

## **Supplemental material**

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Figure S1. *Irg1*<sup>-/-</sup>mice are susceptible to *Aicl1 Mtb* and infection of WT-*Mtb* or *Mtb*-GFP in WT and *Irg1*<sup>-/-</sup> mice and BMDMs. Related to Figs. 1 and 2. (A-E) WT and *Irg1*<sup>-/-</sup> mice were infected with 100-200 CFU of aerosolized WT or isogenic *Aicl1 Mtb*. Mice were monitored for survival (A; n = 6-7), weight change (B; n = 6-7), and bacterial burden in the lung and spleen at 21 dpi (C; n = 8). Each point represents data from one mouse, and bars indicate median values. (D) Absolute numbers of myeloid cell populations in the lung were determined by flow cytometry at 21 dpi (n = 8). M $\varphi$ , macrophages. (E) Cytokine and chemokine levels at 21 dpi in lung homogenates were measured by a Bioplex-Pro cytokine assay (n = 8). (F) Comparison of WT-*Mtb* and *Mtb*-GFP burden in the lung and the spleen at 21 dpi (n = 6-8). Each point represents data from one mouse, and bars represent the median. No statistically significant differences were detected (Mann-Whitney test). (G) CFU analysis on *Mtb*-infected (multiplicity of infection of 1) WT and *Irg1*<sup>-/-</sup> BMDMs or IFN- $\gamma$  pretreated WT and *Irg1*<sup>-/-</sup> BMDMs at 4, 24, and 96 h postinfection. Data in panel G are representative of three independent experiments. Statistical differences were determined by log-rank test (B), Mann-Whitney test (C-E), or one-way ANOVA with Tukey's correction (F and G; \*, P < 0.05; \*\*\*, P < 0.001; ns, not significant).

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Figure S2. Defining cellular infiltrates in the lungs of WT and  $Irg1^{-/-}$  mice after *Mtb* infection and depletion of neutrophils. Related to Figs. 2 and 4. (A) Gating strategy for analysis of innate immune cell populations in the lungs of *Mtb*-infected WT and  $Irg1^{-/-}$  mice. (B) Absolute number of CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells, and B cells in the lungs of WT and  $Irg1^{-/-}$  mice at 21 dpi (n = 6). Each point represents data from one mouse, and bars represent the median. (C) TNF- $\alpha$  concentrations were measured from lungs of WT and  $Irg1^{-/-}$  mice by Bioplex-Pro cytokine assay at time points 10 dpi (n = 6), 14 dpi (n = 10), and 21 dpi (n = 6). (D) Mice were infected with *Mtb*-GFP. Representative histograms and flow cytometry plots for MitoSOX red<sup>+</sup> and Nos2<sup>+</sup> lung neutrophils in WT and  $Irg1^{-/-}$  mice at 21 dpi. The percentage and number of MitoSOX red<sup>+</sup> and Nos2<sup>+</sup> lung neutrophils in WT and  $Irg1^{-/-}$  mice at 21 dpi. The percentage and number of MitoSOX red<sup>+</sup> and Nos2<sup>+</sup> lung neutrophils in WT and  $Irg1^{-/-}$  mice at 21 dpi is shown. Each point represents data from one mouse, and bars represent the median. (B–D) Statistical differences were determined by the Mann-Whitney test (\*\*, P < 0.01; \*\*\*\*, P < 0.0001; rs, not significant). SSC, side scatter. (E) Representative flow cytometry plots from two experiments confirming the extent of neutrophil depletion in the lungs of WT and  $Irg1^{-/-}$  mice treated with anti-Ly6G mAb or an isotype control mAb at 21 dpi. All cells are CD45<sup>+</sup>CD11c: P1-CD11b<sup>hi</sup> Ly6C<sup>hi</sup> inflammatory monocytes; and P3-CD11b<sup>low</sup>LyC<sup>-</sup>-infiltrating macrophages. Numbers on gates represent frequencies of CD45<sup>+</sup>CD11c<sup>-</sup> cells.





Figure S3. **Radiosensitive hematopoietic cells promote Irg1-mediated responses against** *Mtb* and assessment of *Irg1* exon 4 deletion in conditional **knock-out mice.** Related to Fig. 4. (**A**) Scheme for generation of bone marrow chimeric mice. (**B**) Survival of WT  $\rightarrow$  WT, *Irg1<sup>-/-</sup>*  $\rightarrow$  WT, and WT  $\rightarrow$  *Irg1<sup>-/-</sup>* bone marrow chimeric mice infected with 100–200 CFU of aerosolized *Mtb*. The number of evaluated mice (*n*) is indicated in parentheses. (**C**) *Mtb* burden in the lung and spleen of bone marrow chimeric mice was measured at 21 dpi. (**D**) Absolute cell counts for WT (CD45.1) and *Irg1<sup>-/-</sup>* (CD45.2) innate immune cell populations within the lungs of chimeric mice as determined by flow cytometry at 21 dpi (*n* = 7–8). (**C–D**) Each point represents data from one mouse and bars indicate median values. All data are pooled from at least two independent experiments. Statistical differences were determined by log-rank test (\*\*, P < 0.01; B) and Mann-Whitney tests (\*, P < 0.05; \*\*\*, P < 0.001; ns, not significant; C and D). M $\phi$ , macrophages. (**E–G**) Cells were isolated from *Irg1*<sup>fl/fl</sup> and *Irg1*<sup>fl/fl</sup> Cre–expressing mice. mRNA was then extracted and reverse transcribed, and a section of the *Irg1* gene spanning the exon 4 was amplified by PCR. Nonrecombined transcript yields a 490 bp amplicon, whereas Cre recombinase–mediated excision of exon 4 results in a 290 bp amplicon. (**E**) Recombined *Irg1* exon 4 in bone marrow neutrophils of *Cre<sup>-</sup> Irg1*<sup>fl/fl</sup> and *Mrp8*-Cre<sup>+</sup> *Irg1*<sup>fl/fl</sup> mice. (**F**) Recombined *Irg1* exon 4 in bone marrow-derived neutrophils, alveolar macrophages isolated from BAL fluid of *Cre<sup>-</sup> Irg1*<sup>fl/fl</sup> and *CD11c*-Cre<sup>+</sup> *Irg1*<sup>fl/fl</sup> mice. (**G**) Recombined *Irg1* exon 4 in bone marrow-derived neutrophils, alveolar macrophages isolated from BAL fluid, monocytes, and total CD11c<sup>+</sup> cells from spleens of *Cre<sup>-</sup> Irg1*<sup>fl/fl</sup> mice.