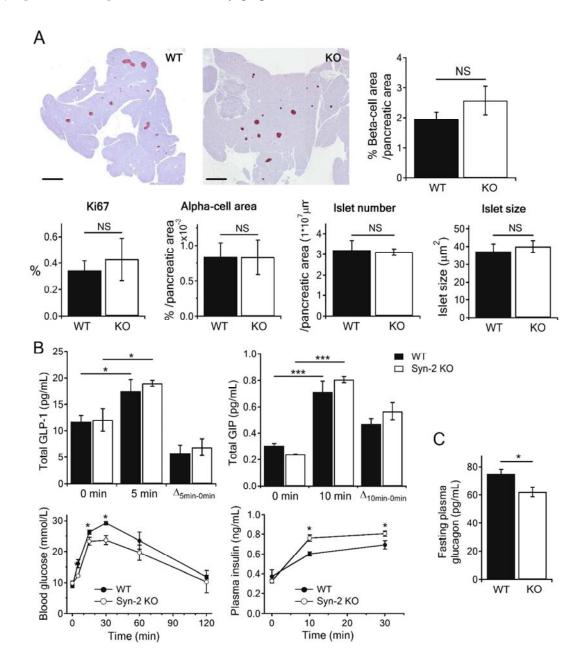
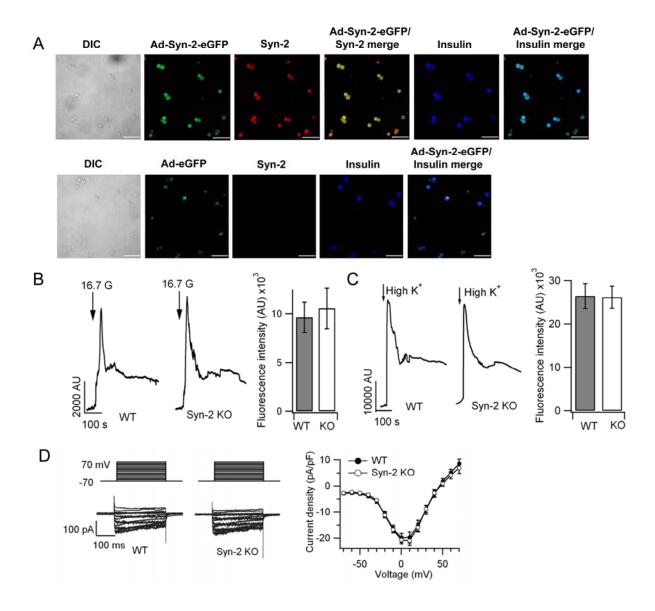
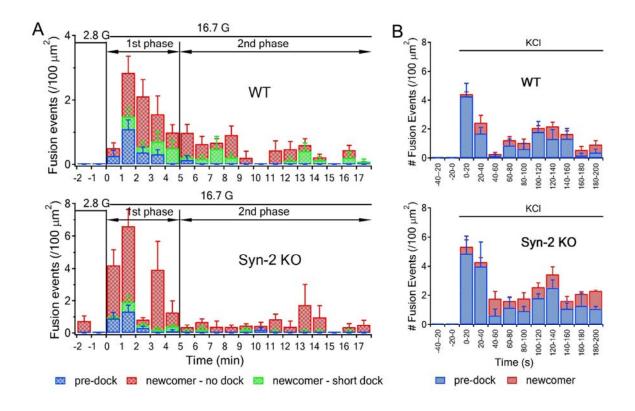
Suppplementary Figure S1. Syn-2 deletion does not affect islet morphology, β-cell mass, or GLP-1/GIP secretion. A: Left: Insulin-immunostained pancreatic sections. Scale bars, 600 μm; Right: Ratios of β-cell area per pancreatic area on mouse pancreatic sections. Bottom from left to right: Ki67-positive islet β-cells as percentage of total islet β-cells on mouse pancreatic sections; Ratios of α-cell area per pancreatic area; islets numbers per pancreatic area; islet sizes on pancreatic sections. N=8-10 mice for each. B: Circulating blood levels of total GLP-1 (*left*) and GIP (*Right*) immunoreactivity (*top*) were measured before and after oral glucose administration, along with determination of blood glucose and insulin levels (*bottom*). N = 6 for each group. C: Fasting plasma glucagon levels of WT and Syn-2 mice (N = 3). \*p < 0.05; \*\*\*p < 0.001. Summary graphs shown as means ± SEMs.



**Supplementary Figure S2-related to Figure 2.** *A*: Confocal images of Syn-2-KO mouse islet β-cells whose Syn-2 expression was restored by Ad-Syn-2-eGFP infection. Top: Syn-2-KO islet β-cells infected with Ad-Syn-2-eGFP; Bottom: Syn-2 KO islet β-cells infected with Ad-eGFP as control. Scale bar: 50 μm. *B-C*: Calcium imaging with fluo-4 showed there was no difference in intracellular  $Ca^{2+}$  concentration increase between WT and Syn-2-KO mouse β-cells stimulated with high glucose (B, 16.7 mM) or with high K<sup>+</sup> (C, 40 mM) stimulation. *D*: Deletion of endogenous Syn-2 in mouse β-cells did not affect voltage-gated calcium channel currents. Representative traces showing Cav currents recorded in whole-cell mode from -70 to 70 mV with 10-mV increment in WT and Syn-2 KO mice (Left). Current-voltage relationship of Cavs from WT (n=15) and Syn-2 KO (n=11) β-cells (*right*). Current were normalized to cell capacitance to yield current density. Values are means ± SEMs.



Supplementary Figure S3. Syn-2 deletion increases newcomer SG exocytosis stimulated by 16.7 mM glucose and pre-dock SG exocytosis stimulated by 50 mM KCl. A: Histogram of fusion events in first (first 5 min after 16.7 mM glucose stimulation) and second phases (5–18 min) in WT (top) versus Syn-2-KO  $\beta$  cells (bottom). Three patterns of fusion events (pre-dock, newcomer-no dock, newcomer-short dock) are indicated by blue, red and green bars, respectively. Data obtained from five independent experiments (2-4 cells from each experiment). B: Histogram of fusion events during 200 seconds of acquisition in WT (top) versus Syn-2-KO  $\beta$  cells (bottom). Pre-dock and newcomer SG fusion events are indicated by blue and red bars, respectively. Data obtained from three independent experiments (3-5 cells from each experiment).



**Supplementary Figure S4-Data analysis for Figure 4.** *A*: Western blot analysis of Syn-2 knockdown expression in INS-1 cells. INS-1 cells were transfected with Syn-2 siRNA and the scrambled siRNA were used as controls. *B*: Analysis of Western blots in **Fig. 4***F* of siRNA knockdown of Syn-2 expression in INS-1 cells at 48 and 72 hrs. *C*: Glucose (as indicated) with or without the presence of 10 nm GLP-1 stimulated insulin secretion were performed on Syn-2 siRNA knockdown and scrambled control INS-1 cells. Data from 4 independent experiments. *D-F*: Quantitative analysis of indicated co-IPed proteins by Syn-2 (*D* for **Fig. 4***E*, untreated INS-1), Syn-1A (*E* for **Fig. 4***G*, Syn-2 siRNA-treated INS-1) and Syn-3 (*F* for **Fig. 4***H*, Syn-2 siRNA-treated INS-1) antibodies. Summary graphs shown are means ± SEMs, N=3. \**p*< 0.05 per Student's *t* test; NS, no significant difference.

