

Figure S1. shRNA screen to find epigenetic dependencies in synovial sarcoma. **Related to Figure 1.** (A) shRNA screen strategy. A library against 400 genes encoding chromatin remodelers was screened in triplicate in two cell lines: mouse synovial sarcoma (M5SS1) and mouse myoblasts (C2C12). shRNA representation was evaluated by next generation sequencing three days after transduction of the shRNA library (T_0) and following serial passages at day 16 (T final). (B) Validation of hits in M5SS1 (upper graph) and C2C12 (lower graph) cell lines. Changes in shRNA representation (Screen) and relative % of GFP⁺ (shRNA-expressing) cells, relative to T_0 (Validation) were plotted. shRNAs against SS18 and KDM2B are highlighted. The dashed line corresponds to 2-fold depletion. (C) Bright field images of M5SS1 cells showing changes in morphology upon SS18-SSX knockdown (SS18 shRNAs are against the human gene and therefore do not target the wild-type mouse Ss18) and KDM2B knockdown. GFP-low colonies that escape shRNA expression do not exhibit the same morphological changes. Scale bar=50 µm. (**D**) Western blot analysis of KDM2B in M5SS1 protein lysates (KDM2B shRNAs 433, 2848) and 3145). (E) Rescue experiments showing comparison of knockdown of SS18-SSX or of KDM2B in M5SS1 cells transduced with an empty vector control (MSCV) or a KDM2B cDNA (mouse long isoform). Percentage of GFP⁺ cells at day 16 relative to T0 is plotted.



Figure S2. Human synovial sarcoma cells depend on KDM2B. Related to Figure 2 and 3. (A) Quantitative RT-PCR in IMR90 human fibroblasts, human macrophages (hMac) (black), human synovial sarcoma cells positive for SS18-SSX2 (red), or SS18-SSX1 (blue) gene fusions and other cancer cells lines (gray). not detected (nd). Error bars correspond to mean + s.d. of technical replicates (n=3). (B) c-MYC, KRAS and KDM2B CRISPR/Cas9 screen data from project Achilles in 33 solid cancer cell lines (Pancreas. Lung, Colon, Ovary and Bone). Color shading corresponds to sgRNAs scoring levels as described in https://portals.broadinstitute.org/achilles with an increased green intensity indicating greater depletion. (C) Bright field images of HS-SY-II transduced with the indicated shRNAs 8 days following shRNA induction. Scale bar=30 µm (D) Schematics for evaluating reversibility of effects induced by SS18-SSX or KDM2B depletion (Top panel). HS-SY-II cells were transduced with the indicated TRE-driven shRNAs linked to GFP. Following 10 days of shRNA expression, GFP positive cells were sorted and replated. On Day 12 cells were seeded and maintained in the presence (+Dox) or absence (-Dox) of doxycycline for further analysis. Colony forming assays forming assays for cells maintained in the presence (+Dox) or absence (-Dox) of doxycycline (lower panel). (E) Quantitative q-PCR for HS-SY-II cells grown in the presence or absence of doxycycline, at day 10 after Dox withdrawal. Error bars correspond to mean + s.d. of two independent replicates. (F) GFP IHC staining of tumors generated by subcutaneous injection of SYO-1 cell line transduced with the indicated shRNAs. Scale bar=150 µm A., E. Unpaired t-test *p value<0.05, **p value<0.005***, p value<0.0005.



Figure S3. The DNA binding domain of KDM2B is critical for synovial sarcoma maintenance. Related to Figure 4. (A) T7 assay showing efficient gene editing using guide RNAs against the different *KDM2B* genomic regions. (**B**) Depletion assays based on the number of GFP⁺ cells at day 3 and at days 21-24 following sgRNA infection in Yamato-SS, SYO-1 and FUJI human synovial sarcoma lines. Error bars correspond to mean + s.d. (n=2). (C) Clonogenic assay of HS-SY-II cells transduced with the indicated shRNAs and MSCV-neo empty vector control, MSCV-Neo expressing wild type mouse KDM2B cDNA (KDM2B^{WT}), a JmjC-deficient mutant (KDM2B^{H211A/H222A}) and a ZF-CxxCdeficient mutant (KDM2B^{C600A/C603A}). (D) Crystal violet quantification of the clonogenic assays presented in (C). Data are represented as mean + s.d. (n=2). (E) Immunoblot analysis of total KDM2B levels and exogenous KDM2B (Myc-tag) levels (* indicates an unspecific band). (F) Immunoblot analysis of total KDM2B levels in cells over expressing KDM2B's short isoform. (G) Clonogenic assay of HS-SY-II cells transduced with the indicated shRNAs and MSCV-hygro empty vector control and the short isoform of KDM2B (KDM2B short IF). (H) Clonogenic assay of IMR90 normal human diploid fibroblasts transduced with the indicated shRNAs (I) T7 assays showing comparable and efficient editing by all guide RNAs against PCGF1. (J) Bright field images of HS-SY-II cells transduced with a control sgRNA (GFP) and an sgRNA against the RAWUL domain of PCGF1. Scale bar=25 µm. D. Unpaired t-test *p value<0.05, **p value<0.005.



SSX (SSXRD domain)

Figure S4. SS18-SSX1 interacts with KDM2B via the SSX repressor. Related to Figure 5. (A) Clonogenic assay for *HA-SS18-SSX1* tagged HS-SY-II clone described in Figure 5A-C, transduced with the indicated shRNAs. (**B**, **C**). Proximity ligation assay images and respective quantification verifying KDM2B and SS18-SSX1/2 *in situ* colocalization using an SS18 specific antibody in MFC7 cells and indicated synovial sarcoma lines (B); or a SS18 antibody in HS-SY-II cells upon SS18-SSX1 knockdown (C). Data correspond to mean + s.d. after quantification of three independent fields. Scale bar=25 µm. (**D**) Co-Immunoprecipitation using an anti-KDM2B antibody in 293T cells transiently expressing HA-tagged versions of C-terminal SS18-SSX1 deletions mutants. Unpaired t-test, **p value<0.005, ***p value<0.0005.



Figure S5. SS18-SSX1/KDM2B bind and activate synovial sarcoma-signature genes. Related to Figure 6. (A) Heat maps showing KDM2B, HA-SS18-SSX1 and BRG1 ChIP-Seq signals over 11,345 KDM2B-enriched regions. Rows correspond to ±5 Kb regions across the midpoint of each KDM2B-enriched peak, ranked by increasing KDM2B ChIP signal. Color shading corresponds to KDM2B, HA-SS18-SSX1, and BRG1 ChIP-Seq read counts in each region. (B) ChIP-qPCR validation of HA-SS18-SSX1 binding at the EN2, LHX3 and UNCX loci using two sets of primers designed to amplify genomic regions corresponding to CGIs. Primers for the promoter of HBB (coding for B-Globin) and an intronic region of BCL6 were used as negative controls. Data are represented as mean + s.d. of three technical replicates. (C) Methylation (beta) values across the MNX1 locus in synovial sarcomas compared with normal tissue (Fat) and all other sarcoma subtypes described in Fig 6E. (D) Methylation (beta) values in all 10,533 SS18-SSX1/KDM2B cooccupied regions, comparing the top 500 regions with highest (red), top 500 with the lowest (green) and medium (blue) SS18-SSX1/KDM2B binding. The central horizontal line inside the box plots corresponds to the median. The top and bottom of the boxes correspond to the third and first quartiles, respectively. The upper whisker extends from the top of the box to the largest value no further than 1.5 IQR from it (where IQR is the inter-quartile range, or distance between the first and third quartiles). The lower whisker extends from the bottom of the box to the smallest value at most 1.5 IQR away from the bottom of the box. Data points beyond the end of the whiskers are considered outliers and are plotted individually. Violin plots represent a kernel density estimate of the probability distribution of the data points along the vertical axis. *** p value< 0.001. (E, F) Gene set enrichment analysis (GSEA) comparing the expression of genes associated with the top 500 regions occupied by SS18-SSX1 in synovial sarcoma when compared to other sarcoma types (E) and upon knockdown of SS18-SSX or of KDM2B (F). (G) Gene set enrichment analysis comparing expression of genes differentially expressed in HS-SY-II cells transduced with KDM2B. 4395 and SSX. 1274. (H) Gene ontology analysis of genes commonly down regulated by SS18-SSX or KDM2B knockdown.



Figure S6. Gene repression is a less prominent feature mediated by SS18-SSX1. Related to Figure 6. (A) Plotted RNA-Seq fold changes of genes downregulated by KDM2B or SS18-SSX1 knockdown. (B) Quantitative RT-PCR validating gene expression results obtained by RNA-Seq for downregulated genes. Error bars correspond to mean + s.d. of two independent replicates. (C) Percentage of genes identified as SS18-SSX1 targets by ChIP in upregulated genes ($log_2 FC \ge 1$), downregulated genes ($log_2 FC \le -1$) and unchanged genes in response to SSX.1274 shRNA expression in HS-SY-II cells. (D) Gene set enrichment analysis comparing levels of HA-SS18-SSX1 ChIP signal for downregulated (left) or upregulated (right) genes as a result of SS18-SSX1 or KDM2B knockdown. (F) Plotted RNA-Seq fold changes of genes upregulated by KDM2B or SS18-SSX1 knockdown.



Figure S7. SS18-SSX1 and KDM2B inhibition induce changes in gene accessibility and BRG1 chromatin occupancy. Related to Figure 7. (A) Scatterplot showing correlation between differential SS18-SSX1 occupancy (HA ChIP signal) upon knockdown of SS18-SSX1 or of KDM2B at 10,533 SS18-SSX1/ KDM2B co-occupied regions. (B) Box plots of fold change difference in HA ChIP signal in 10,533 KDM2B cooccupied regions (KDM2B⁺ peaks) and 451 KDM2B non-occupied regions (KDM2B⁻ peaks) upon KDM2B knockdown (KDM2B. 4395). KDM2B knockdown primarily affects SS18-SSX occupancy at KDM2B bound regions. (C) Box plots of BRG1 ChIP signals at 8,488 BRG1-Loss regions and 3,424 BRG1-Gain regions (in response to SS18-SSX knockdown) in HS-SY-II cells transduced with the indicated shRNAs. (D) Venn diagrams showing overlap between BRG1 losses and gains in SSX.1274 and KDM2B.4395 and respective gene ontology analysis using GREAT. (E) Box plots showing fold change differences upon SSX.1274 and KDM2B.4395 at 4,091 ATAC-Seq high confidence gained peaks in response to SSX.1274 expression. (B, C, E) The line in the boxes corresponds to the median. The top and bottom of the boxes correspond to the third and first quartiles, respectively. The lines above and below the box correspond to the 10th and 90th percentile (B) or to the 1st and 99th percentile (C, E). B., C. Unpaired t-test *** p value < 0.0005