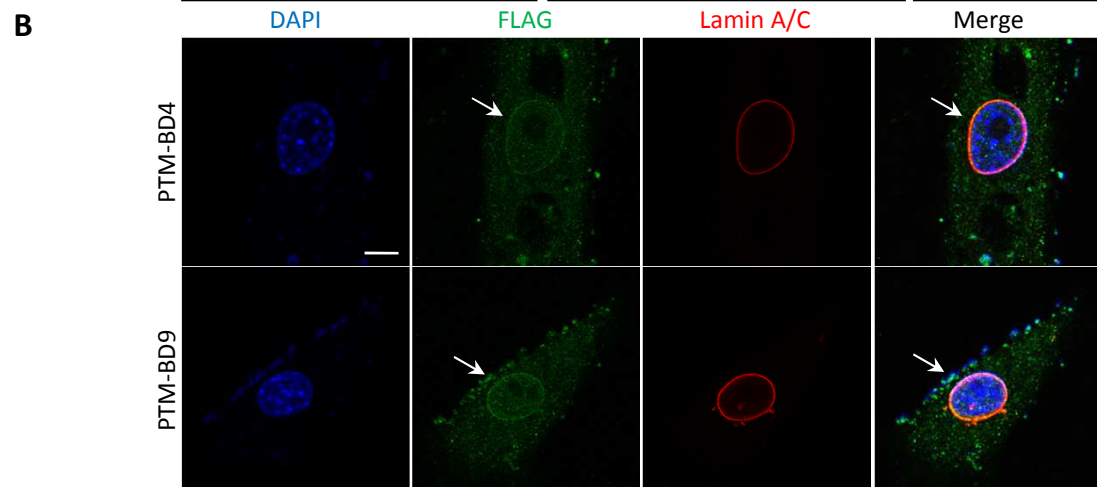
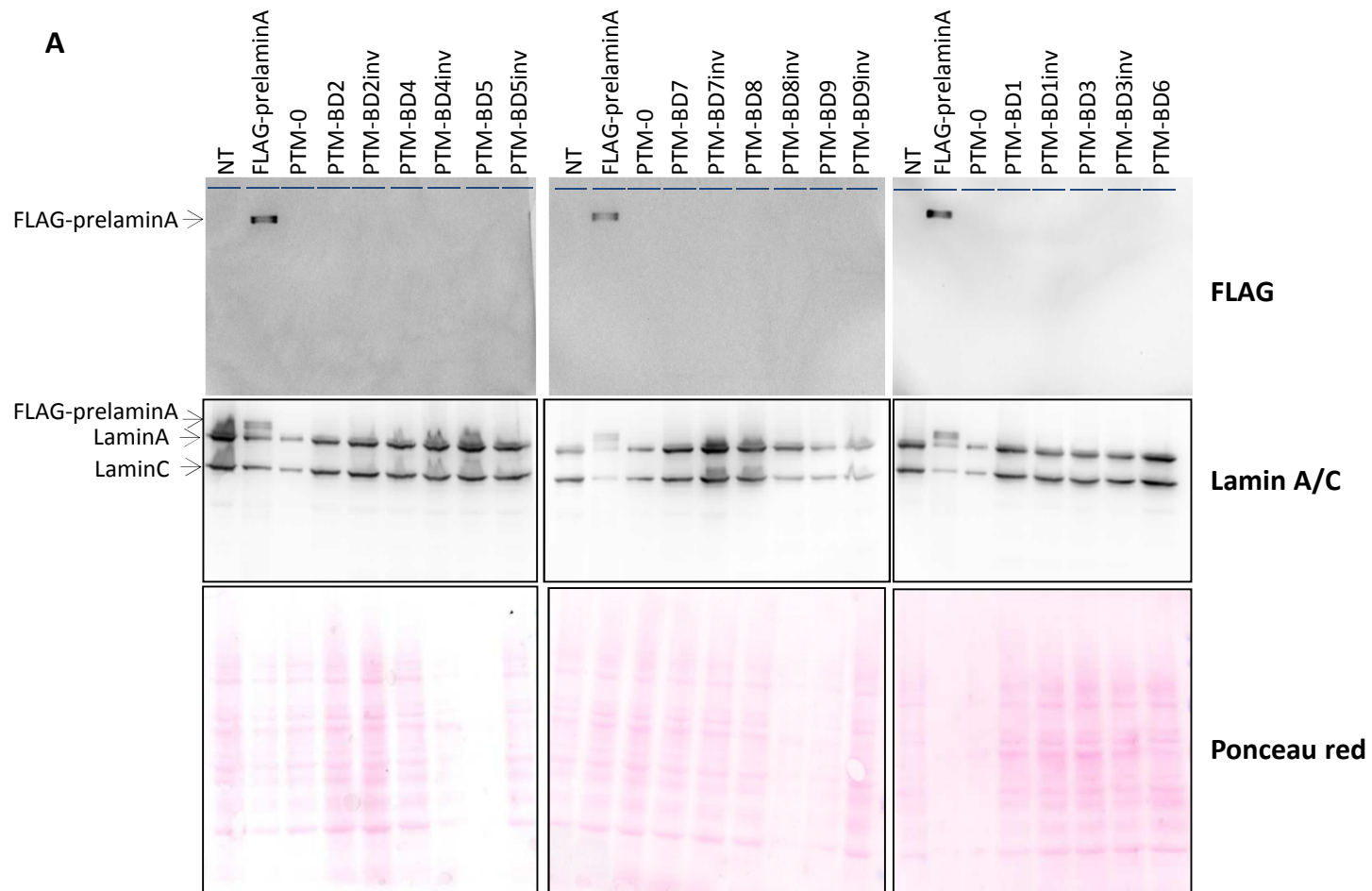


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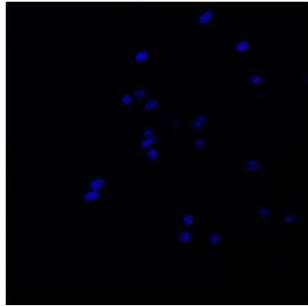
Supplemental Information

Gene Therapy via *Trans*-Splicing for *LMNA*-Related Congenital Muscular Dystrophy

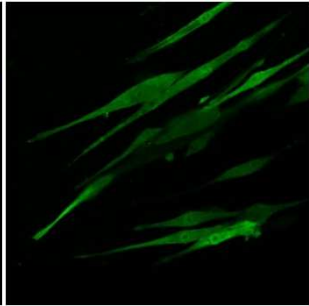
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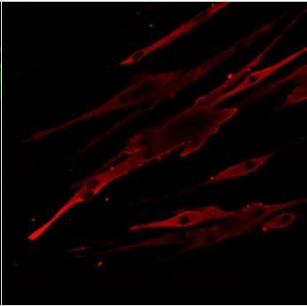
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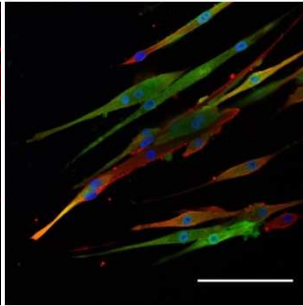
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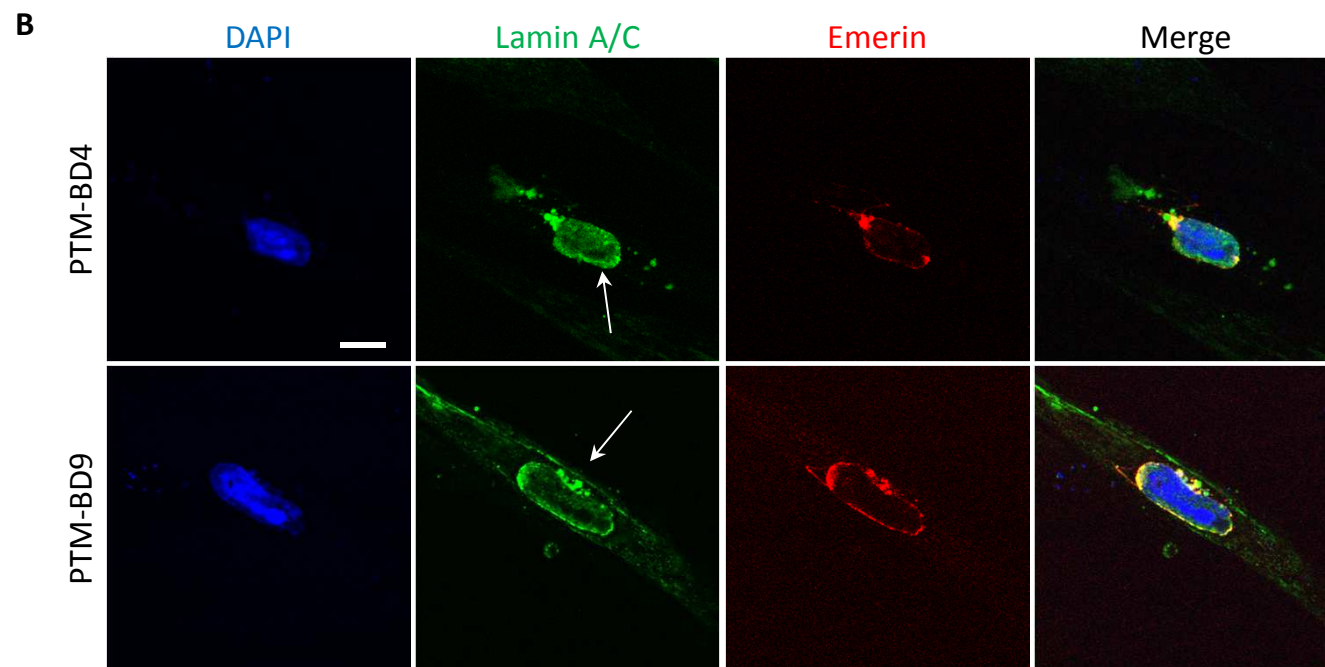
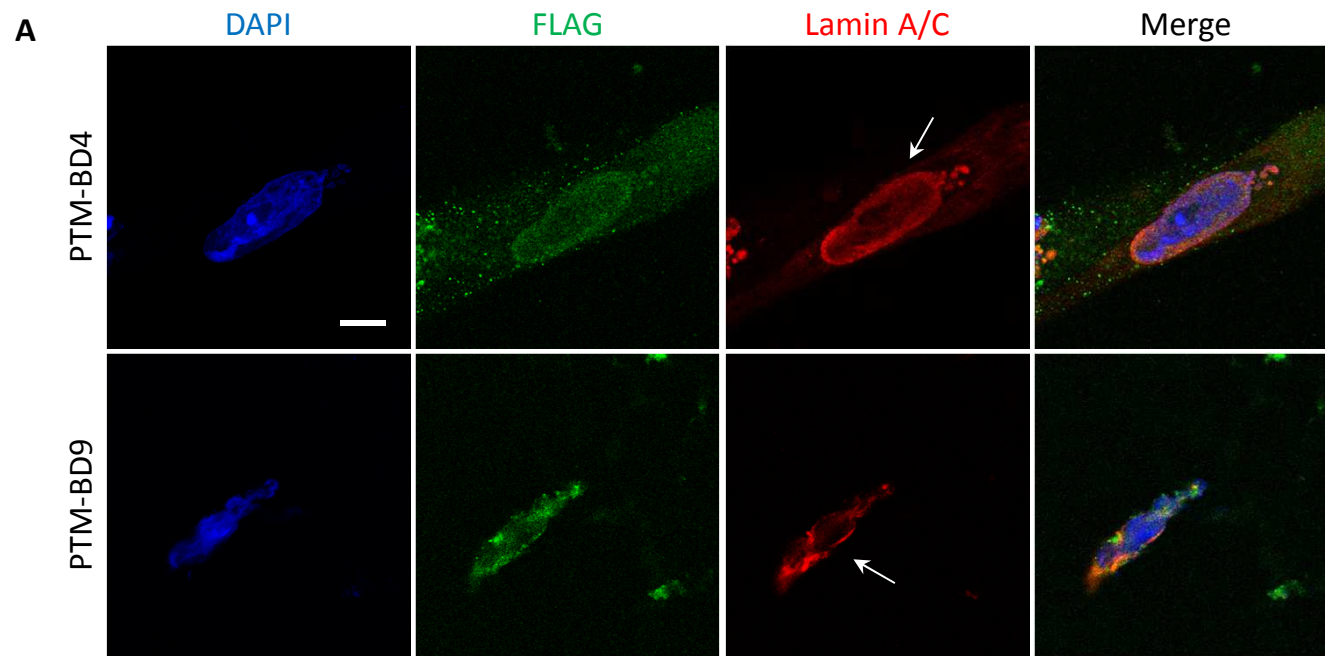


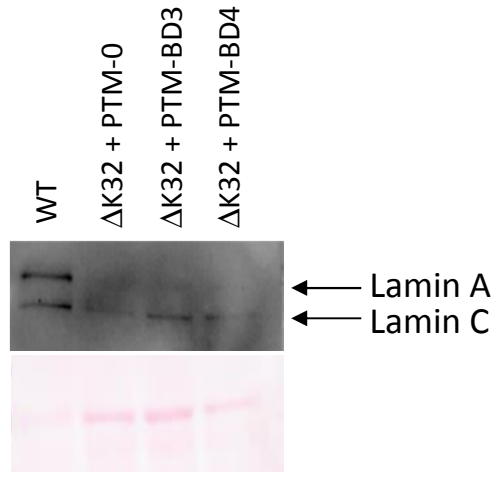
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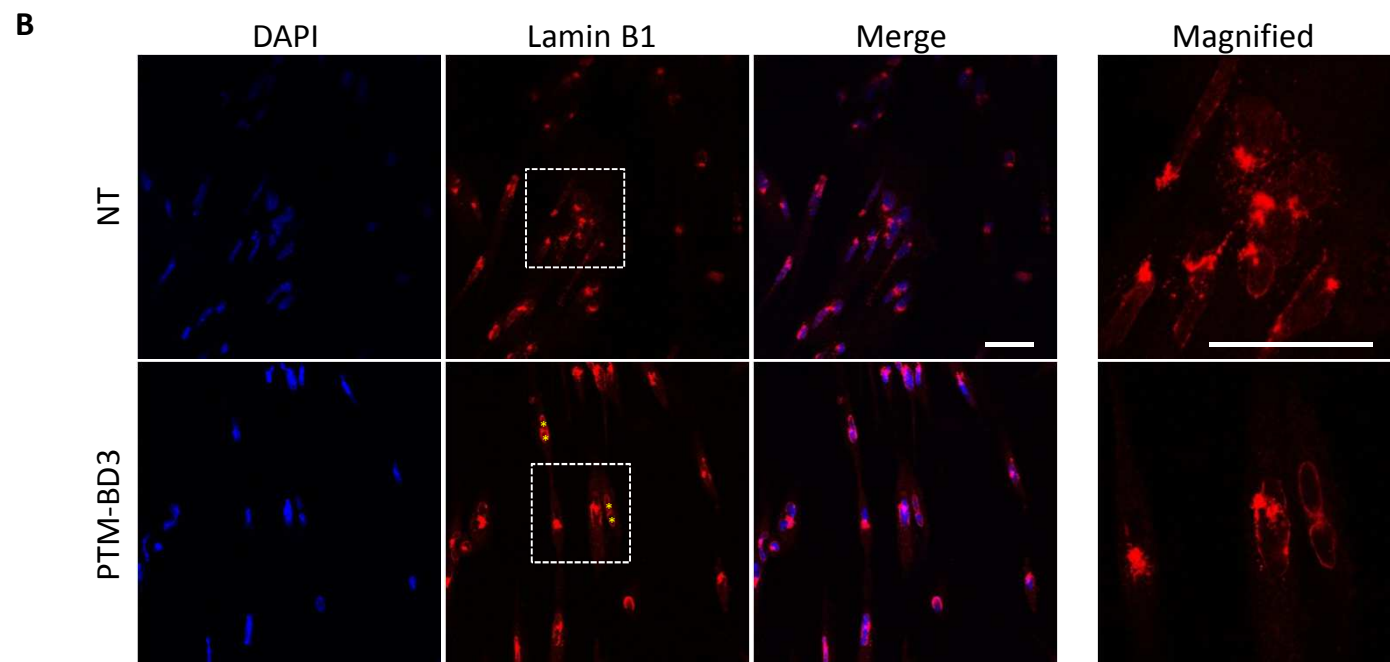
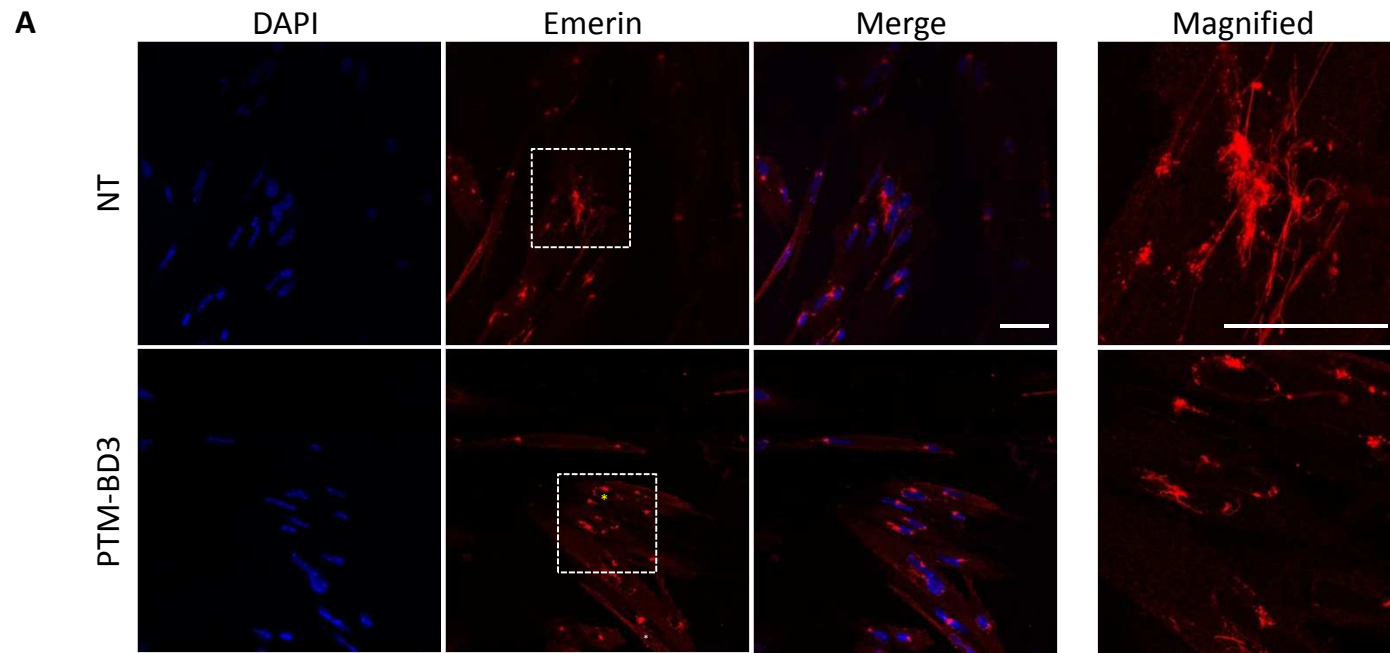


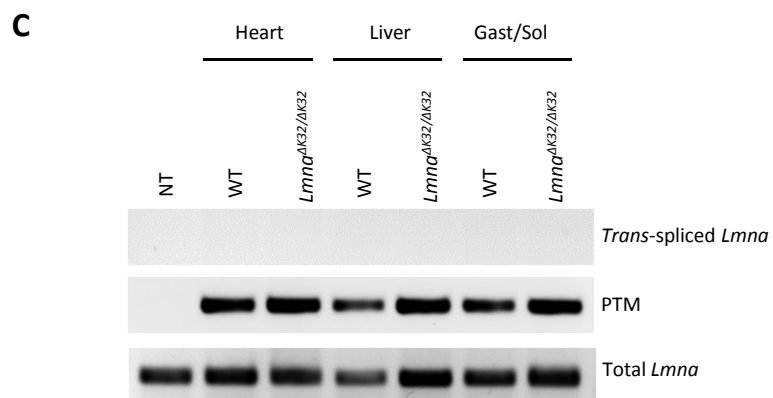
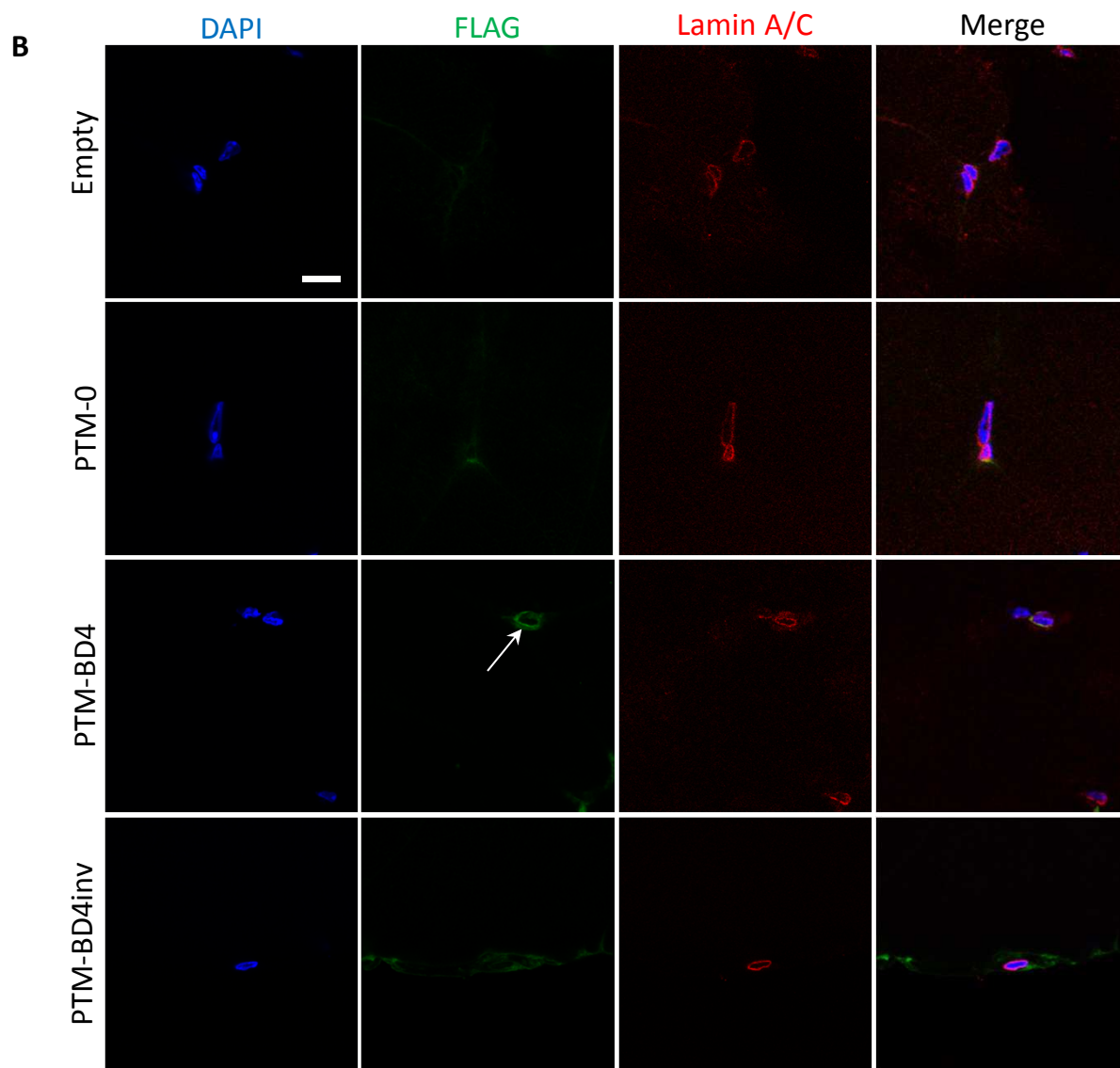
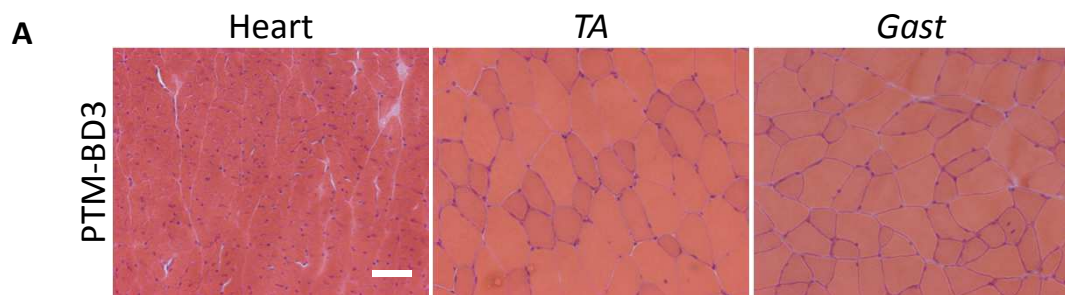
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Supplemental information

Figure S1: Protein evaluation of *trans*-splicing *in vitro*.

(A) Representative western blot analysis for FLAG and lamin A/C expression relative to Ponceau red from C2C12 cells transfected or not (NT) with FLAG-prelaminA, various PTM-BD and PTM-BDinv. No trans-splicing event is detected by western blot. (B) Immunofluorescence analysis of C2C12 transfected with PTM-BD4 and PTM-BD9. Cells were double-stained with anti-FLAG (green) and anti-Lamin A/C (red) antibodies. Nuclei were stained with DAPI (blue). Arrows point on FLAG+ nuclei. (Scale bar: 10 μ m).

Figure S2: Evaluation of the transduction rate in mouse primary myotubes.

Immunofluorescent confocal micrographs of WT mouse primary myotubes transduced with AAV2/9-CMV-GFP and stained for GFP (green) and Desmin (red). Nuclei were stained with DAPI (blue). (Scale bar: 100 μ m).

Figure S3: Detection of repaired *Lmna* mRNA induced by 5' *trans*-splicing in mouse

***Lmna* ^{Δ K32/ Δ K32} primary myotubes.**

Immunofluorescence analysis of *Lmna* ^{Δ K32/ Δ K32} myotubes transduced with AAV2/9-PTM-BD4 and PTM-BD9. Cells were double-stained with anti-FLAG (green) and anti-lamin A/C (red) antibodies (A) or with anti-lamin A/C (green) and anti-emerin (red) antibodies (B). Nuclei were stained with DAPI (blue). Arrows show relocalization of lamin A/C to nuclear envelope. (Scale bar: 10 μ m).

Figure S4: In vitro validation of *Lmna* 5' trans-splicing in primary mouse mutant myotubes.

Western blot analysis for lamin A/C expression relative to Ponceau red from primary mouse WT myotubes and mutant (Δ K32) myotubes transduced either with AAV2/9-PTM-0, -BD3 or -BD4. Transduction with AAV2/9-PTM-BD3 and -BD4 induces a small increase in lamin A/C protein level.

Figure S5: Improve in emerin and lamin B1 localization after *Lmna* 5' trans-splicing in vitro.

Immunofluorescence against emerin (**A**) or lamin B1 (**B**) in non-transduced (NT) or PTM-BD3-transduced primary mutant (Δ K32) myotubes. Nuclei are counterstained with DAPI. Stars highlighted nuclei with corrected emerin or lamin B1 localization. Pictures on the right are a magnification of areas delimited with white dotted squares. (Scale bar: 50 μ m).

Figure S6: Evaluation of trans-splicing efficiency in vivo.

(**A**) Hematoxylin eosin staining of striated muscles sections from WT mice 50 days post-systemic injection with AAV2/9 expressing PTM-BD3. (Scale bar: 50 μ m). *TA*: *Tibialis anterior*; *Gast*: *Gastrocnemius*. (**B**) Immunofluorescence analysis of *Tibialis anterior* sections from WT mice 50 days post-systemic injection with AAV2/9-PTM-0, -PTM-BD4 and PTM-BD4inv. Cells were double-stained with anti-FLAG (green) and anti-Lamin A/C (red) antibodies. Nuclei were stained with DAPI (blue). (Scale bar: 10 μ m). (**C**) PCR analysis was performed with primers specific for trans-spliced *Lmna* (F-FLAG and R-*Lmna* exon 7), PTM (F-FLAG and R-*Lmna* exon 1) or total *Lmna* (F-*Lmna* exon 1 and R-*Lmna* exon 1) on RNA extracts from 15 days-old mouse heart, *Gastrocnemius* and liver after AAV2/9-PTM-BD4.

Supplemental Table 1: Sequence of the different binding domains targeting intron 5 of *Lmna* pre-mRNA.

BD1	CCCTGGAAAGCCAGACAGGCATCAGATTCAGAACCATAGCATGTGGTCCTGGAGTCCCTCCCCACCC CCTTCTCAGTGGGCTTCTGCAGTATCTGCTTAGACCAGAGCCAAGGTGGGAGGGGTAAACTGAGGGTC ACCTG
BD2	CATGTCCAGTCCAAGGGAAGCTCTATCTCCAAGTAGGGAATGCGGCTGGGTTTGTAACTCAGTGAGAA AGCACCTATCTACCAAGCAAGAGGCCTCTGGTTTCAGTCTCCGGCAGGCACAGAAGAAACATGAAAAAC CCTTCATGCACA
BD3	TCAGTGAGAAAGCACCTATCTACCAAGCAAGAGGCCTCTGGTTTCAGTCTCCGGCAGGCACAGAAGAAACATG AAAAACCCCTTCATGCACAGTAAATAGAGGCCTCTGAGGATCAGGGCTAGCTAAAGGAGAGCCCTGGAAAGCC AGACAG
BD4	AGTTAAGGGTGACCTTGAATCTCTGATACTCAACCTGAGATAAAAGGCATGCACCACCCAGTTCCTGTGG TCCTGGGGATAGAAGCCAGGCCATCGTGTATAATAAGCAAGCACTGTACCAACTGAGCTACATGTCCAGTCC AAGGGA
BD5	CTGCCTCCCGAGTGCTGGGATTAAGGCCTGTGCTATTACAACCTGGCTTCTTTTCTTTCTTTTTGAGATA AGGTCTGGTAGTCTAGGCTGGTCTCAAACCTGCTGTATAGTTAAGGGTGACCTTGAATCTCTGATACTTCAA CCTGAG
BD6	TGTTTGTGTTGTGGTATTTTGGTTTGGTTTGGTTTGGTTTTTCAAGACAGGGTTTCTCTGTATAGCCCTGG CTGTCCGGAACTCACTTTGTAGACCAGGCTGGCCTCGAACTCAGAAATCCTCCTGCCTCTGCCTCCCGAGT GCTGGG
BD7	CACTGCAGTGTGCATGACAGACCTGGGAGTCCCGGAGGACAAGAATGTTTCATCATTTTCCACTTTTTTTTTTA AGGATTTTTATTTATGTGTTGTTTGTGTTTGTGTTTGTGGTATTTTGGTTTGGTTTTTGGTTTTTCA AGACAG
BD8	TCTGTCTGAAACCTCTCATTCCGAGGCTAGACCCCATCTTCTTTTGTACAAAATAGTGTAGCTATTGTGAC AAATAATATCGACACAAGTAACATTTCACTGCAGTGTGCATGACAGACCTGGGAGTCCCGGAGGACAAGAA TGTTCA
BD9	AAGGGCTGATGTCGATGAAGAGGGAGGGGCACAGGGGCAGGCAGAGGGACACTCAGGATCTGTAGGCTCTCT CTAGAGCCACCATGGCCTTATCCAGTGACCAGGAAGCCCTTCTAATGTCTGTCTGAAACCTCTCATTCCGA GGCTAG

Supplemental Table 2: PCR primers used to amplify trans-spliced *Lmna* mRNA, total *Lmna* mRNA and PTM mRNA.

Primer	Sequence
F-FLAG	ATGGACTACAAGGACGACGA
F- <i>Lmna</i> ex1	GCCAGCTCTACCCCACTGT
R- <i>Lmna</i> ex1	CAGACTCAGTGATGCGAAGG
F- <i>Lmna</i> ex2	GGGACTTGTTGGCTGCGCA
R- <i>Lmna</i> ex6-1	TCCAGGGCCAGCTTGATGTCCAG
R- <i>Lmna</i> ex9-2	CCATCTCTCGCTCTTTCTCA
R- <i>Lmna</i> ex7	CCGCACGAACTTTCCTCTT