Intramolecular ¹³C analysis of tree rings provides multiple plant ecophysiology signals covering decades - Supporting Information

Thomas Wieloch, Ina Ehlers, Jun Yu, David Frank, Michael Grabner, Arthur Gessler, Jürgen Schleucher

Site and samples - Additional information

Table S1 summarises spatial and temporal information for all samples included in this study. Sampling sites cover a wide range of ecological conditions. Important ecological features of the *Pinus nigra* sampling site include an "open canopy with low competition among trees", "shallow, very dry soils", and "limited or completely absent forest management disturbances"¹. Dominant trees with umbrella-shaped crowns indicating regular water scarcity were selected for sampling. Two 5-mm cores of each of 19 trees (approximate ages, 92-156 years) were taken at breast height. These samples have been used in several former studies¹⁻⁵, for which their annual rings were carefully dated by standard dendrochronological methods⁶. To preclude growth-related signals in our data, we focused the subsampling on tree rings formed during 1961 to 1995 when all trees had reached a comparably stable canopy position. Thus, isotopic shifts due to the contribution of soil-respired CO₂ or related mechanisms should be absent or negligible.

	Species	Site	Tree rings [years]	Lat. [°]	Long. [°]	Altitude [m AMSL]
	Acer saccharum	Wisconsin, USA	1997-2009	45.7N	89.5W	480
perms	Fagus sylvatica	Austria	2005-2011	47.15N	14.28E	840
	Juglans regia	Kyrgyzstan	1850-1859	41.78N	71.93E	1400
gios	Liquidambar styraciflua	Tennessee, USA	2000-2005	35.9N	84.33W	230
An	Quercus robur	England	1989-1994	52.83N	0.5E	25
	Shorea superba	Borneo, Malaysia	1970-1980	5.02N	117.82E	170
	Juniperus zeravshanica	Uzbekistan	1850-1859	39.62N	68.49E	2200
Gymnosperms	Phyllocladus aspleniifolius	Average of three samples:				
		Tasmania, Australia	2005-2011	43.13S	146.692E	225
		Tasmania, Australia	1980-1989	42.21S	145.43E	250
		Tasmania, Australia	1990-1999	42.21S	145.43E	250
	Picea abies	Austria	1910-1919	47.18N	14.16E	1300
	Pinus nigra	Austria	1961-1995	48.13N	16.23E	350
	Pinus ponderosa	Arizona, USA	1850-1860	32.41N	110.72W	2423
	Thuja plicata	Idaho, USA	1850-1859	47.19N	116.29W	1380

Table S1. Spatial and temporal information for all samples included in this study.

¹³C EA-IRMS and ¹³C NMR Spectroscopy - Additional information

For Elemental Analysis - Isotope Ratio Mass Spectrometry (EA-IRMS) measurements we used a Flash EA 2000 coupled to a DeltaV system (both from Thermo Fisher Scientific Inc., Bremen, Germany). Repeated measurements (n \geq 2) confirmed that the equipment provided high accuracy and precision (long-term standard deviation of instrumental measurements: $\pm 0.15\%$ SD).

For quantitative ¹³C NMR, samples of 1,2-O-isopropylidene- α -D-glucofuranose were dissolved in 160 µl deionised water and D-acetonitrile (CAS: 2206-26-0, filling height=35 mm). Either 5 or 6 mg of the relaxation agent Cr(acac)₃ (CAS: 21679-31-2; 5 mg for > 80 mg samples, 6 mg for 20 to 80 mg samples) were added. Samples of 3,6-anhydro-1,2-O-isopropylidene- α -D-glucofuranose were dissolved in D-acetonitrile (filling height=35 mm), and 8 mg Cr(acac)₃ was added. We determined longitudinal relaxation times (T₁) by inversion recovery experiments and adjusted the recycle delay to allow for complete relaxation (\geq 10 T₁) of all relevant ¹³C nuclei. Quantitative ¹³C NMR spectra were acquired at 298 K using a 90° pulse of 8.07 to 8.7 µs, following published procedures⁷.

By increasing the measurement time with decreasing sample amounts, we achieved signal-tonoise ratios (accumulated over all the spectra) of 900 to 1670 (average: 1270) per sample. Samples <20 mg were excluded from analysis, because too much measurement time would have been required for sufficient precision. All free induction decays were processed by the same operator and protocol: exponential apodization with a line broadening of 1 Hz; automatic phase correction plus subsequent manual phase correction if required; automatic baseline correction of the 10-130 ppm spectral range by a 3rd order polynomial. Lorentzian line shape fitting was applied for signal deconvolution. We calculated average areas under ¹³C NMR signals, S_i, by averaging integrals over all replicate spectra.

Intramolecular ¹³C distributions of tree-ring glucose



Figure S1. Intramolecular ¹³C distributions of tree-ring glucose. Data were acquired for six angiosperm and six gymnosperm species from ecologically different sites with global coverage (Table S1). Solid line, average over all angiosperms and gymnosperms, ±SE. In the biochemical literature, intramolecular ¹³C distributions are commonly expressed as $\Delta \delta^{13}$ C_i, the deviations of positional ¹³C abundances from the molecular average. In contrast, in the ecophysiological literature the Δ scale representing fractionation against ¹³C distributions in the biochemical literature, we here express ¹³C distributions of tree-ring glucose in terms of $\Delta \delta^{13}$ C_i. Consequently, the distributions here and in the main text (Fig. 1) appear vertically mirrored around the molecular average.

Test for autocorrelation in ¹³C time-series

A tree ring formed in a particular year may have had significant input of stored glucose monomers from previous years. If so, ¹³C time-series should exhibit autocorrelation signals. To test this possibility, we correlated each ¹³C time-series with three temporally lagged versions of itself. No significant autocorrelation (r=0.37 to 0.40 for p≤0.05 with n=28 to 25) was detected (Tables S2, S3). Therefore, our analyses does not consider conditions of previous years.

Table S2. Pearson correlation coefficients of $\Delta_i(t)$ versus $\Delta_i(t+\tau)$ and $\Delta(t)$ versus $\Delta(t+\tau)$, with t and $(t+\tau)$ denoting ¹³C time-series without and with temporal lag, respectively.

τ [years]	Δ_1	Δ_2	Δ_3	Δ_4	Δ_5	Δ_6	Δ
-1	0.22	0.29	0.10	-0.11	-0.23	0.25	0.07
-2	0.11	0.22	0.09	0.02	-0.01	-0.10	0.09
-3	-0.04	0.00	-0.30	-0.15	-0.09	-0.11	-0.19

Table S3. Pearson correlation coefficients of $\Delta_i'(t)$ versus $\Delta_i'(t+\tau)$, with t and $(t+\tau)$ denoting ¹³C time-series without and with temporal lag, respectively.

τ [years]	Δ_1 '	Δ_2 '	Δ3'	Δ_4 '	Δ5'	Δ_6 '
-1	0.23	0.33	0.12	-0.10	-0.25	0.28
-2	0.07	0.26	0.07	-0.03	-0.01	-0.24
-3	-0.05	0.10	-0.27	-0.08	0.03	-0.18

Determination of growing season length

We define the growing season as the months with a median number of days ≥ 10 , over the observation period 1961-1995, with an average air temperature ≥ 5 °C. Figure S2 shows the number of days per month with ≥ 5 °C at Hohe Warte (Vienna, Austria, WMO ID: 1103500) during the period 1961-1995 (n=35). According to the applied definition, the growing season at our site extends from March to November.



Figure S2. Number of days per month with average air temperatures ≥5°C during the period 1961-1995 at the Hohe Warte meteorological station (Vienna, Austria, WMO ID: 1103500).

Effects of air vapour pressure deficit on ¹³C discrimination



Figure S3. Effects of growing season air vapour pressure deficit (VPD) on whole-molecule ¹³C discrimination, Δ , and on positional ¹³C discrimination, Δ_i '. Data were acquired for treering glucose of *Pinus nigra* laid down from 1961 to 1995 at a moisture-limited site in the Vienna basin. Bars represent slopes of ordinary least squares regressions, b and b_i' ±SE.

ANCOVA

Growing season air vapour pressure deficit (VPD) is significantly correlated with both Δ and Δ_1 ' (r=-0.72, p=5.4*10⁻⁶ and r=-0.68, p=3*10⁻⁵, respectively, n=31). However, the slope of the Δ ~VPD regression is only half as steep as the slope of the Δ_1 '~VPD regression (Fig. 2, -0.011±0.002SE ‰ Pa⁻¹ and -0.023±0.005SE ‰ Pa⁻¹, respectively). According to ANCOVA, this slope difference is statistically significant (p=0.02, Table S4).

Table S4. Summary of iso~VPD*method ANCOVA modelling output, with iso denoting combined Δ and Δ_1 ' data, VPD denoting growing season air vapour pressure deficit, and method as a categorical variable assigning iso data to either Δ or Δ_1 '. Asterisks denote the significance of model terms (*, p≤0.05; ***, p≤10⁻³). The interaction term vpd:method contributes significantly to the model (p=0.02), showing that responses of Δ and Δ_1 ' to VPD differ significantly.

	Sum of squares	Df	F value	Pr(>F)	Significance
vpd	67.315	1	45.24	8.45E-09	***
method	121.947	1	81.95	1.086E-12	***
vpd:method	8.092	1	5.44	0.0232	*

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