

SUPPLEMENTARY MATERIAL

**Functional analysis of cancer-associated DNA polymerase ϵ variants in
*Saccharomyces cerevisiae***

Stephanie R. Barbari, Daniel P. Kane¹, Elizabeth A. Moore and Polina V. Shcherbakova

Eppley Institute for Research in Cancer and Allied Diseases, Fred & Pamela Buffett Cancer
Center, University of Nebraska Medical Center, Omaha, NE 68198, USA

¹ *Present address:* Daniel P. Kane, Department of Biological and Environmental Sciences, Le
Moyne College, Syracuse, NY 13214

Table S1 Genotypes of yeast strains used for construction of *pol2-x* mutants and mutation rate measurements

Strain	Relevant chromosomal mutation	Genotype
Haploid strains		
TM30	WT	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2</i>
TM44	WT	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP</i>
DK039a/DK039b	<i>pol2-F139L</i>	<i>MATa ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-F139L</i>
SB042/SB044	<i>pol2-R252H</i>	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-R252H</i>
SB046/SB051	<i>pol2-R252H</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-R252H</i>
SI022/SI023	<i>pol2-D290V</i>	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-D290V</i>
SI024/SI026	<i>pol2-D290V</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-D290V</i>
DK046/DK047	<i>pol2-P301H</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-P301H</i>
SB112/SB113	<i>pol2-P301H</i>	<i>MATa ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X can1Δ::loxP pol2-P301H</i>
DK048	<i>pol2-F382S</i>	<i>MATα ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-X can1Δ::loxP pol2-F382S</i>
DK049/DK050	<i>pol2-F382S</i>	<i>MATa ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-F382S</i>
SB052/SB053	<i>pol2-V426L</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-V426L</i>
SB054/SB055	<i>pol2-V426L</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-V426L</i>
SI030/SI031	<i>pol2-L439V</i>	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-L439V</i>
SI032/SI033	<i>pol2-L439V</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-L439V</i>
SB063/SB065	<i>pol2-P451R</i>	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-P451R</i>
SB066/SB068	<i>pol2-P451R</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-P451R</i>
SB069/SB071	<i>pol2-S474F</i>	<i>MATa ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-S474F</i>
SB073/SB074	<i>pol2-S474F</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-S474F</i>

Table S1 Genotypes of yeast strains used for construction of *pol2-x* mutants and mutation rate measurements (Continued)

Strain	Relevant chromosomal mutation	Genotype
DK051a/DK051b	<i>pol2-R778W</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-R778W</i>
SB075/SB077	<i>pol2-A979V</i>	<i>MATα ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-A979V</i>
SB079/SB081	<i>pol2-A979V</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-A979V</i>
DK055a/DK055b	<i>pol2-D1757N</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-D1757N</i>
SB083/SB084	<i>mlh1Δ</i>	<i>MATα ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 mlh1Δ::hphMX4</i>
SB086/SB087	<i>mlh1Δ</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP mlh1Δ::hphMX4</i>
SB088/SB089/ SB090/SB091	<i>pol2-R252H mlh1Δ</i>	<i>MATα ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-R252H mlh1Δ::hphMX4</i>
SB092/SB093/ SB094	<i>pol2-R252H mlh1Δ</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-R252H mlh1Δ::hphMX4</i>
SI003/SI004/ SI005/SI006	<i>pol2-R778W mlh1Δ</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-X CAN1::LEU2 pol2-R778W mlh1Δ::hphMX4</i>
SB095/SB096/ SB097/SB098	<i>pol2-A979V mlh1Δ</i>	<i>MATα ade5-1 lys2-Tn5-13 trp1-289 his7-2 leu2-3,112 ura3-4 CAN1::LEU2 pol2-A979V mlh1Δ::hphMX4</i>
SB099/SB100/ SB101/SB102	<i>pol2-A979V mlh1Δ</i>	<i>MATα ade5-1 lys2-InsE_{A14} trp1-289 his7-2 leu2-3,112 ura3-52 can1Δ::loxP pol2-A979V mlh1Δ::hphMX4</i>
Diploid strains		
TM63/SB539/ SB540	WT	<i>MATα/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP</i>
SB555/SB556	<i>pol2-R252H/POL2</i>	<i>MATα/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-R252H/POL2</i>
SI531/SI533	<i>pol2-D290V/POL2</i>	<i>MATα/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-D290V/POL2</i>

Table S1 Genotypes of yeast strains used for construction of *pol2-x* mutants and mutation rate measurements (Continued)

Strain	Relevant chromosomal mutation	Genotype
SI527/SI528/ SI529/SI530	<i>pol2-D290V/pol2-D290V</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-D290V/pol2-D290V</i>
SI505/SI507	<i>pol2-P301H/POL2</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-InsE_{A14}/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-X/ura3-X CAN1::LEU2/can1Δ::loxP pol2-P301H/POL2</i>
SB557/SB558/ SB559	<i>pol2-P301H/pol2-P301H</i>	<i>MATa/MATα ade5-1/ade5-1 lys2::InsE_{A14}/lys2::InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-X/ura3-X CAN1::LEU2/can1Δ::loxP pol2-P301H/pol2-P301H</i>
SI509/SI511	<i>pol2-F382S/POL2</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-InsE_{A14}/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-X/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-F382S/POL2</i>
SI514/SI516	<i>pol2-F382S/pol2-F382S</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-InsE_{A14}/lys2-Tn5-13 trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-X/ura3-X CAN1::LEU2/can1Δ::loxP pol2-F382S/pol2-F382S</i>
SI522/SI526	<i>pol2-L439V/POL2</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-L439V/POL2</i>
SI518/SI519/ SI520/SI521	<i>pol2-L439V/pol2-L439V</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-L439V/pol2-L439V</i>
SB542/SB545	<i>pol2-P451R/POL2</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-P451R/POL2</i>
SB547/SB548	<i>pol2-P451R/pol2-P451R</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-P451R/pol2-P451R</i>
SB549/SB551	<i>pol2-S474F/POL2</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-S474F/POL2</i>
SB552/SB553	<i>pol2-S474F/pol2-S474F</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-S474F/pol2-S474F</i>
SB560/SB561	<i>mlh1Δ::hphMX4/mlh1Δ::hphMX4</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP mlh1Δ::hphMX4/mlh1Δ::hphMX4</i>
SB562/SB563	<i>pol2-R252H/POL2 mlh1Δ::hphMX4/mlh1Δ::hphMX4</i>	<i>MATa/MATα ade5-1/ade5-1 lys2-Tn5-13/lys2-InsE_{A14} trp1-289/trp1-289 his7-2/his7-2 leu2-3,112/leu2-3,112 ura3-4/ura3-52 CAN1::LEU2/can1Δ::loxP pol2-R252H/POL2 mlh1Δ::hphMX4/mlh1Δ::hphMX4</i>

Table S2 Plasmid, restriction site, and method used to construct *pol2* mutants

Human POLE variant	Yeast Pol2 mimic	Plasmid	Restriction site	Mutation in truncated or full-length <i>POL2</i> after integration	Construction method ^a
F104L	F139L	YIpDK1	<i>Bam</i> HI	Full-length	1
R231H	R252H	YIpDK1	<i>Bgl</i> II	Truncated	2
D275V	D290V	YIpDK1	<i>Bam</i> HI	Full-length	1
P286H	P301H	YIpDK1	<i>Bam</i> HI	Full-length	1
N336S	N351S	YIpDK1	<i>Bam</i> HI	Full-length	1
F367S	F382S	YIpDK1	<i>Bam</i> HI	Full-length	1
V411L	V426L	YIpDK1	<i>Bgl</i> II	Truncated	2
L424V	L439V	YIpDK1	<i>Bam</i> HI	Full-length	1
P436R	P451R	YIpDK1	<i>Bgl</i> II	Truncated	2
S459F	S474F	YIpDK1	<i>Bgl</i> II	Truncated	2
R762W	R778W	p173	<i>Age</i> I	Full-length	1
A966V	A979V	p173	<i>Bsr</i> GI	Truncated	2
D1752N	D1757N	p174	<i>Age</i> I	Full-length	1

^aRefers to the two methods described in the Materials and Methods section.

Table S3 Spontaneous mutation rates in haploid yeast strains with *pol2* mutations mimicking human *POLE* variants

Human POLE variant	Yeast <i>POL2</i> allele	Can ^R mutation		His ⁺ reversion		Lys ⁺ reversion	
		Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase
WT	<i>POL2</i>	23 (17-32)	1.0	1.3 (0.76-1.9)	1.0	31 (25-49)	1.0
F104L	<i>pol2-F139L</i>	21 (18-23)	0.91	1.2 (0.94-1.5)	0.92	26 (24-34)	0.84
R231H	<i>pol2-R252H</i>	18 (16-21)	0.78	0.78 (0.58-0.95)	0.60	36 (33-46)	1.2
D275V	<i>pol2-D290V</i>	53 (44-55)	2.3	4.9 (4.2-5.9)	3.8	42 (36-44)	1.4
P286H	<i>pol2-P301H</i>	290 (250-440)	13	29 (25-37)	22	51 (40-63)	1.6
N336S	<i>pol2-N351S</i>	21 (19-25)	0.91	1.6 (1.3-2.4)	1.2	32 (30-36)	1.0
F367S	<i>pol2-F382S</i>	380 (300-490)	17	16 (8.7-22)	12	56 (30-140)	1.8
V411L	<i>pol2-V426L</i>	28 (25-32)	1.2	2.4 (2.0-3.3)	1.8	30 (28-35)	0.97
L424V	<i>pol2-L439V</i>	120 (82-140)	5.2	11 (9.6-16)	8.5	32 (27-36)	1.0
P436R	<i>pol2-P451R</i>	120 (98-150)	5.2	12 (11-16)	9.2	40 (36-49)	1.3
S459F	<i>pol2-S474F</i>	680 (560-800)	30	35 (30-47)	27	71 (62-87)	2.3
R762W	<i>pol2-R778W</i>	21 (17-25)	0.91	1.0 (0.59-1.6)	0.77	26 (22-29)	0.84
A969V	<i>pol2-A979V</i>	21 (19-28)	0.91	1.4 (0.87-1.7)	1.1	31 (27-34)	1.0
D1752N	<i>pol2-D1757N</i>	21 (15-32)	0.91	1.0 (0.78-1.4)	0.77	22 (20-27)	0.71

^aMutation rates are given as the median of at least 18 independent cultures, with 95% confidence intervals in parentheses. Fold increase is shown relative to the wild-type haploid (WT). Bold text indicates $p < 0.05$ by Wilcoxon-Mann-Whitney compared to WT. Data from this table were used to generate Figure 2.

Table S4 Summary of published tumor sequencing data used to calculate the frequency of DNA binding cleft variants

Human <i>POLE</i> variant	Exon	No. times reported ^a	No. tumors analyzed ^b	Incidence per 10,000 tumors
D275V	9	1	13,283	0.75
P286H	9	1	13,283	0.75
P286R	9	160	13,159	120
F367S	11	5	7,079	7.1
L424V	13	3	13,424	2.2
P436R	13	6	13,424	4.5
S459F	14	18	11,831	15

^aNumber of times the variant has been reported in sporadic CRC and EC.

^bTotal number of tumors from studies where the corresponding exon was sequenced.

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Table S5 P values (Fisher's exact test) for comparison of frequencies of individual *POLE* variants

POLE Variant	D275V	P286H	L424V	P436R	F367S	S459F
P286H	1					
L424V	0.62	0.62				
P436R	0.12	0.12	0.51			
F367S	0.022	0.022	0.13	0.53		
S459F	<0.0001	<0.0001	0.0003	0.0069	0.14	
P286R	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001

Data used for analysis are in Supplementary Table S4. P values indicating statistically significant differences are in bold.

Table S6 Spontaneous mutation rates in diploid yeast strains with *pol2* mutations mimicking human *POLE* variants

Human POLE variant	Yeast <i>POL2</i> alleles	Can ^R mutation		His ⁺ reversion		Lys ⁺ reversion	
		Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase
WT	<i>POL2/POL2</i>	20 (15-24)	1.0	1.1 (0.70-1.7)	1.0	22 (17-26)	1.0
D275V	<i>pol2-D290V/POL2</i>	21 (19-23)	1.0	2.7 (2.0-3.1)	2.5	16 (13-18)	0.73
	<i>pol2-D290V/pol2-D290V</i>	40 (37-43)	2.0	5.3 (4.4-5.9)	4.8	22 (19-28)	1.0
P286H	<i>pol2-P301H/POL2</i>	130 (110-150)	6.5	10 (9.4-13)	9.1	30 ^b (27-38)	0.68 ^b
	<i>pol2-P301H/pol2-P301H</i>	270 (240-280)	14	24 (21-27)	22	49 ^b (40-52)	1.1 ^b
F367S	<i>pol2-F382S/POL2</i>	230 (190-310)	12	24 (17-37)	22	59 ^b (49-76)	1.3 ^b
	<i>pol2-F382S/pol2-F382S</i>	530 (420-710)	27	50 (32-69)	45	66 (37-83)	3.0
L424V	<i>pol2-L439V/POL2</i>	68 (53-82)	3.4	5.1 (4.5-6.4)	4.6	17 (16-22)	0.77
	<i>pol2-L439V/pol2-L439V</i>	120 (100-140)	6.0	13 (11-15)	12	20 (18-22)	0.91
P436R	<i>pol2-P451R/POL2</i>	82 (74-97)	4.1	7.0 (6.5-9.6)	6.4	20 (17-25)	0.91
	<i>pol2-P451R/pol2-P451R</i>	180 (150-200)	9.0	16 (14-19)	15	24 (21-31)	1.1
S459F	<i>pol2-S474F/POL2</i>	500 (370-640)	25	27 (24-37)	25	30 (26-31)	1.4
	<i>pol2-S474F/pol2-S474F</i>	1000 (850-1300)	50	92 (70-120)	84	93 (80-120)	4.2

^aMutation rates are given as the median of at least 18 independent cultures, with 95% confidence intervals in parentheses. Fold increase is shown relative to the wild-type diploid (WT). Bold text indicates $p < 0.05$ by Wilcoxon-Mann-Whitney compared to WT. Data from this table were used to generate Figure 5.

^bStrain contains two copies of the *lys2-InsE_{A14}* allele (Supplementary Table 1). The fold increase shown is the fold increase in the Lys⁺ reversion rate over wild-type divided by two.

Table S7 Spontaneous mutation rates in yeast strains with *pol2-R252H*, *-R778W* and *-A979V* mutations in MMR-proficient or MMR-deficient (*mlh1Δ*) background

Ploidy	<i>POL2</i> allele	<i>MLH1</i> allele	Can ^R mutation		His ⁺ reversion		Lys ⁺ reversion	
			Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase	Mutation rate (x10 ⁻⁸) ^a	Fold increase
<i>n</i>	<i>POL2</i>	<i>MLH1</i>	15 (14-20)	1.0	0.92 (0.64-1.3)	1.0	26 (18-30)	1.0
	<i>pol2-R252H</i>	<i>MLH1</i>	23 (15-31)	1.5	0.90 (0.69-2.1)	0.98	32 (28-34)	1.2
	<i>pol2-R778W</i>	<i>MLH1</i>	15 (13-19)	1.0	0.59 (0.48-0.73)	0.64	14 (14-15)	0.54
	<i>pol2-A979V</i>	<i>MLH1</i>	17 (13-34)	1.1	0.93 (0.65-1.2)	1.0	21 (18-24)	0.81
	<i>POL2</i>	<i>mlh1Δ</i>	440 (380-490)	29	97 (57-110)	110	130000 (120000-170000)	5000
	<i>pol2-R252H</i>	<i>mlh1Δ</i>	970 (930-1100)	65	150 (140-160)	160	160000 (140000-260000)	6200
	<i>pol2-R778W</i>	<i>mlh1Δ</i>	530 (440-650)	35	100 (91-110)	110	140000 (130000-160000)	5400
	<i>pol2-A979V</i>	<i>mlh1Δ</i>	410 (390-470)	27	99 (90-110)	110	170000 (140000-190000)	6500
<i>2n</i>	<i>POL2/POL2</i>	<i>MLH1/MLH1</i>	17 (16-23)	1.0	1.2 (0.87-1.6)	1.0	18 (16-22)	1.0
	<i>pol2-R252H/POL2</i>	<i>MLH1/MLH1</i>	18 (15-21)	1.1	1.2 (1.0-1.7)	1.0	20 (17-24)	1.1
	<i>POL2/POL2</i>	<i>mlh1Δ/mlh1Δ</i>	450 (410-520)	26	110 (98-140)	92	120000 (100000-130000)	6700
	<i>pol2-R252H/POL2</i>	<i>mlh1Δ/mlh1Δ</i>	820 (710-1200)	48	150 (140-170)	130	140000 (120000-160000)	7800

^aMutation rates are given as the median of at least 18 independent cultures, with 95% confidence intervals in parentheses. Fold increase is shown relative to the wild-type haploid or diploid. Bold text indicates $p < 0.05$ by Wilcoxon-Mann-Whitney compared to *mlh1Δ* haploids or *mlh1Δ/mlh1Δ* diploids. Data from this table were used to generate Figure 6.

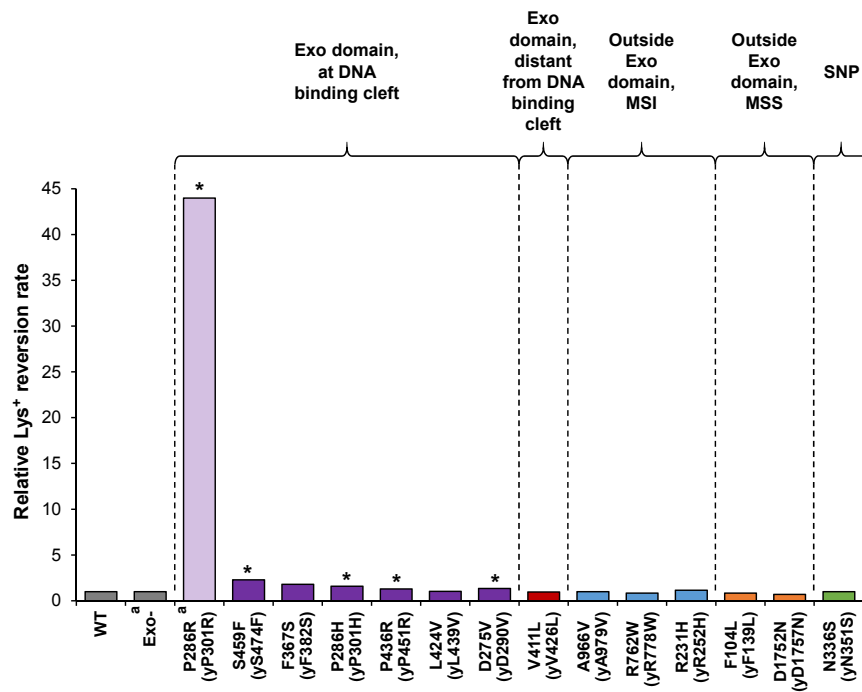


Figure S1 Effect of cancer-associated Polε variants on the rate of *lys2-InsE_{A14}* reversion in haploid yeast strains. Relative mutation rates are shown compared to wild-type (WT). All experimental details and symbols are as in Figure 2. Asterisks indicate $p < 0.05$ by Wilcoxon-Mann-Whitney compared to WT. Data are from Supplementary Table S3. ^aData from Kane and Shcherbakova 2014.

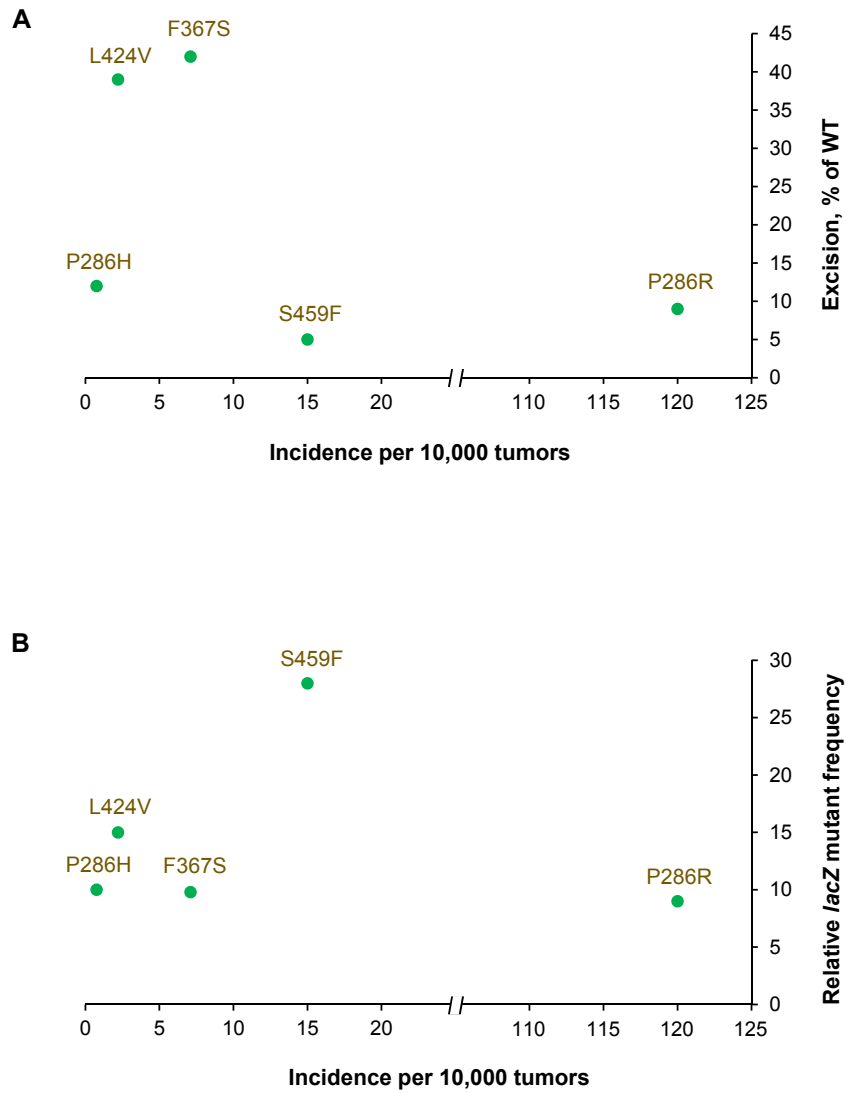


Figure S2 Lack of correlation between *in vitro* effects of Pole mutations and their frequency in tumors. [A] Relationship between exonuclease activity defect and variant frequency. [B] Relationship between *in vitro* error rate and variant frequency. Exonuclease activity and *in vitro* error rates were measured in Shinbrot *et al.* 2014 using the N-terminal fragment of human Pole containing the exonuclease and DNA polymerase domains. The variant frequency in tumors was determined as described in Figure 4.

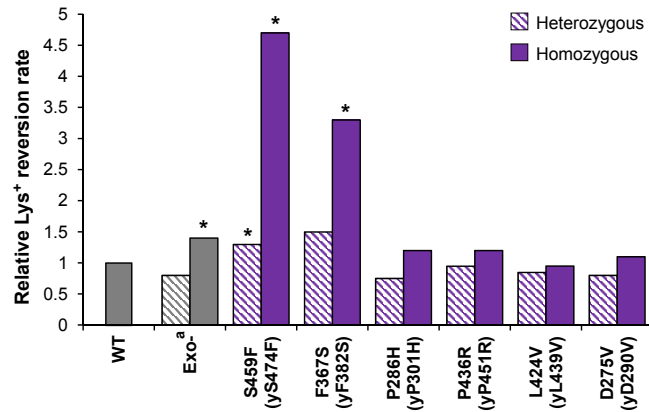


Figure S3 Effect of cancer-associated Pol ϵ variants on the rate of *lys2-InsE_{A14}* reversion in diploid yeast strains. The strains contained one copy of the *lys2-InsE_{A14}* reporter allele and a nonreversible mutation at the same position of the *LYS2* gene in the homologous chromosome. All other experimental details and symbols are as in Figure 5. Relative mutation rates are shown compared to wild-type (WT). Asterisks indicate $p < 0.05$ by Wilcoxon-Mann-Whitney compared to WT. Data are from Supplementary Table S6. ^aData from Kane and Shcherbakova 2014.

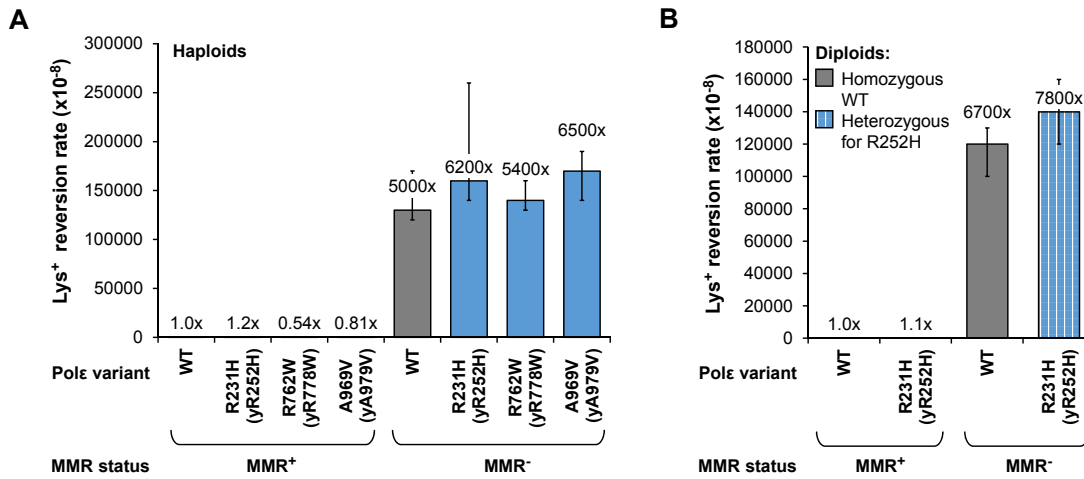


Figure S4 PolE variants found in MMR-deficient tumors do not affect the rate of *lys2-InsE_{A14}* reversion in MMR-proficient or MMR-deficient background. The reversion rate was measured in haploid [A] or diploid [B] yeast strains containing indicated chromosomal *pol2* mutations. The MMR defect was mimicked by deleting the yeast *MLH1* gene. Mutation rates are given as the median for at least 18 independent cultures, with error bars indicating 95% confidence intervals. Fold increase in mutation rate relative to the strain with wild-type *POL2* and *MLH1* genes is shown above each bar. Data are from Supplementary Table S7.