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## Life Sciences Reporting Summary

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Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. For final submission: please carefully check your responses for accuracy; you will not be able to make changes later.

## Experimental design

1. Sample size

Describe how sample size was determined.

No statistical methods were used to predetermine sample sizes. Due to radiation damage, X-ray diffraction data collection of the protein crystals was limited to 5-10 degree per crystal. To collect a complete data set for structure determination, diffraction data from multiple crystals were integrated and scaled using HKL2000. By calculating completeness of the data set, diffraction data from 20 crystals were used to ensure the completeness was close to 100%.

2. Data exclusions

Describe any data exclusions.

No data were excluded.

3. Replication

Describe the measures taken to verify the reproducibility of the experimental findings.

All attempts at replication were successful.

4. Randomization

Describe how samples/organisms/participants were allocated into experimental groups.

Randomization is not relevant to this study, as protein and crystal samples are not required to be allocated into experimental groups in protein structural studies, and no animals or human research participants are involved in this study.

5. Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis.

Blinding is not relevant to this study, as protein and crystal samples are not required to be allocated into experimental groups in protein structural studies, and no animals or human research participants are involved in this study.

Note: all in vivo studies must report how sample size was determined and whether blinding and randomization were used.

6.	Statistical parameters	
	For all figures and tables that use statistical methods, conf Methods section if additional space is needed).	irm that the following items are present in relevant figure legends (or in the
n/a	Confirmed	
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)	
	A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly	
	A statement indicating how many times each experiment was replicated	
	The statistical test(s) used and whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.	
	A description of any assumptions or corrections, such as an adjustment for multiple comparisons	
	Test values indicating whether an effect is present  Provide confidence intervals or give results of significance tests (e.g. P values) as exact values whenever appropriate and with effect sizes noted.	
	A clear description of statistics including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)	
	Clearly defined error bars in <u>all</u> relevant figure captions (with explicit mention of central tendency and variation)	
	See the web collection on stat	tistics for biologists for further resources and guidance.
•	Software	
Ро	licy information about availability of computer code	
7.	Software	
	Describe the software used to analyze the data in this study.	Graphpad Prism5 and Pymol. For structure refinement, HKL3000, PHASER, REFMAC and COOT. For homology modeling, PROMALS3D and MODELLER-9v15. For molecular docking, DOCK3.7, AMBER, QNIFFT, Marvin (version 15.11.23.0, ChemAxon, 2015; http://www.chemaxon.com), Corina(Molecular Networks GmbH), Omega (OpenEye Scientific Software) and AMSOL.
		entral to the paper but not yet described in the published literature, software must be made burage code deposition in a community repository (e.g. GitHub). <i>Nature Methods</i> guidance for rinformation on this topic.
•	Materials and reagents	
Ро	licy information about availability of materials	
	Materials availability	
	Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a third party.	All constructs made are freely available without restrictions for use to investigators.
9.	Antibodies	
	Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).	No antibodies were used in this study.
10	. Eukaryotic cell lines	
	a. State the source of each eukaryotic cell line used.	Spodoptera frugiperda (Sf9) cells (Expression Systems) and HEK293 T cells (ATCC CRL-11268)
	b. Describe the method of cell line authentication used.	All cells used in this study are commercial and were obtained from vendors as indicated in the manuscript. HEK293T were certified mycoplasma free and authenticated by ATCC. Cells were also validated by analysis of short tandem repeat (STR) DNA profiles and these profiles showed 100% match at the STR database from ATCC
	c. Report whether the cell lines were tested for mycoplasma contamination.	HEK293T (ATCC CRL-11268; 59587035) were certified mycoplasma free by ATCC.
	d. If any of the cell lines used are listed in the database	No commonly misidentified cell lines were used.

d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use.

## ▶ Animals and human research participants

Policy information about studies involving animals; when reporting animal research, follow the ARRIVE guidelines

11. Description of research animals

Provide all relevant details on animals and/or animal-derived materials used in the study.

No animals were used.

Policy information about studies involving human research participants

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

The study didn't involve human research participants.