### Citations for the species trees used in this analysis

The topology of the species tree for each of the 12 datasets used in this analysis. The rooted versions of these trees were taken as true and used for evaluation of STRIDE. The unrooted version of these species trees were used as input for STRIDE.

Birds (Jarvis, E.D., Mirarab, S., et al. 2014), Flies (Wiegmann, B.M., Trautwein, M.D., et al. 2011),
Fish (Betancur, R.R., Broughton, R.E., et al. 2013), Fungi (James, T.Y., Kauff, F., et al. 2006),
Hymenoptera (Mao, M., Gibson, T., et al. 2015), Kinetoplastids (Seward, E.A. and Kelly, S. 2016),
Laurasiatheria (Meredith, R.W., Janecka, J.E., et al. 2011), Metazoa (Dunn, C.W., Hejnol, A., et al. 2008), Nematoda (Zhou, Y. and Holmes, E.C. 2007), Primates (Perelman, P., Johnson, W.E., et al. 2011), Rodents (Meredith, R.W., Janecka, J.E., et al. 2011) and Plants (Bremer, B., Bremer, K., et al. 2009).

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Seward EA, Kelly S. 2016. Dietary nitrogen alters codon bias and genome composition in parasitic microorganisms. Genome Biol, 17.

Wiegmann BM, Trautwein MD, Winkler IS, Barr NB, Kim JW, Lambkin C, Bertone MA, Cassel BK, Bayless KM, Heimberg AM, *et al.* 2011. Episodic radiations in the fly tree of life. Proceedings of the National Academy of Sciences of the United States of America, 108:5690-5695.

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# Supplemental Text 1

Prior to analysis of the gene trees, the unrooted species tree was analysed to determine the species sets in which genes would be expected to occur following a gene duplication event along any branch of the species tree. For each direction along each branch the sets of species in the child clades (X & Y) and in the grandchild clades  $(x_1, x_2, y_1 \& y_2)$  immediately following that branch were identified (Fig. 8A). Then each gene tree was analysed in turn to identify well-supported gene duplication events within the gene tree as follows: each node, n, in the tree was considered in turn, if the node was an unresolved polytomy it was excluded as such nodes correspond to either a higher-order multiplication events (e.g. triplication) or an unresolved event in the gene tree (e.g. an amalgamation

of several weakly supported bipartitions). Each analysed node therefore had three branches incident on it, and any pair of branches could potentially represent duplicate genes (Fig. 8B). For each pair of branches,  $b_1$  and  $b_2$ , the sets of species,  $S_1$  and  $S_2$ , below each branch were used to identify the locations in the species tree corresponding to these branches in the gene tree. This was done by identifying the smallest block,  $B_i$  in the species tree that contains all the species in  $S_i$  (i=1,2), thus making the method robust in the case of subsequent gene loss (Fig. 8B). If there was more than one block satisfying this criteria, each of these possible blocks were considered. A node, n, was considered as a putative gene duplication event if  $B_1=B_2$ .

Nodes that were identified as putative gene duplication events were further examined to reduce the possibility that their existence or location had been misidentified due to errors in gene tree inference. The criteria were: 1) There must be at least one gene from each of the expected grandchild clades in both  $S_1$  and  $S_2$  (Fig. 8C). 2) The branching structure immediately after the gene duplication event on branches  $b_1$  and  $b_2$  must match the expected branching structure (Fig. 8D), i.e. the first node for each duplicate split the descendent species into the expected sets X and Y, or subsets thereof. Note that it would not be valid to check the topology to the level of grandchild clades in step 2 since this would fail to identify gene duplication events if there were also a subsequent gene duplication event one branch lower in the species tree. In this case, the observed grandchild clades would both be subsets of one of the expected child clades rather than grandchild clades. Steps 1 and 2 check that the observed clades are subsets of the expected clades (rather than requiring they be equal to) as this is necessary to make the method robust to subsequent gene loss events.

# Supplemental Figure Legends

#### Supplementary Figures 1 -12

STRIDE analysis applied to the test datasets gene trees 1) Simulated metazoa dataset 2) Simulated fly dataset 3) Simulated primate dataset 4) Diptera gene trees 5) Fish gene trees 6) Hymenoptera gene trees 7) Kinetoplastid gene trees 8) Metazoa gene trees 9) Nematoda gene trees 10) Primate gene trees 11) Laurasiatheria gene trees 12) Rodent gene trees. A) Numbers of identified gene duplication events are marked on the branches they are observed on and arrows indicate which

block of the bipartition the duplicate genes occur in. Gene duplication events are in agreement with the maximum parsimony root of the tree if the arrow points away from the root, and are in green. Gene duplication events are in disagreement with the maximum parsimony root if the arrow points towards the root (thus suggesting that the indicated clade, that spans the marked root, is monophyletic) and are in blue. The maximum parsimony root is circled in red. B) The probabilities for the location of the root calculated by STRIDE coloured according to the displayed heat map. The correct root is marked with an asterisk.

#### Supplemental Figure S13

Factors affecting the accuracy of STRIDE. A) Number of duplications divided by number of species versus the probability assigned to the correct root. B) Number of duplications conflicting with the maximum parsimony root versus the probability assigned to the correct root. C) Number of conflicting duplications versus the number of species.

#### **Supplemental Figure S14**

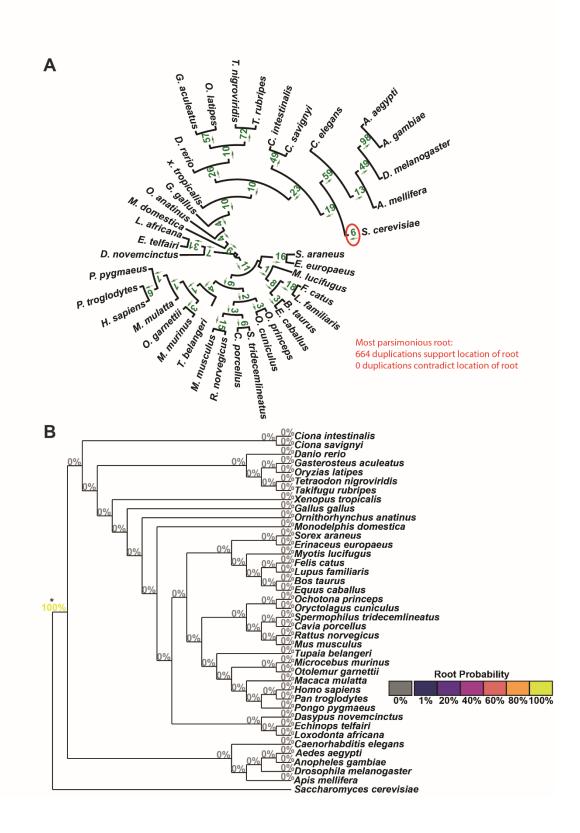
Identification of well-supported gene duplication events for an example gene tree. Upper case letters M,N,O,P & Q are species, lower case m,n,o,p & q are genes from the corresponding species. A) The unknown, rooted species tree (left) and the observed, unrooted species tree (right). Black dot and arrow show the location of a single hypothetical gene duplication event on a branch with time flowing in the direction indicated by the arrow. The branch location and direction is uniquely identified by the block, B, of species whose common ancestor would have inherited the duplicate genes. The expected species in the child clades (X & Y) and grandchild clades ( $x_1$ ,  $x_2$  & Y) after this hypothetical duplication are highlighted with light/dark grey ellipses respectively. B) The unknown, rooted gene tree (left) and the observed, unrooted gene tree (right) for a hypothetical gene family with three gene duplication events (marked by \*) and two gene loss events (grey, dotted line). The node currently being analysed is n and  $b_r$  is the current, tentative direction towards the root. For these n and  $b_r$ ,  $b_1$  and  $b_2$  are analysed to see if they are well-supported gene duplication branches. Si is the set of species below branch  $b_i$ ,  $B_i$  is the smallest block of a bipartition containing  $S_i$  (i=1,2) C) The check that genes from each of the expected grandchild clades are present on each duplicate branch D)

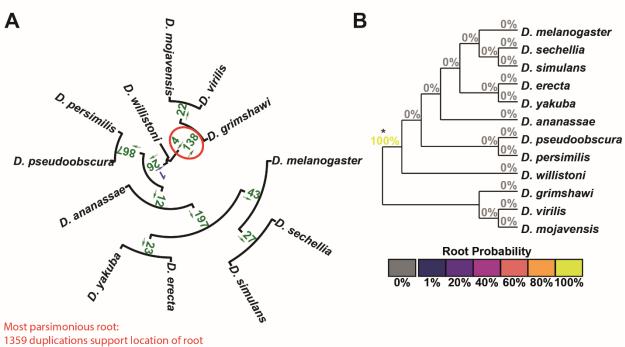
The check that the local topology for each duplication branch agrees with expected topology.  $U_i$  and  $V_i$  are the observed child clades on branch  $b_i$ . The observed child clades should not contain genes from any species not in the expected child clades

### Supplemental Figure S15

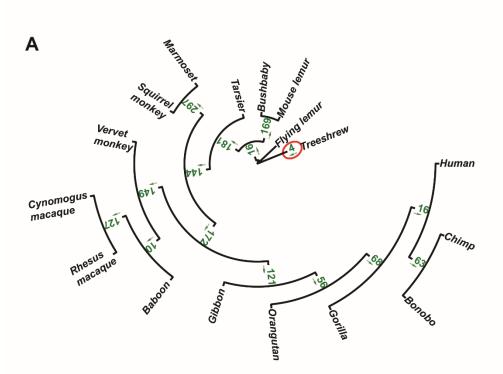
The dependence of the probability for the root of species tree assigned by STRIDE on the parameter,  $\alpha$ , that specifies the ratio of the expected number of false-positive duplications to true-positive duplications on a branch. For each plot,  $\alpha$  ranges from one-tenth of the actual value used by STRIDE ( $\alpha'/10$ ) to ten times the value used by STRIDE ( $10\alpha'$ ).

Fig. S1

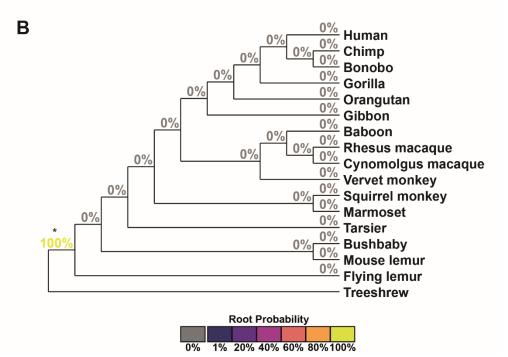


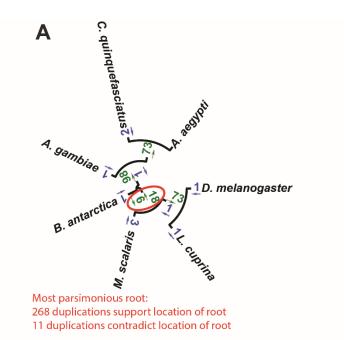


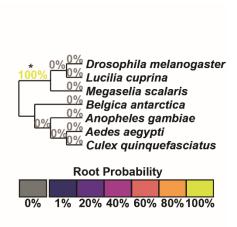
1 duplications contradict location of root



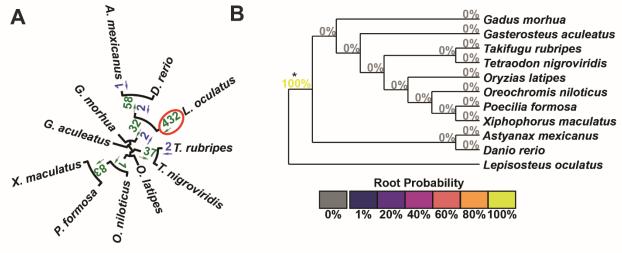
Most parsimonious root: 1593 duplications support location of root 0 duplications contradict location of root



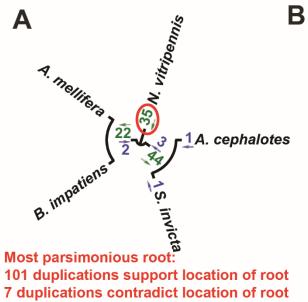


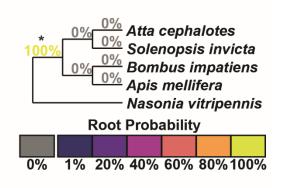


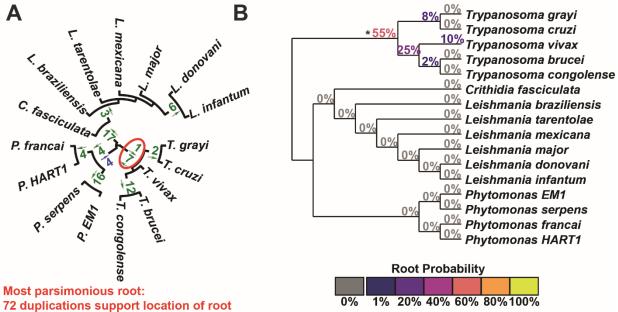
В



Most parsimonious root: 643 duplications support location of root 7 duplications contradict location of root

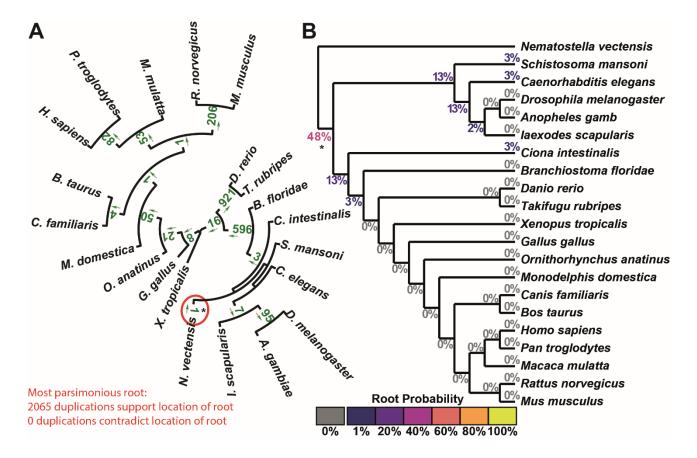


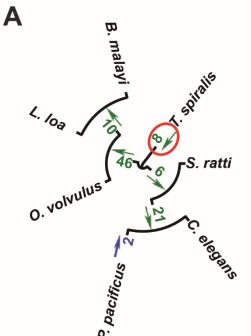


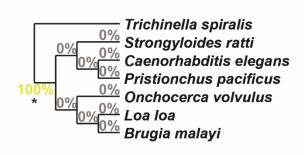


4 duplications contradict location of root









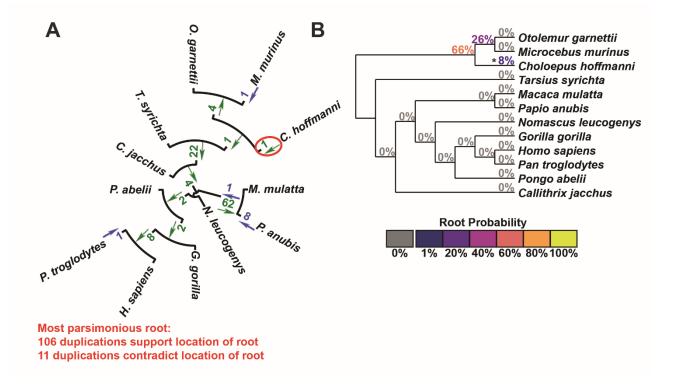
**Root Probability** 

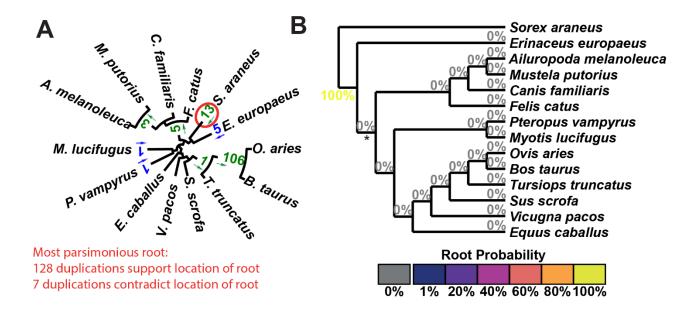
0%

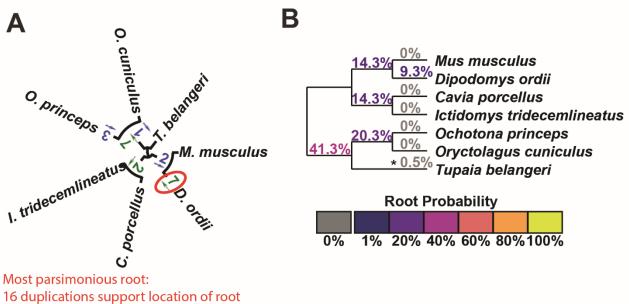
1% 20% 40% 60% 80% 100%

Β

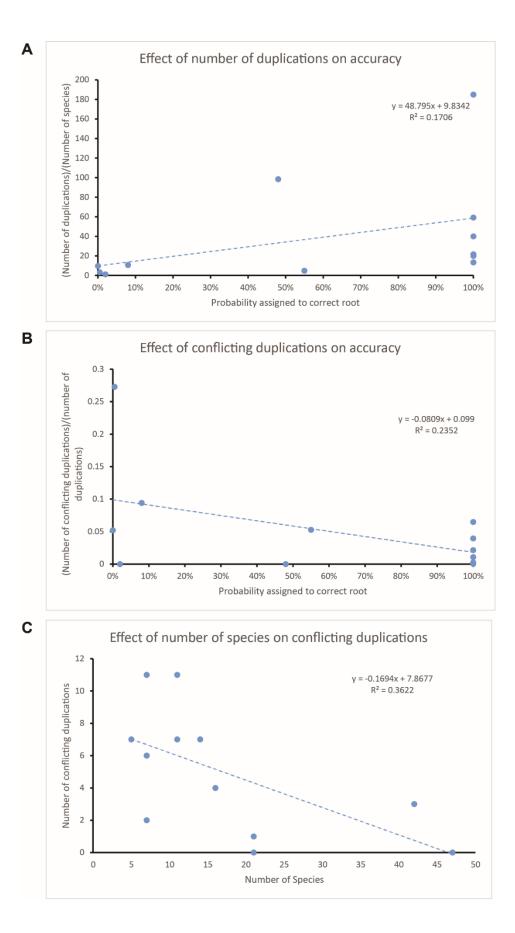
P. pa	
Most parsimonious root:	
91 duplications support location of root	
2 duplications contradict location of root	

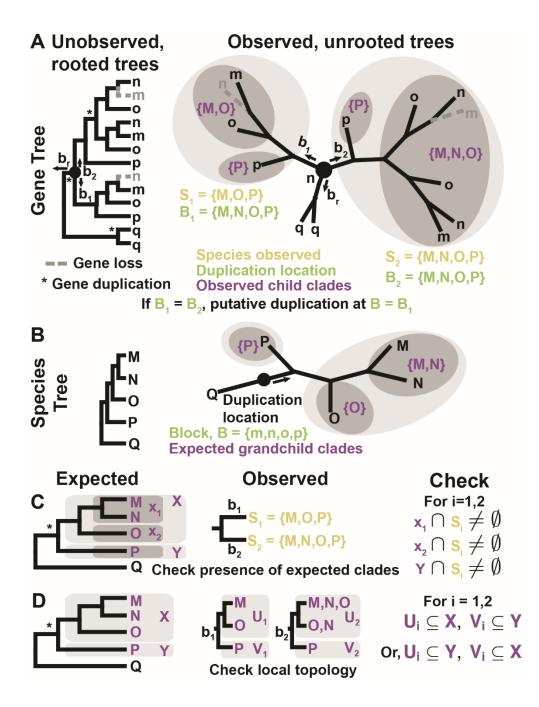




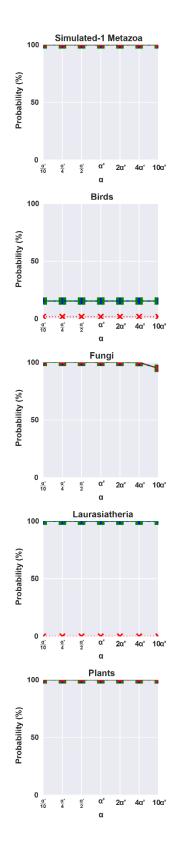


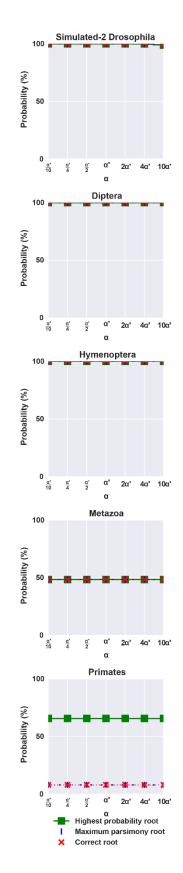
6 duplications contradict location of root











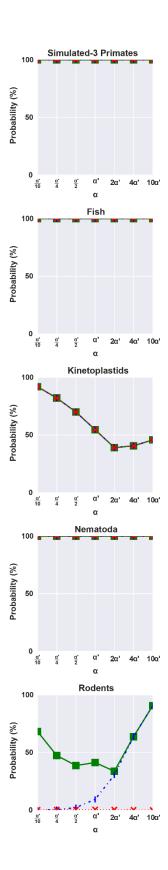


Table S1: Summary of gene duplication prediction accuracy on simulated datasets

		STRIDE				Notung				DLCpar_search			
	nDups	ТР	FP	P (%)	R (%)	ТР	FP	P (%)	R (%)	ТР	FP	P (%)	R (%)
Metazoa	1440	664	0	100.0	46.1	1347	4216	24.2	93.5	1207	85	93.4	83.8
Flies	4628	1359	1	99.9	29.3	3613	20531	15.0	78.1	3479	8624	28.7	75.2
Primates	4317	1592	1	99.9	36.8	3763	7996	32.0	87.2	3133	1620	65.9	72.6
Total	10385	3615	2	99.9	34.8	8723	32743	21.0	84.0	7819	10329	43.0	75.3

nDups = number of gene duplication events in the simulated dataset. TP = true positive. FP = false positive. P = precision. R = recall.