DNA IS A NEW TARGET OF PARP3

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Figure S1. The cleavage of the ligated product by APE1. A) Ligation by DNA ligase IIIa with the followed cleavage by APE1 of the initial one-window gapped (lines 4-8) or nicked (lanes 11-15) DNA substrates. The reactions were performed using consequential addition of DNA ligase IIIa and APE1. At the first step, 0.02 µM [³²P]-DNA substrate in HDB buffer with 10 mM MgCl₂ were treated by 0.1 µM DNA ligase IIIa for 30 min at 37°C, and an aliquot was selected (lanes 4 and 11). Then the mixtures were treated with 1 nM human APE1 at 37°C for 15, 30, 60 and 600 sec. Subsequently, an aliquot of all samples were selected at each time-point. In control experiments 0.02 µM [³²P]-DNA substrate in HDB buffer with 10 mM MgCl₂ was treated by 1 nM APE1 for 600 sec at 37°C or 10 µM Nudix for 20 h at 37°C. All mixtures were resolved on a 20% polyacrylamide gel containing 7 M urea and 10% formamide in 1x TBE buffer. The gels were dried and subjected to autoradiography and/or phosphorimaging for quantitation using the Typhoon imaging system from GE Healthcare Life Sciences and analysed using OriginPro7.5, Microcal Software, USA. Here, lane 1 shows the DNA electrophoretic mobility of 18 nt oligonucleotide; lanes 2 show the initial mobility of [32P]-labelled ADP-DNA substrate. Lanes 5-8 and 12-15 represented the kinetic assay for APE1. Lanes 3 and 10 contained corresponding DNA substrates treated by APE1. Substrates gap-mod and nick-mod represent onewindow gapped or nicked DNA duplexes assembled by the same whole DNA strand and contained the ADP-ribose residue at the 5'-end of the [32P]-labelled oligonucleotide. "M" inside the circle marks the modification induced by PARP3. The asterisk marks the [32P]-label. (B) Quantitative analysis of the assay reactions. The bars in the charts correspond to the respective lanes on the upper autoradiogram.

