

Supplemental Information

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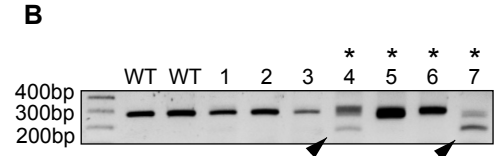
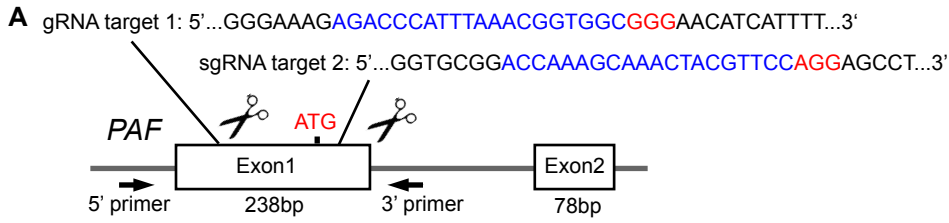
Figure S5. Analysis of Regenerating *PAF* KO intestine and ISCs (related to Figure 4 and Figure 5)

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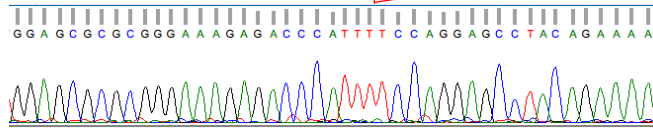
Figure S7. Reduced CRC cell Stemness by *PAF* KO (related to Figure 7)

Supplemental Table

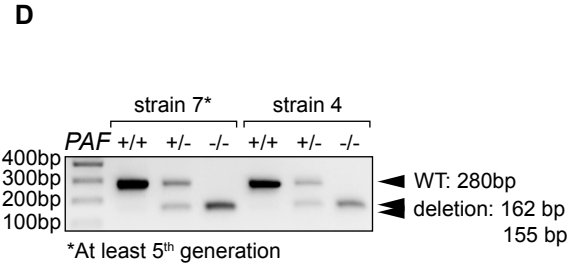
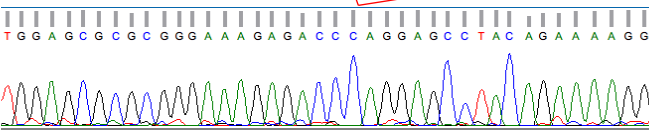
Table S3. Primer sequence information (related to STAR Methods)



C ...AGCGCGCGGGAAAGAGACCCATTTAAACGGT-----ACTACGTTCCAGGAGCCTACA....



...GAGCGCGCGGGAAAGAGACCCATTTAAA-----TACGTTCCAGGAGCCTACAGAAAAG...



E Sequencing results of potential off-target regions

Sequence	Mismatches	Gene	Locus	Mutation (-30~+30)
<u>GTTCCCGGCAC TTTTAAATTAAG</u>	4MMs [8:12:13:19]	NM_001099298	chr2:+65604588	No
GCTCCCGCCACCGCTTGAACAAG	4MMs [2:14:17:20]	NM_026577	chr16:+62846516	No
GCCAGAGCAAAATATGTTCCAAG	4MMs [1:5:12:15]	NM_146283	chr13:-21628884	No
ACCAAATCCCACCACGTTCCAGG	4MMs [7:9:10:13]	NM_175177	chr16:+31458336	No
ACCTAAGCATCCTACCTCCAAG	4MMs [4:10:11:16]	NM_001177565	chr12:+88858284	No
ACCTAAGCATGCTACCTCCAAG	4MMs [4:10:11:16]	NM_010120	chr18:+46767791	No
ACGAAAGGAAACTACGGCCTAG	4MMs [3:8:17:18]	NM_019750	chr9:+107482963	No
ACCAAATCACACTATGTTCTTGG	4MMs [7:10:15:20]	NM_172925	chr9:+77504809	No
ACCATAGCAAAATATGCTCCAG	4MMs [5:12:15:17]	NM_001029934	chr11:-84839164	No
ACAAAAGCAAACCACCTTACCAG	4MMs [3:13:16:19]	NM_011916	chr9:-95948855	No
GCCAAAAGCAAAC TCTTTCGCGG	4MMs [1:14:16:20]	NM_016974	chr7:-52960515	No
ACCAAAGCACAGTTCTTCCAGG	4MMs [10:12:14:16]	NM_177863	chr4:+82546380	No
AGCAAAGCAAAC TGTATTCCAG	4MMs [2:14:15:16]	NM_001048192	chr5:+31193044	No
ACCAAAGCAAAC TACTGAGCAAG	4MMs [16:17:18:19]	NM_001166065	chr13:+97705	No

gRNA 1 potential off-target

gRNA 2 potential off-target

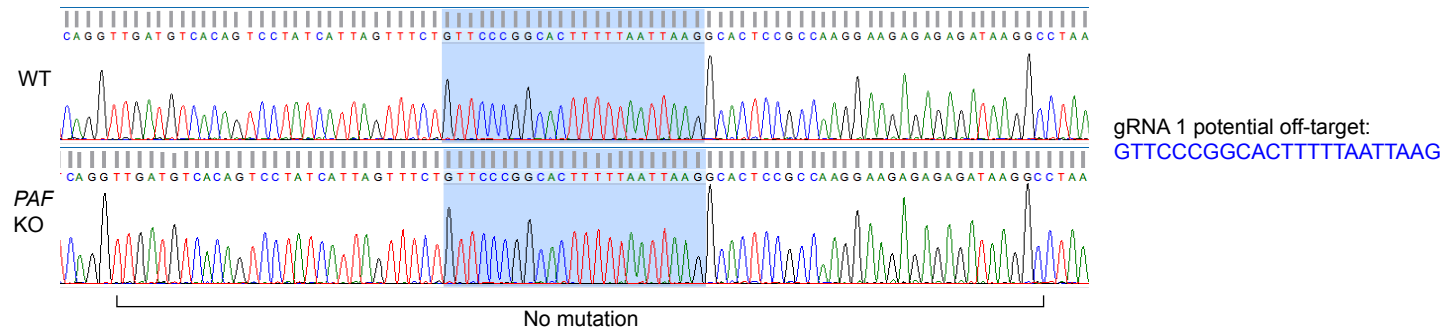


Figure S1. Establishment of *PAF* KO Mice (related to Figure 2)

(A) Schematic view of targeting Exon 1 of *PAF* gene by two guide RNAs. Arrows indicate the 5'- and 3'- primer pairs for genotyping.

(B) Detection of *PAF* deleted chromosome by PCR genotyping. Four pups showed band shift among the seven pups from zygotes injection. Asterisks indicate pups harboring deleted alleles. Strain 4 and 7 showed the small size PCR bands (large size of deletion).

(C) Sequencing analysis of strain 4 and 7. Strain 4 and 7 carried 118bp and 125bp deletion between the two gRNAs targeted regions, respectively. Because deleted region contained first ATG sequences, chromosomes harboring deletion in strain 4 and 7 are null.

(D) Generation of *PAF* KO founder from strain 4 and 7. At least 5 successive breeding with C57BL/6 were conducted for experiments.

(E) Sequencing result of off-target regions. Predicted off-targets of gRNAs (MIT CRISPR Designer) were sequenced using the founder strains. No mutation was detected in +30~-30 region of off-targets. One example of potential off-target sequencing was shown (the lower panel).

PAF (Abcam ab)

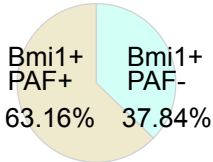
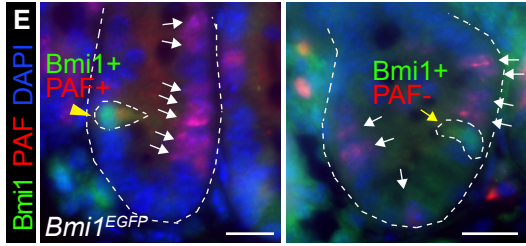
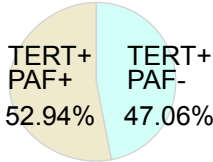
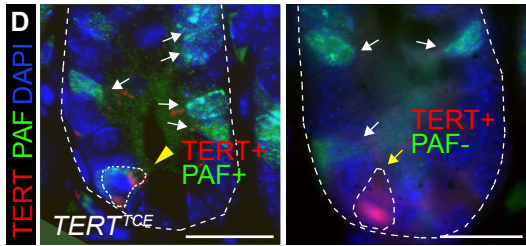
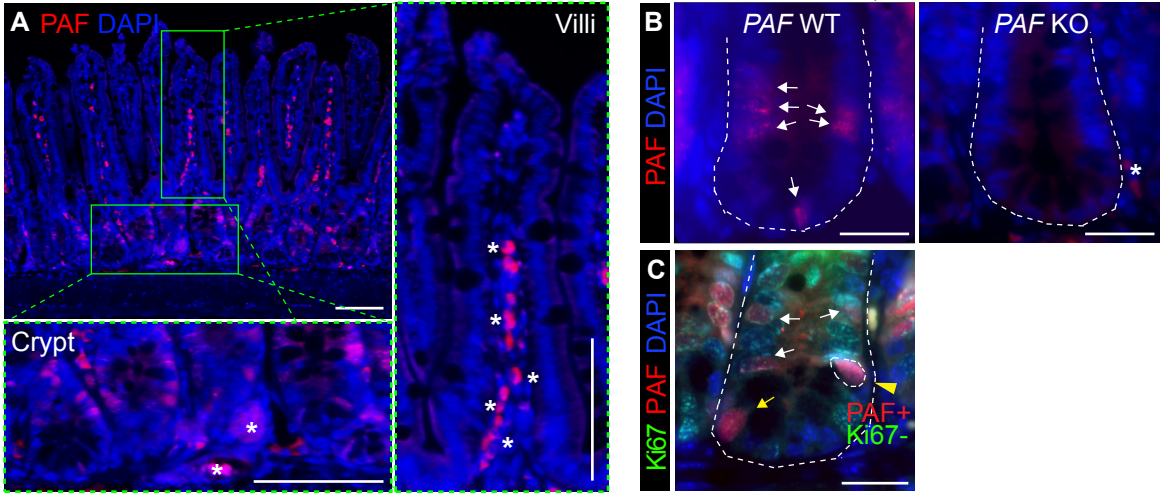


Figure S2. Expression of PAF in Intestinal Crypts (related to Figure 2)

(A) PAF⁺ cell in the mouse small intestine. Mice were analyzed for immunofluorescent (IF) of PAF (red) staining. Green boxes indicate the magnification view of Villi and Crypt. Asterisks mark non-specific staining signals. Scale bars=100 μ m.

(B) Validation of PAF expression by the alternative anti-PAF antibody. Anti-PAF [ab56773] antibody showed the same pattern of PAF expression in *PAF* WT and KO mice as shown in Figure 2A using Anti-PAF [G-11].

(C) PAF⁺:Ki67⁻ cells in the crypt. Co-immunostaining of mouse small intestine (*PAF* WT) for PAF and Ki67. White arrows: PAF⁺:Ki67⁺ cells; yellow arrows: PAF⁺:Ki67⁺ CBC cells; arrowhead: PAF⁺:Ki67⁻ cell.

(D, E) PAF expression in TERT⁺ and Bmi1⁺ cells. Co-immunostaining of *TERT-Tdtomato-CreERT2* knockin mouse for PAF and tdTomato (D); or *Bmi1-EGFP* knockin mouse intestine samples for PAF and Bmi1 (E); Arrowheads: PAF⁺:TERT⁺ (D) and PAF⁺:Bmi1⁺ (E) cells; Arrows: PAF⁻:TERT⁺ (D) and PAF⁻:Bmi1⁺ (E) cells; The numbers indicate % of PAF⁺:TERT⁺ (D) and PAF⁺:Bmi1⁺ (E) cell from the total numbers (TERT⁺ or Bmi1⁺ cells). Asterisks mark non-specific staining signals. Scale bars=20 μ m.

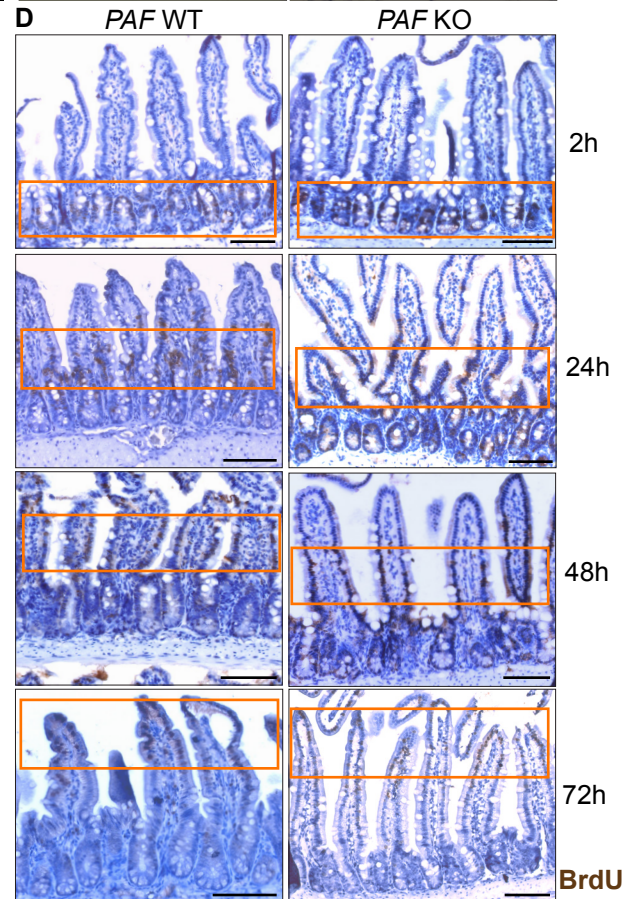
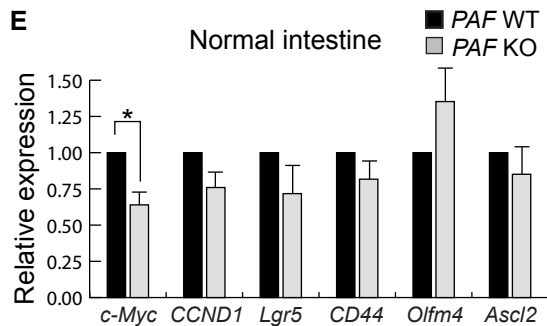
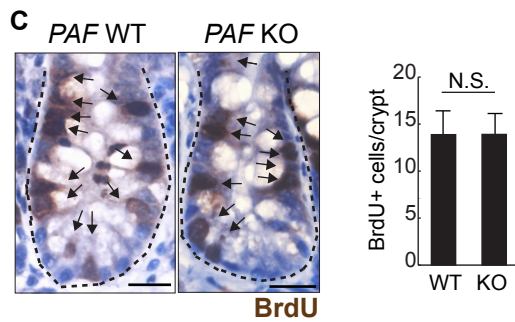
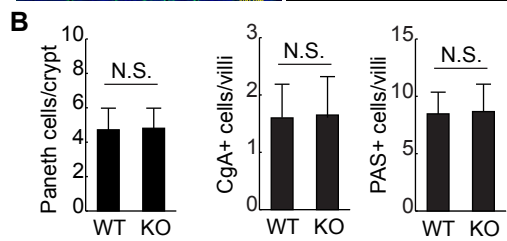
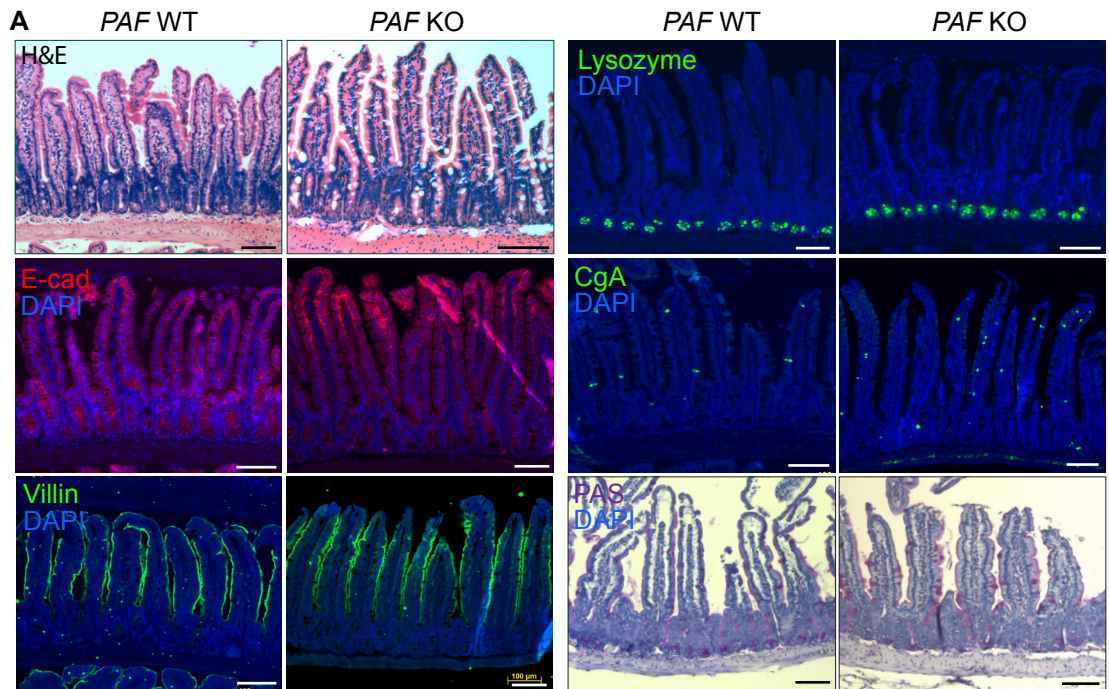


Figure S3. Analysis of *PAF* KO Intestine (related to Figure 3)

(A) Comparison of WT and *PAF* KO intestine in the normal condition. H&E staining and immunostaining for E-cad (epithelial cell marker), Villin (intestinal epithelial marker), Lysozyme (the Paneth cell marker), CgA (Neuroendocrine cell marker), and PAS (Goblet cell marker) in WT and *PAF* KO intestines. Scale bars=100 μ m.

(B) Analysis of Lysozyme+ (the Paneth cell) in the crypt. Quantification of CgA+ (Neuroendocrine cell), and PAS+ (Goblet cell marker) in the intestine. At least 40 fields were counted.

(C) Quantification of proliferation cell in WT and *PAF* KO intestinal crypts. BrdU staining after 2h BrdU induction. The right graph: Quantification of BrdU+ cell in the crypts of *PAF* WT and KO mice. Scale bar=20 μ m.

(D) Tracing the migration of BrdU+ cells in the crypts. BrdU migration assays of *PAF* WT and KO mice. Green boxes indicate the migrating BrdU+ cells. Scale bars=100 μ m.

(E) Comparison of gene expression levels relating intestinal homeostasis in *PAF* WT and KO mice. qRT-PCR was conducted using crypt fraction from *PAF* WT and KO mice. Of note, the slight decrease in *c-Myc* expression was detected. Asterisks: $P < 0.05$.

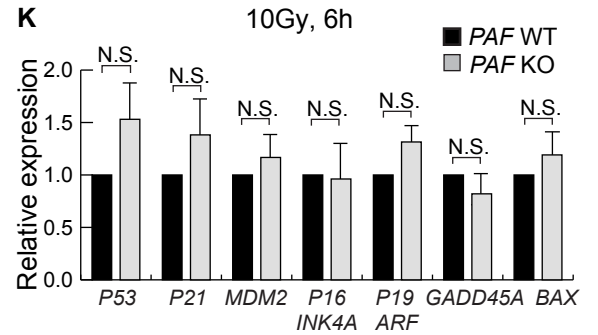
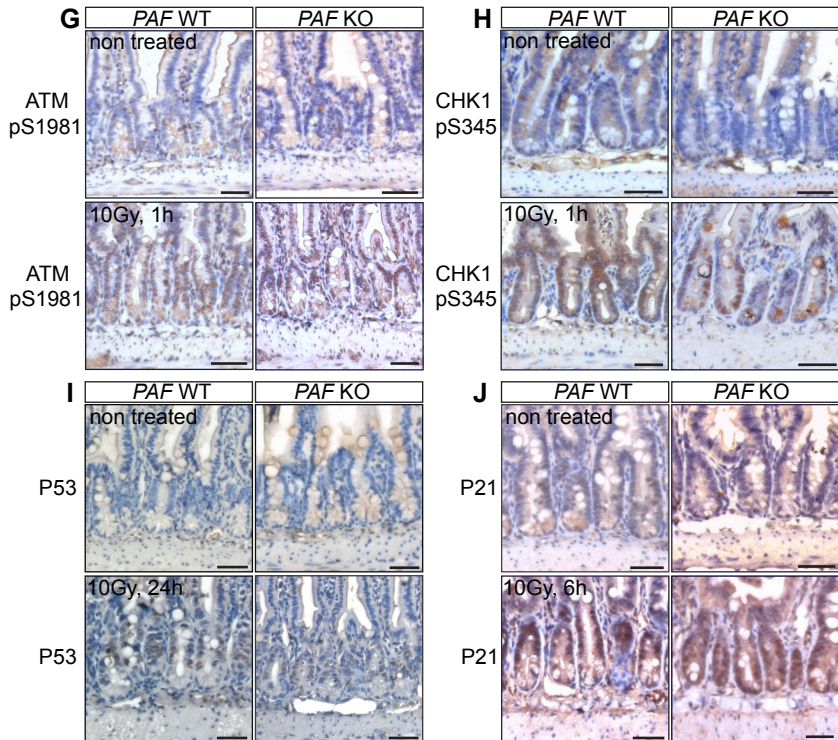
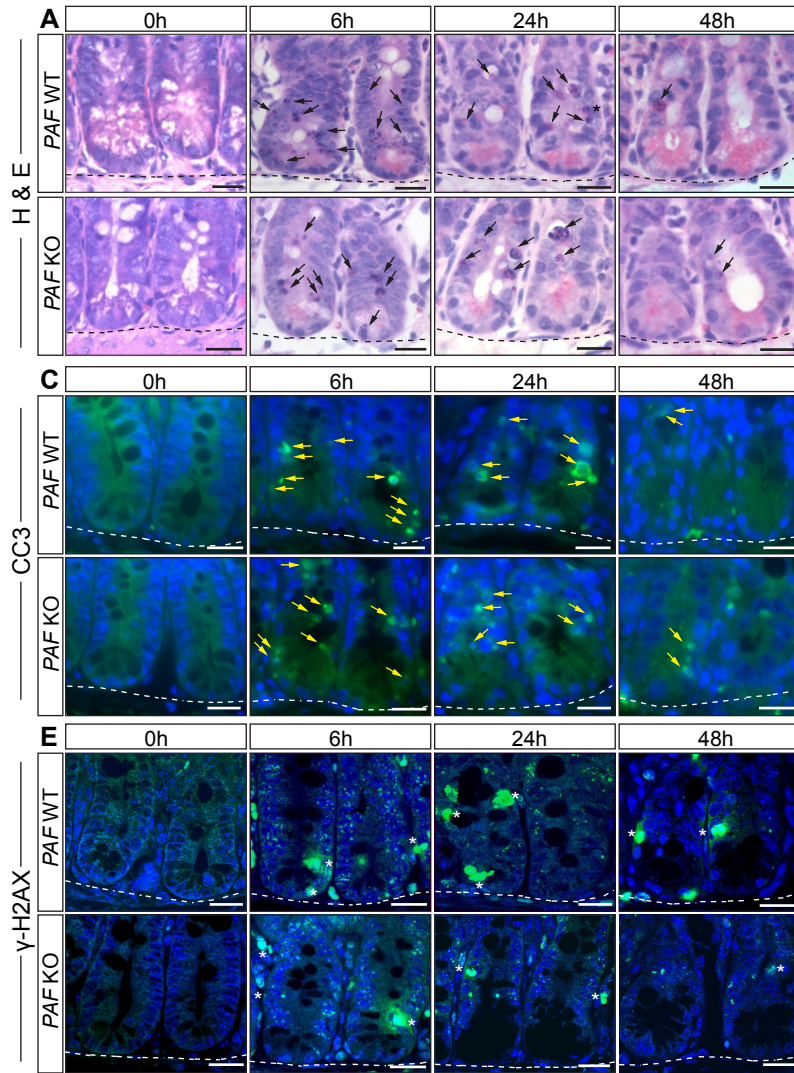


Figure S4. *PAF* KO Does Not Affect Intestinal Cell Death and DNA Damage Responses (related to Figure 3)

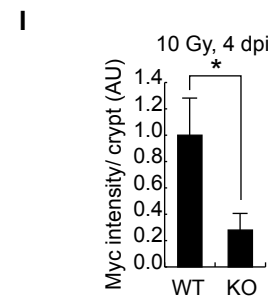
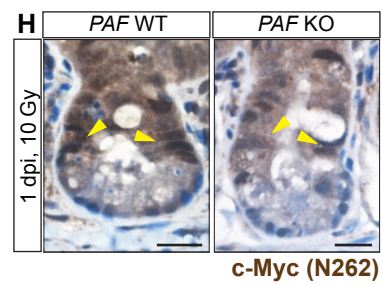
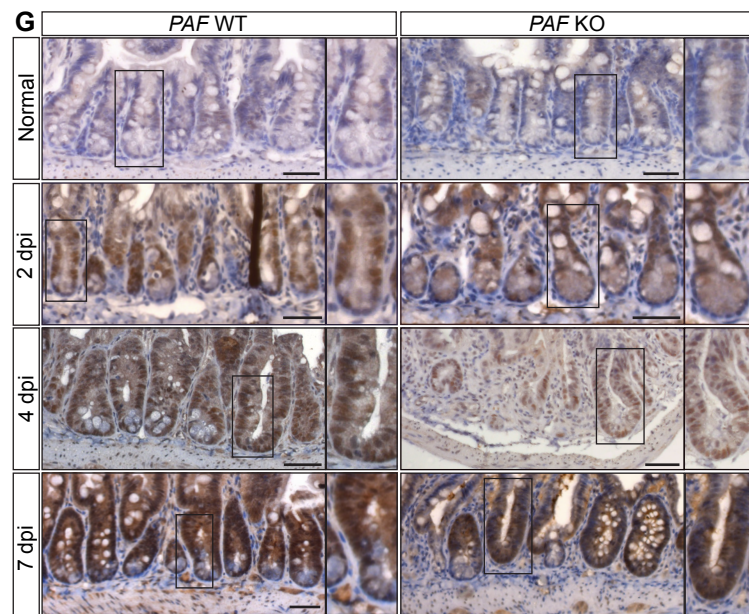
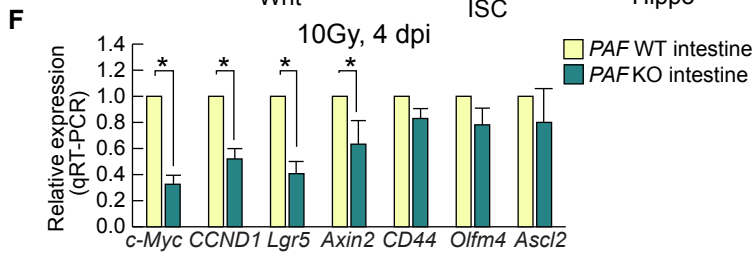
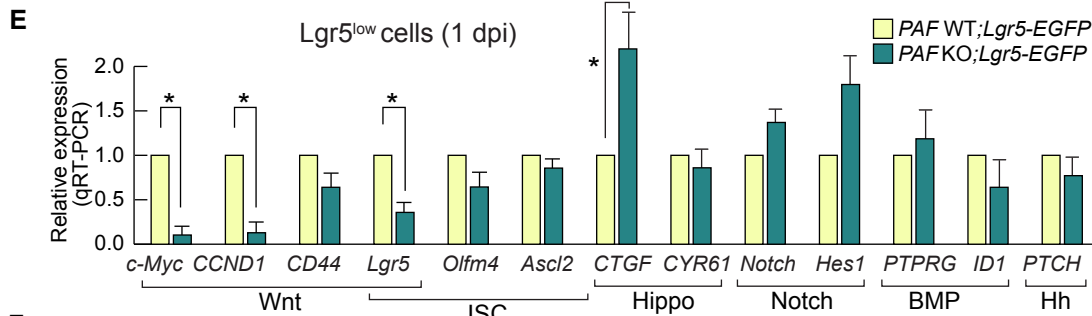
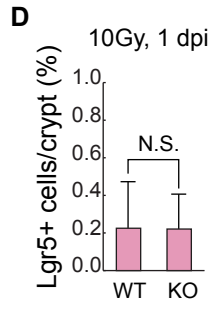
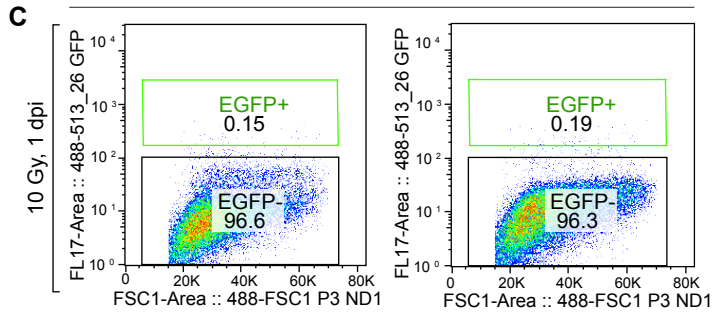
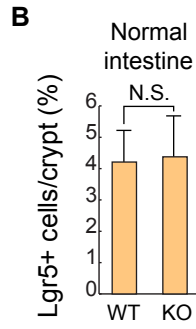
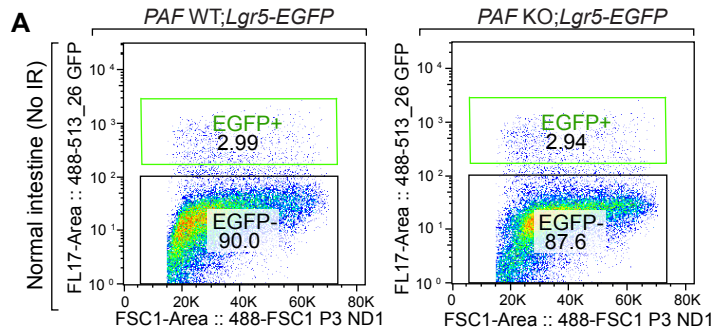
(A-D) No differences in cell death after IR injury between *PAF* WT and KO mice. Apoptotic bodies in the crypts were analyzed at the indicated time points (0h, 6h, 24h, 48h) (A); Quantification of apoptotic bodies in the crypt (B); IF staining of *PAF* WT and KO mice were performed for cleaved-Caspase 3 (CC3), a marker for apoptosis (C) at the indicated time points (0h, 6h, 24h, 48h); Quantification of CC3 in the crypt (D). Scale bars=20 μ m. The representative images were shown from at least three mice.

(E, F) No differences in DNA damage (γ -H2AX) response after IR injury between *PAF* WT and KO mice. DNA double-strand break was analyzed by IF staining using γ -H2AX between the *PAF* WT and KO mice crypts at the indicated time points (0h, 6h, 24h, 48h) (E); Quantification of γ -H2AX in the crypt from *PAF* WT and KO mice (F). Scale bars=20 μ m. The representative images were shown from at least three mice.

(G, H) The early time DNA damage response was not changed in *PAF* KO crypt. IHC for ATM pS1981 in the *PAF* WT and KO mice crypts after irradiation injury (1h, 10Gy)(G); IHC for CHK1 pS345, a downstream target of ATR/ATM DNA damage response, in the *PAF* WT and KO mice crypts after irradiation injury (1h, 10Gy)(H). Scale bars=50 μ m.

(I, J) Myc-related DNA damage response pathways were not changed in *PAF* KO crypt. IHC for P53 (I) (24h, 10Gy) and P21 (J) (6h, 10Gy) in the *PAF* WT and KO mice crypts after irradiation injury. Scale bars=50 μ m.

(K) Gene expression analysis of Myc-related DNA damage response. *PAF* WT and *PAF* KO crypts (6h, 10Gy) were analyzed for qRT-PCR. At least three experiments were conducted. qRT-PCR was conducted using indicating primers.



c-Myc (N262)

Figure S5. Analysis of Regenerating *PAF* KO Intestine and ISCs (related to Figure 4 and Figure 5)

(A, B) No differences in the Lgr5⁺ cell population of the normal intestines between *PAF* WT and KO mice. Enzymatically dissociated cells isolated from *PAF* WT;*Lgr5-EGFP* and *PAF* KO;*Lgr5-EGFP* crypts were assessed by FACS (A); Quantification of the ratio of Lgr5⁺ cells in *PAF* WT and KO mice from at least three independent experiments (B). The representative images were shown from three independent experiments.

(C, D) No differences in Lgr5⁺ cell populations after IR-injury between *PAF* WT and KO mice. FACS analyzed images from *PAF* WT;*Lgr5-EGFP* and *PAF* KO;*Lgr5-EGFP* crypts (C). Of note, after IR injury, the small number of Lgr5^{low} cells were detected in both *PAF* WT;*Lgr5-EGFP* and *PAF* KO;*Lgr5-EGFP* crypts; Quantification of Lgr5⁺ cells in the IR-injured crypts (1 dpi, 10 Gy) from at least three independent experiments (D). The representative images were shown from three independent experiments.

(E) Target gene analysis of Wnt, Hippo, Notch, BMP, and Hh signaling pathways in the sorted Lgr5⁺ cells. FACS-isolated Lgr5⁺ cells from *PAF* WT;*Lgr5-EGFP* and *PAF* KO;*Lgr5-EGFP* crypts (1 dpi, 10Gy) were analyzed for qRT-PCR. At least three experiments were conducted.

(F) Target gene analysis of Wnt signaling pathways in whole intestine. Whole intestinal crypts (4 dpi, 10Gy) were analyzed for qRT-PCR. At least three experiments were conducted.

(G) Decreased c-Myc expression in normal and regenerating intestinal crypts of *PAF* KO. c-Myc immunostaining (brown, N-262) of the mouse small intestine (*PAF* WT and KO) at the indicated time points. Hematoxylin for nuclear counterstaining (blue). Scale bars=50μm.

(H) Weak expression of Myc in regenerating crypt of *PAF* KO at 1 dpi. Yellow arrowheads indicated that the cells showed high Myc expression in the crypts after IR injury. Scale bars=20μm.

(I) Quantification of Myc intensity in *PAF* WT and *PAF* KO regenerating intestine (10Gy, 4dpi). For quantifying the signal intensity, at least 20 crypts from the indicated genotypes (n=3) were analyzed by image J.

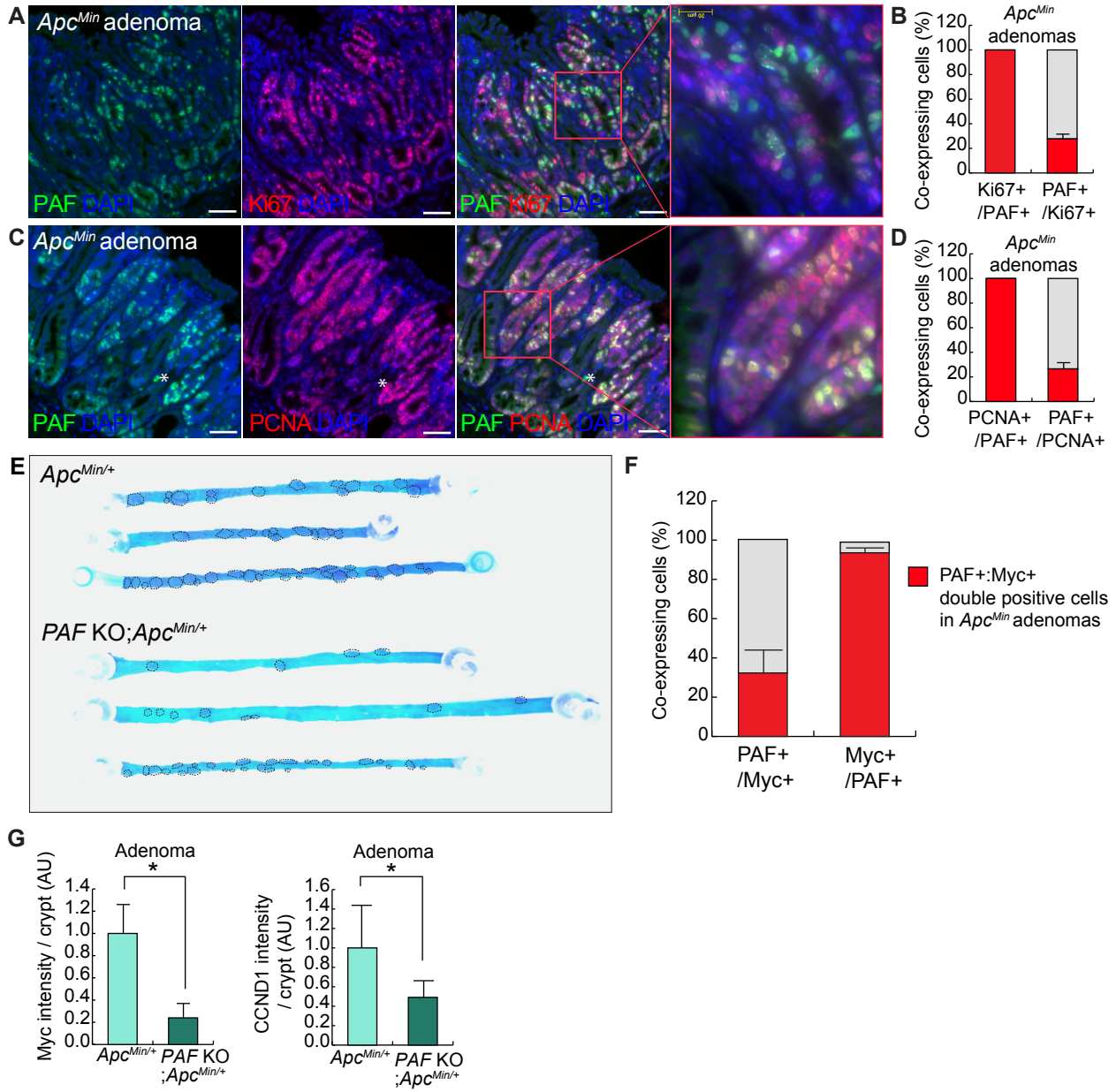


Figure S6. Reduced Intestinal Tumorigenesis by *PAF* KO (related to Figure 6)

(A, B) *PAF* and Ki67 co-expression analysis in *Apc^{Min}* adenomas. The representative images of *PAF* and Ki67 co-immunostaining of *Apc^{Min}* (16 weeks) adenomas (A); Quantification of co-expressing cells (B). 100% of *PAF*+ cells were colocalized with Ki67 staining (N≥500). 27.68% of Ki67 expressing cells (Ki67+) were colocalized with *PAF* expression (N≥500). Scale bars=50μm.

(C, D) *PAF* and PCNA co-expression analysis of *Apc^{Min}* adenomas. Representative images of *PAF* and Ki67 co-immunostaining of *Apc^{Min}* (16 weeks) adenomas (C); Quantification of co-expressing cells (D). 100% of *PAF*+ cells were co-localized with PCNA staining (N≥500). 26.49% of PCNA expressing cells (PCNA+) were co-localized with *PAF* expression (N≥500). Asterisk mark non-specific staining signals. Scale bars=50μm

(E) Decreased the development of *Apc* mutation-driven adenomas in *PAF* KO mice. The representative image of the whole small intestine from *Apc^{Min/+}* and *PAF* KO;*Apc^{Min/+}* mice.

(F) *PAF* and Myc co-expression analysis in *Apc^{Min}* adenomas. 33.96% of Myc-expressing cells (Myc+) were co-localized with *PAF*+ cells (N≥400). 97.25% of *PAF*+ cells were co-localized with Myc+ cells (N≥150).

(G) Quantification of *Myc* and *CCND1* intensity in *PAF* WT; *Apc^{Min}* and *PAF* KO; ; *Apc^{Min}* adenomas (16 weeks old). For quantifying the signal intensity, at least 15 adenomas from the indicated genotypes (n=3) were analyzed by image J.

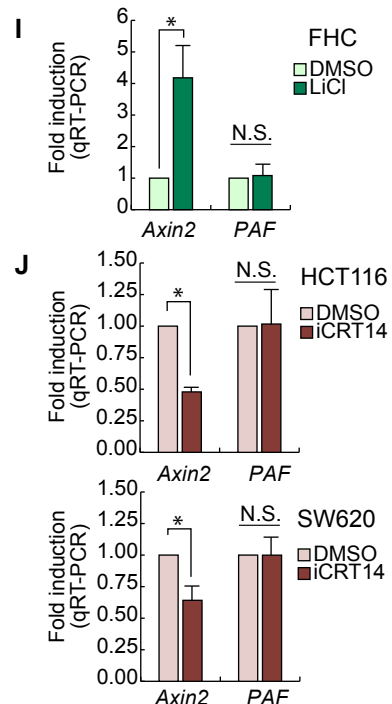
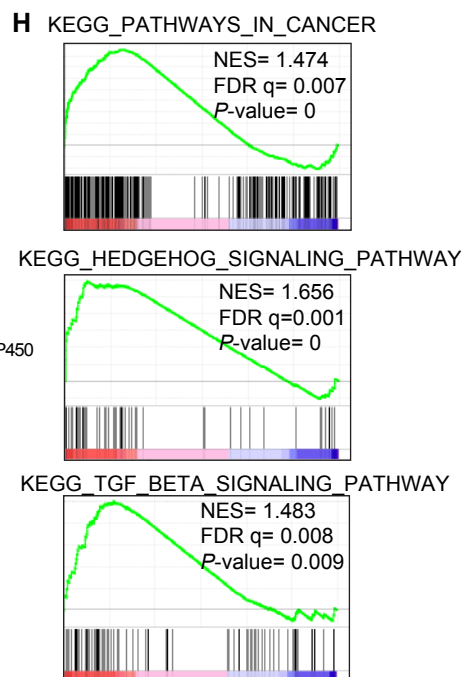
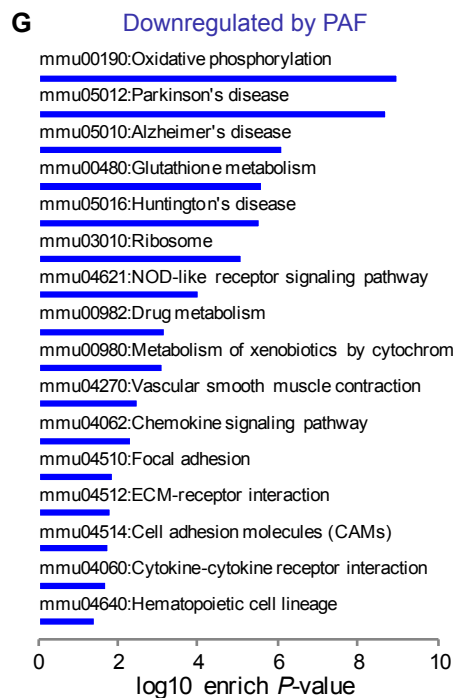
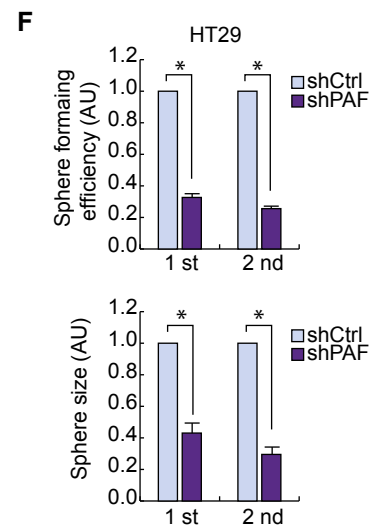
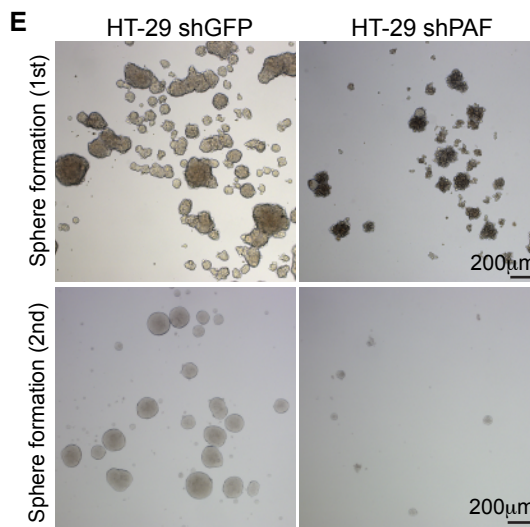
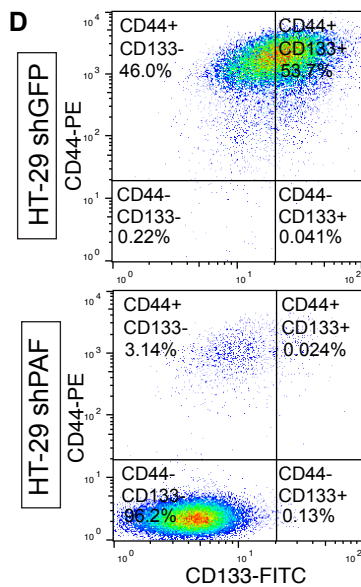
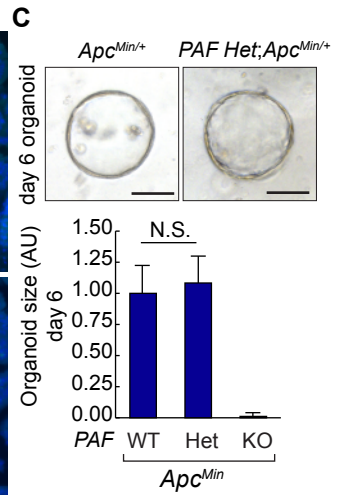
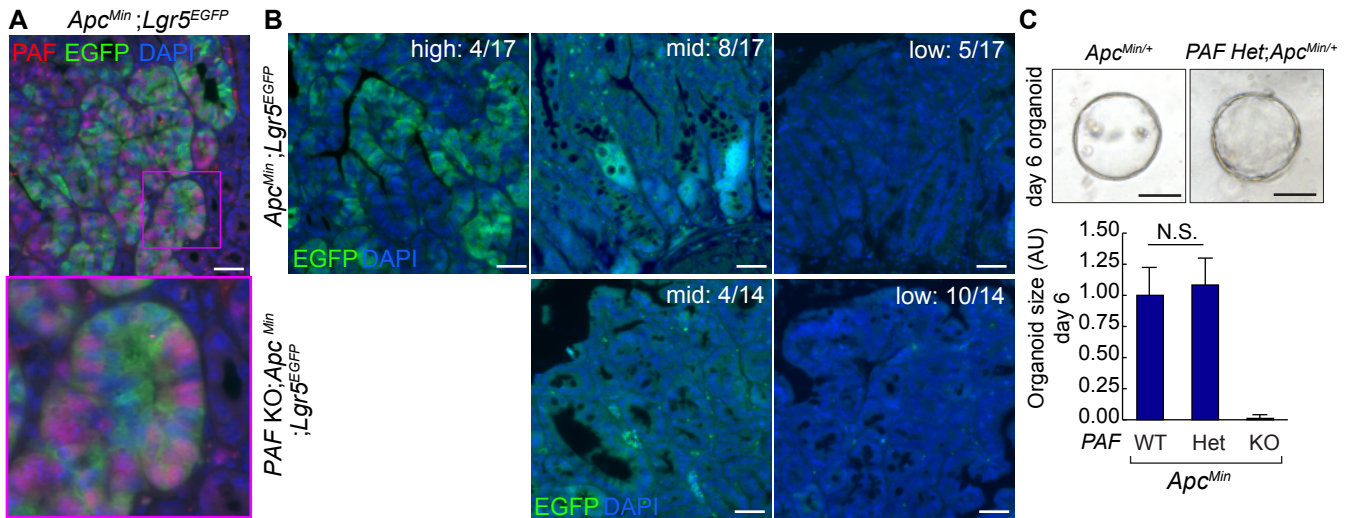


Figure S7. Reduced CRC Cell Stemness by PAF KO (related to Figure 7)

(A) PAF and Lgr5 (GFP) immunostaining in *Apc^{Min}* adenomas. Scale bars=100 μ m.

(B) Analysis of Lgr5 expression in adenomas from *Apc^{Min/+};Lgr5-GFP* and *PAF KO;Apc^{Min/+};Lgr5-EGFP* (16 wk old). Scale bars=100 μ m.

(C) No differences in the cystic organoid growth between *Apc^{Min/+}* and *PAF Het;Apc^{Min/+}* ($N \geq 30$ cystic organoids were analyzed). Scale bars=100 μ m.

(D) PAF depletion reduced expression of CD44 and CD133 in CRC cell line. FACS analysis for CD44 and CD133 using shGFP (control) and shPAF of HT-29 cells.

(E, F) PAF depletion reduced sphere formation in CRC cell line. Representative images of sphere formation assay (1st and 2nd) using shGFP (control) and shPAF of HT-29 cells (E); Quantification of the number and the size of spheres (F). Scale bar=200 μ m.

(G) Significantly PAF downregulated signaling pathways identified by GSEA analysis (KEGG) using RNA-seq result. $P < 0.05$.

(H) Gene set enrichment analysis (GSEA) for PAF activated signaling pathways using KEGG method. NES, normalized enrichment score; FDR, False detection rate; P-value, Nominal P-value.

(I) No differences in PAF expression upon Wnt activation. FHC (normal intestinal epithelial cell line) cells were treated with LiCl, an inhibitor of GSK3 for β -catenin stabilization (Wnt agonist)(25 mM, 24h) and analyzed for qRT-PCR. Expression of Axin2 was used as a positive control.

(J) No differences in PAF expression upon Wnt inactivation. HCT116 and SW620 (CRC cell lines) cells were treated with iCRT14 (Wnt signaling inhibitor) (100 μ M, 24h) and analyzed for qRT-PCR.

Table S3. Primer sequence information (Related to STAR Methods)

List of all primer sequences used for qRT-PCR and ChIP assays

qRT-PCR primer		
Murine gene	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>PAF</i>	TGTGATCAGGTTGCAAAGGA	TTCAGGCTGTCCCCTAAAGA
<i>c-Myc</i>	ATGCCCTCAACGTGAACTTC	GTCGCAGATGAAATAGGGCTG
<i>CCND1</i>	GCGTACCCTGACACCAATCTC	CTCCTCTTCGCACTTCTGCTC
<i>Axin2</i>	TGACTCTCCTTCCAGATCCCA	TGCCCACACTAGGCTGACA
<i>CD44</i>	CACCATTGCCTCAACTGTGC	TTGTGGGCTCCTGAGTCTGA
<i>CD133</i>	TCGTACTGGTGGCTGGGTGGC	ACCACAAGGATCATCAATATC
<i>Lgr5</i>	ACCCGCCAGTCTCCTACATC	GCATCTAGGCGCAGGGATTG
<i>Olfm4</i>	TGGCCCTTGAAGCTGTAGT	ACCTCCTTGGCCATAGCGAA
<i>Ascl2</i>	AAGCACACCTTGACTGGTACG	AAGTGGACGTTTGCACCTTCA
<i>CTGF</i>	AGCCTCAAACCTCAAACACC	CAACAGGGATTTGACCAC
<i>CYR61</i>	GATGACCTCCTCGGACTCGAT	CGTGCAGAGGGTTGAAAAGAA
<i>Notch</i>	TCAGTGTGACCCAGACCTTG	CAAAGGCCAGAAAGAGCTG
<i>Hes1</i>	GGTATTTCCCAACACGCT	GGCAGACATTCTGGAAATGA
<i>PTPRG</i>	CGGAGGTTACTGGAACCGTG	CAGGGTCCCACATAGCCT
<i>ID1</i>	CTGAACGGCGAGATCAGTG	TTTTCTCTTGCCTCCTGAA
<i>PTCH</i>	GCAAGTTTTTGGTTGTGGGT	TTCTCGACTCACTCGTCCAC
<i>HPRT1</i>	TCAGTCAACGGGGGACATAAA	GGGGCTGACTGCTTAACCAG
<i>18S</i>	AAGTCCCTGCCCTTTGTACACA	GATCCGAGGGCCTCACTAAAC
Human gene		
Murine gene	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>PAF</i>	CCAGGGTAAACAAGGAGACG	CAGGAAGCAGTGGCTTAGGA
<i>Axin2</i>	AGCTTACATGAGTAATGGGG	AATTCCATCTACACTGCTGTC
<i>HPRT1</i>	ACCCTTTCCAAATCCTCAGC	GTTATGGCGACCCGCAG
Primers for ChIP assays		
Murine gene	Forward primer (5' to 3')	Reverse primer (5' to 3')
<i>c-Myc-TBE</i>	ATCTGATCAGGGCCGATTT	GAGCCTGCAGAGACCCTAGAA
<i>CCND1-TBE</i>	CTTTTCTCTGCCCGGCTTT	TCTGGAGGCTGCAGGACTT
<i>GAPDH</i>	CCTTAGCCCTGAGCTGTGTC	ATGTTTTCTGGGGTGCAAAG