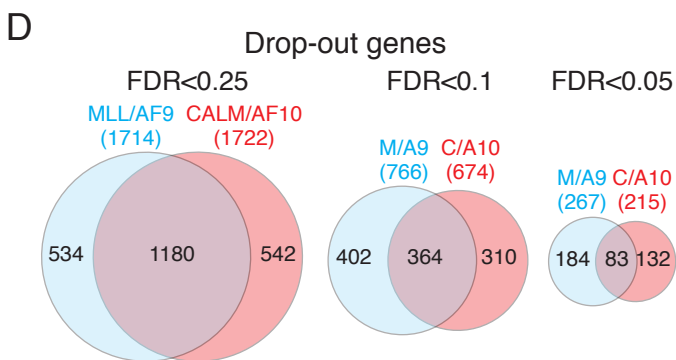


C

Sample	Mapped reads	Library sgRNA	Zero counts	Gini Index	Coverage
CALM-AF10_LibraryA_Day3	41,606,471	67,405	773	0.11	x617
CALM-AF10_LibraryA_Day18	40,699,845	67,405	6,006	0.2	x603
CALM-AF10_LibraryB_Day3	42,997,399	62,804	419	0.11	x685
CALM-AF10_LibraryB_Day18	44,087,417	62,804	6,641	0.21	x702
MLL-AF9_LibraryA_Day3	43,433,781	67,405	734	0.1	x644
MLL-AF9_LibraryA_Day18	50,203,365	67,405	6,711	0.21	x745
MLL-AF9_LibraryB_Day3	44,561,317	62,804	1,002	0.11	x710
MLL-AF9_LibraryB_Day18	44,572,208	62,804	5,614	0.2	x710



E

MLL/AF9	p value	CALM/AF10	p value
Node name (KEGG pathway)		Node name (KEGG pathway)	
HSA03010_RIBOSOME	2.44E-17	HSA03010_RIBOSOME	4.69E-21
HSA00970_AMINOACYL_TRNA_BIOSYNTHESIS	3.50E-15	HSA00190_OXIDATIVE_PHOSPHORYLATION	4.22E-13
HSA00240_PYRIMIDINE_METABOLISM	9.28E-11	HSA00970_AMINOACYL_TRNA_BIOSYNTHESIS	6.79E-12
HSA03020_RNA_POLYMERASE	2.78E-10	HSA03050_PROTEASOME	9.19E-09
HSA03050_PROTEASOME	2.66E-08	HSA03020_RNA_POLYMERASE	1.47E-08
HSA00190_OXIDATIVE_PHOSPHORYLATION	8.14E-08	HSA00240_PYRIMIDINE_METABOLISM	3.13E-08
HSA00230_PURINE_METABOLISM	1.08E-06	HSA04110_CELL_CYCLE	3.39E-06
		HSA04120_UBIQUITIN_MEDIATED_PROTEOLYSIS	6.31E-05

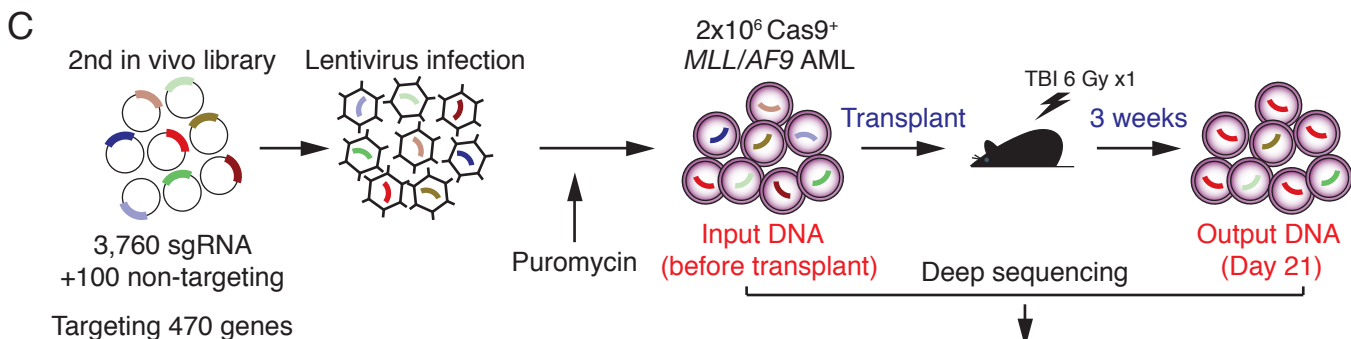
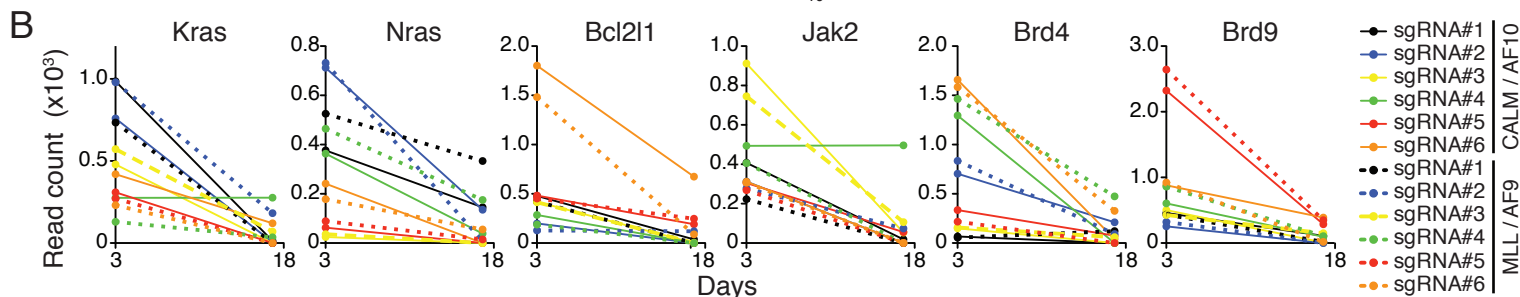
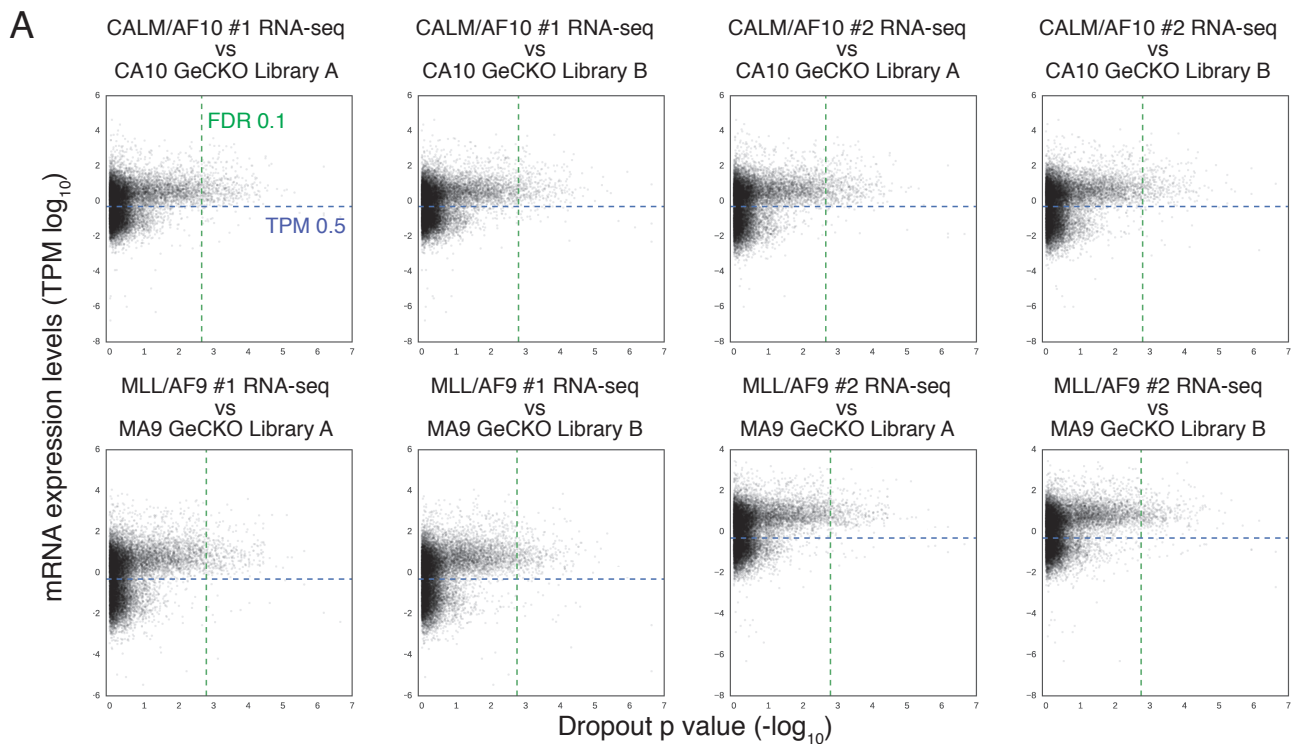
Figure S1. Related to Figure 1. CRISPR-Cas9 screens in mouse AML cell lines with normal karyotypes.

(A) Representative results of karyotype analysis. **(B)** Schematic representations of the GeCKO screen. **(C)**

Sequencing read counts of each sample. Read mapping and gini index calculations were performed using the

MAGeCK count program. **(D)** Venn diagrams show numbers of dropout genes at indicated FDR values. **(E)**

Pathway analysis of dropout genes shows enrichment of signatures representing basal cellular machineries.



D

MAGeCK analysis

Sample	Mapped reads	Library sgRNA	Zero counts	Gini Index	Coverage
MLL-AF9 before transplant	4,966,871	3,860	11	0.05	$\times 1287$
MLL-AF9 3wks after transplant	3,235,660	3,860	2,359	0.76	$\times 838$

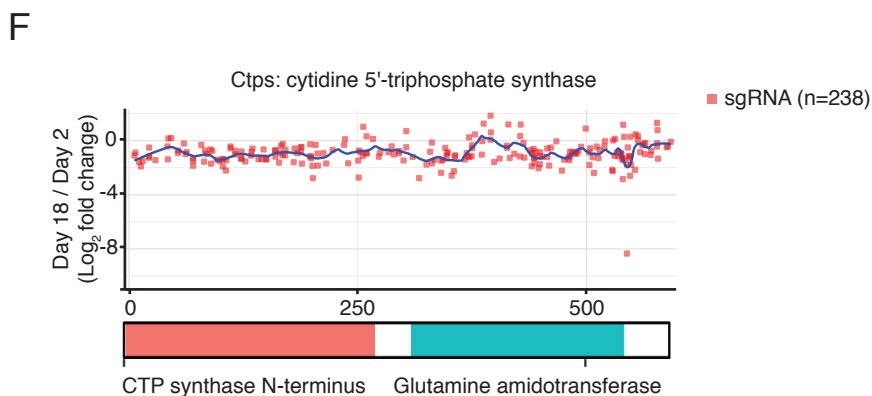
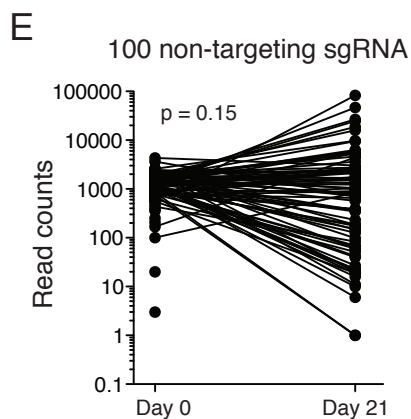


Figure S2. Dropout genes and validation screens: additional information. Related to Figure 1.

(A) RNA-seq experiments were performed using 4 primary mouse AML lines established independently in vivo (CALM/AF10 #1, CALM/AF10 #2, MLL/AF9 #1 and MLL/AF9 #2). mRNA expression levels (y-axis: TPM values) and the dropout p value (x axis: minus \log_{10} values in GeCKO screens) of each gene were plotted. **(B)** Graphs show read counts of individual sgRNAs targeting well-known genes essential for AML, before and after a 16-day incubation. **(C)** Workflow of second screen in vivo. Cas9-expressing MLL/AF9 cells were transduced with a pool of sgRNAs targeting 470 selected genes (8 sgRNA per gene) and 100 non-targeting sgRNAs and then transferred to sub-lethally irradiated recipients. Change in abundance of each sgRNA between the initial and final cell population was analyzed using the MAGeCK program. **(D)** Sequencing results of DNA samples obtained before and after transplant. **(E)** Read counts of each non-targeting sgRNA before and after transplant were plotted. **(F)** Results of CTPS domain-mapping.

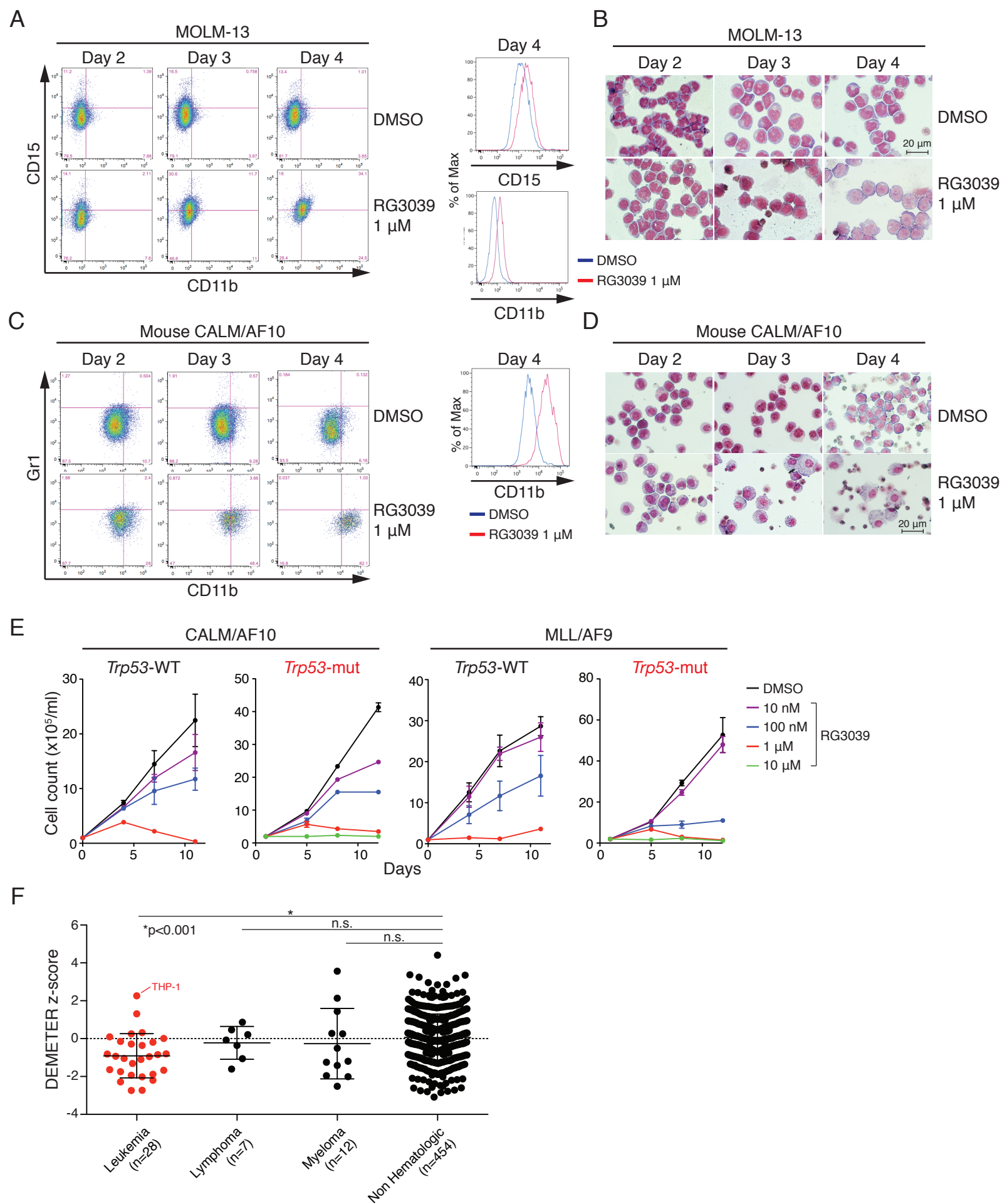
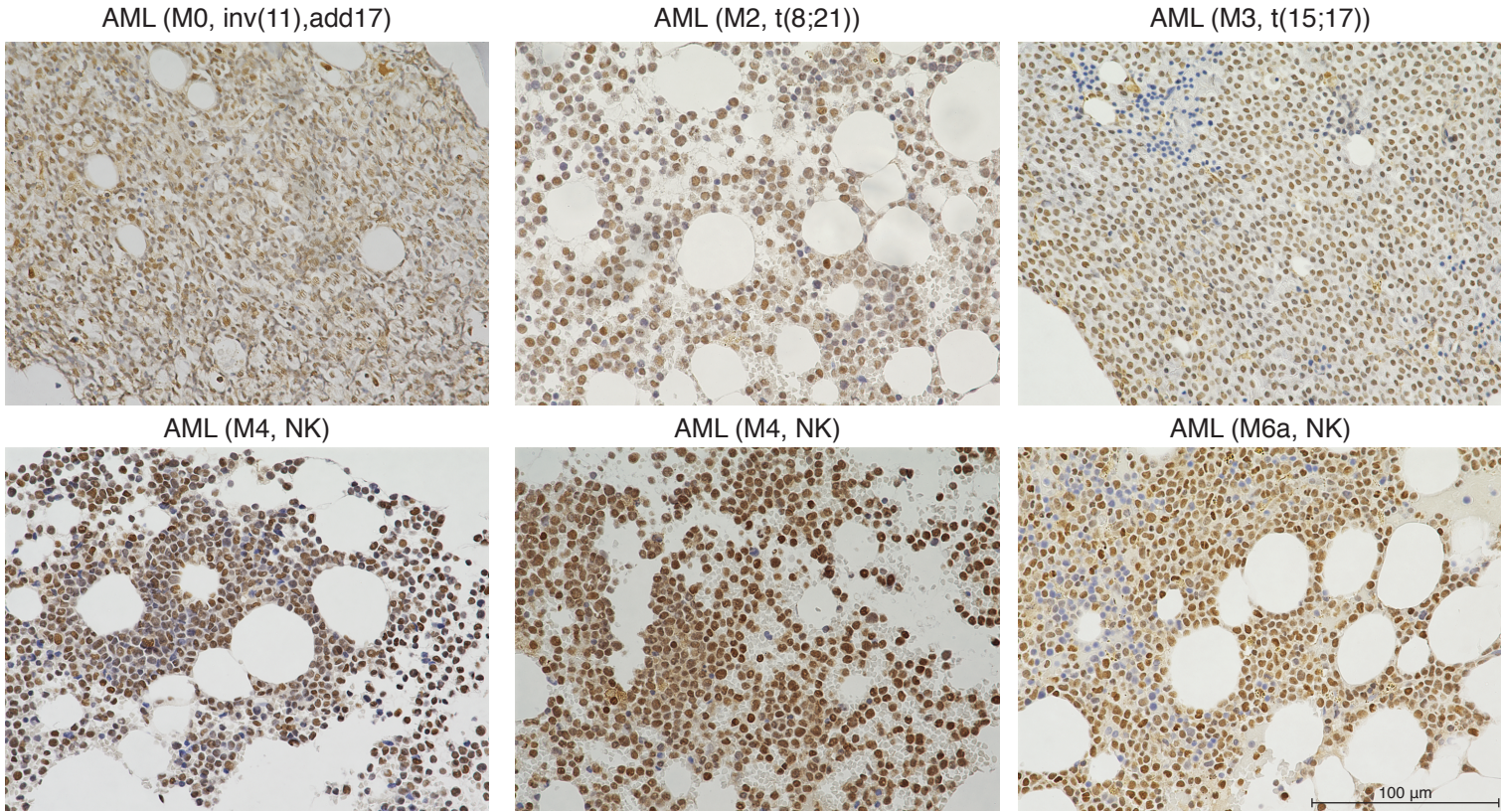
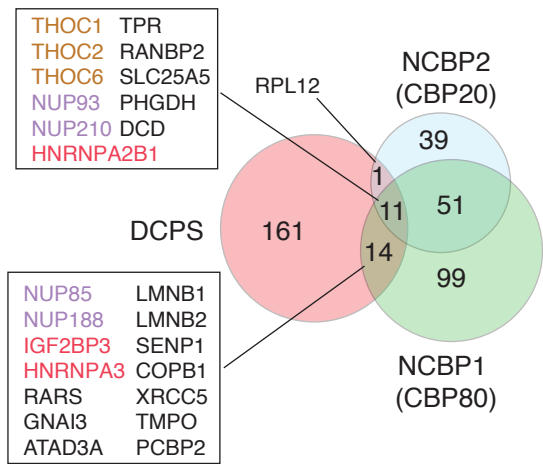


Figure S3. RG3039 slows proliferation and induces AML cell differentiation via p53-independent mechanisms. Related to Figure 2. (A) Representative FACS plots of MOLM-13 cells treated with either DMSO (vehicle) or RG3039. (B) May-Giemsa stain of MOLM-13 cells treated with either DMSO or RG3039. (C) Representative FACS plots of mouse CALM/AF10 cells treated with either DMSO or RG3039. (D) May-Giemsa stain of CALM/AF10 cells. (E) *Trp53*-WT or mutant mouse AML cells were treated with RG3039, and growth curves generated as described in Figure 2I. Data are represented as means \pm SD. (F) Dot graphs show DCPS dependency of 501 cancer cell lines representing leukemia, lymphoma, multiple myeloma and non-hematologic tumors. Data are shown as means \pm SD. The Y-axis represents DEMETER z-score of shRNA targeting DCPS (<https://depmap.org/rnai/genedeps?gene=DCPS>).

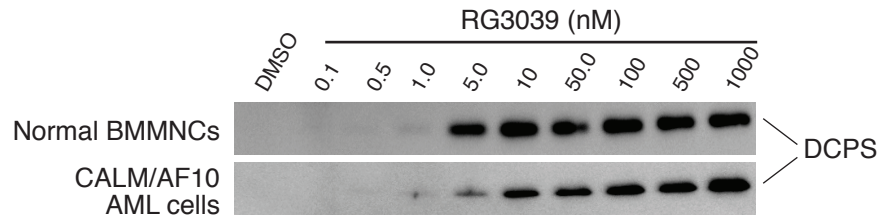
A



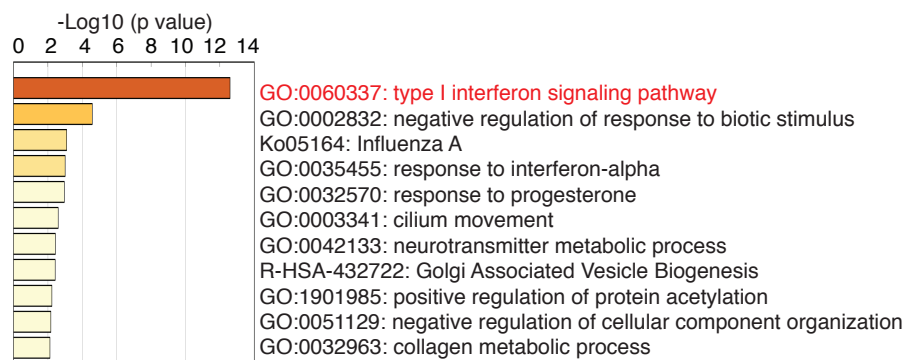
B



C



D



E

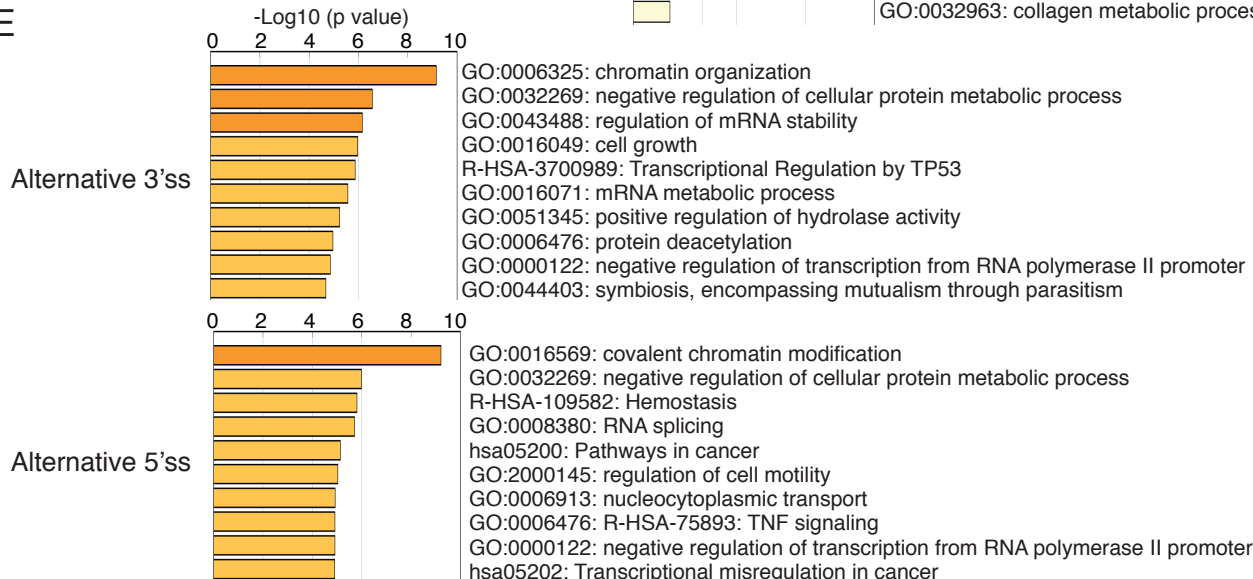


Figure S4. DCPS protein localizes to the nucleus and associates with factors involved in pre-mRNA metabolic pathways. Related to Figures 3 and 4. (A) Immunohistochemistry (IHC) for DCPS protein in BM sections of six human AML cases. Shown are FAB subtype and karyotype in each case. Brown staining: DCPS protein. DCPS is predominantly nuclear in AML cells. NK: normal karyotype. **(B)** CBC, which consists of NCBP1 and NCBP2, and DCPS share some interacting proteins. NCBP1 and NCBP2 binding proteins were described previously (Andersen et al., 2013). **(C)** Affinity between RG3039 and DCPS was assessed by CETSA. **(D)** Results of gene expression signature analysis of RG3039-treated AML cells using the Metascape program (<http://metascape.org/>). **(E)** Metascape analysis of aberrantly-spliced genes following RG3039 treatment of CALM/AF10 AML cells.

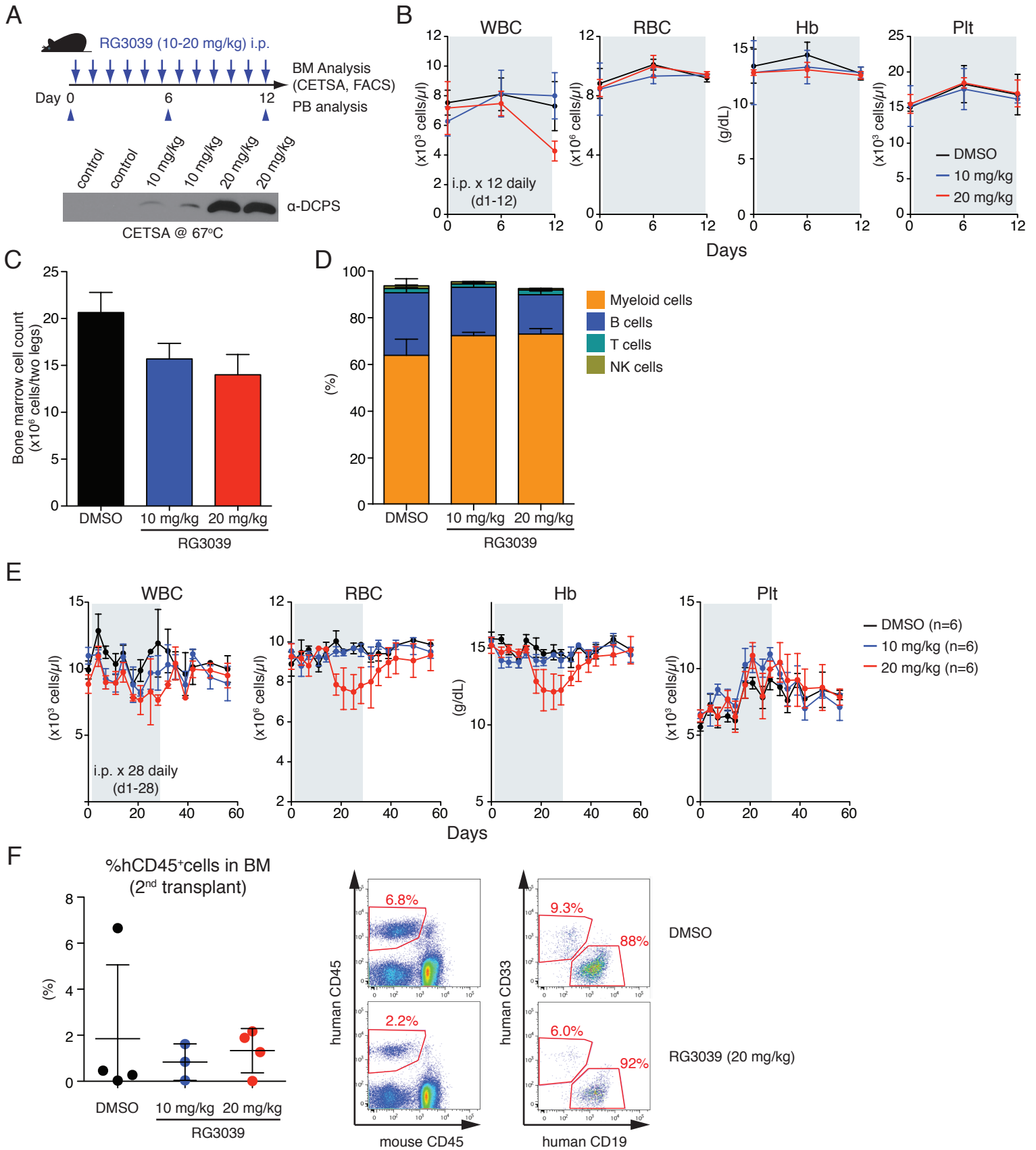
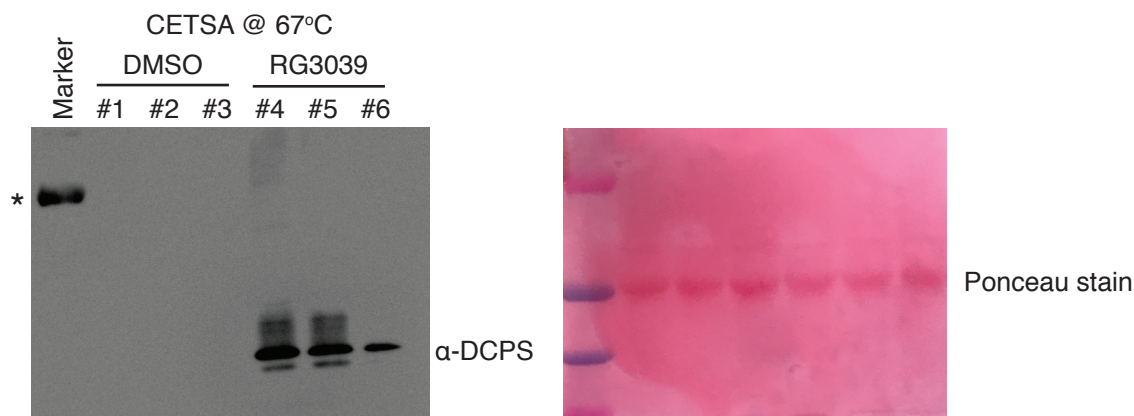


Figure S5. Effects of DCPS inhibition in vivo. Related to Figure 5. (A) DMSO or RG3039 (10 mg/kg or 20 mg/kg) was administered intraperitoneally daily for 12 days, and peripheral blood counts were analyzed before and after treatment (day 6 and day 12). CETSA was performed using BM cells the day after the last RG3039 (or DMSO) injection. **(B)** Peripheral blood counts were analyzed by a hematology analyzer before and after RG3039 treatment. Data are represented as means \pm SD. WBC: white blood cell; RBC: red blood cell; Hb: hemoglobin; Plt: platelet. **(C)** BM cell counts were measured the day after the last RG3039 (or DMSO) injection. Data are represented as means \pm SD. **(D)** Proportions of myeloid, B, T and NK cells in BM mononuclear cells were analyzed by FACS. Data are represented as means \pm SD. **(E)** DMSO (vehicle) or RG3039 (10 mg/kg or 20 mg/kg) was administered intraperitoneally daily for 28 days, and peripheral blood counts were analyzed every 5-7 days. Treatment periods are depicted in gray. Data are represented as means \pm SD. **(F)** Engraftment of human hematopoietic cells upon a second transplant. The Y-axis shows proportions of hCD45⁺ cells in BM 6 weeks after the second transplant. Data are represented as means \pm SD. Representative BM FACS plots are shown.

A

PDX	DFAM-68555-V1	DFAM-15354-V2	DFAM-71927-V3
WHO classification	Therapy-related AML	AML-MRC	AML-NOS
FAB classification	M5a	N/A	N/A
Karyotype	46,XX,t(9;11)(p22;q23)[3]/47,ider,+8[8]/50,ider,+6,+8,+9,der(9)t(9;11)(p22;q23),+13[cp5]	47, XY, +11 Trisomy 11	46,XX,dup(2)(q21q33),t(8;16)(p11;p13),psu,dic(22;1)(p11;p11)[10]/46,XX,dup(1)(q32q42),t(8;16),psu,dic(9;1)(q34;p11)[4]/46,XX,t(8;16),psu,dic(19;1)(p13;p11)[4]/46,XX[2]t(8;16)(p11;p13) KAT6A-CREBBP
Molecular details	t(9;11)(p22;q23) <i>MLL-AF9</i> Hyperdiploid, Trisomy 8 <i>FLT3</i> NM_004119 c.2027A>C p.N676T - in 79.6% of 314 reads Gain of <i>PIM1</i> (on 6p), gain of <i>RAD21</i> (on 8q), gain of <i>FLT3</i> and <i>PDS5B</i> (on 13q), gain of <i>CALR</i> , <i>NOTCH3</i> , <i>JAK3</i> and <i>MEF2B</i> (on 19p), gain of <i>CEBPA</i> , <i>CNOT3</i> and <i>U2AF2</i> (on 19q), gain of <i>RUNX1</i> and <i>U2AF1</i> (on 21q).	<i>DNMT3A</i> NM_175629 c.1868_1868insT p.Y623fs* - in 4.0% of 375 reads <i>FLT3</i> NM_004119 c.2027A>C p.N676T - in 88.7% of 309 reads (<i>FLT3</i> -ITD) Gain of <i>PIM1</i> (on 6p), gain of <i>RAD21</i> (on 8q), gain of <i>FLT3</i> and <i>PDS5B</i> (on 13q).	<i>NOTCH1</i> NM_017617 c.7230_7230insTG p.P2411fs* - in 52.1% of 121 reads Gain of <i>PIM1</i> (on 6p) <i>TP53</i> mutation (p.V173M)

B



C

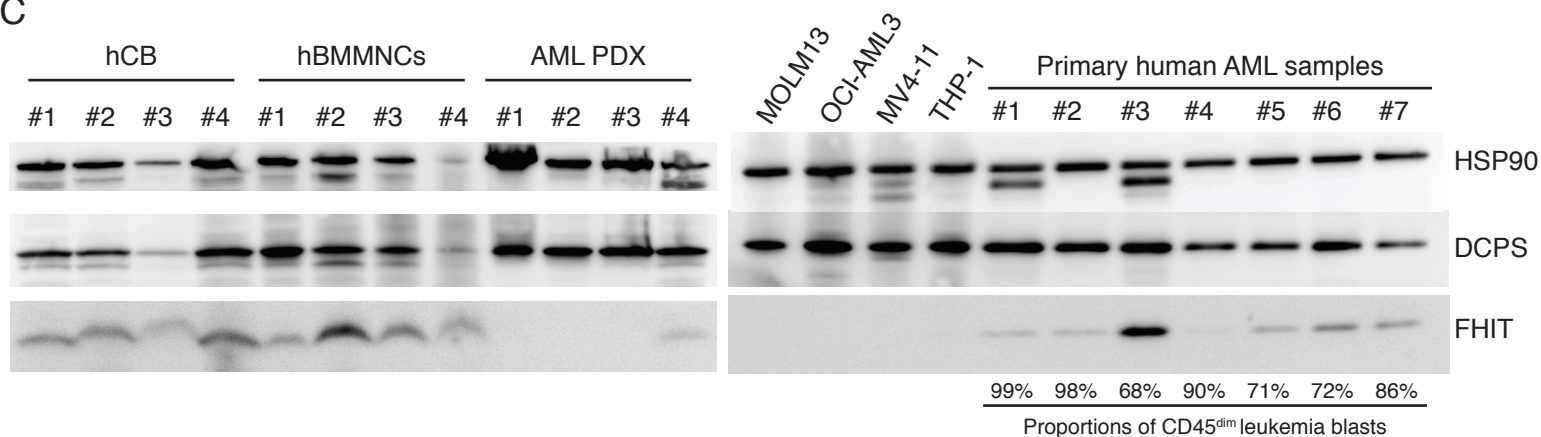


Figure S6. Characteristics of AML PDX lines and Western blot analysis for FHIT. Related to Figure 6. (A) Characteristics of three PDX lines. Information regarding WHO clarification, FAB classification and karyotype was obtained from the ProXe website. Mutational status as assessed by the Rapid Heme Panel (DFCI). (B) Representative results of CETSA using human AML cells harvested from BM of DMSO- or RG3039-treated PDX mice. For CETSA analysis AML cells in BM (DFAM-15354) were harvested the day after the last RG3039 (or DMSO) injection. Asterisk denotes non-specific signal. Equal protein loading was validated by Ponceau-S staining. (C) Western blot analysis of FHIT using anti-FHIT antibody.