

# Supporting Information

## Mussel-Inspired Multifunctional Hydrogel Coating for Prevention of Infections and Enhanced Osteogenesis

*Hao Cheng,<sup>a,b,c,#</sup> Kan Yue,<sup>a,b,#</sup> Mehdi Kazemzadeh-Narbat,<sup>a,b,#</sup> Yanhui Liu,<sup>a,b,d</sup> Akbar Khalilpour,<sup>a,b</sup> Bingyun*

*Li,<sup>e</sup> Yu Shrike Zhang,<sup>a,b</sup> Nasim Annabi,<sup>a,b,f,\*</sup> Ali Khademhosseini<sup>a,b,g,h,\*</sup>*

<sup>a</sup> Biomaterials Innovation Research Center, Division of Biomedical Engineering, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Cambridge, MA, 02139, USA

<sup>b</sup> Harvard-MIT Division of Health Sciences and Technology, Massachusetts Institute of Technology, Cambridge, MA, 02139, USA

<sup>c</sup> Orthopaedic Department, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, China

<sup>d</sup> College of Textiles, Donghua University, Shanghai, 201620, China

<sup>e</sup> Department of Orthopaedics, School of Medicine, West Virginia University, Morgantown, WV 26506, USA

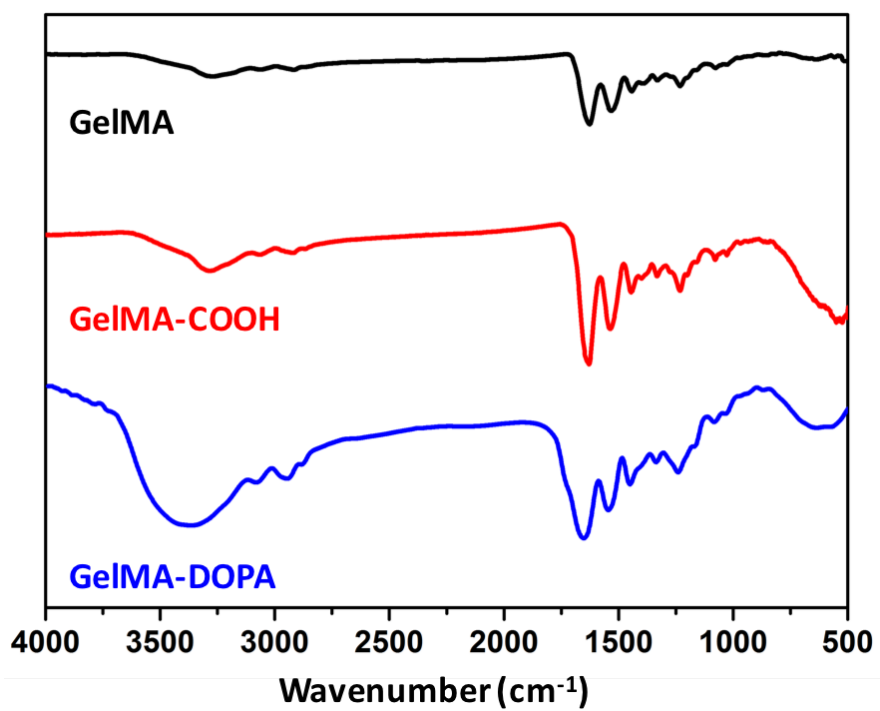
<sup>f</sup> Department of Chemical Engineering, Northeastern University, Boston, MA, 02115-5000, USA

<sup>g</sup> Department of Bioindustrial Technologies, College of Animal Bioscience and Technology, Konkuk University, Seoul, 143-701, the Republic of Korea

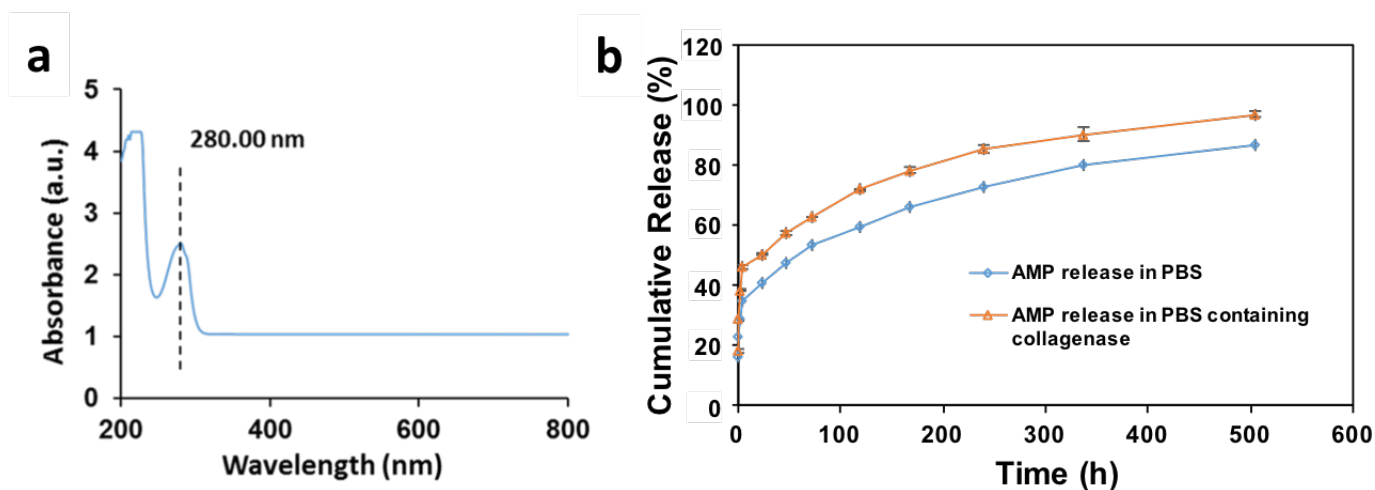
<sup>h</sup> Department of Physics, King Abdulaziz University, Jeddah, 21569, Saudi Arabia

<sup>#</sup> These authors contributed equally to this work.

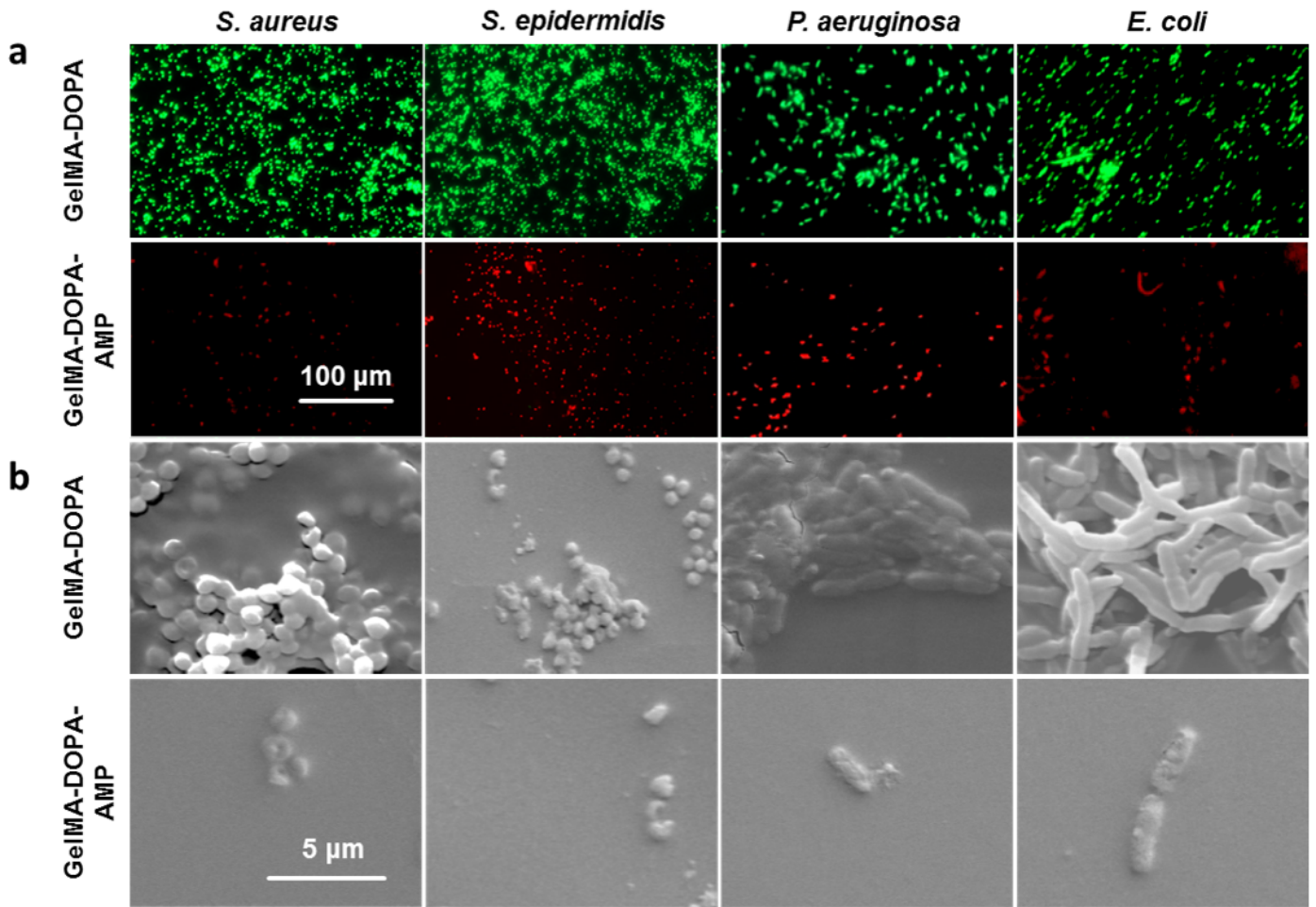
<sup>\*</sup> Corresponding authors: Prof. Ali Khademhosseini ([alik@bwh.harvard.edu](mailto:alik@bwh.harvard.edu)), Prof. Nasim Annabi ([n.annabi@neu.edu](mailto:n.annabi@neu.edu))



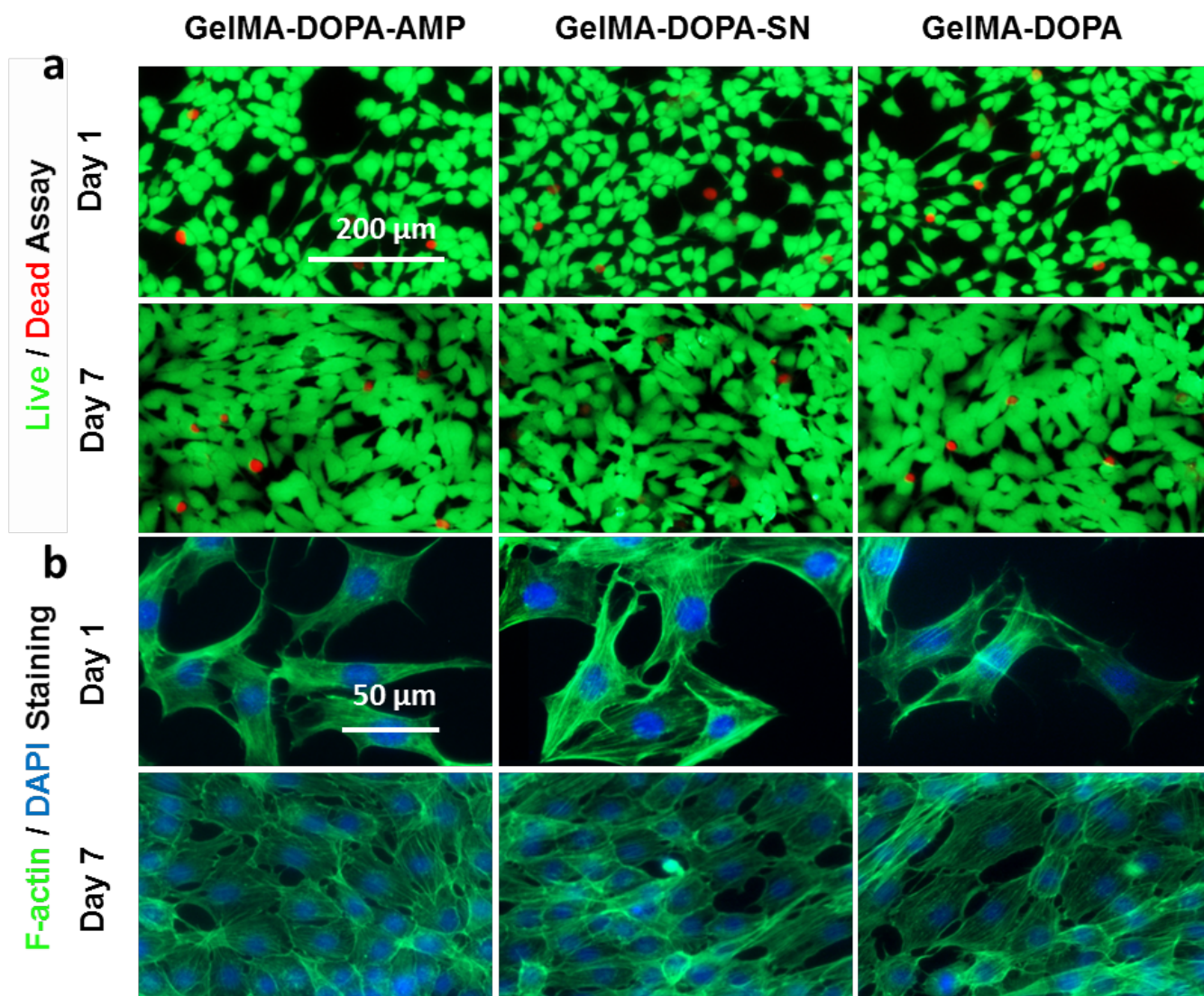
**Figure S1.** FT-IR spectra of GelMA (black curve), GelMA-COOH (red curve), and GelMA-DOPA (blue curve).



**Figure S2.** *In vitro* AMP release from 20% (w/v) GelMA-DOPA-AMP-SN hydrogels. **(a)** UV-Vis spectrum of AMP in PBS. Absorbance at 280 nm was monitored to determine the released amount of AMP from hydrogel matrix. **(b)** Cumulative release kinetics of AMP released from the GelMA-DOPA AMP-SN composite hydrogels coated on Ti substrates immersed in PBS or PBS containing 2U/mL collagenase in 21 days.



**Figure S3.** Antimicrobial activity of GelMA-DOPA hydrogels at 20% (w/v) prepolymer concentration with different additives. **(a)** AMP released from GelMA-DOPA-AMP samples demonstrated efficient antimicrobial ability to kill the tested bacteria, *S. aureus*, *S. epidermidis*, *P. aeruginosa*, and *E. coli*. Compared to GelMA-DOPA samples without AMP loading, no live bacteria were observed on the GelMA-DOPA-AMP samples after 24 h culture (green: live bacteria; red: dead bacteria; scale bar: 100 μm). **(b)** SEM images of surfaces of GelMA-DOPA and GelMA-DOPA-AMP hydrogels incubated overnight with *S. aureus*, *S. epidermidis*, *P. aeruginosa*, and *E. coli*. Very few bacteria were observed on the AMP-loaded samples (scale bar: 5 μm).



**Figure S4.** Cytotoxicity and cellular evaluation of different hydrogel formulations at 20% (w/v) prepolymer concentration. **(a)** Images of hMSCs seeded on the surface of GeIMA-DOPA-AMP, GeIMA-DOPA-SN and GeIMA-DOPA hydrogels with Live/Dead staining on days 1 and 7 (scale bar: 200  $\mu\text{m}$ ). **(b)** Images of hMSCs seeded on the surface of GeIMA-DOPA-AMP, GeIMA-DOPA-SN, and GeIMA-DOPA hydrogels with staining for f-actin/cell nuclei on days 1 and 7 (scale bar: 50  $\mu\text{m}$ ).

**Table S1.** Primer sequences for qPCR experiments.

<i>Genes</i>	<i>Forward</i>	<i>Reverse</i>
<b>Osteocalcin</b>	ATGAGAGCCCTCACACTCCTCG	GTCAGCCAACTCGTCACAGTCC
<b>Osteopontin</b>	TTCCAAGTAAGTCCAACGAAAG	GTGACCAGTTCATCAGATTCAT
<b>ALP</b>	GAGTATGAGAGTGACGAGAA	AGTGGGAGTGCTTGTATC
<b>RUNX2</b>	CGGAATGCCTCTGCTGTTAT	ACTCTTGCCTCGTCCACTCC