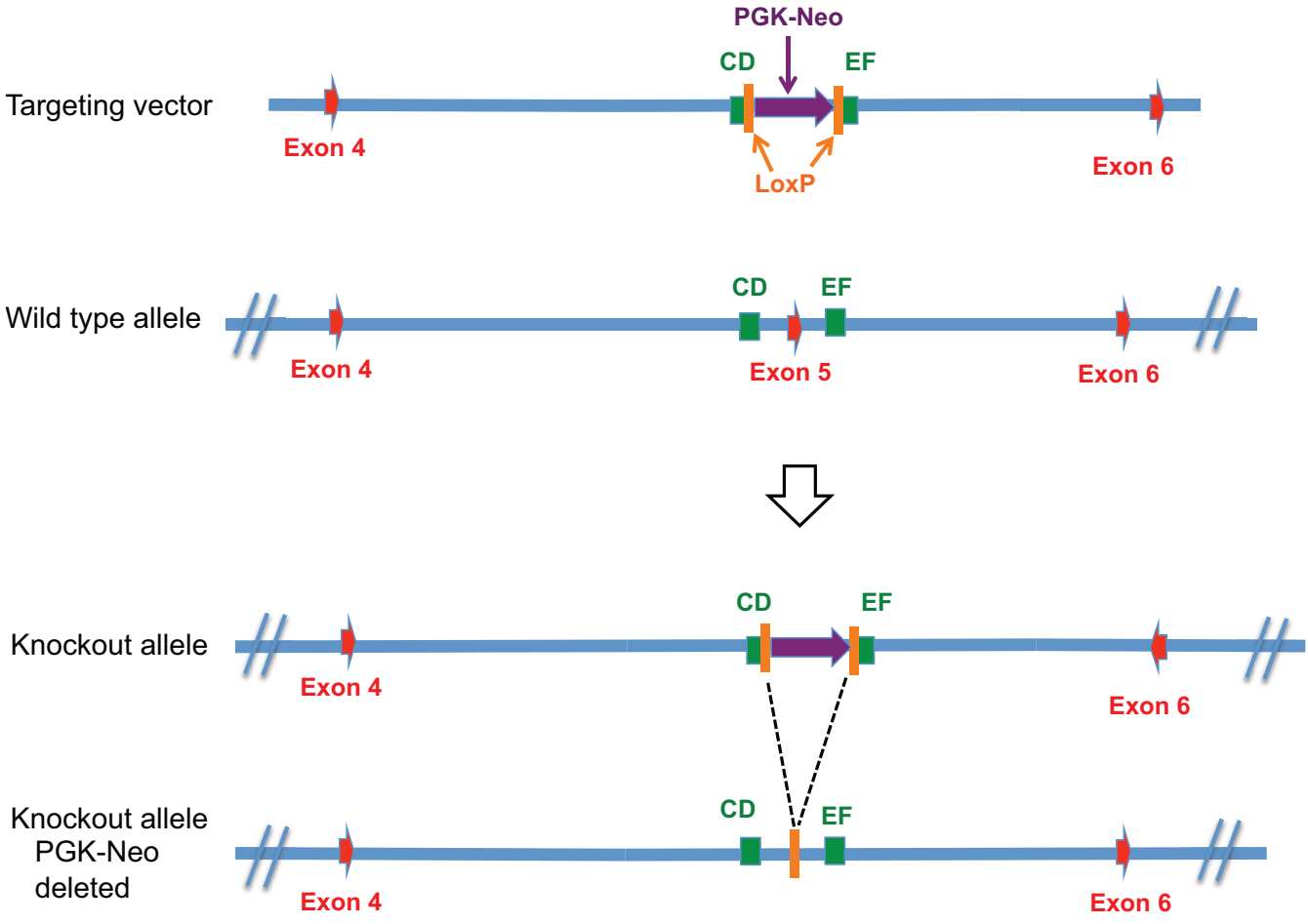

Figure S1. Genetic engineering of the *Slc25a13* targeted knockout mutation. A 10,696 bp genomic fragment of wildtype *Slc25a13*, including exons 4, 5, and 6, was retrieved from a C57BL/6J bacterial artificial chromosome (BAC) and cloned into a plasmid targeting vector. Homology arms CD and EF (each ~100 bp) were used to replace exon 5 and surrounding sequences (821 bp) of the targeting vector with a floxed PGK-Neo cassette (1906 bp, purple arrow). The completed targeting vector was introduced into ES cells by electroporation, and homologous recombination resulted in replacement of exon 5 with the PGK-Neo selection cassette. Properly targeted ES cells, identified by G418 selection and confirmed by Southern blot and loss-of-allele assays, were used to produce mice with the mutant allele, which were then mated with mice from a cre-deleter strain to remove the floxed PGK-Neo cassette.