

# **A neuronal basis for fear discrimination in the lateral amygdala**

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## **Supplementary Information**

## Supplementary Note 1

### Figure 4

(e) New Tone (7 kHz, pre-injection) and Tone (7 kHz, post-injection): A 3×2 mixed ANOVA model showed a significant main effect of group ( $F_{(2, 19)} = 7.25, P < 0.01$ ), a significant main effect of trial ( $F_{(1, 19)} = 9.23, P < 0.01$ ) and a significant group×trial interaction ( $F_{(2, 19)} = 20.07, P < 0.001$ ). Simple main effects analysis indicated a significant difference between groups in post-injection trial ( $F_{(2, 19)} = 24.13, P < 0.001$ ) but not in pre-injection trial ( $F_{(2, 19)} = 0.90, P > 0.05$ ). In post-injection trial, freezing levels of Daun02-injected rats were significantly higher than freezing levels in the other two conditions ( $P < 0.001$  in both cases). CS (1 kHz, post-injection): A one-way ANOVA indicated no significant differences between groups ( $F_{(2, 19)} = 0.24, P > 0.05$ ).

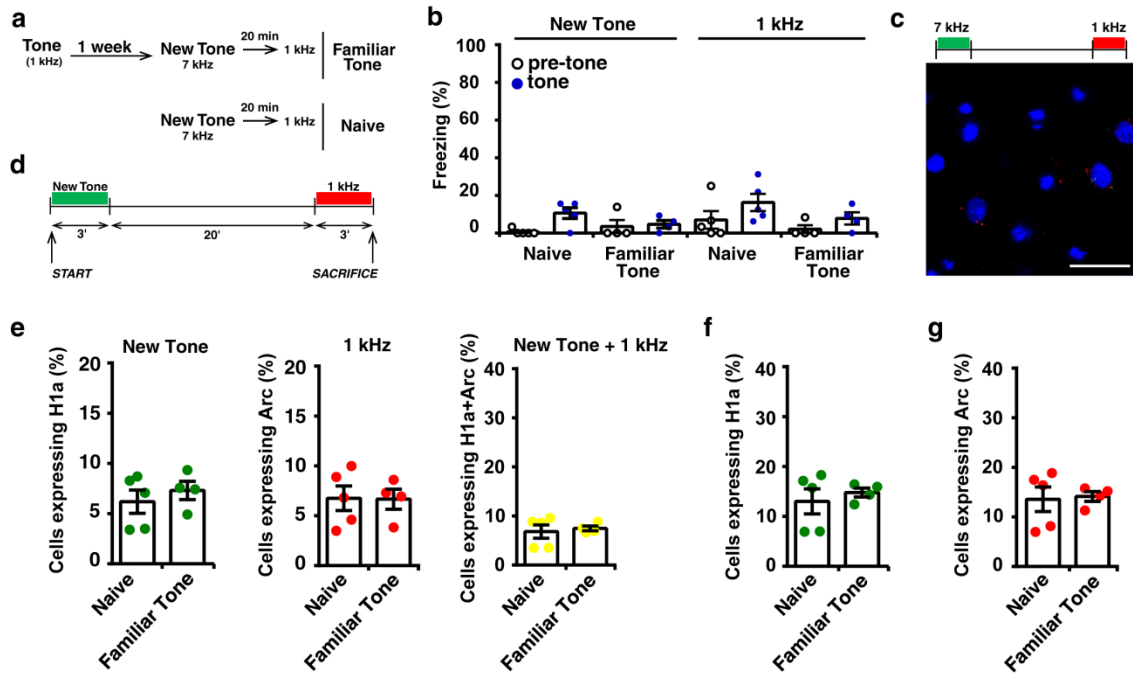
(h) Tones 1-7 (Extinction session - Day 1): A 2×7 mixed ANOVA yielded a not significant main effect of group ( $F_{(1, 10)} = 0.31, P > 0.05$ ) and a not significant group×trial interaction ( $F_{(2, 70, 26, 99)} = 0.21, P > 0.05$ ). The main effect of trial was significant ( $F_{(2, 70, 26, 99)} = 3.13, P < 0.05$ ), indicating a general decrease in freezing response levels. Tones 30-36 (Extinction session - Day 1): A 2×7 mixed ANOVA model showed a not significant main effect of group ( $F_{(1, 10)} = 4.01, P > 0.05$ ), a not significant main effect of trial ( $F_{(6, 60)} = 0.33, P > 0.05$ ) and a not significant group×trial interaction ( $F_{(6, 60)} = 0.21, P > 0.05$ ). Tones 30-36 (Extinction session - Day 2): A 2×7 mixed ANOVA model showed a not significant main effect of group ( $F_{(1, 10)} = 0.004, P > 0.05$ ), a not significant main effect of trial ( $F_{(2, 90, 29, 03)} = 1.30, P > 0.05$ ) and a not significant group×trial interaction ( $F_{(2, 90, 29, 03)} = 1.44, P > 0.05$ ).

(i) CS (1 kHz, pre-injection) and CS (1 kHz, post-injection): A 3×2 mixed ANOVA revealed a significant main effect of group ( $F_{(2, 13)} = 15.36, P < 0.001$ ) and a significant group×trial interaction ( $F_{(2, 13)} = 14.11, P = 0.001$ ). The main effect of trial was not significant ( $F_{(1, 13)} = 0.80, P > 0.05$ ). Simple main effects analysis indicated a significant difference between groups in post-injection trial ( $F_{(2, 13)} = 20.57, P < 0.001$ ) but not in pre-injection trial ( $F_{(2, 13)} = 2.83, P > 0.05$ ). In post-injection trial, freezing levels of Daun02-injected rats were significantly lower than freezing levels in the other two

conditions ( $P < 0.001$  in both cases). New Tone (7 kHz, post-injection): A one-way ANOVA indicated no significant differences between groups ( $F_{(2, 13)} = 0.99, P > 0.05$ ).

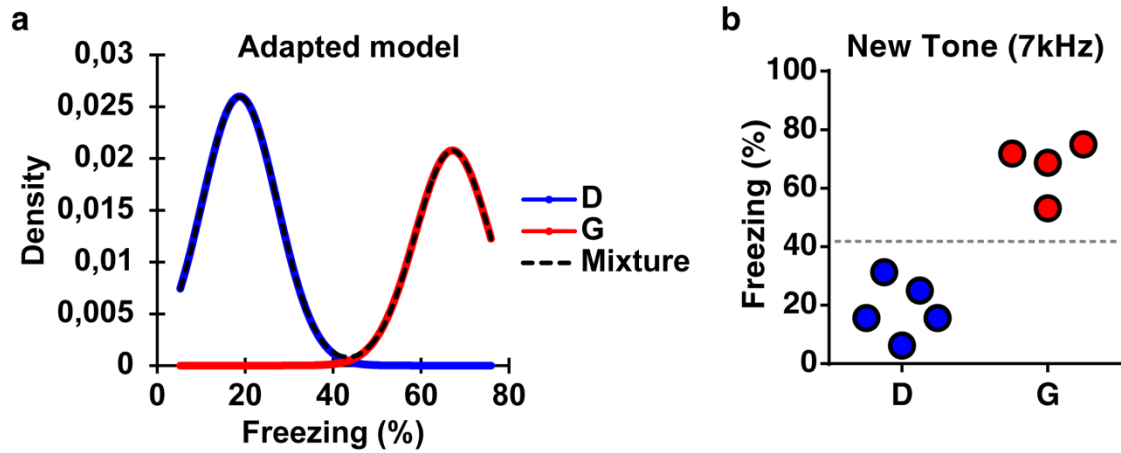
(j) New Tone (15 kHz, pre-injection) and Tone (15 kHz, post-injection): A 2×2 mixed ANOVA model showed a not significant main effect of group ( $F_{(1, 16)} = 0.74, P > 0.05$ ), a not significant main effect of trial ( $F_{(1, 16)} = 0.01, P > 0.05$ ) and a not significant group×trial interaction ( $F_{(1, 16)} = 1.75, P > 0.05$ ).

## Supplementary Figures and Legends

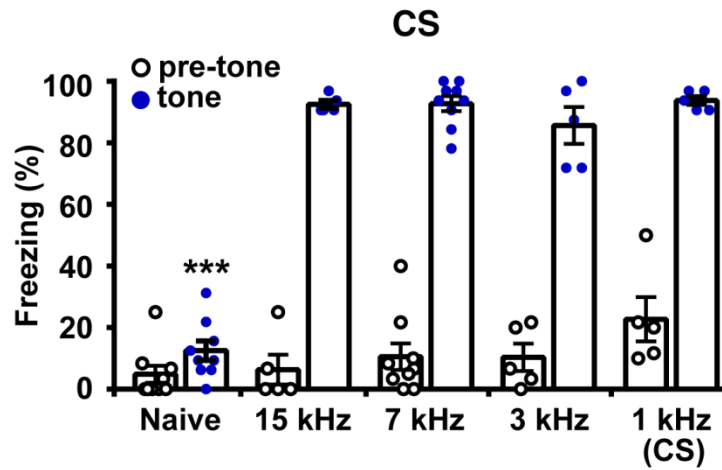


**Supplementary Figure 1. LA activity in naïve and familiar tone groups following the presentation of 7 kHz and 1 kHz tones.** (a) Experimental design of behavioral and catFISH experiments of familiar tone group and totally naïve group. In the familiar tone group, during training rats were presented with the 1 kHz tone unaccompanied by any painful stimulation. One week later, rats were presented with a new tone (7 kHz) followed 20 min later by the 1 kHz tone. Totally naïve animals were presented with the two tones only during the test trial. (b) The percentage of freezing during 7 kHz tone presentation was similar between naïve and familiar tone groups ( $t_{(7)} = 1.59$ ,  $P > 0.05$ ). Similar results were obtained during the presentation of the 1 kHz tone ( $t_{(7)} = 1.43$ ,  $P > 0.05$ ). (c) Representative image showing *H1a*- and *Arc*-expressing neurons in the familiar tone group ( $n = 4$ ). (d) Experimental design of catfish experiment. (e) The percentages of neurons expressing *H1a* ( $t_{(7)} = -0.73$ ,  $P > 0.05$ ), *Arc* ( $t_{(7)} = 0.05$ ,  $P > 0.05$ ) or both ( $t_{(7)} = -0.41$ ,  $P > 0.05$ ) were similar in the naïve and familiar tone groups. (f,

**g)** No differences were detected between groups in the total rate of *H1a* ( $t_{(7)} = -0.6$ ,  $P > 0.05$ ) and in the Arc total rate ( $t_{(7)} = -0.19$ ,  $P > 0.05$ ). All data are mean and SEM.

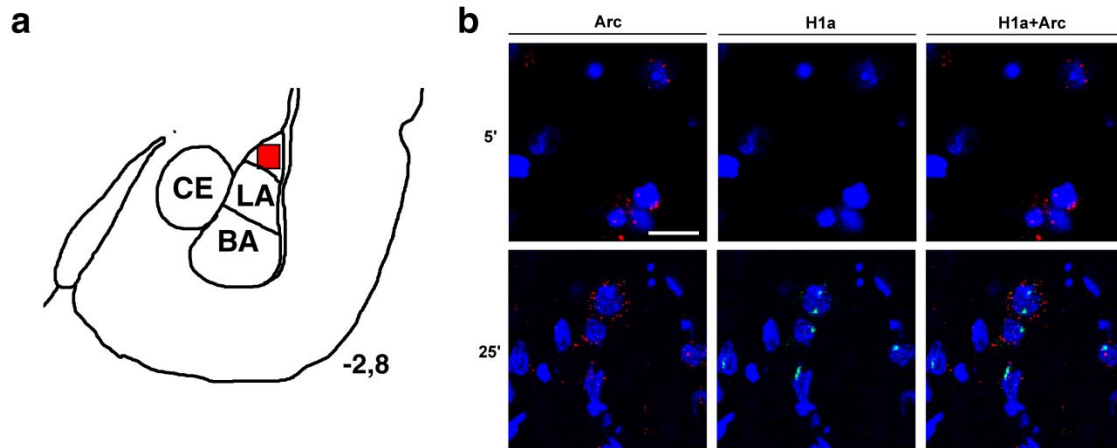


**Supplementary Figure 2. Cluster analysis of behavioral response to the New Tone.** (a) Two-component Gaussian mixture model (GMM) showing the distribution of discriminator (blue) and generalizer (red) animals presented with the New 7 kHz tone. The threshold estimated through the expectation-maximization (EM) algorithm is ~43%. (b) Dot plot showing freezing levels of animals presented with the New 7 kHz tone. Rats are plotted in blue (discriminators) and red (generalizers). Grey dashed line represents the threshold (43%) generated by the EM-GMM algorithm.



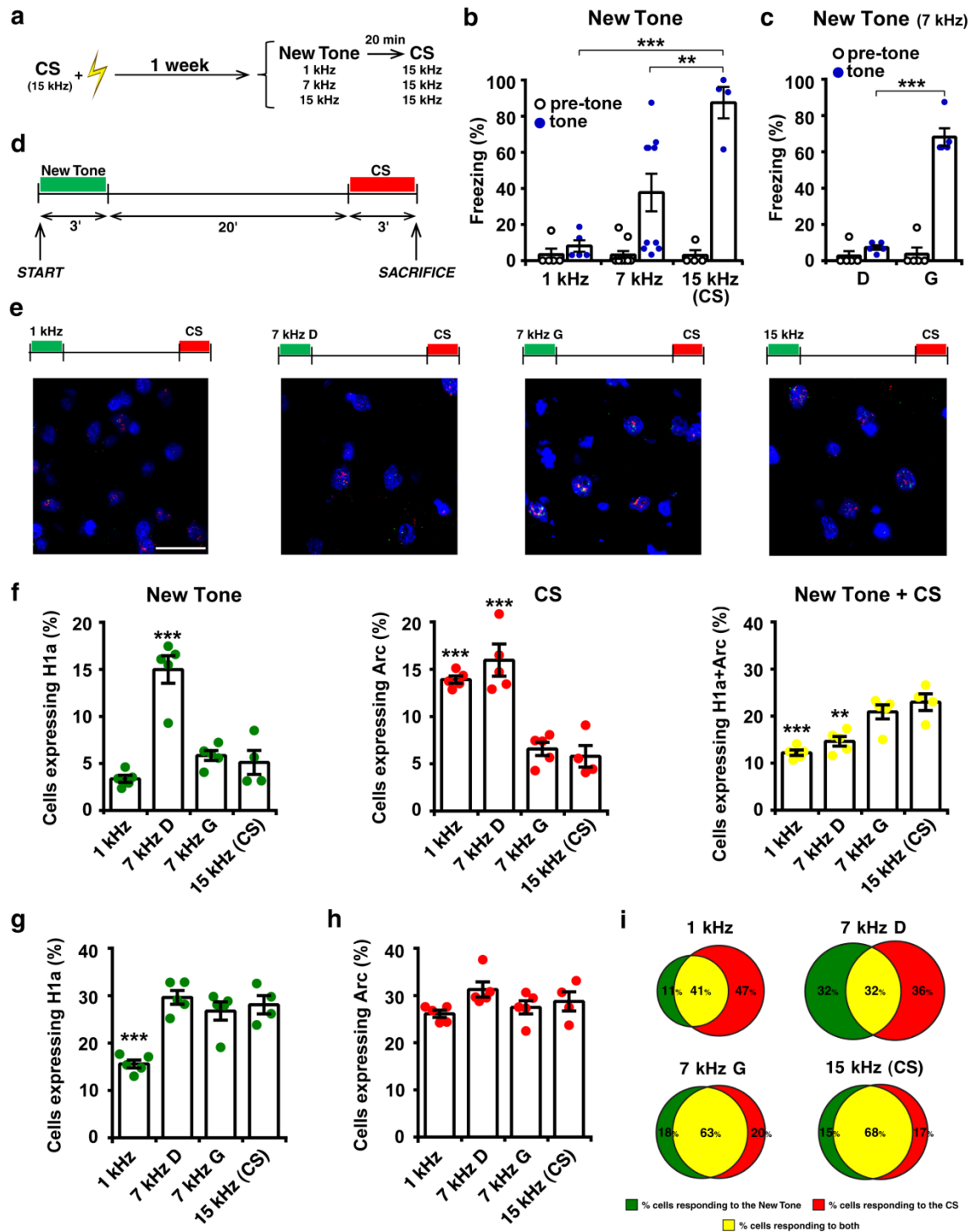
**Supplementary Figure 3. Freezing response to CS in each behavioral group.**

There were significant differences between groups in freezing response to the conditioned stimulus (CS) ( $F_{(4, 28)} = 149.50$ ,  $P < 0.001$ ). *Post-hoc* multiple comparisons revealed that naïve animals were significantly lower than 15 kHz, 7 kHz, 3 kHz and 1 kHz groups ( $P < 0.001$  in each comparison), while there were no differences among all other conditions ( $P > 0.05$  in each comparison). One-way ANOVA with Newman-Keuls *post-hoc* tests. All data are mean and SEM.



**Supplementary Figure 4. Temporal profiles of *Arc* and *H1a* RNA expression detected using catFISH.** a) A catFISH analysis was performed in the lateral nucleus of the amygdala (LA). The section diagram was drawn on the basis of our DAPI-stained sections. b) Time-dependent expression of *Arc* and *H1a* mRNA following CS presentation in the LA. Rats were sacrificed immediately or 22 minutes after the behavioral test. Immediately after the behavioral procedure, *Arc* was expressed in nucleus, whereas *H1a* expression appeared in the nucleus only 25 minutes after the session. At this time point, *Arc* was already detectable in the cytoplasm. Scale bar, 20  $\mu\text{m}$ .

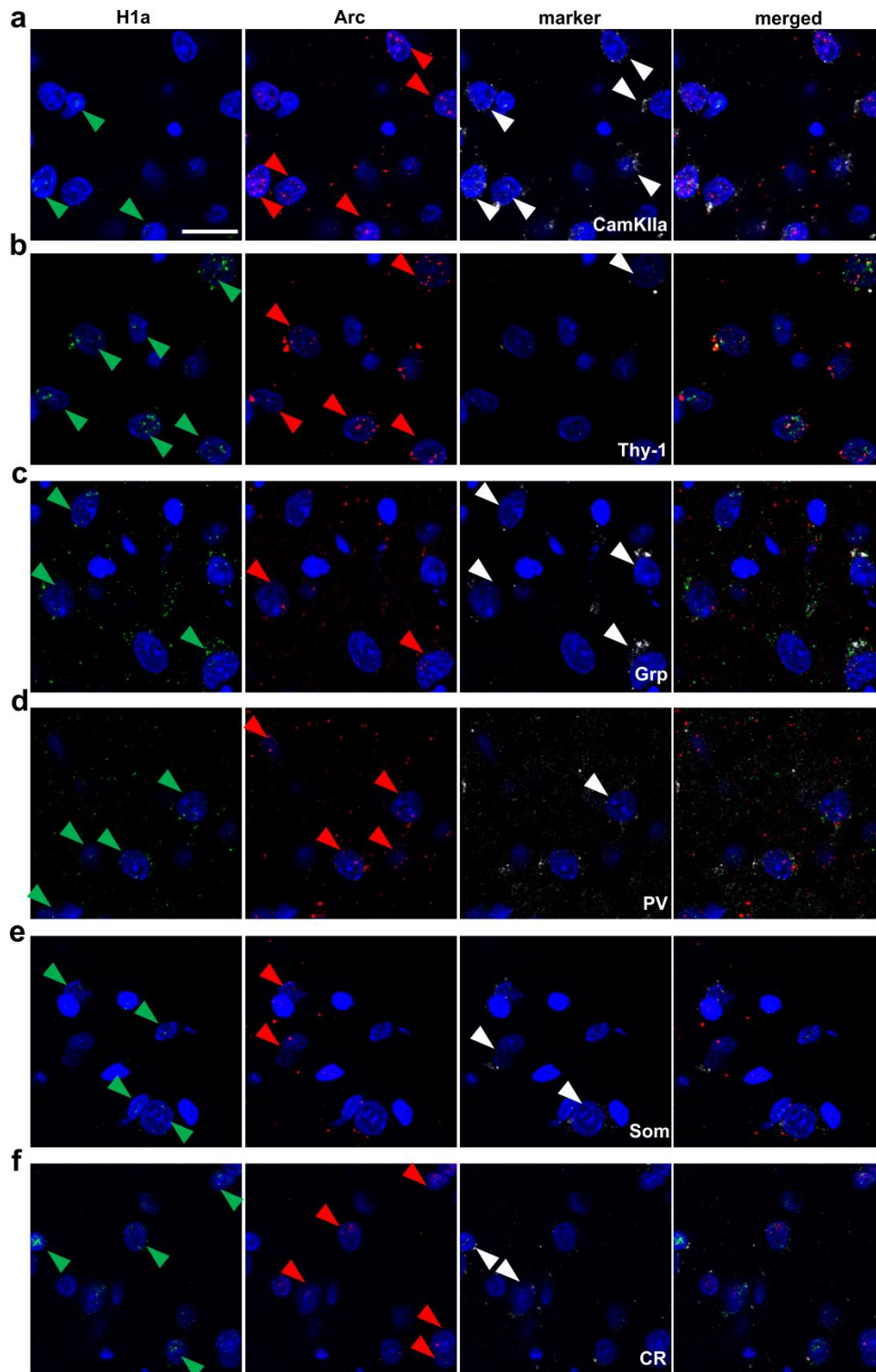




**Supplementary Figure 5. catFISH analysis of LA activity in animals in which the frequency of tones was counterbalanced.** a) Experimental design of behavioral and catFISH experiments. Training was performed by using 15 kHz tone as CS. One week

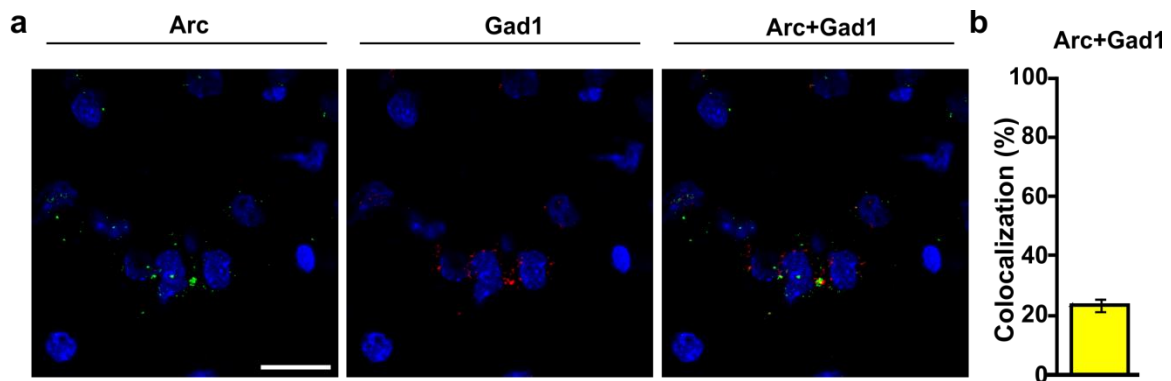
after training, rats were presented with a new tone that was different in each behavioral group (i.e., 1 kHz,  $n = 5$ ; 7 kHz,  $n = 10$  and 1 kHz,  $n = 4$ ). After 20 min, they were exposed to the CS (15 kHz). **(b)** There were significant differences between groups in freezing response during the presentation of tones of different frequencies ( $F_{(2, 16)} = 10.42$ ,  $P < 0.01$ ). Freezing levels of animals exposed to the 15 kHz tone (CS) were significantly higher than 7 kHz ( $P < 0.01$ ) and 1 kHz conditions ( $P < 0.001$ ). **(c)** Freezing in “discriminator” animals (D,  $n = 5$ ) was lower than that observed in “generalizer” (G,  $n = 5$ ) animals during the 7 kHz tone delivery ( $t_{(8)} = 12.07$ ,  $P < 0.001$ ). **(d)** Time course of catFISH experiments. **(e)** Representative images showing neurons expressing single nuclear *H1a* (green arrows) and *Arc* (red) mRNA and double-labeled cells (yellow) in the 1 kHz, 7 kHz (discriminators and generalizers) and 15 kHz groups. Scale bar, 20  $\mu\text{m}$ . **(f)** In the “discriminator” group, dot-plots graphs showed an increase in *H1a*- (new tone) expressing neurons with respect to the other conditions ( $P < 0.001$ ). Both 7 kHz discriminators and 1 kHz groups showed an increase in *Arc*- (CS) expressing neurons with respect to 7 kHz generalizers and 15 kHz (CS) group ( $P < 0.001$ ) and a decrease in double-labeled cells in comparison to 7 kHz generalizers and 15 kHz (CS) group (1 kHz:  $P < 0.001$ ; 7 kHz D:  $P < 0.01$ ). (One-way ANOVA:  $F_{(3, 15)} = 28.63$ ,  $P < 0.001$  (*left*);  $F_{(3, 15)} = 21.59$ ,  $P < 0.001$  (*middle*);  $F_{(3, 15)} = 16.18$ ,  $P < 0.001$  (*right*)). **(g)** The total rate of *H1a* was lower in the 1 kHz group in comparison to other groups ( $F_{(3, 15)} = 17.09$ ,  $P < 0.001$  in each comparison). **(h)** The total rate of *Arc* was high and similar between groups ( $F_{(3, 15)} = 2.35$ ,  $P > 0.05$ ). **(i)** Scaled Venn diagrams showing the percentage of *H1a*- (green), *Arc*- (red) and *H1a*+*Arc*- (yellow) labeled cells in the different experimental conditions. In 7 kHz D group the neuronal populations activated during both new tone or

CS presentation were less overlapped with respect to other groups. \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ . All data are mean and SEM. One-way ANOVA with Newman-Keuls test [(b), (f), (g), (h)]; unpaired  $t$  test (c).

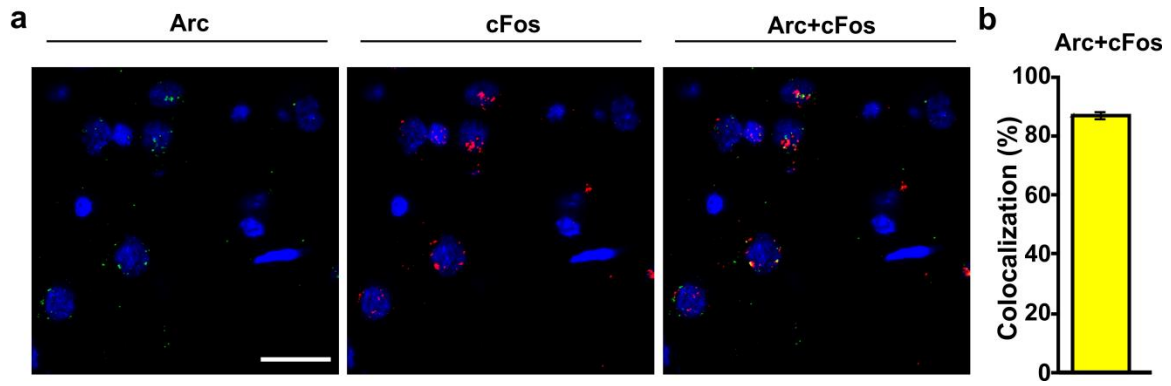


**Supplementary Figure 6. Representative images of triple catfish analysis performed in showing discriminator rats following a new tone or CS presentation within the LA. (a-f) Additional representative photomicrographs of triple catfish**

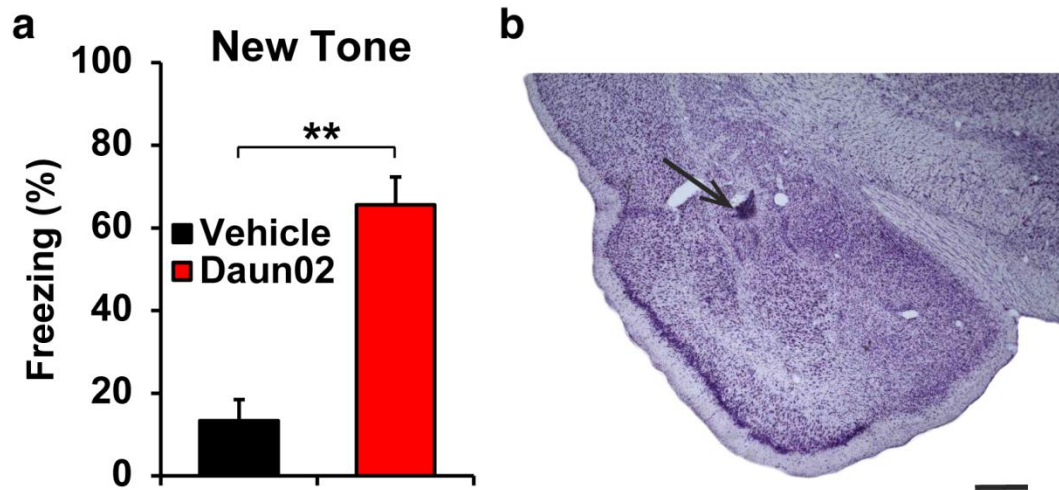
showing neurons expressing *H1a* (green arrows), *Arc* (red arrows) and *CamKIIa* (**a**), *Thy-1* (**b**), *Grp* (**c**), *PV* (**d**), *Som* (**e**) and *CR* (**f**) (white arrows). Each image was acquired in a different field with respect to regions showed in Fig. 3. Scale bar: 20  $\mu\text{m}$ .



**Supplementary Figure 7. In LA, some *Arc*- expressing neurons displayed an inhibitory phenotype. (a)** Representative images showing *Arc* and *Gad1* mRNA expression in LA. Scale bar: 20  $\mu$ m. **(b)** catFISH analysis revealed the expression of *Gad1* in  $23.35 \pm 2.28\%$  of LA neurons expressing *Arc*. All data are mean and SEM.



**Supplementary Figure 8. In LA, the majority of *Arc*- expressing neurons expressed also *cFos* mRNA.** (a) Representative images showing *Arc* and *cFos* mRNA expression in LA. Scale bar: 20  $\mu\text{m}$ . (b) catFISH analysis showed that  $87.23\% \pm 1.25$  of *Arc*-labeled neurons expressed also *cFos* mRNA. All data are mean and SEM.



**Supplementary Figure 9. Validation of Daun02 experiments.** (a) Freezing responses during new tone presentation were significantly higher in Daun02-injected rats was significantly higher than in Vehicle-injected animals ( $t_{(4)} = -6.17$ ,  $P < 0.01$ ). (b) Representative micrographs of Nissl-stained tissues obtained from animals injected with Daun02 within the LA. The arrow indicates the position of the needle track. Scale bar, 500  $\mu\text{m}$ . \*\*  $P < 0.01$ ; unpaired  $t$  test (a). All data are mean and SEM.



<b>Group</b>	<b><i>n</i></b>	<b>Mean Predicted Reactivation Ratio</b>	<b>Mean Observed Reactivation Ratio</b>	<b><i>t</i></b>	<b><i>P</i></b>
15 kHz	5	0.30	0.85	18.77	< 0.001
7 kHz D	5	0.27	0.53	6.07	< 0.01
7 kHz G	4	0.31	0.78	34.59	< 0.001
3 kHz	5	0.37	0.82	38.09	< 0.001
1 kHz	5	0.32	0.85	18.35	< 0.001

**Supplementary Table 1. Comparison between predicted and observed reactivation ratio in each behavioral group.** For each behavioral group the observed reactivation ratio is significantly higher than the reactivation ratio predicted by chance (paired *t* test).