Supplementary	information
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Purinergic dysregulation causes hypertensive glaucoma-like optic neuropathy

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Supplementary Figures

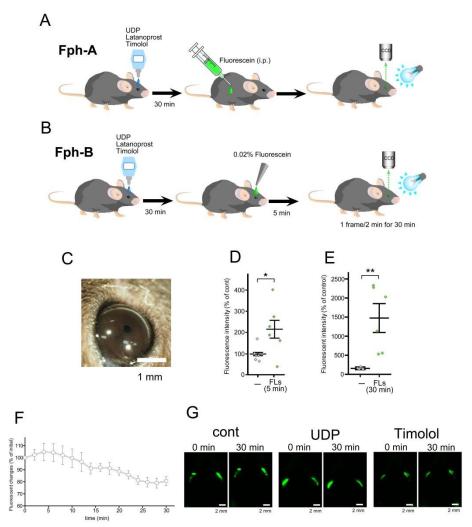


Figure S1. Aqueous humor dynamics assays.

(A) Fph-A procedures. Eye drops were administrated for 30 mins followed by fluorescein injection (i.p.). The fluorescence levels in the anterior chamber were monitored every 2 mins. (B) Fph-B procedures. Eye drops were administrated for 30 min followed by topical administration of fluorescein for 5 mins. After washing eyes with 500 μ l of saline five times, fluorescence images of the anterior chamber were observed. (C) Pupil dilations were not induced for the assays to observe fluorescence limited to the anterior chamber. (D, E) Fluorescence levels of isolated aqueous humor at (D) 5 and (E) 30 mins after i.p. injection of fluorescein. Fluorescence levels in isolated aqueous humor showed a significant increase (n = 5, *p < 0.05 and **p < 0.01, unpaired *t*-test). (F) Fluorescence changes in control mice. During the first 4–6 minutes, the fluorescence level was slightly increased followed by a gradual reduction. (G) Representative fluorescence images at 0 and 30 mins. Data are expressed as the mean \pm SEM.

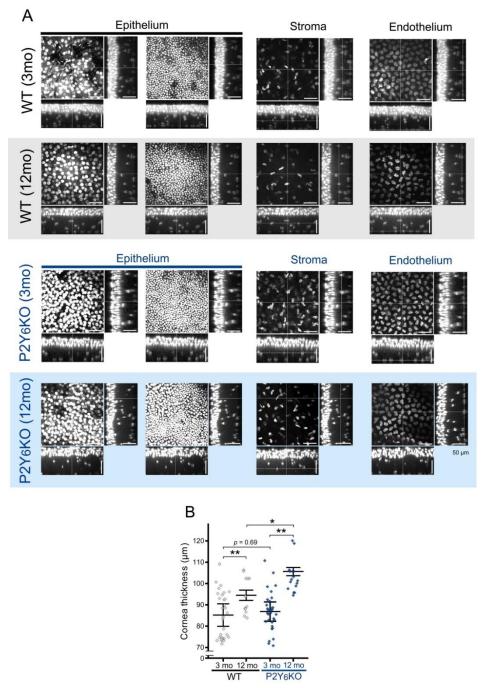


Figure S2. Comparison of central cornea thickness between WT and P2Y₆KO mice.

(A) Layer structure of the cornea. No significant structural differences were observed in the epithelial, stromal and endothelial cell layers between WT and P2Y₆KO mice. (B) Central cornea thickness (CCT). No significant differences in CCT at 3 months of age were observed. At 12 months, CCTs in both WT and P2Y₆KO mice were increased compared with 3-month-old groups, and P2Y₆KO had a thicker CCT than that of WT mice (n = 20, *p < 0.05, **p < 0.01, one-way ANOVA followed by Fisher's LSD test). Values are the mean \pm SEM. Scale bars: 50 μ m.

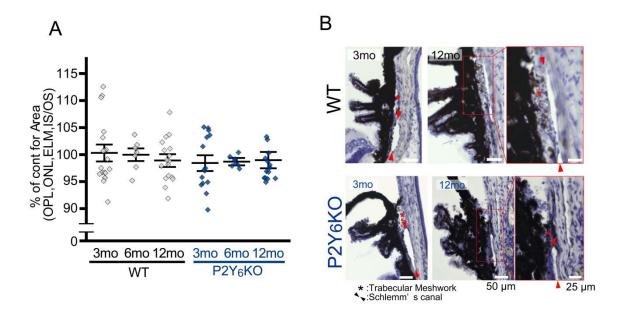


Figure S3. Histological differences in WT and P2Y₆KO mice.

(A) No changes in area size of the outer retina (OPL, ONL, ELM, IS/OS) were observed by OCT analysis (n = 10–20, one-way ANOVA followed by Fisher's LSD test). (B) No significant abnormalities were observed in anterior chamber structures in $P2Y_6KO$ mice. Values are the mean \pm SEM. Scale bars, 25 μ m in (B, insert) and 50 μ m in (B).

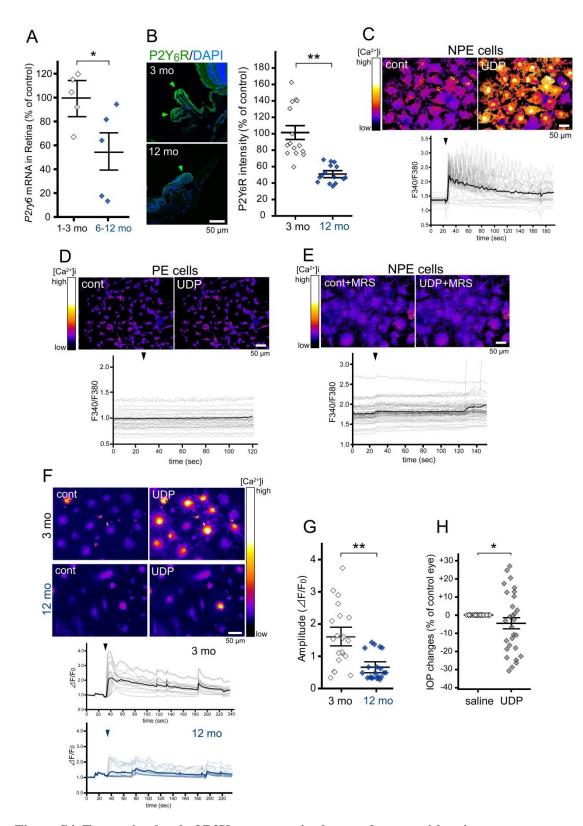


Figure S4. Expression level of P2Y₆ receptors in the eye changes with aging.

(A) The level of P2ry6 mRNA in the retina was significantly reduced with age (n = 10, *p = 0.05, unpaired t-test). Retinal P2ry6 mRNA levels in WT mice at 6–12 months were

significantly lower compared with mice aged 1–3 months. (**B**) Immunoreactivity for P2Y₆R in NPE cells of the ciliary body was decreased with age. Fluorescence intensities for P2Y₆R signals in NPE cells of 3-month-old mice were significantly lower compared with those of 12-month-old mice (n = 10, **p < 0.01, unpaired t-test). (**C**) Nonpigmented epithelial (NPE) cells showed clear Ca²⁺ responses evoked by UDP (100 μ M, arrow). (**D**) Pigmented epithelial (PE) cells did not respond to UDP. (**E**) UDP-evoked responses in NPE cells were clearly blocked by MRS2578 (30 μ M). Ca²⁺ imaging in (**B–D**) were performed in epithelial cells from 3-month-old WT mice. (**F** and **G**) Age-associated reduction in UDP-evoked Ca²⁺ responses in NPE cells. NPE cells from 12-month-old WT mice showed smaller amplitudes of Ca²⁺ transients evoked by UDP (100 μ M, arrow) compared with NPE cells from 3-month-old mice (n = 20, **p < 0.01, unpaired t-test). (**H**) IOP-lowering effect of UDP on WT mice aged 12 months. Instillation of UDP (500 μ M, 5 μ l/eye for 1.5 h) on 12-month old WT mice showed a 50% reduction in efficacy compared with 3-month-old WT mice (4.6±3.1 and 10.1±2.8% in 3- and 12-month-old WT mice, respectively). Scale bars, 50 μ m in (B)-(F).