Channel 4 recorded the timing for the UV light stimulation. (c) Average number of spikes per 1 sec stimulation of all the neurons ( $n = 15$ ) at different UV light intensity. Data are presented as mean and S.E.M.



**Supplementary Figure 13 | Schematic diagram of pupillary light reflex experiment.**