

Supplementary Material and Methods

Food intake

Food intake was measured at time 0, 1, 3, 6, 12, 24 and 36 months of treatment, for a period of 5–14 days. Animals are separated for their morning meal from 09:30 to 11:00 h, and again from 14:00 to 15:00 h, then allowed paired access from 15:00 to 17:00 h. Individual food intake from the morning was summed across pairs and combined with afternoon paired intake to give daily kcal/pair. Daily caloric intake per pair was averaged across the 5–14 days sampling period for each time point.

Hormone assays

Insulin concentrations in monkey plasma were determined using a chemiluminescence-based automatic clinical platform (Roche Diagnostics Cobas e411, Indianapolis, IN); this assay was used previously in non-human primates (Varlamov *et al.*, 2010). The range of the insulin assay is 0.2–1000 uIU/ml. The intra- and inter-assay variations were <7%.

Leptin concentrations were measured by radioimmunoassay following the manufacturer's instructions (Millipore HL-81K, Billerica, MA). This assay was described previously for the nonhuman primate (Power *et al.*, 2013). The assay range was 0.78–100 ng/ml and intra- and inter-assay variations were 13.9% and 24.7%, respectively.

Total adiponectin levels were measured by ELISA following the manufacturer's instructions (Alpco 80-ADPHY-E01, Salem, NH) and was previously validated in nonhuman primates (Swarbrick *et al.*, 2009). The assay range was 0.075–4.8 ng/ml. Intra- and inter-assay variations for total adiponectin were 1.9% and 10.9%, respectively.

Plasma ghrelin levels were measured by RIA following the manufacturer's instructions (Phoenix RK-031-31, Burlingame, CA) and this

assay was described previously in humans (Weigle *et al.*, 2003). The assay range was 10–1280 pg/ml and intra- and inter-assay variations were 3.2% and 12.2%, respectively.

C-reactive protein was measured by ELISA following the manufacturer's instructions (Alpco 30–9710 s). This assay was described previously in humans (Harris *et al.*, 2013), and the assay range was 1.9–150 ng/ml and intra- and inter-assay variations were 4.2% and 21.6%, respectively.

Supplementary Material References

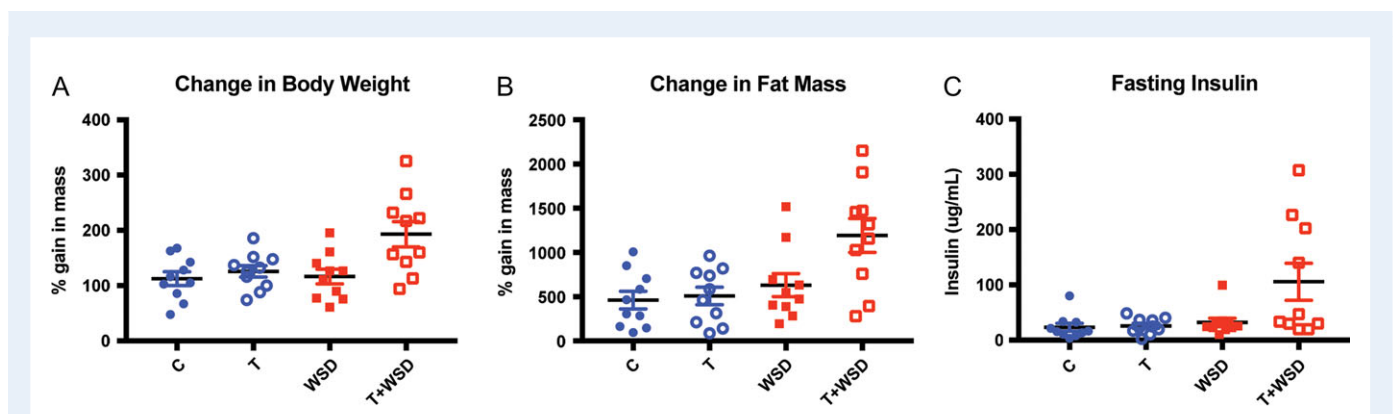
Harris CB, Chowanadisai W, Mishchuk DO, Satre MA, Slupsky CM, Rucker RB. Dietary pyrroloquinoline quinone (PQQ) alters indicators of inflammation and mitochondrial-related metabolism in human subjects. *J Nutr Biochem* 2013;**24**:2076–2084.

Power ML, Ross CN, Schulkin J, Ziegler TE, Tardif SD. Metabolic consequences of the early onset of obesity in common marmoset monkeys. *Obesity (Silver Spring)* 2013;**21**: E592–E598.

Swarbrick MM, Havel PJ, Levin AA, Bremer AA, Stanhope KL, Butler M, Booten SL, Graham JL, McKay RA, Murray SF *et al.* Inhibition of protein tyrosine phosphatase-1B with antisense oligonucleotides improves insulin sensitivity and increases adiponectin concentrations in monkeys. *Endocrinology* 2009;**150**: 1670–1679.

Varlamov O, Somwar R, Cornea A, Kievit P, Grove KL, Roberts CT Jr. Single-cell analysis of insulin-regulated fatty acid uptake in adipocytes. *Am J Physiol Endocrinol Metab* 2010;**299**: E486–E496.

Weigle DS, Cummings DE, Newby PD, Breen PA, Frayo RS, Matthys CC, Callahan HS, Purnell JQ. Roles of leptin and ghrelin in the loss of body weight caused by a low fat, high carbohydrate diet. *J Clin Endocrinol Metab* 2003;**88**: 1577–1586.



Supplementary Figure S1 Individual variability in metabolic measurements following Control (C), Testosterone (T), western-style diet (WSD), and T + WSD treatment. Horizontal lines represent the mean with $n = 10$ /group. Change in body weight (A), fat mass (B) and fasting insulin (C) are shown for the 36-month treatment time point.