## Supplemental figures



FIG S1. HPLC-MS analysis of authentic compounds.

(A, C, E, G, I, K, and M) HPLC chromatograms of vanillic acid (retention time, 3.1 min), VAL-Tris (2.9 min), VAA (4.7 min), acetovanillone (5.8 min), syringic acid (2.6 min), HPV (4.1 min), and HPS (2.9 min). (B, D, F, H, J, L, and N) ESI-MS spectra of vanillic acid (negative mode), VAL-Tris (positive mode), VAA (negative mode), acetovanillone (positive mode), syringic acid (negative mode), HPV (positive mode), and HPS (negative mode).



FIG S1—Continued.



### FIG S2. Expression of *hpvZ* in *E. coli*.

Proteins (10  $\mu$ g) were separated on SDS-12% polyacrylamide gels and stained with Coomassie Brilliant Blue. Lanes: 1–3, *E. coli* BL21(DE3) harboring pCold I (vector) and pTf16; 4–6, *E. coli* BL21(DE3) harboring pCold12830 and pTf16; 1 and 4, cell extracts; 2 and 5, soluble fractions; 3 and 6, membrane fractions; M, molecular mass markers.





Schematic representations of the disruptions of SLG\_12830 (*hpvZ*) (A) and SLG\_07270 (C) by the insertion of a kanamycin resistance gene (*kan*) and the disruption of SLG\_20400 (E) by deletion. (B) Southern hybridization analysis of SME059. Total DNA of SYK-6 and SME059 digested with XhoI were hybridized with a *hpvZ* probe and a *kan* probe. (D) Southern hybridization analysis of SME092. Total DNA of SYK-6 and SME092 digested with PstI were hybridized with a 07270 probe and a *kan* probe. (F) Colony PCR analysis of SME061 using primers shown in Table 3.



FIG S4. Optimum pH (A) and temperature (B) for HpvZ.

(A) The HPV-oxidizing activity of HpvZ was determined using 50 mM GTA buffer (pH 7.0 to 9.0; open circles) and 50 mM CHES (*N*-cyclohexyl-2-aminoethanesulfonic acid) buffer (pH 8.6 to 10.0; closed circles) at 30°C. (B) The HPV-oxidizing activity of HpvZ was determined using 50 mM Tris-HCl buffer (pH 7.5) at 25 to 50°C. The data represent the averages  $\pm$  standard deviations of three independent experiments.



FIG S5. Chemical structures of substrates used to examine the substrate range of HpvZ. Abbreviations: HPV,  $\beta$ -hydroxypropiovanillone; HPS,  $\beta$ -hydroxypropiosyringone; GGE, guaiacylglycerol- $\beta$ -guaiacyl ether; MPHPV,  $\alpha$ -(2-methoxyphenoxy)- $\beta$ -hydroxypropiovanillone; DCA, dehydrodiconiferyl alcohol; DCA-C, 3-(2-(4-hydroxy-3-methoxyphenyl)-3-(hydroxymethyl)-7-methoxy-2,3-dihydrobenzofuran-5-yl)acrylic acid.

GxGxxG/A motif	
MVDVRTVDYVIVGAGSAGCVLANRLSADRHTEVV MASKTPDFIVIGSGSSGAVIAARLSEDPDASVL MNHHELPNNSMIVIAPSGRDKINWRAALSTEKFDYVIVGAGSAGAVLAARLTEDPAVRVL 	34 33 60 29 31
putativo activo sito	
QQPAINKYLAGPDPFGETDDQMFHYAQVAGGTLYHAVGTCRMGSDPKAVVDARLR ATAPLSEVLTGEVMPGPDVRTPDEIDAWVRNAINTALHPVGTCRMGSDDMAVVDARLR ATAPQSEITGAEIAPGIDVQSDEALDEHIRATTTTTQHPLGTCRMGSGPMAVVDPQLR QAPSMAKHFKHEVVPGQAVKTDDEIIEDIRRRAETIYHPVGTCRMGKDPASVVDPCLK QAPAFDEIR-GKPVYATASNNDDELIEDIRNRADTIYHPVGTCKMGPDSDPMAVVDSSLR :: * * ::*** **. :*** *:	504 494 520 489 489
	GRGxxG/A motif   MVDVRTVDVVIVGAGSAG CVLANRLSADRHTEVV   MSKTPDFIVIGAGSAG CVLANRLSADRHTEVV

# FIG S6. Comparison of the amino acid sequence of HpvZ with those of other GMC oxidoreductase family enzymes.

Enzymes: HpvZ (SLG\_12830), HPV oxidase; PhcC (SLG\_09480) and PhcD (SLG\_09500), DCA-C oxidases of *Sphingobium* sp. SYK-6; AlkJ (CAB51051), alcohol dehydrogenase of *Pseudomonas putida* GPo1; PegA (BAE96591), polyethylene glycol dehydrogenase of *Sphingopyxis terrae*. The GxGxxG/A motif located at N-terminal region (A) and the putative active-site histidine located at C-terminal region (B) are shown.



### FIG S7. Expression of the ALDH genes of SYK-6 in E. coli.

Cell extracts (10  $\mu$ g) of *E. coli* BL21(DE3) carrying each of the 23 ALDH genes were separated by SDS-12% polyacrylamide gels and stained with Coomassie Brilliant Blue. Lane M, molecular mass markers.



FIG S8. Conversion of VAA by cell extract of *desV ligV* double mutant (SME077). A SME077 cell extract (>10 kDa; 500  $\mu$ g of protein/ml) was incubated with 100  $\mu$ M VAA in the presence of CoA, MgSO<sub>4</sub>, and ATP. Portions of the reaction mixtures were collected at the start (A), and after 2 h (B) and 16 h (C) of incubation, and analyzed by HPLC.



#### FIG S9. Expression of *ferA*, *ferB*, and *ferB2* in *E. coli*.

Proteins (10  $\mu$ g) were separated on SDS-12% polyacrylamide gels and stained with Coomassie Brilliant Blue. Lanes: M, molecular mass markers; 1–4, cell extracts of *E. coli* BL21(DE3) harboring pET-16b (lane 1), pE16FA (lane 2), pE16FB (lane 3), and pE16FB2 (lane 4).





VAA (100  $\mu$ M) was incubated with cell extracts of *E. coli* BL21(DE3) harboring pE16FA (A and B), cell extracts of *E. coli* BL21(DE3) harboring pE16FA and *E. coli* BL21(DE3) harboring pE16FB (C), and cell extracts of *E. coli* BL21(DE3) harboring pE16FA and *E. coli* BL21(DE3) harboring pE16FB2 (D) in the presence of CoA, MgSO<sub>4</sub>, and ATP. Portions of the reaction mixtures were collected at the start (A) and after 60 min (B–D) of incubation, and analyzed by HPLC-MS. Negative-ion ESI-MS spectrum of compound IX is shown in panel E.



FIG S11. Conversion of VAA by *ferA* mutant (SME009).

Resting cells (OD<sub>600</sub> of 1.0) of SYK-6 (open circles) and SME009 (closed circles) were incubated with 100  $\mu$ M VAA. Portions of the reaction mixtures were collected, and the amount of VAA was determined by HPLC. All the experiments were performed in triplicate, and the data represent the averages ± standard deviations.