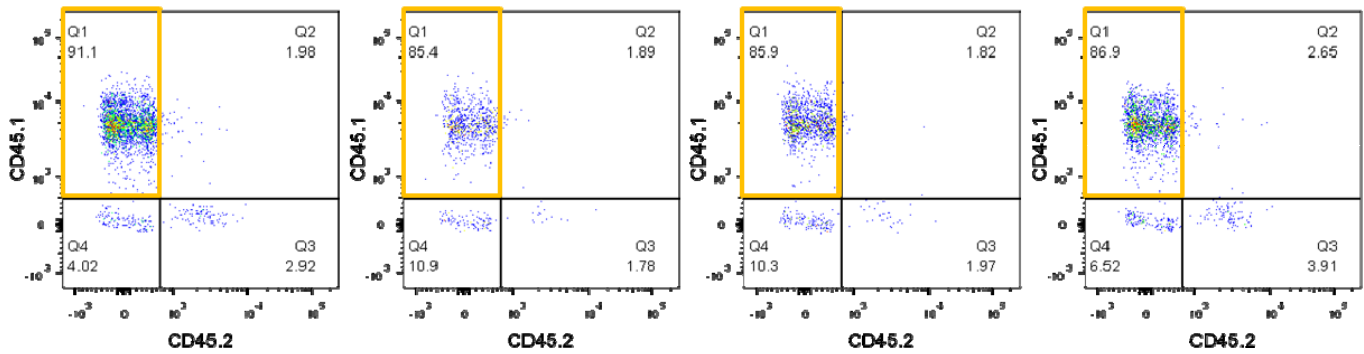


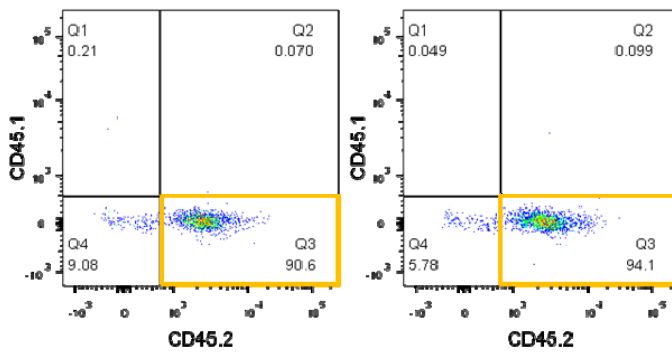
Supplemental Figure 1.

Chimeras:

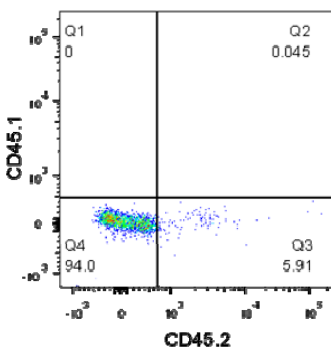
CD45.1 (C57Bl/6, Pepboy) → CD45.2 (C57Bl/6, WT)



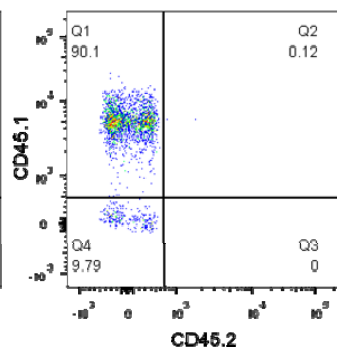
CD45.2 (C57Bl/6, WT) → CD45.2 (C57Bl/6, WT)



FMO: CD45.1

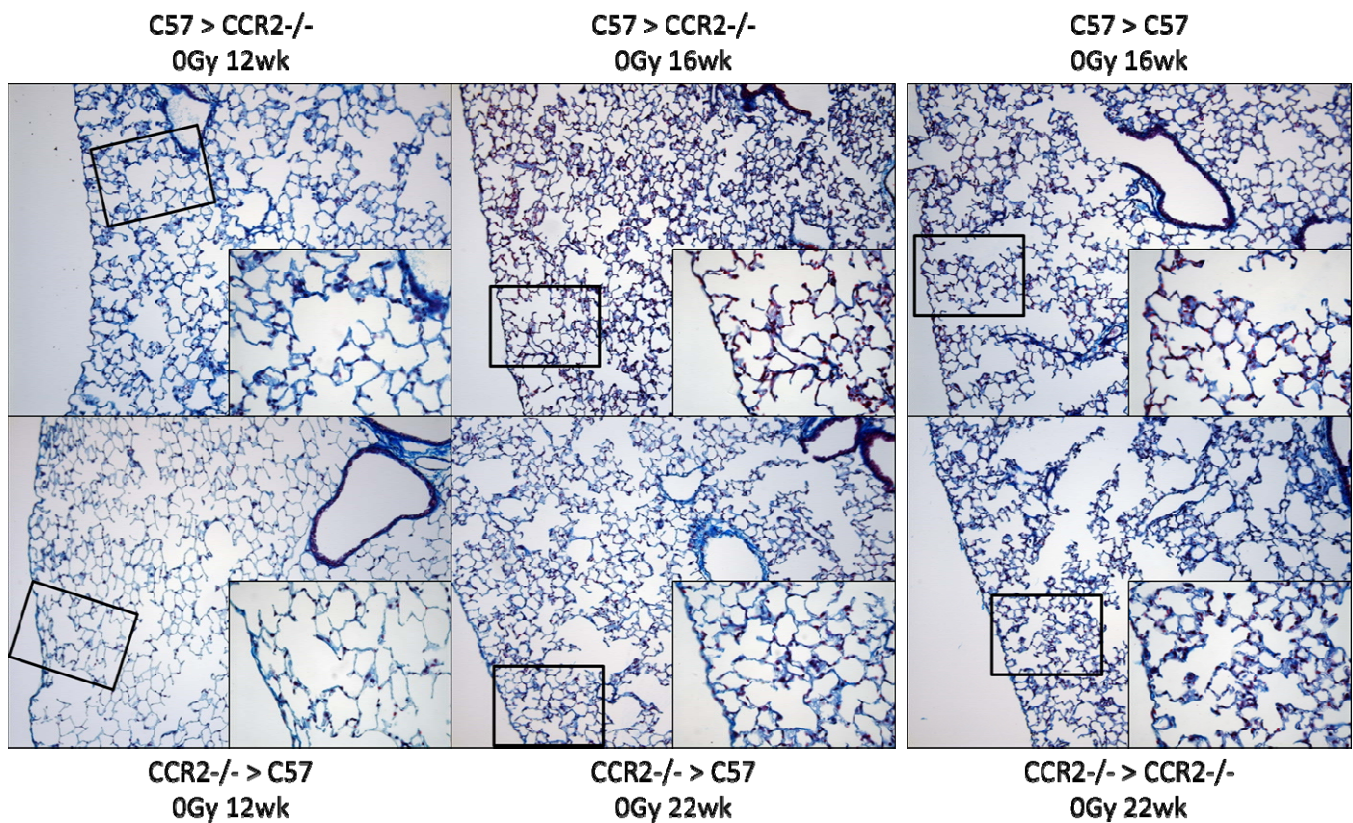


FMO: CD45.2



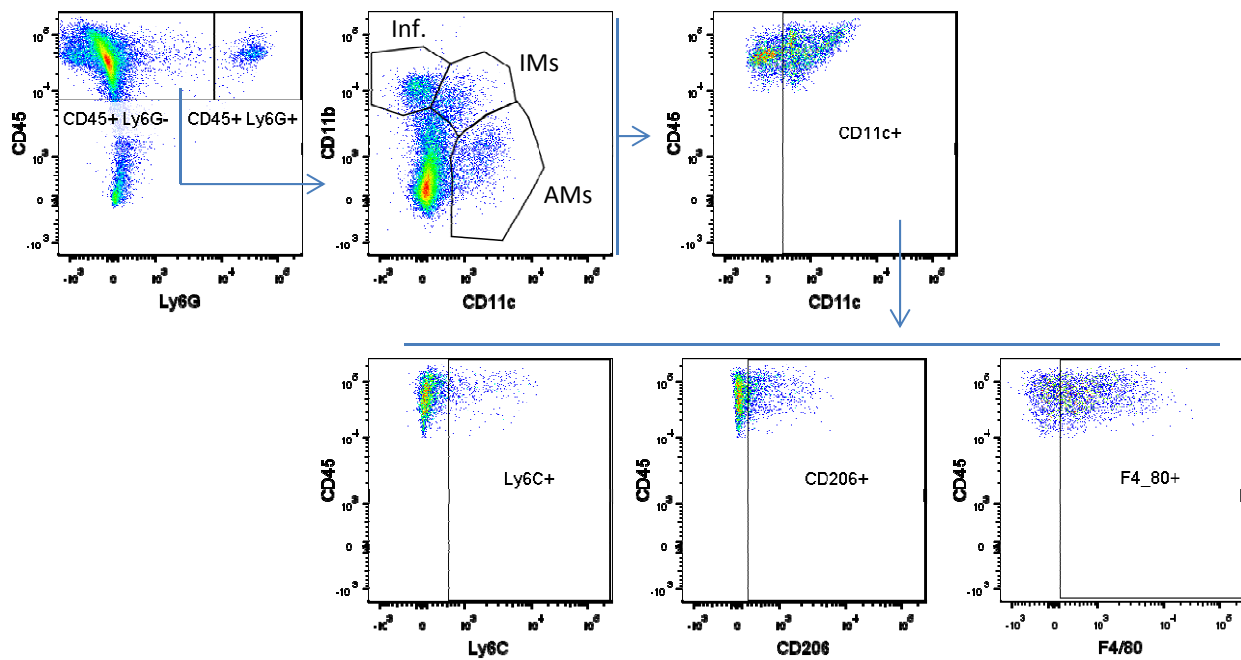
Supplemental Figure 1. Bone marrow engraftment validation in CCR2^{-/-} chimeric mice. Engraftment was verified via flow analysis 8 wk following generation of chimeras by identifying CD45.1 donor derived cells in blood samples collected from CD45.2 recipient mice.

Supplemental Figure 2.



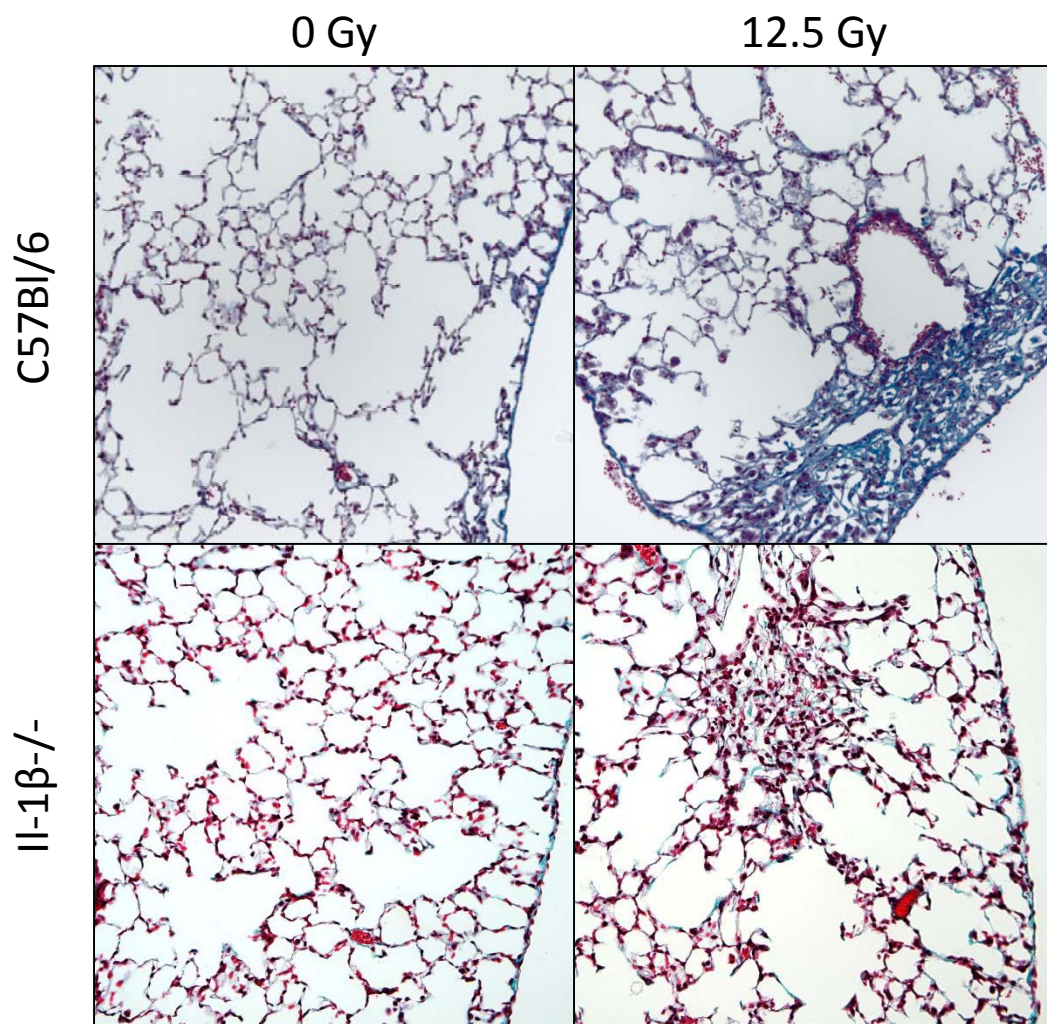
Supplemental Figure 2. Lung histology from CCR2^{-/-} chimeric mice. Gomori Trichrome stained lung sections were prepared between 12 and 22 wk following exposure to 0 or 12.5 Gy thoracic irradiation (n = 3-4 mice/treatment group). Original magnification x200 (inset x400).

Supplemental Figure 3.



Supplemental Figure 3. Gating scheme for characterization of pulmonary macrophage subsets. In CCR2^{-/-} chimeric mice, between 12 and 18 wk following exposure to 0 Gy or 12.5 Gy thoracic irradiation, CD45⁺ cells were enriched from lung digests using MACS and analyzed by flow cytometry. Following doublet and dead cell discrimination, CD45⁺, Ly6G⁻ cells were gated. CD11b and CD11c were used to discern alveolar (AMs; CD11b intermediate, CD11c⁺), interstitial (IMs; CD11b⁺, CD11c⁺), and infiltrating (Inf; CD11b⁺, CD11c⁻) macrophages for analysis of population dynamics. To analyze resident macrophages, CD11c⁺ cells were gated and the percentages of this population expressing Ly6C, CD206 and F4/80 were determined.

Supplemental Figure 4.



Supplemental Figure 4. Lung histology from C57Bl/6 and IL-1 β ^{-/-} mice. Gomori Trichrome stained lung sections were prepared 32 wk following exposure to 0 or 12.5 Gy thoracic irradiation. Original magnification x200.