

**ELECTRONIC SUPPLEMENTARY MATERIALS**  
*Proceedings of the Royal Society Biology*

**Olfaction written in bone: Cribriform plate size parallels olfactory receptor gene repertoires in Mammalia**

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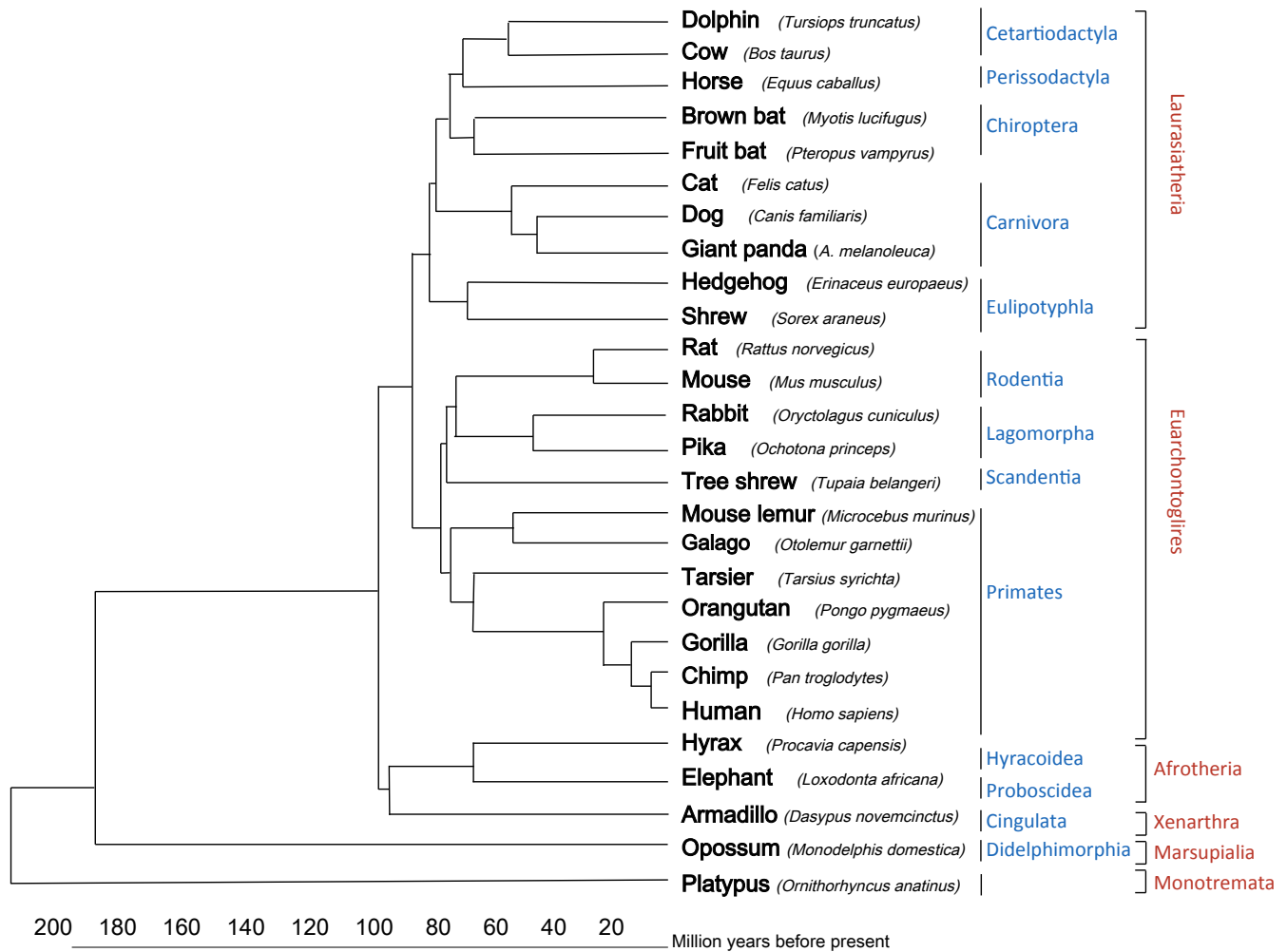
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**Supplementary movie S1. Movie of cribriform plate and nasal anatomy within skull.**

Animated 3D model of an Arctic fox (*Vulpes lagopus*) skull featuring the cribriform plate and general nasal anatomy. The cribriform plate (red) model clearly displays the perforations, or foramina, through which axon bundles from olfactory sensory neurons pass en route to the olfactory bulb of the brain. Sensory neurons in the olfactory epithelium are located primarily on the ethmoturbinal (green) and nasoturbinal (yellow) bones. Maxillary turbinals, which carry respiratory epithelium are shown in blue. The Arctic fox is not among the sample species but its model is used here because of its particular clarity.

**Supplementary figure S2. Time-calibrated phylogeny of sample species.** Tree compiled from published molecular phylogenies [32, 33] (See Main References).



**Supplementary table S3: Key resource table for sample specimens, CT scanning machines and facilities, and digital imaging software.**

Resource	Source	Identifier
<b>Skulls</b>		
<i>Ailuropoda melanoleuca</i>	Institute of Zoology Chinese Academy of Sciences	IZCAS6072
<i>Bos taurus</i>	Museum of Vertebrate Zoology	MVZ114370
<i>Bos taurus</i>	UCLA Dept. Ecology and Evolutionary Biology	UCLAEEB110_1
<i>Canis familiaris</i>	Natural History Museum Los Angeles County	LACM031106
<i>Canis familiaris</i>	Natural History Museum Los Angeles County	LACM022825
<i>Canis familiaris</i>	Natural History Museum Los Angeles County	LACM30539
<i>Canis familiaris</i>	Donald R. Dickey Collection UCLA	UCLA3054
<i>Canis lupus</i>	National Museum of Natural History	USNM291011
<i>Canis lupus</i>	National Museum of Natural History	USNM507338
<i>Dasyypus novemcinctus</i>	Natural History Museum Los Angeles County	LACM46145
<i>Dasyypus novemcinctus</i>	Donald R. Dickey Collection UCLA	UCLA1697
<i>Equus caballus</i>	Museum of Vertebrate Zoology	MVZ223018
<i>Equus caballus</i>	UCLA Dept. Ecology and Evolutionary Biology	UCLAEEB110_2
<i>Erinaceus europaeus</i>	Natural History Museum Los Angeles County	LACM58376
<i>Erinaceus europaeus</i>	Natural History Museum Los Angeles County	LACM58375
<i>Erinaceus europaeus</i>	Museum of Vertebrate Zoology	MVZ183386
<i>Felis catus</i>	UCLA Dept. Ecology and Evolutionary Biology	UCLAEEB110_3
<i>Felis catus</i>	UCLA Dept. Ecology and Evolutionary Biology	UCLAEEB110_4
<i>Gorilla gorilla</i>	Museum of Vertebrate Zoology	MVZ174521
<i>Gorilla gorilla</i>	Donald R. Dickey Collection UCLA	UCLA1788
<i>Gorilla gorilla</i>	Donald R. Dickey Collection UCLA	UCLA2367
<i>Homo sapiens</i>	Museum of Vertebrate Zoology	MVZ106555
<i>Homo sapiens</i>	University of Texas Austin	UTO_HS01
<i>Homo sapiens</i>	UCLA Dept. Ecology and Evolutionary Biology	UCLAEEB110_5
<i>Loxodonta africana</i>	Natural History Museum Los Angeles County	LACM52471
<i>Microcebus murinus</i>	Museum of Vertebrate Zoology	MVZ132535
<i>Microcebus murinus</i>	Duke Lemur Center	DLC7006
<i>Microcebus murinus</i>	Duke University Primate Center	DUPC098
<i>Monodelphis domestica</i>	Museum of Vertebrate Zoology	MVZ144306
<i>Monodelphis domestica</i>	Museum of Vertebrate Zoology	MVZ144307
<i>Mus musculus</i>	Texas Memorial Museum	TMM3196
<i>Myotis lucifugus</i>	American Museum of Natural History	AMNH232239
<i>Myotis lucifugus</i>	American Museum of Natural History	AMNH232244
<i>Ochotona princeps</i>	Donald R. Dickey Collection UCLA	UCLA17230
<i>Ochotona princeps</i>	Museum of Vertebrate Zoology	MVZ42613
<i>Ornithorynchus anatinus</i>	Museum of Vertebrate Zoology	MVZ126977
<i>Ornithorynchus anatinus</i>	Museum of Vertebrate Zoology	MVZ126978
<i>Oryzctolagus cuniculus</i>	Natural History Museum Los Angeles County	LACM7601
<i>Oryzctolagus cuniculus</i>	Natural History Museum Los Angeles County	LACM7603

<i>Otolemur garnettii</i>	Cleveland Museum of Natural History	CMNH B0748
<i>Pan troglodytes</i>	National Museum of Natural History	USNM395820
<i>Pongo pygmaeus</i>	Museum of Vertebrate Zoology	MVZ65532
<i>Pongo pygmaeus</i>	University of Texas Austin	UTO_49859
<i>Procavia capensis</i>	Natural History Museum Los Angeles County	LACM90782
<i>Procavia capensis</i>	Texas Memorial Museum	TMM4351
<i>Pteropus vampyrus</i>	Donald R. Dickey Collection UCLA	LACM91185
<i>Pteropus vampyrus</i>	Donald R. Dickey Collection UCLA	LACM91183
<i>Pteropus vampyrus</i>	Museum of Vertebrate Zoology	MVZ116834
<i>Rattus norvegicus</i>	Donald R. Dickey Collection UCLA	UCLA6994
<i>Rattus norvegicus</i>	Donald R. Dickey Collection UCLA	UCLA9452
<i>Smilodon fatalis</i>	Rancho La Brea Tar Pits	LACMRLP_R37376
<i>Sorex araneus</i>	Museum of Vertebrate Zoology	MVZ179796
<i>Tarsius syrichta</i>	Duke Lemur Center	DLC1406_82
<i>Tupaia belangeri</i>	Natural History Museum Los Angeles County	LACM008157
<i>Tupaia belangeri</i>	Museum of Vertebrate Zoology	MVZ119721
<i>Tursiops truncatus</i>	San Diego Society of Natural History	SDSNH 21212
<i>Tursiops truncatus</i>	Natural History Museum Los Angeles County	LACM84269
<i>Tursiops truncatus</i>	Natural History Museum Los Angeles County	LACM95828

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#### High resolution CT scanners

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GE Phoenix V tome x s	General Electric Applications Technology	<a href="https://gemeasurement.com/">https://gemeasurement.com/</a>
GE Phoenix nanotom s	General Electric Applications Technology	<a href="https://gemeasurement.com/">https://gemeasurement.com/</a>
GE Phoenix nanotom m	General Electric Applications Technology	<a href="https://gemeasurement.com/">https://gemeasurement.com/</a>
North Star Imaging ACTIS	The University of Texas High-Resolution X-ray Computed Tomography Facility	<a href="http://www.ctab.geo.utexas.edu">http://www.ctab.geo.utexas.edu</a>
Nikon Metrology XT H 225 ST	Molecular Imaging Center, Univ. of Southern California	<a href="http://mic.usc.edu/">http://mic.usc.edu/</a>
Xradia microXCT	The University of Texas High-Resolution X-ray Computed Tomography Facility	<a href="http://www.ctab.geo.utexas.edu">http://www.ctab.geo.utexas.edu</a>
Siemens SOMATOM Definition AS64	Ronald Reagan Medical Center UCLA	<a href="https://usa.healthcare.siemens.com">https://usa.healthcare.siemens.com</a>

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#### Imaging software

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Mimics v. 15.0-18.0	Materialise; Leuven, Belgium	<a href="http://www.materialise.com">http://www.materialise.com</a>
3-Matics v. 7.0.1	Materialise; Leuven, Belgium	<a href="http://www.materialise.com">http://www.materialise.com</a>
Rhinoceros v. 4	Robert McNeel and Associates	<a href="https://www.rhino3d.com/">https://www.rhino3d.com/</a>

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#### CONTACT FOR RESOURCE SHARING.

Further information and requests for resources, scanning parameters, CT scan files, and 3D skull models should be directed to and will be fulfilled by the Lead Contact, Deborah Bird ([dbirdseed@gmail.com](mailto:dbirdseed@gmail.com)).

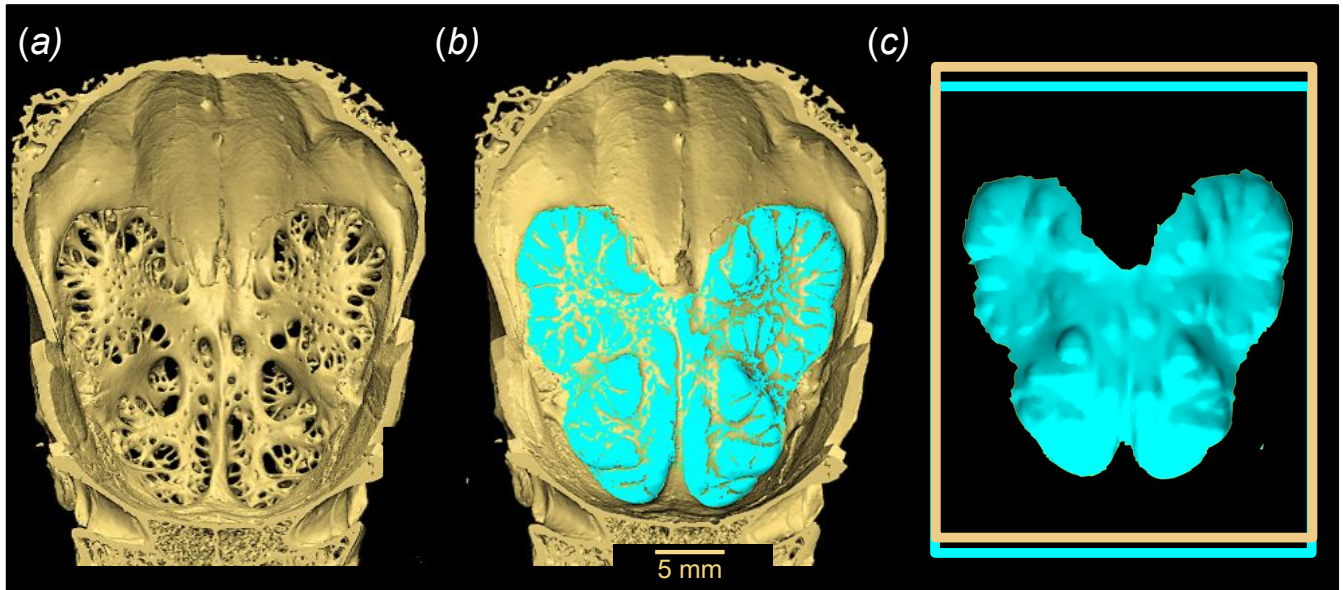
**Supplementary table S4. Cribriform plate surface area data for individual specimens.**

Abbreviations, AMNH: American Museum of Natural History. CMNH: Cleveland Museum of Natural History. DLC: Duke Lemur Center. DUPC: Duke University Primate Center. IZCAS: Institute of Zoology of the Chinese Academy of Sciences. LACM: Natural History Museum of Los Angeles County. LACMLRP: Rancho La Brea Tar Pits. MVZ: Museum of Vertebrate Zoology. SDSNH: San Diego Society of Natural History. UCLA: Dickey Collection at the University of California Los Angeles. UCLAEEB: UCLA Department of Ecology and Evolutionary Biology. USNM: National Museum of Natural History. UTO: University of Texas Austin. TMM: Texas Memorial Museum. \*The presence of a cribriform plate in the ethmoid bone of the bottlenose dolphin (*Tursiops truncatus*) could not be established in this study.

<b>Species</b>	<b>Specimen</b>	<b>Sex</b>	<b>Cribriform plate surface area (mm<sup>2</sup>)</b>
<i>Ailuropoda melanoleuca</i>	IZCAS6072	U	741.58
<i>Bos taurus</i>	MVZ114370	F	2482.09
<i>Bos taurus</i>	UCLAEEB110_1	U	2406.12
<i>Canis familiaris</i>	LACM031106	M	382.21
<i>Canis familiaris</i>	LACM022825	U	601.60
<i>Canis familiaris</i>	LACM30539	M	851.57
<i>Canis familiaris</i>	UCLA3054	U	446.28
<i>Canis lupus</i>	USNM291011	M	1045.53
<i>Canis lupus</i>	USNM507338	F	946.09
<i>Dasypus novemcinctus</i>	LACM46145	M	425.00
<i>Dasypus novemcinctus</i>	UCLA1697	M	456.73
<i>Equus caballus</i>	MVZ223018	M	2321.06
<i>Equus caballus</i>	UCLAEEB110_2	U	2327.43
<i>Erinaceus europeus</i>	LACM58376	M	93.44
<i>Erinaceus europeus</i>	LACM58375	F	81.04
<i>Erinaceus europeus</i>	MVZ183386	M	104.79
<i>Felis catus</i>	UCLAEEB110_3	M	188.86
<i>Felis catus</i>	UCLAEEB110_4	U	134.59
<i>Gorilla gorilla</i>	MVZ174521	F	165.18
<i>Gorilla gorilla</i>	UCLA1788	M	261.12
<i>Gorilla gorilla</i>	UCLA2367	F	135.21
<i>Homo sapiens</i>	MVZ106555	F	208.91
<i>Homo sapiens</i>	UTO_HS01	M	171.03
<i>Homo sapiens</i>	UCLAEEB110_5	F	161.65
<i>Loxodonta africana</i>	LACM52471	F	17922.00
<i>Microcebus murinus</i>	MVZ132535	F	12.82
<i>Microcebus murinus</i>	DLC7006	M	11.02
<i>Microcebus murinus</i>	DUPC098	U	13.18

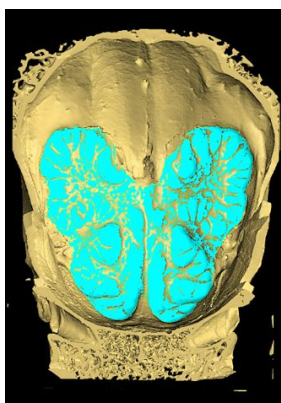
<i>Monodelphis domestica</i>	MVZ144306	F	24.01
<i>Monodelphis domestica</i>	MVZ144307	M	23.03
<i>Mus musculus</i>	TMM3196	M	10.21
<i>Myotis lucifugus</i>	AMNH232239	U	5.83
<i>Myotis lucifugus</i>	AMNH232244	U	4.83
<i>Ochotona princeps</i>	UCLA17230	M	19.29
<i>Ochotona princeps</i>	MVZ42613	M	19.29
<i>Ornithorynchus anatinus</i>	MVZ126977	M	22.42
<i>Ornithorynchus anatinus</i>	MVZ126978	M	24.16
<i>Oryztolagus cuniculus</i>	LACM7601	M	124.79
<i>Oryztolagus cuniculus</i>	LACM7603	M	109.74
<i>Otolemur garnettii</i>	CMNH_B0748	U	39.33
<i>Pan troglodytes</i>	USNM395820	U	125.47
<i>Pongo pygmaeus</i>	MVZ65532	M	129.65
<i>Pongo pygmaeus</i>	UT49859	M	149.27
<i>Procavia capensis</i>	LACM90782	M	77.33
<i>Procavia capensis</i>	UCLATMM4351	M	68.16
<i>Pteropus vampyrus</i>	LACM91185	U	44.92
<i>Pteropus vampyrus</i>	LACM91183	U	47.67
<i>Pteropus vampyrus</i>	MVZ116834	M	74.87
<i>Rattus norvegicus</i>	UCLA6994	F	35.61
<i>Rattus norvegicus</i>	UCLA9452	M	35.86
<i>Smilodon fatalis</i>	LACMRLP_R37376	U	1012.00
<i>Sorex araneus</i>	MVZ179796	F	10.57
<i>Tarsius syrichta</i>	DLC1406_82	F	3.22
<i>Tupaia belangeri</i>	LACM008157	M	57.93
<i>Tupaia belangeri</i>	MVZ119721	M	55.30
<i>Tursiops truncatus</i>	SDSNH 21212	U	NA*
<i>Tursiops truncatus</i>	LACM84269	M	NA*
<i>Tursiops truncatus</i>	LACM95828	U	NA*

**Supplementary figure S5. Digitally quantifying CP surface area.** (a), Three-dimensional CP model from the French bulldog (*Canis familiaris*) is reconstructed from CT scan data in Mimics imaging program. (b), CP foramina are digitally filled to create a generalized continuous surface within the perforate region of the bone. (c), Surface area is isolated and calculated with imaging software 3-matic.

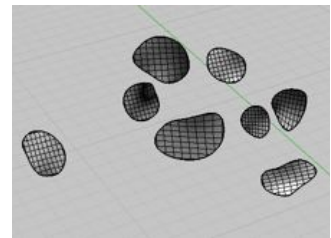
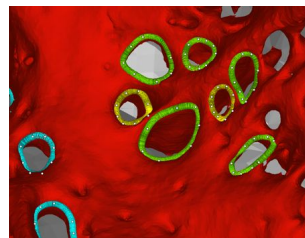
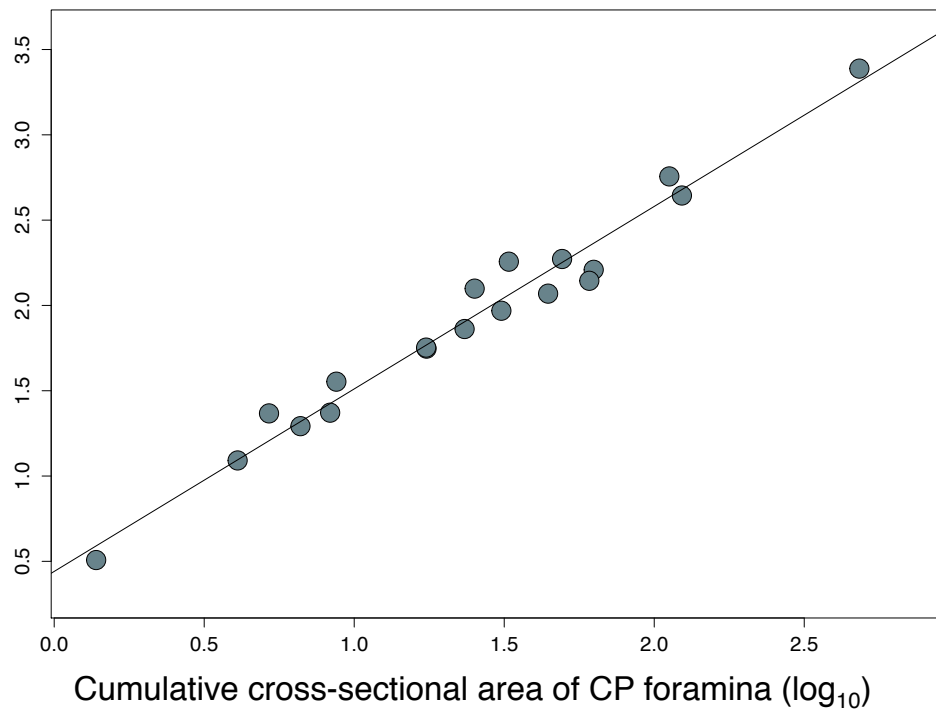




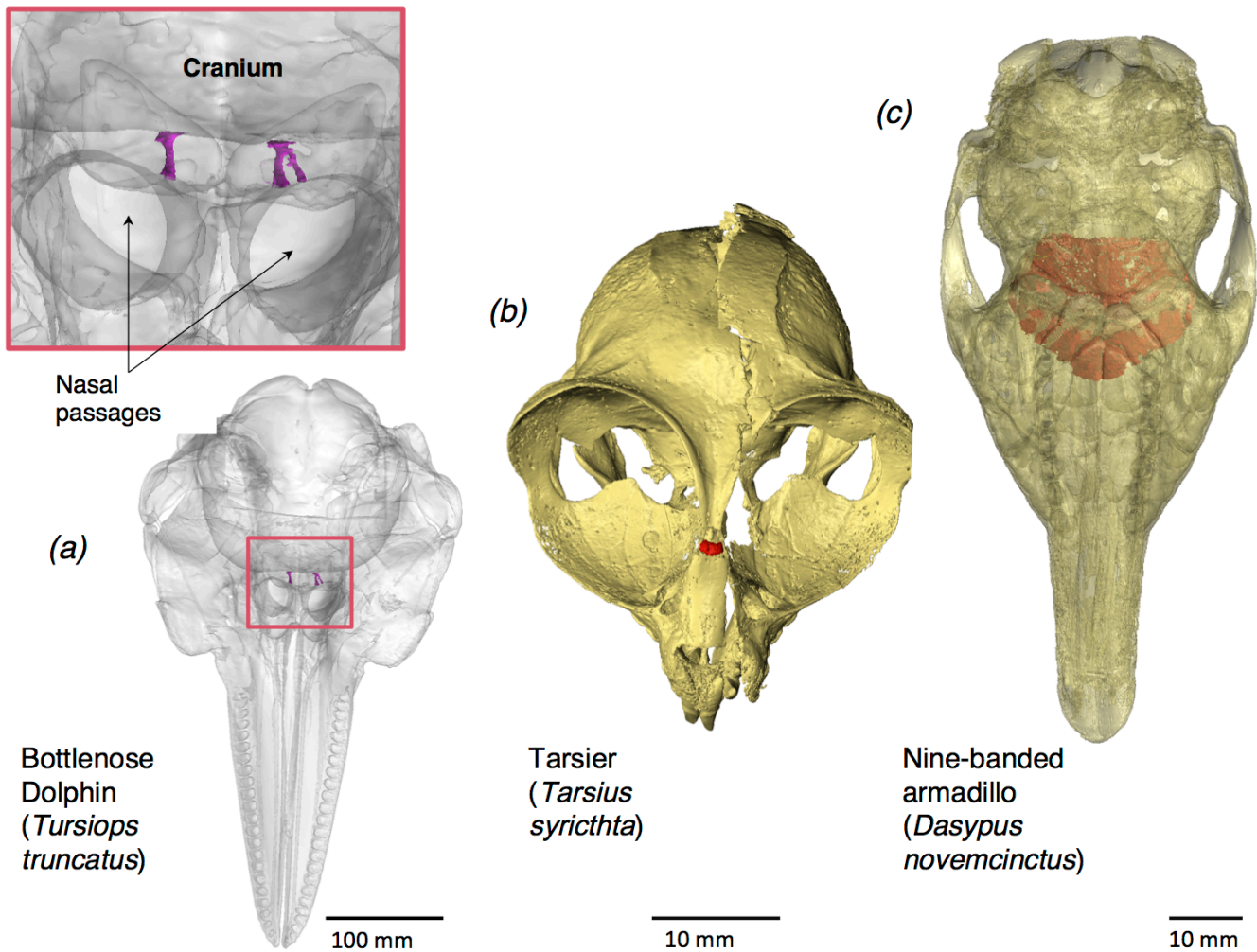
**Supplementary figure S6. Strong linear correlation between cribriform plate (CP) surface area (perforate area) and cumulative cross-sectional area of the CP foramina (log-log) ( $r^2 = 0.91$ ,  $P < 0.0001$ ,  $n = 19$ ).** The more exacting metric of total CP foramina area, representing the imprint of olfactory axon bundles in the ethmoid bone, serves to validate CP surface area as a metric of relative olfactory innervation. CP surface area is the preferable metric for future studies, as it is easily quantifiable, even in skulls with some CP damage, such as fossils. CP surface area was measurable here in 26 sample species, while foramina area was measurable in only 19 species. Image on y axis: taken from figure S5. Images on the x axis: left, section of CP bone (red) with splines, (rings of coordinate points) assigned to the perimeters of several foramina in Mimics; right, imaging software Rhinoceros creates non-planar surfaces from the splines then calculates and tallies all foramina areas.



Absolute cribriform plate (CP) surface area ( $\log_{10}$ )



**Supplementary figure S7: 3D skull models showing marked reduction in CP morphology in taxa heavily reliant on non-olfactory sensory modalities. (a), CP loss viewed in nasal morphology of bottlenose dolphin (*Tursiops truncatus*), which uses echolocation. Pink: Single pair of foramina connecting ethmoid area of the nasal passage to the cranium. Inset, magnification of foramina. (b-c), Impact of orbit size on CP size constraints, contrasting the tarsier (*Tarsius syrichta*) and armadillo (*Dasypus novemcinctus*). (b), Tarsier, a visually-specialized nocturnal primate with enlarged, forward facing orbits, has reduced olfactory anatomy. Red: CP squeezed between convergent orbits. (c), By contrast, the armadillo's large CP (red) fills the space between small, laterally oriented, wide set orbits.**



**Supplementary table S8: Morphological and genomic data for sample species.** Abbreviations: CP, cribriform plate; ORG, olfactory receptor gene. Body mass estimates are species means. Where sexual dimorphism was present, we considered the sex of individuals when deriving the species body mass. Individual dog breed body size estimates were averaged to calculate the mean body mass of the dog (*Canis familiaris*). <sup>a</sup>References for body mass and ORG data listed below the table. References for ORG data are found within the reference list in the main text as well. For cribriform plate surface area data of individual specimens, see table S4.

Species	Common name	n	CP surface area (mm <sup>2</sup> )	Body mass (kg)	Body mass references <sup>a</sup>	Relative CP surface area	Total ORGs	Functional ORGs	Pseudo-genes	Percent pseudo-genes	ORG data references <sup>a</sup>
<i>Ailuropoda melanoleuca</i>	Giant panda	1	741.58	111.600	1	0.016	1235	907	328	0.266	22
<i>Bos taurus</i>	Cow	2	2444.10	453.592	2	0.214	2284	1186	1057	0.463	23
<i>Canis familiaris</i>	Dog: French bulldog, saluki, borzoi, dachshund	4	570.42	21.890	3,4,5,4	0.274	1100	811	278	0.253	23
<i>Canis lupus</i>	Gray wolf	2	995.81	34.875	6	0.410	NA	NA	NA	NA	NA
<i>Dasyus novemcinctus</i>	Nine-banded armadillo	2	440.87	5.647	7	0.472	3146	1146	2000	0.636	24
<i>Equus caballus</i>	Horse	2	2324.24	250.000	4	0.328	2658	1066	1569	0.59	23
<i>Erinaceus europaeus</i>	Hedgehog	3	93.09	1.177	8	0.155	625	295	330	0.528	24
<i>Felis catus</i>	Cat	2	161.72	4.750	9	0.081	1052	677	375	0.356	25
<i>Gorilla gorilla</i>	Gorilla	3	187.17	104.167	10	-0.566	597	263	334	0.559	24
<i>Homo sapiens</i>	Human	3	180.53	62.000	11	-0.463	821	396	425	0.518	26
<i>Loxodonta africana</i>	African elephant	1	17922.00	2766.000	12	0.650	4267	1948	2230	0.523	23
<i>Microcebus murinus</i>	Mouse lemur	3	12.34	0.054	13	-0.018	980	576	404	0.412	24
<i>Monodelphis domestica</i>	Opossum	2	23.52	0.071	4	0.198	1492	1188	294	0.197	27
<i>Mus musculus</i>	Mouse	1	10.21	0.015	4	0.192	1366	1130	236	0.173	23
<i>Myotis lucifugus</i>	Little brown bat	2	5.33	0.010	4	0.015	741	460	281	0.379	22
<i>Ochotona princeps</i>	Pika	2	19.60	0.158	4	-0.062	803	489	314	0.391	24
<i>Ornithorhynchus anatinus</i>	Platypus	2	23.29	1.484	4	-0.500	718	265	370	0.515	27
<i>Oryctolagus cuniculus</i>	Rabbit	2	117.26	1.800	14	0.158	1046	768	256	0.245	23
<i>Otolemur garnettii</i>	Galago	1	39.33	0.760	4	-0.119	803	433	370	0.461	24
<i>Pan troglodytes</i>	Chimpanzee	1	125.47	45.000	4	-0.548	813	380	414	0.509	26
<i>Pongo pygmaeus</i>	Orangutan	2	139.46	75.000	15	-0.619	821	296	488	0.594	26
<i>Procavia capensis</i>	Rock hyrax	2	72.74	4.000	16	-0.231	681	319	362	0.532	24
<i>Pteropus vampyrus</i>	Large fruit bat	3	55.82	1.067	17	-0.045	670	343	327	0.488	24
<i>Rattus norvegicus</i>	Rat	2	35.74	0.222	18	0.121	1767	1207	508	0.287	23
<i>Smilodon fatalis</i>	Sabertooth cat	1	1012.00	230.000	19	-0.014	NA	NA	NA	NA	NA
<i>Sorex araneus</i>	Shrew	2	10.57	0.009	20	0.335	1781	1031	750	0.421	24
<i>Tarsius syrichta</i>	Tarsier	1	3.22	0.117	21	-0.778	345	89	256	0.742	24
<i>Tupaia belangeri</i>	Tree shrew	2	56.61	0.200	4	0.344	2170	979	1191	0.549	24
<i>Tursiops truncatus</i>	Bottlenose dolphin	3	NA*	175.000	4	NA	26	12	14	0.538	24

### References for supplementary table S8:

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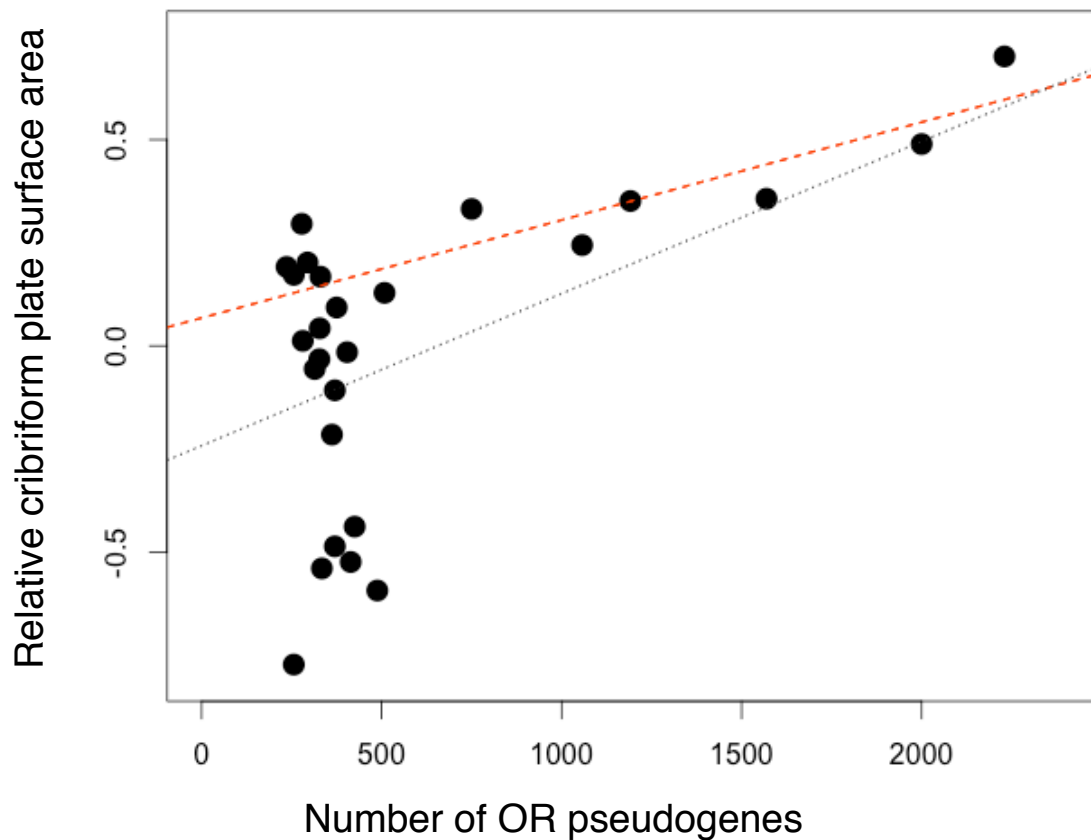
**Supplementary table S9. Assembly statistics for genome assemblies from which olfactory receptor gene counts were extracted.** Abbreviations, BGI: Beijing Genomics Institute; BCM: Baylor College of Medicine; Broad: Broad Institute; ICGSC: International Cat Genome Sequencing Consortium; WU: Washington University. Sources of data: NCBI Assembly, University of California Santa Cruz Genome Browser, Genome Reference Consortium.

Species	Common name	Assembly name	Coverage x	Contig N50	Scaffold N50	Gaps btw scaffolds	Data generation center	Genbank accession number
<i>Ailuropoda melanoleuca</i>	Giant panda	AilMel_1.0	60	39,886	1,281,781	0	BGI	GCA_000004335.1
<i>Bos taurus</i>	Cow	Btau_4.0	7.1	76,397	1,949,504	2,301	BCM	GCF_000003205.2
<i>Canis familiaris</i>	Dog	canFam2	7.6	179,651	45,337,677	86	Broad	GCA_000002285.1
<i>Dasypus novemcinctus</i>	Nine-banded armadillo	Dasnos2.0	2.3	2,428	55,360	0	BCM	GCA_000208655.1
<i>Equus caballus</i>	Horse	EquCab2	6.8	112,381	46,749,900	51	Broad	GCA_000002305.1
<i>Erinaceus europeus</i>	European hedgehog	eriEur1	2	2,753	NA	0	Broad	GCA_000181395.1
<i>Felis catus</i>	Cat	felCat8	14	45,189	18,072,971	303	ICGSC	GCA_000181335.3
<i>Gorilla gorilla</i>	Gorilla	gorGor3.1	2.1	11,661	913,458	7,001	WSI	GCA_000151905.1
<i>Homo sapiens</i>	Human	hg18	10	38,509,590	38,509,590	301	multiple	GCF_000001405.12
<i>Loxodonta africana</i>	African elephant	Loxaf3.0	7	69,023	46,401,353	0	Broad	GCA_000001905.1
<i>Microcebus murinus</i>	Gray mouse lemur	micMur1	1.93	3,511	140,884	0	Broad	GCA_000165445.1
<i>Monodelphis domestica</i>	Gray short-tailed opossum	monDom4	6.5	108,014	59,809,810	209	Broad	GCF_000002295.2
<i>Mus musculus</i>	Mouse	NCBIIm36	7	24,800	16,900,000	NA	multiple	CAAA01000000
<i>Myotis lucifugus</i>	Little brown bat	myoLuc1	1.8	3,100	93,000	NA	Broad	AAPE01000000
<i>Ochotona princeps</i>	American pika	ochPri2	1.9	3,265	88,760	0	Broad	GCA_000164825.1
<i>Ornithorhynchus anatinus</i>	Platypus	ornAna2	6	11,544	991,605	0	WU	GCA_000002275.2
<i>Oryctolagus cuniculus</i>	European rabbit	OryCun2.0	7.48	64,648	35,972,871	83	Broad	GCA_000003625.1
<i>Otolemur garnettii</i>	Northern greater galago	otoGar1	1.87	3,172	NA	NA	Broad	AAQR01000000
<i>Pan troglodytes</i>	Chimpanzee	panTro2	6	30,554	8,803,938	3,108	multiple	GCF_000001515.3
<i>Pongo pygmaeus</i>	Bornean orangutan	ponAbe2.0	6	15,648	747,367	17,860	WU	GCA_000001545.3
<i>Procavia capensis</i>	Rock hyrax	proCap1	2.4	3,379	24,297	0	BCM	GCA_000152225.1
<i>Pteropus vampyrus</i>	Large fruit bat	pteVam1	2.9	8,527	124,060	0	BCM	GCA_000151845.1
<i>Rattus norvegicus</i>	Brown rat	rn4	7	36,847	18,621,810	270	BCM	GCF_000001895.3
<i>Sorex araneus</i>	Common shrew	sorAra1	2	3,175	NA	0	Broad	GCA_000181275.1
<i>Tarsius syrichta</i>	Phillipine tarsier	tarSyr1	2.1	2,876	12,214	0	WU	GCA_000164805.1
<i>Tupaia belangeri</i>	Northern treeshrew	tupBell	2	2,974	NA	0	Broad	GCA_000181375.1
<i>Tursiops truncatus</i>	Bottlenose dolphin	Turtru1.0	2.82	9,719	166,056	0	BCM	GCA_000151865.1

**Supplementary table S10. Summary statistics.** Abbreviations: CP, Cribriform plate; RelCP, Relative cribriform plate size; ORG, Olfactory receptor genes; PGLS, Phylogenetic generalized least squares.

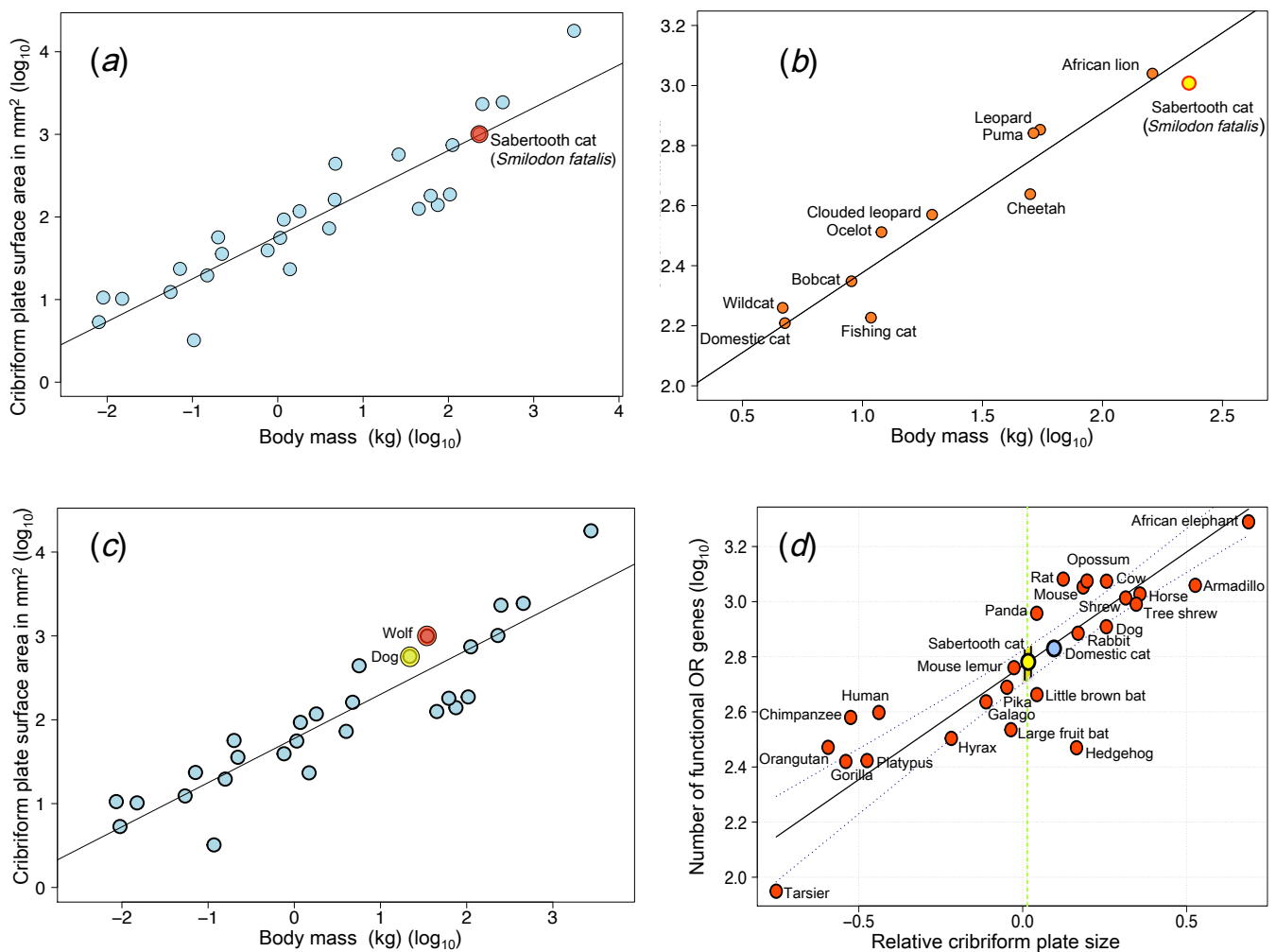
Regression	n	r <sup>2</sup>	P value	PGLS r <sup>2</sup>	P value	Pearson's r	P value
CP surface area (log <sub>10</sub> ) vs body mass (log <sub>10</sub> )	26	0.823	<0.0001	0.875	<0.0001	0.91 (.84, .96)	<0.0001
RelCP vs functional ORG (log <sub>10</sub> )	26	0.763	<0.0001	0.7468	<0.0001	0.87 (.73, .95)	<0.0001
RelCP vs functional ORG (log <sub>10</sub> ) (species with ≥ 6x coverage genomes)	14	0.89	<0.0001	0.815	<0.0001	0.94 (.85, .98)	<0.0001
RelCP vs functional ORG (log <sub>10</sub> ) (ORG count in ≤ 2x coverage species adjusted to predicted count for higher-coverage assemblies )	26	0.841	<0.0001	0.81	<0.0001	0.92 (.81, .97)	<0.0001
RelCP vs total ORG (log <sub>10</sub> )	26	0.68	<0.0001	0.682	<0.0001	0.76 (.63, .86)	<0.0001
RelCP vs total ORG (log <sub>10</sub> ) (ORG count in ≤ 2x coverage species adjusted to predicted count for higher-coverage assemblies)	26	0.71	<0.0001	0.684	<0.0001	0.77 (.64, .86)	<0.0001
RelCP vs number of pseudogenes (all species)	26	0.358	0.001	0.41	0.0004	0.6 (.3, .77)	0.0013
RelCP vs number of pseudogenes (6 species with > ~750 pseudogenes)	6	0.74	0.027	0.723	0.032	0.86 (.31, .99)	0.009
RelCP vs number of pseudogenes (20 species with < ~550 pseudogenes)	20	0.09	0.19	0.009	0.68	-0.31 (-.82, .2)	0.19
RelCP vs number of pseudogenes (7 species with > ~500 pseudogenes)	7	0.825	0.004	0.82	0.005	0.91 (.59, .995)	0.041
RelCP vs number of pseudogenes (19 species with < ~500 pseudogenes)	19	0.225	0.04	0.029	0.485	-0.47 (-.87, .06)	0.0019
RelCP vs percentage of pseudogenes	26	0.114	0.09	0.09	0.135	-0.84 (-.95, -.63)	0.074
Number functional ORG vs pseudogenes	26	0.465	0.0001	0.54	<0.0001	0.68 (0.32, .87)	0.0005
Number functional ORG vs percent pseudogenes	26	0.119	0.084	0.066	0.2	-0.35 (-.75, .14)	0.084
Absolute CP surface area (log <sub>10</sub> ) vs functional ORG (log <sub>10</sub> )	26	0.22	0.015	0.27	0.006	0.47 (-.01, 0.74)	0.013
Absolute CP surface area (log <sub>10</sub> ) vs functional ORG (log <sub>10</sub> ) elephant omitted	25	0.128	0.079	0.16	0.047	0.36 (-.12, 0.66)	0.078

**Supplementary figure S11. Linear regressions in partitioned and non-partitioned data: RelCP vs absolute number of OR pseudogenes.** Among the six species with the largest pseudogene counts, the correlation is  $r^2 = 0.744$ ,  $P = 0.026$ , PGLS- $r^2 = 0.723$ ,  $P = 0.032$  (red dashed line). There is no significant correlation among the 20 species with the lowest pseudogene counts. A significant linear correlation exists across the entire data,  $r^2 = 0.36$ ,  $P = 0.001$ , PGLS- $r^2 = 0.41$ ,  $P = 0.004$  (gray dotted line).





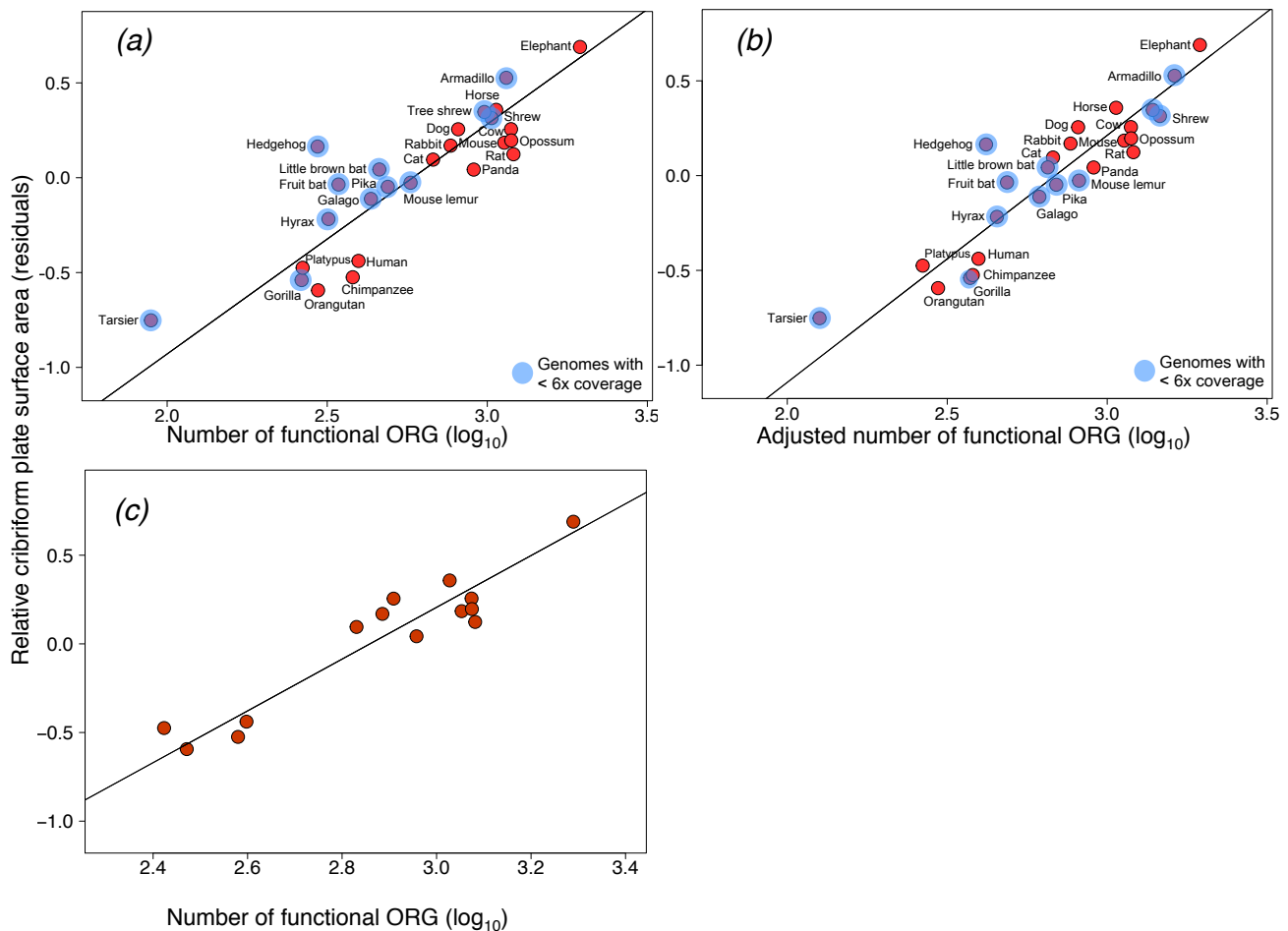
**Supplementary figure S12. Estimating relative CP size (RelCP) to predict the likely position of two non-genome species, the sabertooth cat (*Smilodon fatalis*) and the gray wolf (*Canis lupus*), on our olfactory scale. (a), Log-log plot of absolute CP surface area regressed against body mass for all sample species plus the extinct felid (*Smilodon fatalis*). Residual value for *Smilodon* was used to estimate RelCP for the sabertooth cat. (b), *Smilodon* CP surface area is smaller for its body size than most of the ten living felid species for which CP data is known. (c), In a regression of CP surface area vs. body size among the sample species plus the gray wolf (*Canis lupus*), the wolf has a larger RelCP than the domestic dog (species mean of four dog breeds), predicting a larger ORG repertoire. (d) The identical regression plot in main figure 2d, here with species labels added.**



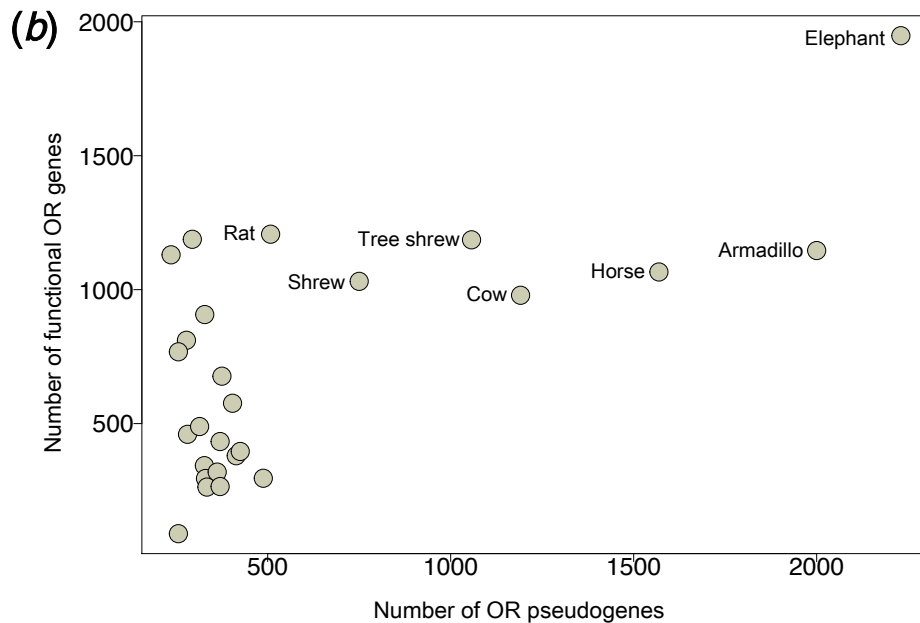
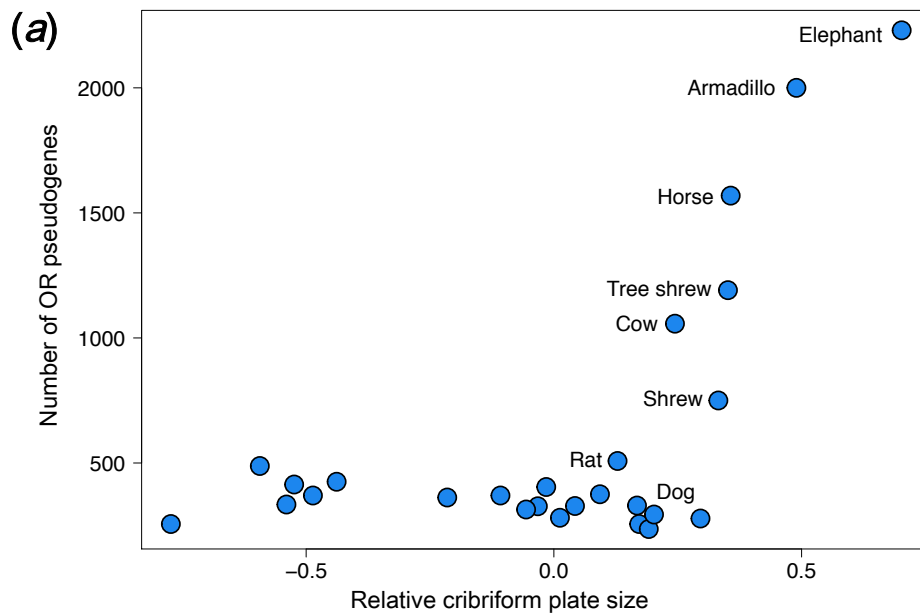
**Supplementary table S13. Sources and morphological data for felid species.** Museums and collections are as follows: FMNH: Field Museum of Natural History; LACM: Museum of Natural History of Los Angeles County; LACMRLP: Rancho La Brea Tar Pits; MMNH: James Ford Bell Museum University of Minnesota, Minneapolis; UCLAEEB: University of California Los Angeles Department of Ecology and Evolutionary Biology. <sup>a</sup>Estimated body masses (in kg) are from [1] except for two species, *Felis catus* [9] and *Smilodon fatalis* [19] (see table S8 references).

Species	Sex	Specimen	Body Mass <sup>a</sup>	Cribriform plate surface area (mm <sup>2</sup> )
<i>Acinonyx jubatus</i>	M	FMNH29635	50	473.07
<i>Acinonyx jubatus</i>	F	FMNH127834	50	396.43
<i>Felis catus</i>	M	UCLAEEB110_4	4.65	188.86
<i>Felis catus</i>	U	UCLAEEB110_3	4.65	134.59
<i>Felis silvestris</i>	M	LACM14480	4.65	206.27
<i>Felis silvestris</i>	F	LACM14474	4.65	157.46
<i>Leopardus pardalis</i>	M	FMNH34339	12	296.37
<i>Leopardus pardalis</i>	F	LACM26789	12	353.50
<i>Lynx rufus</i>	M	UCLA10115	9	225.13
<i>Lynx rufus</i>	F	LACM15254	9	220.71
<i>Neofelis nebulosa</i>	M	LACM31155	19.5	371.34
<i>Neofelis nebulosa</i>	M	USNM282124	19.5	429.04
<i>Panthera leo</i>	M	MMNH17537	161.5	1149.92
<i>Panthera leo</i>	F	MMNH17533	161.5	1041.53
<i>Panthera pardus</i>	M	LACM11704	55	712.55
<i>Prionailurus viverrinus</i>	U	LACM90838	10.85	184.97
<i>Prionailurus viverrinus</i>	F	LACM56718	10.85	152.30
<i>Puma concolor</i>	M	LACM87430	51.6	701.50
<i>Puma concolor</i>	F	LACM85440	51.6	686.83
<i>Smilodon fatalis</i>	U	LACMRLP R37376	230	820.05

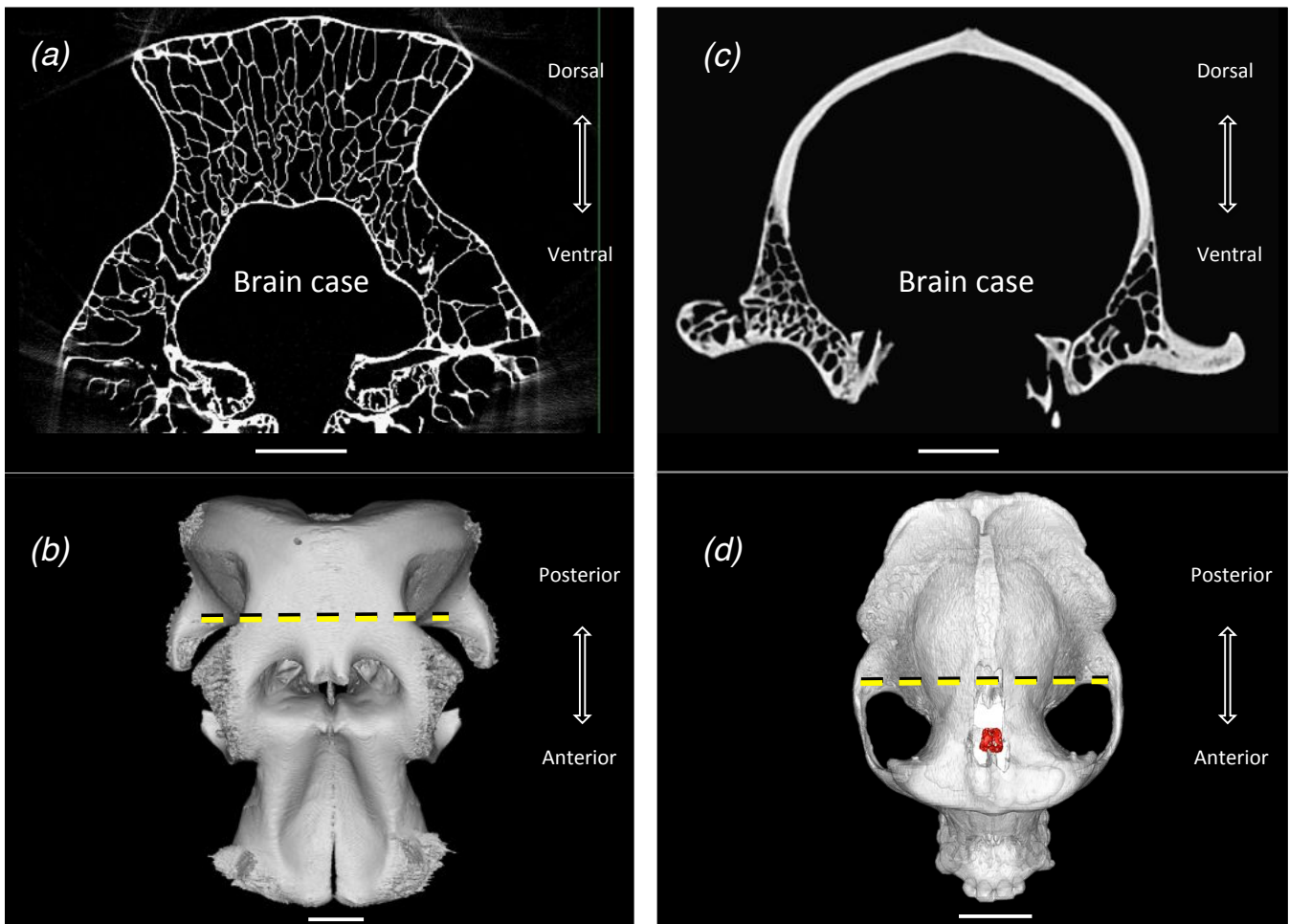
**Supplementary figure S14. Adjusting ORG counts for low coverage genomes, or omitting low coverage species, improves correlation between RelCP and functional ORG number.** (a), Plot from figure 2b; RelCP vs. functional ORGs ( $\log_{10}$ ). Blue circles: species with low coverage ( $\leq 6x$ ) genome assemblies ( $r^2 = 0.76$ ,  $P < 0.0001$ ). (b), Adjusted regression plot. Number of functional ORGs in low-coverage genomes is increased by 39.7%. Percent increase is derived by comparing the number of ORGs extracted from earlier, low-coverage draft genomes to the number from current  $\geq 6x$  coverage assemblies, averaged over four available relevant cases (elephant, dog, cat, rabbit). Adjusted correlation:  $r^2 = 0.84$ ,  $P < 0.0001$ . Any increase in ORG count in low-coverage species strengthens the correlation, as all but two have relatively few ORGs for their RelCP. (c), Omitting species with  $\leq 6x$  genomes strengthens the correlation between RelCP and functional ORGs further ( $r^2 = 0.89$ ,  $P < 0.0001$ ).



**Supplementary figure S15. High number of OR pseudogenes in species with large RelCP and large ORG repertoires.** (a), Inversion of the axes from figure 3a showing that species with largest RelCP tend to have highest numbers of OR pseudogenes. (b), A similar non-linear, dual pattern emerges when the dependent variable in figure 3a is changed from RelCP to number of functional ORG. Species with > ~500 pseudogenes tend to have larger functional ORG repertoires.



**Supplementary figure S16. Skull pneumatization as possible correlate of large CP size; contrast between elephant and gorilla skulls shown in CT scan images and 3D skull models. (a),** Coronal cross-section slice from CT scan of elephant (*Loxodonta africana*) skull showing ~250 mm pneumatized bone surrounding brain. Scale bar: 100 mm. **(b),** Dorsal view of elephant skull model. Yellow line: locale of image in (a). Scale bar: 100 mm. **(c),** Coronal cross-section from CT scan of gorilla (*Gorilla gorilla*) skull; brain surrounded by non-pneumatized cortical bone. Scale bar: 40 mm. **(d),** Dorsal view of gorilla skull model. Yellow line: locale of image in (c). Red: CP.



**Supplementary figure S17. Ethmoturbinal complexity matches large RelCP in African elephant; contrast between turbinal-rich elephant skull and turbinal-poor rock hyrax skull shown in CT scan images and 3D skull models.** Ethmoturbinal bones, which carry the olfactory epithelium, are unusually expansive in the African elephant (*Loxodonta africana*). Quantifying turbinal area is beyond the scope of this paper, but here we visually contrast homologous cross-sections from the nasal chamber at the anterior-most extension of the CP in the elephant and its closest living relative, the rock hyrax (*Procavia capensis*), which has a relatively small CP. **(a)**, Anterolateral view of elephant skull model. **(b)**, Coronal cross-section from CT scan of elephant nasal cavity. **(c)**, Hyrax skull model. **(d)**, Coronal slice from CT scan of hyrax nasal cavity. Orange planes: locales of images (b) and (c). Blue: turbinals. Red: CP. Scale bars, (a): 100mm, (b): 20mm, (c): 5mm.

