

## Supplementary Information

### AgRP neurons regulate bone mass

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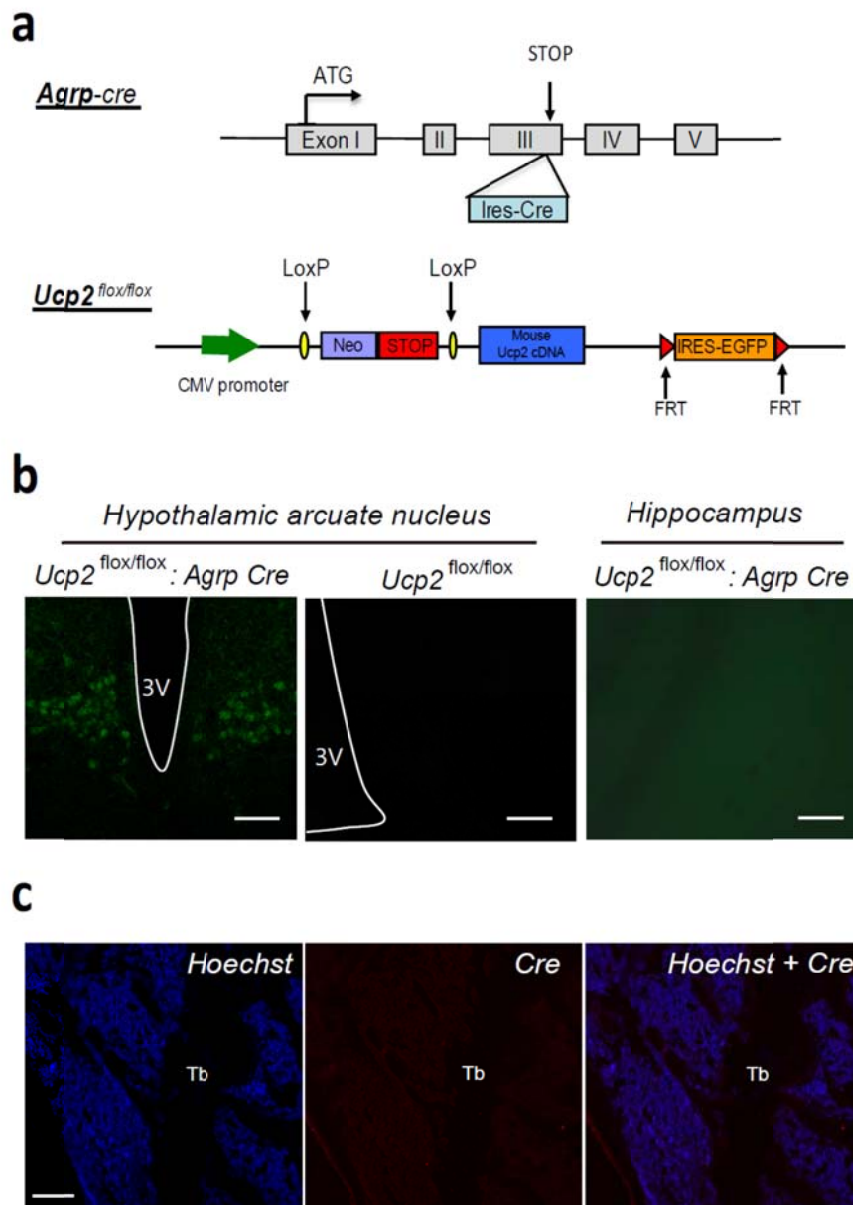
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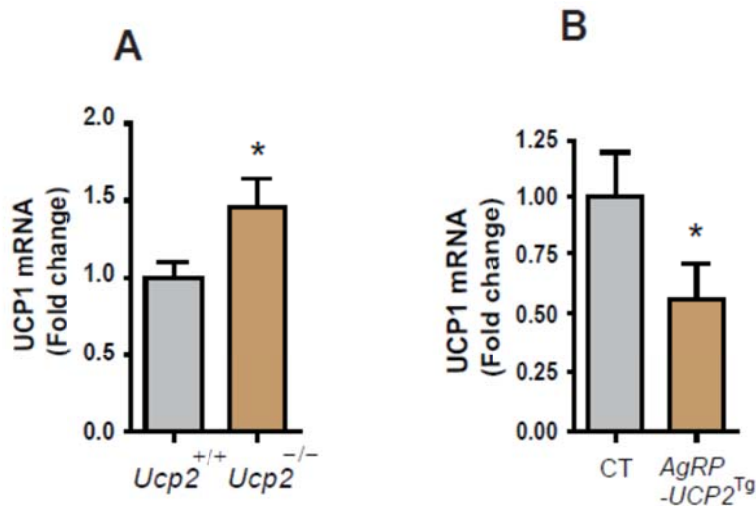
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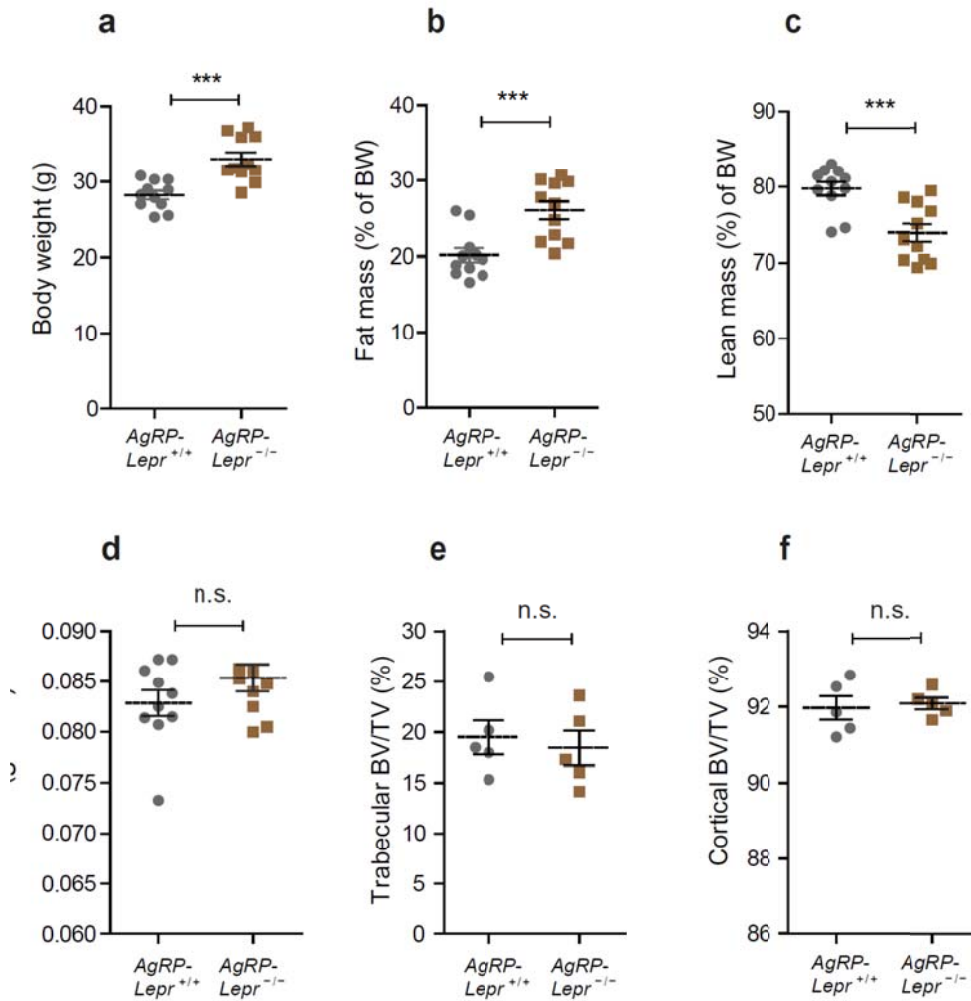
Supplementary Method



**Supplementary Figure 1 (Related to Figure 1). Generation of mice overexpressing *Ucp2* in AgRP neurons. (a) Schematic diagram of the *Agrp cre* allele and the *Ucp2<sup>flox/flox</sup>* transgene. (b) *Agrp-cre* mice were mated with *Ucp2<sup>flox/flox</sup>* mice. Transgenic mice express EGFP protein in the hypothalamic arcuate nucleus following cre-mediated recombination but not in hippocampus. (c) Immunolocalization of Cre protein in the femur of three-month-old *Agrp-cre* mice. Sections were incubated with antibody against *Cre* and counterstained with Hoechst (Blue). Immunoreactive signal of *Cre* (Red) was not observed in the femur of *Agrp Cre* mice. Tb, Trabecular bone. Scale bar = 100  $\mu$ m.**



**Supplementary Figure 2 (Related to Figures 1 and 4). *Ucp1* mRNA expression in brown adipose tissue (BAT) of *Ucp2*<sup>-/-</sup> mice and *AgRP-Ucp2*<sup>Tg</sup> mice.** *Ucp2*<sup>-/-</sup> mice reveals an elevation of *Ucp1* mRNA levels in BAT ( $n=5$  for *Ucp2*<sup>+/+</sup>,  $n=6$  for *Ucp2*<sup>-/-</sup>). *AgRP-Ucp2*<sup>Tg</sup> mice reveals a reduced *Ucp1* mRNA levels in BAT ( $n=6$  for CT,  $n=9$  for *AgRP-Ucp2*<sup>Tg</sup>,  $p<0.05$ ). \*  $p<0.05$ . Data are presented as means  $\pm$  s.e.m. P values for unpaired comparisons were analyzed by Student's *t* test.



**Supplementary Figure 3 (Related to Figures 1, 2 and 3). Impaired leptin signaling in AgRP neurons resulted in increased body weight and fat mass but did not have altered bone phenotypes.** Two-month-old male *Agrp<sup>Lepr<sup>-/-</sup></sup>* mice exhibited increased (a) body weight ( $n=11$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=11$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p<0.001$ ) and (b) fat mass ( $n=11$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=11$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p<0.001$ ) but a decreased (c) lean mass ( $n=11$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=11$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p<0.001$ ). (d) DXA analysis demonstrated that BMD was not altered in *Agrp<sup>Lepr<sup>-/-</sup></sup>* mice ( $n=10$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=10$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p=0.0974$ ). (e, f) Micro-CT analysis showed no differences in trabecular and cortical BV/TV between *Agrp<sup>Lepr<sup>+/+</sup></sup>* and *Agrp<sup>Lepr<sup>-/-</sup></sup>* mice (e:  $n=5$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=5$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p=0.3372$ ; f:  $n=5$  for *Agrp<sup>Lepr<sup>+/+</sup></sup>*,  $n=5$  for *Agrp<sup>Lepr<sup>-/-</sup></sup>*,  $p=0.3787$ ). \*\*\*  $p<0.001$ . Data are presented as means  $\pm$  s.e.m. P values for unpaired comparisons were analyzed by Student's *t* test.

## Supplementary Method

### Immunohistochemical analysis of Cre expression.

Femora were dissected from three-month-old *Agrp Cre* mice. After cleaning, they were fixed overnight in 0.1 M phosphate buffer (PB) containing 4% paraformaldehyde. Decalcification was later performed by immersing bone specimens in 15% wt/vol ethylenediaminetetraacetic acid (EDTA; Sigma) at 25°C for 3 wk. Decalcifying solution was replaced every 3 days. After being embedded in paraffin, bone specimens were cut longitudinally into 7  $\mu$ m sections, which were later incubated at 37°C for 30 min in antigen retrieval solution (0.01 mg/ml proteinase K, 50 mM Tris HCl, pH 8.0, and 5 mM EDTA). To inhibit background endogenous peroxidase activity, sections were incubated for 1h with 10% H<sub>2</sub>O<sub>2</sub>. Nonspecific binding was blocked for 2 h by 4% bovine serum albumin, 10% normal goat serum, and 0.7% Tween-20 in PB buffer. Thereafter, sections were incubated at 4°C overnight with 1:1000 mouse primary antibody against *Cre* (MAB3120, Millipore, Billerica, MA). After being washed with PB buffer, the sections were incubated with avidin-biotinylated HRP-complex (ABC, Vector Laboratories) for 2 h and then reacted with Tyramide Signal Amplification system (NEN Life Science, Boston, MA). After processing, the sections were counterstained with Hoechst and photographed using fluorescence microscopy.