

Figure S1

Figure S1: **A.** Representative immunoblot analysis of *CREBBP*, *BRD2*, *EP300*, *BRD3* and *GAPDH* in normal B-cells and CLL patient-derived B-cells. **B.** Protein expression quantification in B-cells isolated from normal donors (n=9) or CLL patients (n=26). Orange dots represent the four cases used or ChIP-seq studies; **C** *BRD4* transcript level in B-cells isolated from normal donors (n=9) or CLL patients (n=26), normalized to *GAPDH*. Red lines represent averages. Orange dots represents the four CLL cases used for ChIP-analysis; **D.** Boxplot showing the expression level distributions (transcript parts per million, TPM) of *BRD4* in patient's derived primary CLL cells and normal B cells (GEO study GSE42262, sample GSM1036401); **E.** Hockey stick plot for BRD4 load across enhancers of four CLL patient cells and a normal B cell line. SEs are defined as enhancers surpassing the inflection point; **F.** Venn diagram of SEs for primary CLL cells (basal and CpG stimulated conditions); **G.** Heatmap showing BRD4 load (total of reads per million per base pair) of SEs that are common for the four analyzed CLL samples (at least 3 out of 4) under basal or CpG stimulate conditions but not in normal B cell line; **H.** BRD4 load at super-enhancers of selected genes know to be either up-regulated (*IL4R*, *MIR21*, *TCL1A*, *CCR7*, and *PAX5*) or down-regulated (*AICDA*) in CLL compared to normal B cells with known prominent roles in tumor biology; **I.** Gene tracks of H3K27ac (red) and BRD4 (yellow) occupancy in primary CLL cells. The x-axis shows genomic position and y-axis shows signal of ChIP-seq occupancy in units of rpm/bp.

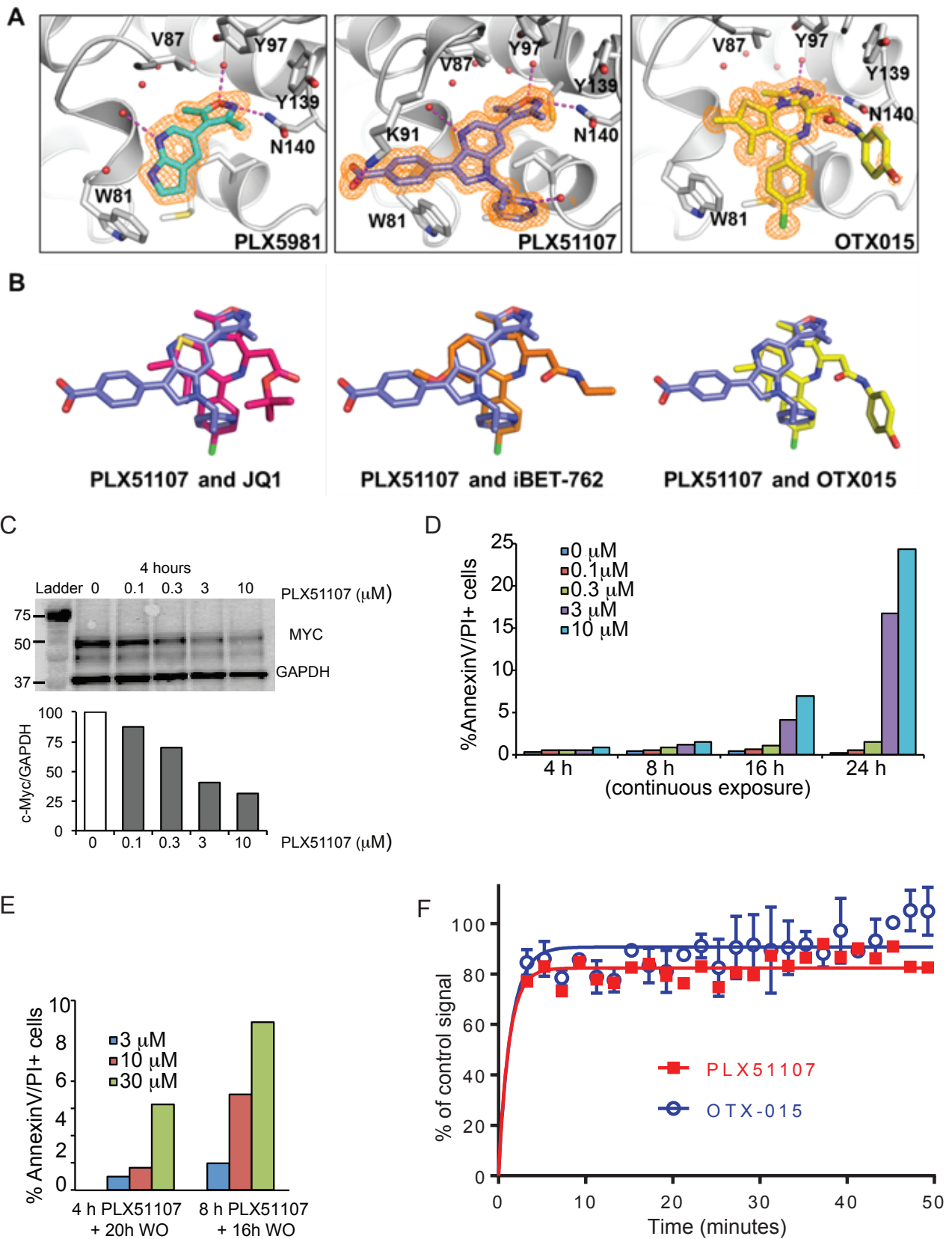


Figure S2

	$t_{1/2}(\text{min})$	Apparent $K_{\text{off}} (\text{min}^{-1})$
PLX51107	< 3.25	> 0.213
OTX-015	< 3.25	> 0.213

Figure S2: Co-structures with different BRD4 inhibitors. A. A F_o-F_c omit map contoured at $+3\sigma$ level for PLX5981, PLX51107 and OTX015. The inhibitors and key residues at the active site are shown in stick model. F_o-F_c omit maps were calculated using the refined coordinates of the structures excluding the ligands; **B.** Overlay of PLX51107 (purple) with JQ1 (red; PDB: 3MXF), i-BET762 (orange; PDB: 3P5O) and OTX015 (yellow). **(C-F):** Effect of exposure time on PLX51107 pharmacodynamic markers and the determination of compound residence times. **C.** Short term (4 hours) PLX51107 treatment at 3 and 10 μM concentrations resulted in robust downregulation of c-Myc in MV4-11 cells (GAPDH was used as loading control). **D.** AnnexinV/PI flow cytometric analysis of MV4-11 cells treated with PLX51107 continuously for 4, 8, 16, and 24 hours. Significant apoptosis was observed at 3 and 10 μM only after prolonged incubation. **E.** AnnexinV/PI flow cytometric analysis of MV4-11 cells treated with PLX51107 for 2, 4 or 8 hours followed by a washout period of 22, 20, or 16 hours (i.e. cytometric analysis was conducted 24 hours after treatment initiation in all cases). Two representative dot plots are shown. **F.** Off-rate measurement of PLX51107 and OTX-015 using a time-resolved fluorescence resonance energy transfer (TR-FRET) assay. Compound incubations were performed with 0.3 μM GST-BRD4 (containing BD1 and BD2), 0.3 μM Eu-anti-GST antibody, and 1.5 μM of the test compounds.

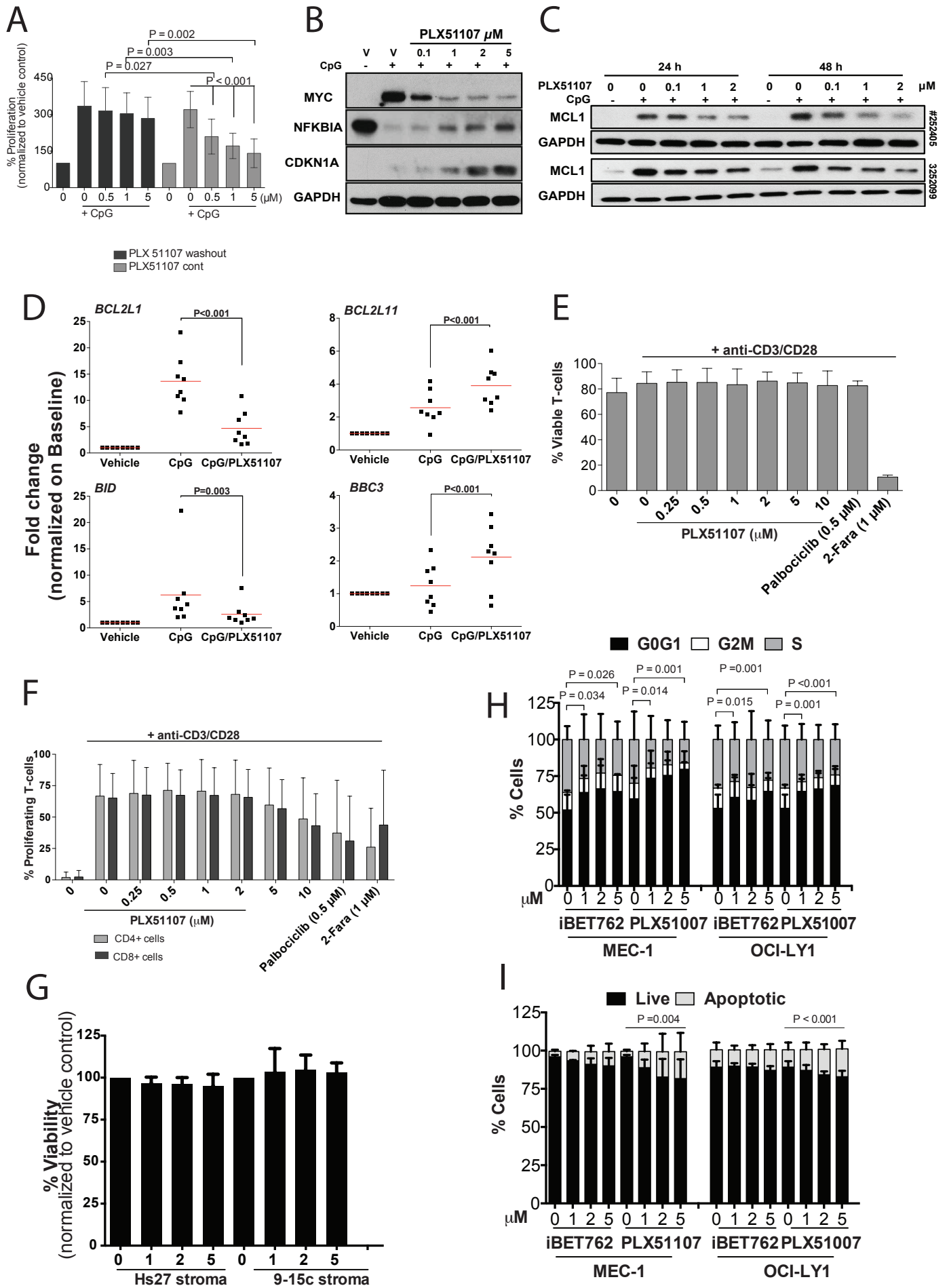
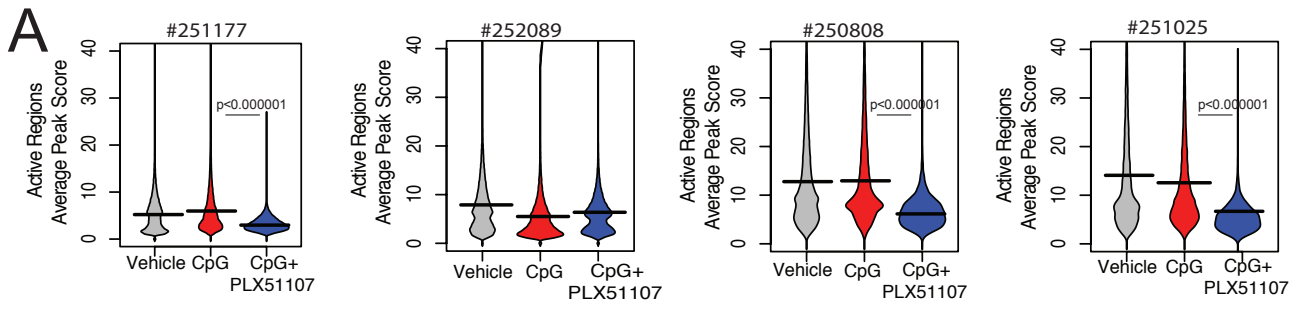
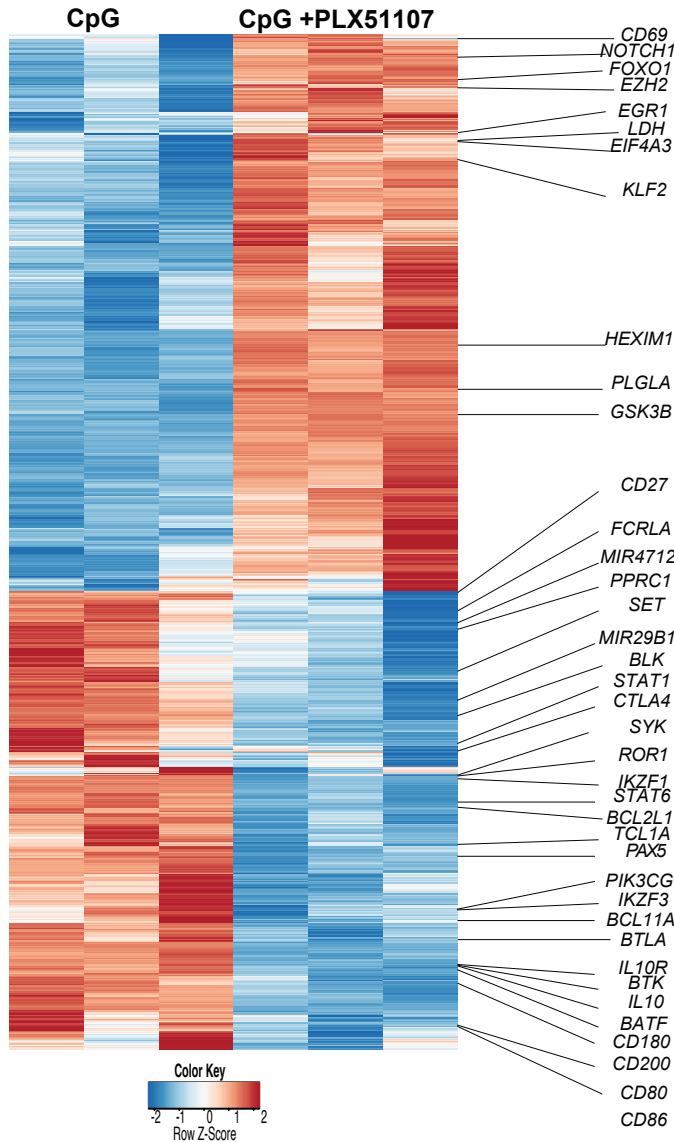


Figure S3

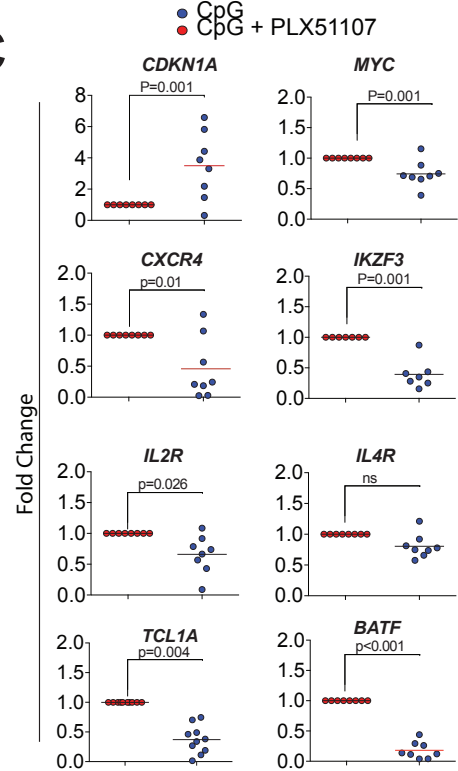
Figure S3: BET inhibition attenuates CpG-induced survival effects in CLL. **A.** Dose-dependent decrease of proliferation of CpG stimulated normal B cells upon BRD4 inhibition with increasing doses of PLX51107 for 4h followed by 44h washout or 48h continuously; (n=5); **B.** Representative immunoblot analysis of cMYC, I κ B α , CDKN1A and GAPDH following PLX51107 treatment (0.1, 1, 2 or 5 μ M) in CpG-stimulated CLL cells (4h, n=3); **C.** Representative immunoblot analyses of the antiapoptotic protein, MCL1 following PLX51107 treatment of CpG stimulated B-CLL cells (24 and 48h, n=5) GAPDH is used as loading control; **D.** Quantitative real time-PCR analysis of the indicated apoptosis-related genes in primary B-CLL cells following PLX51107 treatment (1 μ M, 4 h) under stimulated conditions with 3.2 μ M CpG oligonucleotides (n=8). Red lines represent averages; **E-F.** Flow cytometric analysis of the effect of PLX51107 on T-cell viability and proliferation. Freshly isolated CD3 T-cells from healthy donors were stimulated by plate bound anti-CD3 and soluble anti-CD28 (+ anti-CD3/CD28) and treated with vehicle DMSO or increasing concentrations of PLX51107 as indicated. Palbociclib (0.5-1 μ M) and Fludarabine (2-Fara, 1/5 μ M) were used as control drugs to inhibit proliferation and viability, respectively. (E) T-cell viability was evaluated following a 4-day treatment using the LIVE/DEAD Fixable near-IR stain. Results are shown as Mean \pm SD (n=3-7); (F) CD4 and CD8 T-cell were investigated for CFSE dilution resulting from proliferation at the indicated time points and treatment conditions. Gates were set on viable CD4 or CD8 T-cells that underwent at least 1 cellular division and results are shown as % proliferating T-cells (Mean +) (**F**); **G.** Human Hs27 or murine 9-15c bone marrow-derived stroma cells were treated with increasing concentrations of PLX51107 for 72 h. Effects on proliferation were determined by MTS assay and represented as % of vehicle control. Results shown as mean \pm SD of n=3 independent experiments per cell line; **H.** Cell cycle analysis demonstrates accumulation of cells in G0/G1 phase in malignant B-cell lines following treatment with iBET762 or PLX51107 (1, 2 or 5 μ M) at 72h (n=3). Results shown as Mean \pm SD (n=3-4); **I.** Annexin/PI flow cytometric analysis of malignant B-cell lines treated with BRD4i, iBET-762 or PLX51107 (1, 2 or 5 μ M) at 72h (n=4).



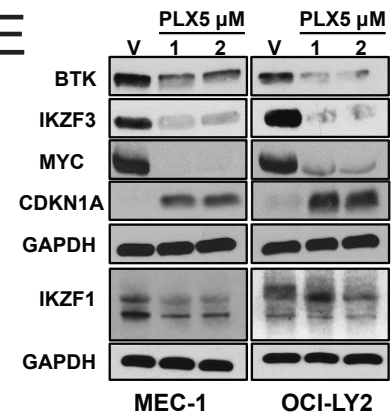
B



C



E



D

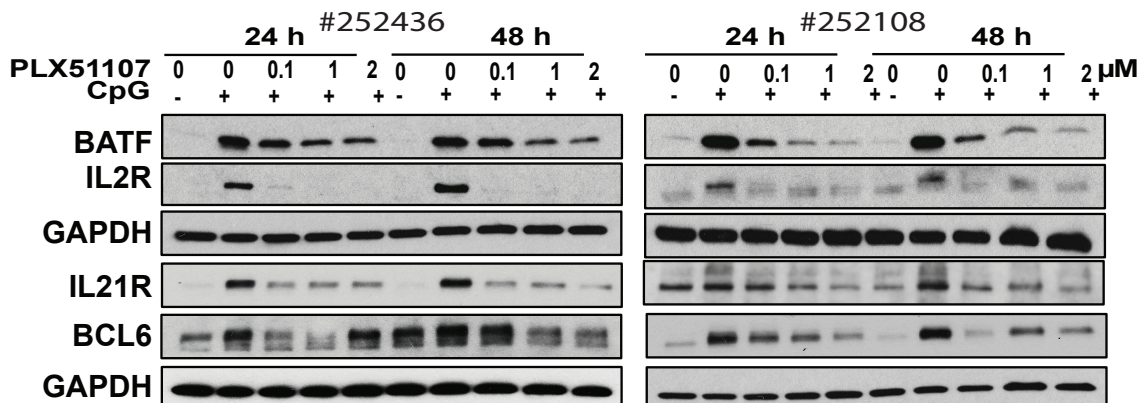


Figure S4

Figure S4: **A.** Bean plots showing global distribution of BRD4 peak scores at active regions following 4h treatment of primary CLL cells (a = vehicle, b = 3.2 μ M CpG stimulation, c = CpG stimulation + PLX51107, horizontal bars mark the mean scores); **B.** Heatmap of 1361 differentially expressed genes (745 up and 616 down regulated) with absolute fold change of 2 or more and p-value less than 0.01 upon BRD4 inhibition in microarray data. Microarray analysis using GeneChip Human Transcriptome Array HTA 2.0 (Affymetrix[®]) was performed in patient-derived CLL cells stimulated with CpG oligonucleotides (3.2 μ M) and treated with either vehicle (VEH/CpG) or 1 μ M PLX51107 (PLX5/CpG) for 4 h. Experiments were performed as triplicates, each representing a pool of 2-3 CLL patient samples each; **C.** Quantitative real time-PCR analysis of the selected genes in PLX51107 treated CpG-stimulated CLL patient-derived B-cells (n=8-10). Red lines represent averages; **D.** of protein levels of BATF, IL2R, IL21R, BCL6, and GAPDH following PLX51107 treatment (0.1, 1, or 2 μ M) in CpG-stimulated patient-derived CLL cells (24 and 48 h, n=5); **E.** Representative immunoblot analyses of relative protein levels of BTK, IKZF3, MYC, CDKN1A, IKZF1 and GAPDH following treatment with vehicle (V) or PLX51107 (PLX5: 1 or 2 μ M, 72 h) in three B-cell malignant cell lines (MEC-1, OCI-LY1).

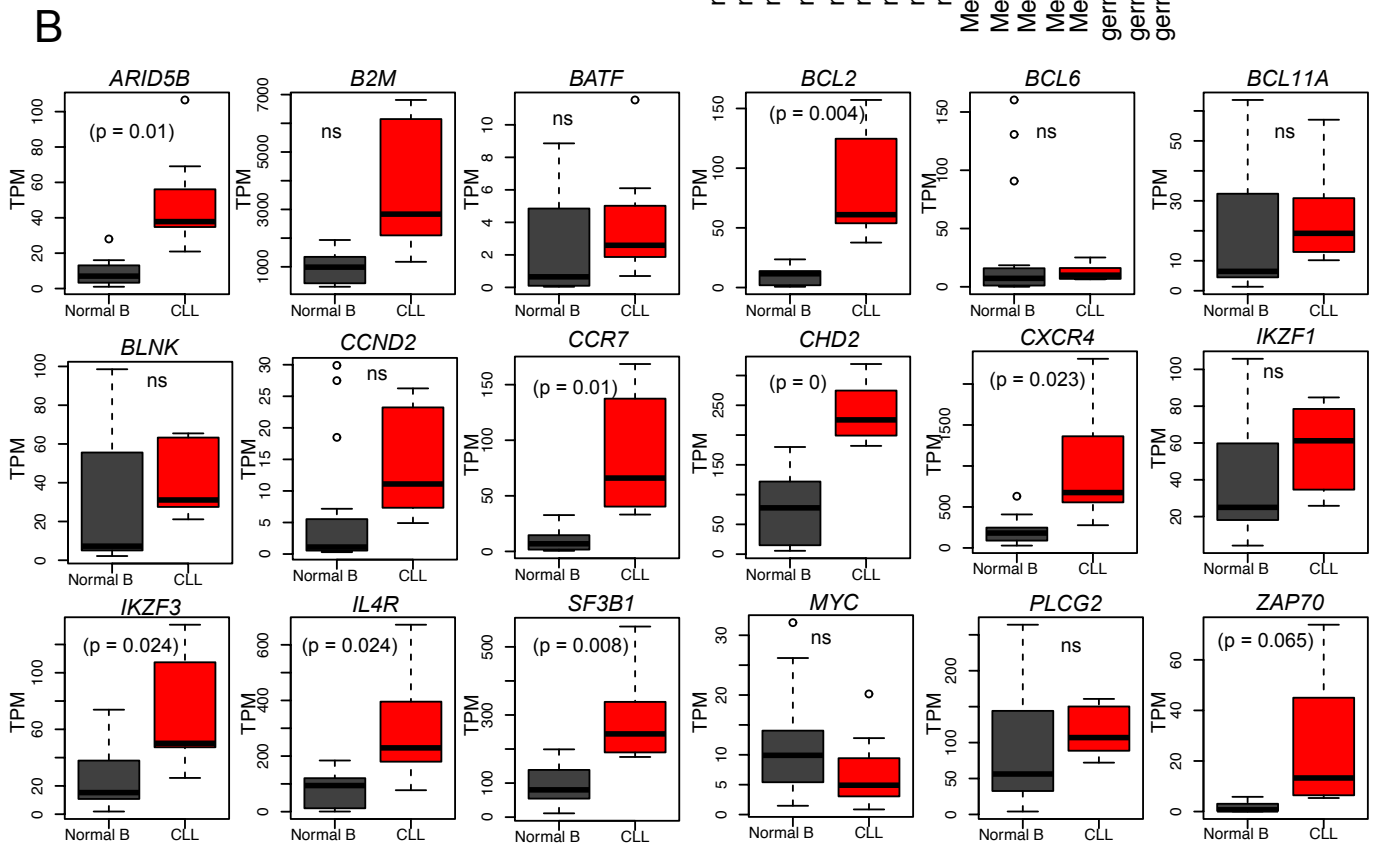
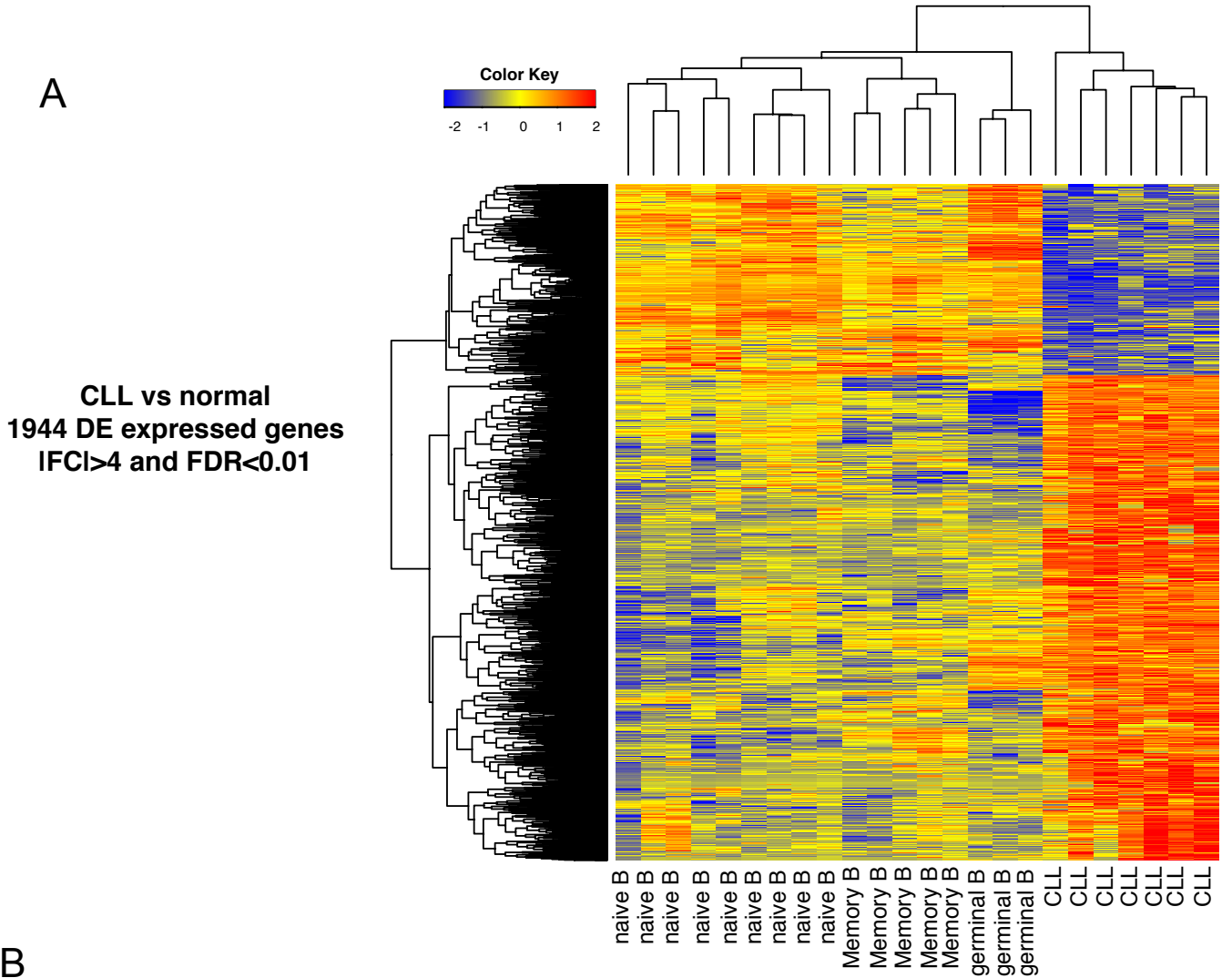


Figure S5

Figure S5. A. Differential gene expression profile in CLL compared to normal B cells. Heatmap of 1944 differentially expressed genes (with absolute fold change of 4 or more and p-value less than 0.01 in CLL compared to normal B cells in RNA-seq data; **B.** Boxplot showing the expression level distributions (transcript parts per million, TPM) for patients derived primary CLL cells and normal B cells for select genes.

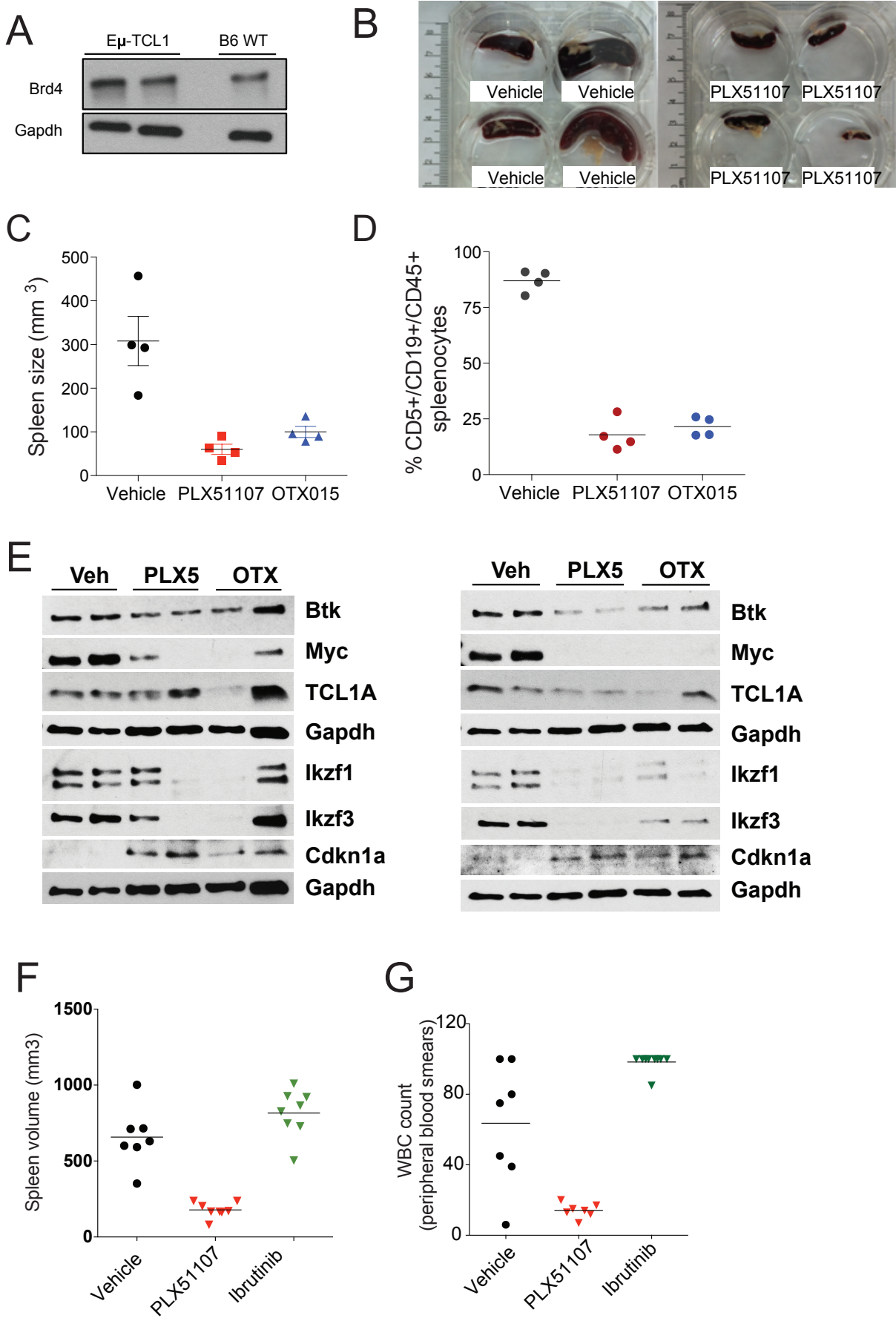


Figure S6

Figure S6: Targeting BRD4 in transgenic mouse models of B-cell malignancies. A. Immunoblot analysis of BRD4 expression in CD19 positive B-cells isolated from spleens of leukemic E μ -TCL1A mice and C56BL/6 wild type mouse (WT, 12 mo); GAPDH is used as loading control. **B.** Splenocytes derived from E μ -TCL1A mice were cultured in vitro with or without increasing concentration of PLX51107 for 48h. Effects on proliferation were determined by MTS assay and represented as % of vehicle control. Results shown as mean \pm SD of n=3 independent experiments; **C.** Splenocytes derived from E μ -TCL1A mice were cultured in vitro with or without increasing concentration of PLX51107 for 8h. *TCL1* mRNA transcript derived from the E μ -TCL1 transgene, *Hexim1* and *Myc* were assessed using qRT-PCR. Endogenous *Tcl1* expression (not shown) was also measured and was undetermined as previously described in this model; **D.** Decreased spleen size of PLX51107-treated E μ -TCL1 mice vs. vehicle control following an 8 day treatment (PLX51107: 20 mg/kg, qd, oral gavage); **E.** Caliper-derived spleen volumes of mice treated with PLX51107 (20 mg/kg, qd, oral gavage), OTX015 (50 mg/kg, qd, oral gavage) or vehicle control for 30 days. Spleen volumes were measured at time of death using a manual caliper (with 1mm error). Splenic volume was then calculated using the standard clinical ellipsoid equation of length \times width \times thickness \times 0.523; **F.** Flow cytometry analysis of CD19/CD5/CD45 positive peripheral blood cells of mice treated for 30 days with PLX51107, OTX15 or vehicle control BL; **G.** Representative immunoblot analysis of relative protein levels of Btk, Myc, TCL1A, Ikaros1, Ikaros3, and Cdkn1a in splenocytes derived from mice treated with PLX51107, OTX15 or vehicle control for 30 days; **H-I.** Using an adoptive transfer model of E μ -TCL1, recipient wild type mice were randomized to receive vehicle Ibrutinib or PLX51107 (20 mg/kg, qd, oral gavage) at leukemia onset. Disease progression was measured by flow cytometry as % CD19/CD5/CD45 positive PBL: PLX51107 reduced spleen mass (F), and decreased % of circulating leukemic PBL as measured at time of death (G).

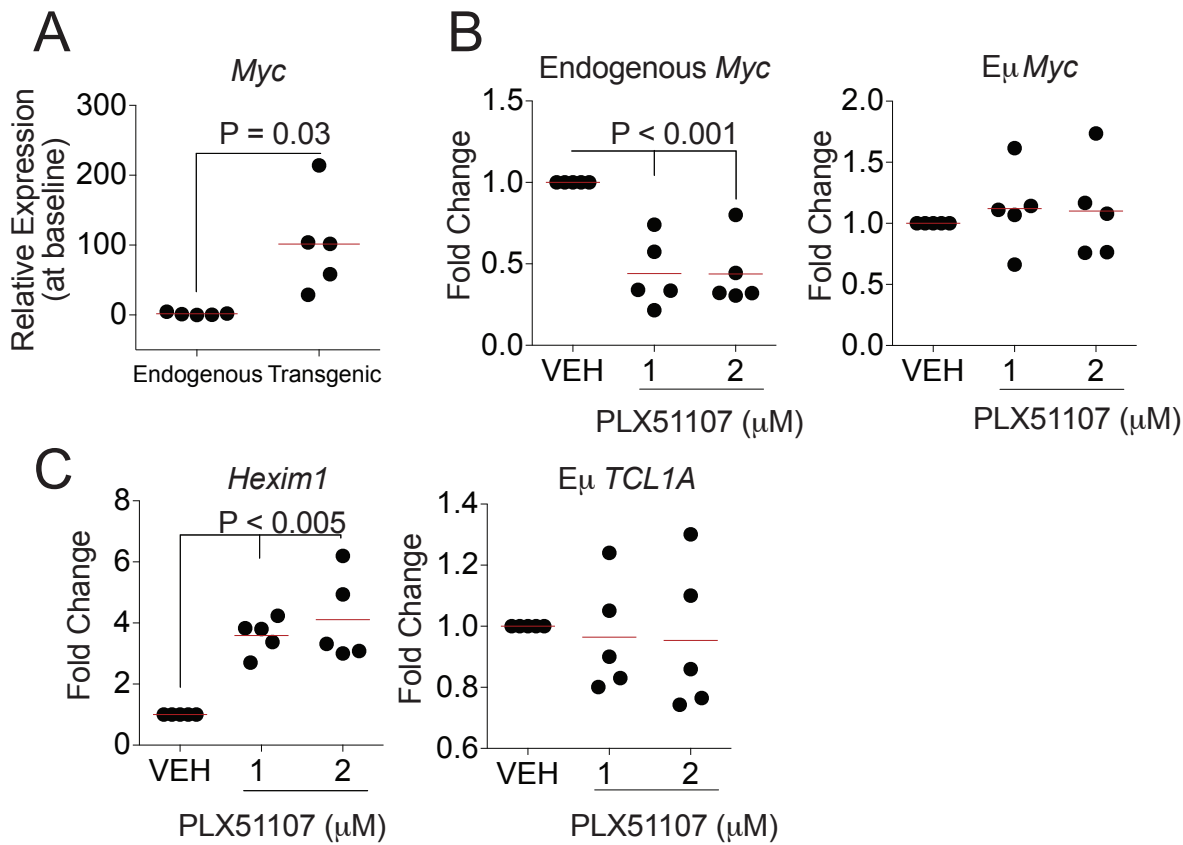


Figure S7

Figure S7: BRD4 target modulation in a disease model of aggressive CLL and Richter's Transformation: A-C) Splenocytes derived from *E μ -Myc/TCL1* mice were cultured in vitro. *Myc* mRNA transcript derived from the *E μ -Myc* transgene or the endogenous *Myc* locus was assessed using qRT-PCR at baseline (A) or following 8h exposure to either DMSO (vehicle) or PLX51107 (B) Transcript levels of *Hexim1* and *E μ -TCL1* transgene following 8h exposure to either DMSO (vehicle) or PLX51107.

SUPPLEMENTARY TABLES

Table S1. Characteristic of patients used for the in vitro studies.

Fig. 1A	IgVH mutation al status (%)	Karyotype	Cytogenetics				Treatment Status
			17p	11q	13q	12q	
250672	U (0)	49,XY,+X,add(1)(q21),dic(1;17)(p12;p11.2),t(2;14)(p13;q32.3),+4,add(4)(p12)x2,+6,add(6)(p21),psu dic(8;6)(q24;q12),der(9)t(1;9)(p22;p13),+11,+12,t(19;22)(q13.3;q11.2)[cp25]/50,sl,+15[5].ish add(6)(D6Z1+,SEC63+),dic(8;6)(D6Z1+,SEC63-)	pos	pos	neg	pos	T
251161	M (10)	46,XX,t(7;8)(p15;q24.1),del(13)(q12q14)[9]/47,XX,+X[2]/46,XX[7]/nonclonal[1]	neg	neg	pos	neg	T
251806	U (0)	46,XX,del(11)(q21q23)[3]/46,XX[16]/nonclonal[1]	neg	pos	pos	neg	T
251868	U (0)	43-46,XX,del(11)(q14q23),del(13)(q12q14)[cp2]/46,XX[12]/nonclonal[1]	neg	post	pos	neg	T
250782	U (2.2)	46,XX,add(10)(q24),add(19)(p13.3)[cp3]/45,sl,-X[cp8]/46,X,del(X)(q22)[cp2]/46,XX,del(13)(q22q34)[cp2]/46,XX[2]/nonclonal[3]	pos	neg	pos	neg	T
250547	M (8.9)	46,XX(19)/nonclonal(1)	pos	neg	pos	neg	Unt

Fig. S1A	IgVH mutation al status (%)	Karyotype	Cytogenetics				Treatment Status
			17p	11q	13q	12q	
250751	M (*)	46,XX[19]/nonclonal[1]	neg	neg	pos	neg	T
251600	U (0)	46,XY[20]	neg	pos	pos	neg	T
251308	M (7.8)	46,XY[20]	neg	neg	pos	neg	Unt
250872	M (4.1)	46,XX[20]	neg	neg	pos	neg	Unt
250622	M (4.4)	46,XX[19]/nonclonal[1]					Unt
250789	U (0)	48,XX,dup(1)(q12q32),inv(7)(p13q22),t(14;19)(q32;q13.3),+15,+21[17]/nonclonal with clonal abnormalities[3].ish t(14;19)(3 (sq) IGH+;3 (sq) IGH+,5 (sq) IGH+)	neg	neg	neg	neg	T
250749	U (0)	44,XY,add(3)(p26),der(11)t(11;15)(q21;q13),del(12)(q22),-15,dic(17;18)(p11.2;p11.2)[cp10]/46,XY[9]/nonclonal[1]Previously described as: 44,XY,add(3)(p26),der(11)t(11;15)(q21;q13),del(12)(q22),-15,der(17;18)(q10;q10)[cp10]/46,XY[9]/nonclonal[1]	pos	pos	pos	neg	T
250967	U (*)	38-41,XX,ider(2)(p10)add(2)(p23),add(3)(p21),-4[8],del(4)(q13)[5],add(7)(q36),-8,-9,der(11)add(11)(p15)add(11)(q21),-13,-	pot	pos	pos	neg	T

		13,-14,-15,- 17,der(19)t(13;19)(q14;p13.3),-21,-22,- 22,+mar1-12,inc[cp19, one is 4n]/46,XX[1]					
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Fig. S4A	IgVH mutational status	Karyotype	Cytogenetics				Treatment Status*
			17p	11q	13q	12q	
250486	M (5)	46,XX,der(13)t(13;17)(q14;q11.2)ins(13;?) (q14;?),der(17t(13;17)(q14;q11.2)[20]	neg	neg	neg	neg	T
250510	M (11.53)	46,XY[20]	neg	neg	pos	neg	Unt
251186	U (0.3)	46,XY,t(1;3)(p34;q12)[2]/46,XY[18]	neg	neg	pos	neg	Unt
251847	U (1.4)	48,XY,+7,add(7)(q22),+12[20]	neg	neg	neg	pos	Unt
251900	U (0)	45,XX,dic(4;18)(p12;p11.3),del(11)(q13.3q23.3)[cp2]/45,sl,add(12)(q24.3)[9]/45, sdl1,add(15)(q26)[2]/45,sdl2,+12,- add(12)[5]/45,sdl3,add(5)(q35)[cp2]	neg	pos	pos	neg	Unt
250236	U (0)	46,XY,dic(11;17)(p11.2;p11.2),+mar1[4]/ 46,sl,- dic(11;17),+11[cp3]/46,XY[10]/nonclonal[3]	pos	neg	pos	neg	T
250547	U (*)_	46,XX[19]/nonclonal[1]	pos	neg	pos	neg	Unt
250720	M (7.14)	46,XY,t(13;17)(q14;p13)[cp6,one is 4n]/46,sl,add(9)(q34)[cp3]/46,XY,der(17) t(2;17)(p11.2;p11.2)[cp5]/45,XY,t(3;4)(q21;q35),del(11)(q21),- 17[3]/46,XY[1]/nonclonal[2]	pos	pos	neg	neg	Unt
251608	M (5.1)	46,X,t(X;8)(q28;q22),del(6)(q15q21),inv(11) (p15q23)[6]/nonclonal w/clonal abnormalities[2]/46,XX[11]/nonclonal[1]	neg	neg	pos	neg	T
252097	M (6.1)	45,X,-Y,del(21)(q21.1q22)[17]/nonclonal w/clonal abnormalities[2]/46,XY[1]	neg	neg	pos	neg	Unt

Fig 4G and S3D	IgVH mutational status	Karyotype	Cytogenetics				Treatment Status*
			17p	11q	13q	12q	
251032	U (0.3)	45,XX,dic(4;17)(p11;p11.2)[12]/46,XX,t(13;16) (q14;q24)[2]/46,XX[6]	pos	neg	pos	neg	T
251201	U (0)	46,XY,add(11)(q22)[6]/46,XY[13]/4n[1]	neg	pos	neg	neg	Unt
251495	M (8)	46,XX[20]	neg	neg	pos	neg	Unt
251725	U (0)	46,XY[20]	neg	neg	neg	neg	Unt
252026	M (3)	44-76,XY,- 15,der(17)t(15;17)(q12;p12)[cp5,one is 4n]/40-45,XY,der(3)t(2;3)(p13;p21),- 6,add(17)(p13)[cp5]/46,XY[4]nonclonal[1]. ish der(3)t(2;3)(REL+),nuc ish(D4Z1x2,D6Z1x1)[40/210]	pos	pos	pos	neg	Unt
251045	U (*)_	46,XY[19]/nonclonal[1]	neg	neg	pos	neg	T
251180	M (4.8)	46,XY[19]/nonclonal[1]	neg	neg	pos	neg	Unt
251749	U (0)	Insufficient Metaphases (46,XY[3]/nonclonal[1])	neg	neg	pos	neg	T
252502	M (6.1)	46,XX[18]/nonclonal[2]	neg	neg	pos	neg	Unt
250810	U (0)	46,XX,del(10)(q23q25)[3]/45,X,- X[cp4]/46,XX[13]	neg	neg	neg	neg	T

ChIP-seq and ATAC-seq	IgVH mutation al status	Karyotype	Cytogenetics				Treatment Status*
			17p	11q	13q	12q	
251177	U (0)	46,XY,del(11)(q14q23)[11]/nonclonal w/clonal abnormalities[6]/46,XY[2]/nonclonal[1]	neg	pos	neg	neg	T
252089	U (0.8)	47,XX,+12[20]	neg	neg	neg	pos	Unt
250808	U (*)	46,XX[20]	neg	neg	pos	neg	Unt
251025	U (*)_	46,XY[30]	neg	neg	pos	neg	Unt

Fig 4G, S3C, and S4C	IgVH mutation al status	Karyotype	Cytogenetics				Treatment Status*
			17p	11q	13q	12q	
252405	U (0)	45,XX,dic(17;18)(p11.2;p11.2)[5]/46,XX[15]/nonclonal[2]	pos	neg	neg	neg	T
252099	M (11.1)	46,XX,del(13)(q12q14)[4]/46,XX[15]/nonclonal[1]	neg	neg	pos	neg	Unt
251692	U (*)_	46,XY[20]	neg	neg	pos	neg	T
252436	U (0)	46,XX[21]	neg	pos	neg	neg	Unt
252108	M (4.7)	45,X,-X[10]/45,sl,t(10;13)(q22;q14)[2]/46,XX[8]	neg	neg	pos	neg	Unt

Fig 3C	IgVH mutation al status	Karyotype	Cytogenetics				Treatment Status*
			17p	11q	13q	12q	
250468	U (*)_	45,X,-Y,t(11;14)(q13;q32.3)[18]/46,XY[2]	neg	pos	neg	neg	Unt
251915	U (1.4)	46,XY,del(11)(q21q23)[16]/nonclonal w/del(11q)[1]/46,XY[2]/nonclonal[1]	neg	pos	pos	neg	Unt
252087	U (0)	47,XX,+12[19]/46,XX[1]	neg	pos	pos	pos	Unt
250169	U (*)_	47,XY,+12[12]/nonclonal w/ clonal abnormalities[1]/46,XY[6]/nonclonal[1]	neg	pos	pos	pos	T
250236	U (0)	46,XY,dic(11;17)(p11.2;p11.2),+mar1[6]/43-45,sl,-dic(11;17),+11[cp2]/46,sdl,-mar1,+mar2[2]/46,XY[10]/nonclonal[8]	pos	neg	pos	neg	T
250529	U (2.36)	46,XY[29]/nonclonal[1]	neg	neg	pos	pos	Unt
250955	M (6)	46,XY[20]	neg	neg	pos	pos	Unt
251013	U (0.7)	47,XX,+12[10]/47,sl,del(6)(q13q25)[6]/nonclonal w/ clonal abnormalities[1]/46,XX[3]	neg	neg	neg	pos	Unt
251383	U (*)_	47,XY,+12[1]/45-46,sl,dic(8;17)(p11.2;p11.2)[14]/nonclonal w/clonal abnormalities[1]/46,XY[4]	pos	neg	neg	pos	Unt
251606	U (0.3)	46,XX,i(17)(q10)[cp1]/45,sl,-9,-10,+mar1[cp1]/46,XX,-14,add(17)(p11.2),-18,-21,+r,+mar2,+mar3[cp18, one is 4n]/46,XX[7]/nonclonal[2]	pos	neg	pos	neg	Unt
251725	U (0)	46,XY[20]	neg	neg	neg	neg	Unt
251975	M (4.8)	47,XY,+12[3]/46,XY[17]	neg	neg	pos	pos	Unt
252099	M (11.1)	46,XX,del(13)(q12q14)[4]/46,XX[15]/nonclonal[1]	neg	neg	pos	neg	Unt

Fig S3A, and	IgVH	Karyotype	Cytogenetics	Treatment
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S3B	mutational status		17p	11q	13q	12q	Status
250262	U (0.3)	46,XY[19]/nonclonal[1]	neg	neg	pos	neg	T
250776	U (0)	46,XY,del(11)(q21q23)[2]/46,XY,dic(8;12)(p11.2;p11.2),+12[cp2]/46,XY[15]/nonclonal[1]	neg	pos	pos	pos	T
251161	M (10.9)	46,XX,t(7;8)(p15;q24.1),del(13)(q12q14)[12]	neg	neg	pos	neg	Unt
251606	U (0.3)	46,XX,i(17)(q10)[2]/45,sl,-9,-10,+mar[12]/90,sdx2[3]/nonclonal w/clonal abnormalities[1]/nonclonal[2]	pos	neg	pos	neg	Unt

Fig 3A, and 3B	IgVH mutational status	Karyotype	Cytogenetics				Treatment Status
			17p	11q	13q	12q	
251476	U (1)	N/A	neg	neg	pos	neg	Unt
251751	U (1)	N/A	neg	neg	pos	neg	Unt
252535	U (*)	47,XY,+2,del(2)(q31q35),del(11)(q13q23)[2,one w/nonclonal abnormalities]/46,sl,-del(2),add(2)(p23),del(13)(q12q14)[5,two w/nonclonal abnormalities]/46,XY[12]/nonclonal[1]	neg	pos	pos	neg	Unt
252053	U (*)	46,XY,del(13)(q12q14)[10]/46,sl,del(11)(q21q23)[3]/46,sdl1,t(5;19)(q11.2;q12)[2]/46,sl,add(17)(q25)[2]/46,XY[3]Previously reported as:46,XY,del(13)(q12q14)[10]/46,sl,del(11)(q13q23)[3]/46,sdl1,t(5;19)(q11.2;q12)[2]/46,sl,add(17)(q25)[2]/46,XY[3]	neg	pos	pos	neg	Unt
251843	U (0.3)	46,XX,t(9;13)(q33;q12)[2]/46,sl,der(11)inv(11)(p15q13)del(11)(q22q23)[5]/46,XX,del(13)(q14.1q14.3)[3]/46,XX[10]	neg	pos	pos	neg	T

Test for IgVH mutational status was performed but no % was documented; U (unmutated IgVH mutational status); M (mutated IgVH mutational status); pos (positive); neg (negative); T (treated patient); Unt (Untreated patient); NA (information not available).

Table S2. Top functional annotations of 764 genes common to super-enhancers (SEs) of at least three out of four samples by DAVID Functional Annotation Tool.

Annotation Term	Genes	p-value	FDR
GO Biological Process Enrichment			
B cell receptor signaling pathway	IGHG2, PTPN6, IGLC7, BLK, IGLC6, TRDC, IGHM, IGHD, BCL2, IGHE, IGHA1, IGHA2, IGKC	4.01E-10	6.78E-07
Positive regulation of B cell activation	IGHG2, IGLC7, IGHD, IGHE, IGLC6, IGHA1, IGHA2, TRDC, IGKC, IGHM	8.43E-10	1.42E-06
Phagocytosis, recognition	IGHG2, IGLC7, IGHD, IGHE, IGLC6, IGHA1, IGHA2, TRDC, IGKC, IGHM	1.80E-09	3.04E-06
Immune response	TNF, IL19, TNFRSF8, TNFSF13, TNFSF12, HLA-DMA, TNFRSF4, IL10, CHIT1, B2M, TNFRSF1B, CXCR5, IL4R, TNFRSF18, IGHA1, NRROS, IGHA2, IGKC, LTB, LTA, CIITA, IL2RA, IL24, HLA-DQA1, SERPINB9, CCR7, IGHD, IRF8, IGHE, TNFSF12-TNFSF13	4.04E-09	6.82E-06
Phagocytosis,	IGHG2, IGLC7, IGHD, IGHE, IGLC6,	1.63E-	2.76E-05

engulfment	IGHA1, IGHA2, TRDC, IGKC, IGHM	08	
Negative regulation of transcription from RNA polymerase II promoter	TNF, USP3, SPI1, PAX5, CBX7, NR1H2, HEXIM2, ATN1, HEXIM1, BCL11A, SOX15, BCL6, SUPT4H1, TCF4, ETV6, BCOR, CIITA, ZFP36, ZBTB20, IKZF1, ARID5B, KLF16, SPEN, TMPRSS6, JUNB, FOXP1, PLK3, BTG2, IRF8, JAZF1, TGIF1, PAF1, ZFPM1, ID3	4.75E-06	0.00803
Negative regulation of transposition	APOBEC3G, APOBEC3H, APOBEC3F, APOBEC3C, APOBEC3D	9.29E-06	0.0157
GO Molecular Function Enrichment			
Immunoglobulin receptor binding	IGHG2, IGLC7, IGHD, IGHE, IGLC6, IGHA1, IGHA2, TRDC, IGKC, IGHM	9.44E-10	1.37E-06
Protein binding	212 Genes	1.66E-05	0.0241
Tumor necrosis factor receptor binding	TRAF1, TNF, TNFSF13, TNFSF12, LTB, LTA, TRAF4	1.82E-05	0.02643
Antigen binding	IGHG2, IGLC7, IGHD, IGHE, IGLC6, IGHA1, IGHA2, FCGRT, TRDC, IGKC, IGHM	3.63E-05	0.05255
Hydrolase activity, acting on carbon-nitrogen bonds, in cyclic amidines	APOBEC3G, APOBEC3H, APOBEC3F, APOBEC3C, APOBEC3D	4.40E-05	0.06374
GO Cellular Component Enrichment			
External side of plasma membrane	IGHG2, IL2RB, TNF, IL2RA, IGLC7, IGLC6, TNFRSF13C, TRDC, IGHM, PDCD1, B2M, CD83, CCR7, ECE1, CXCR5, IGHD, IGHE, IGHA1, IGHA2, IGKC	2.75E-08	3.72E-05
Immunoglobulin complex, circulating	IGHG2, IGLC7, IGHD, IGHE, IGLC6, TRDC, IGKC	1.00E-06	0.00136
Cytoplasmic mRNA processing body	ZFP36, ZFP36L1, PATL2, APOBEC3G, LSM2, APOBEC3H, APOBEC3F, ZC3H12D, APOBEC3D, SAMD4B, DDX6	2.20E-06	0.00297
Blood microparticle	IGHG2, IGLC7, IGLC6, HSPA1A, HSPA1B, TRDC, IGHM, HSPA1L, PFN1, IGHD, IGHE, IGHA1, IGHA2, IGKC	7.01E-06	0.00949
KEGG Pathway Enrichment			
Cytokine-cytokine receptor interaction	IL2RB, TNF, IL2RA, IL21R, TNFRSF13C, TNFRSF8, TNFSF13, TNFSF12, TNFRSF4, IL10, FLT3LG, TNFRSF1B, CCR7, CXCR5, CXCR4, IL10RA, IL4R, TNFRSF18, LTB, LTA	1.66E-06	0.00207

Table S3: Crystallography data collection and refinement statistics

	BRD4-PLX5981 (PDB: 5WMA)	BRD4 ^{D96A} -PLX51107 (PDB: 5WVG)	BRD4 ^{D96A} -OTX015 (PDB: 5WMD)
Data collection			
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁

Cell dimensions			
<i>a, b, c</i> (Å)	39.2, 44.5, 78.3	38.8, 42.2, 92.4	36.9, 44.6, 77.8
Resolution (Å) ^a	78.6-1.40	24.9-1.15	21.2-1.18
	(1.45-1.40)	(1.19-1.15)	(1.22-1.18)
<i>R</i> _{sym} or <i>R</i> _{merge}	0.111 (0.558)	0.04 (0.486)	0.037 (0.239)
<i> σ </i>	9.8 (3.3)	20.8 (2.5)	25.5 (6.7)
Completeness (%)	99.9 (100.0)	94.3 (70.3)	93.6 (79.8)
Redundancy	5.6 (5.6)	5.7 (4.0)	5.9 (5.5)
Refinement			
Resolution (Å)	38.7-1.40	24.1-1.19	21.2-1.27
No. reflections	26,187	48,029	33,012
<i>R</i> _{work} / <i>R</i> _{free}	0.146/0.176	0.130/0.146	0.145/0.170
R.m.s deviations			
Bond lengths (Å)	0.004	0.004	0.012
Bond angles (°)	1.0	1.0	1.5
Most favored region (%) ^b	99.2	98.3	96.7
Additional allowed region	0.8	1.7	3.3
Disallowed region (%) ^b	0.0	0.0	0.0

^aHighest resolution shell is shown in parenthesis.

^bIn the Ramachandran plot

Table S4: Single concentration binding assays and *K*_d measurements for PLX51107 against 32 bromodomains.

Domain ^a	Bromodomain Protein Name	%Inhibition at 1 μM	<i>K</i> _d (nM)
BRD2(1)	bromodomain-containing protein 2, bromodomain 1	100	1.6
BRD2(2)	bromodomain-containing protein 2, bromodomain 2	100	5.9
BRD3(1)	bromodomain-containing protein 3, bromodomain 1	100	2.1
BRD3(2)	bromodomain-containing protein 3, bromodomain 2	100	6.2
BRD4(1)	bromodomain-containing protein 4, bromodomain 1	100	1.7
BRD4(2)	bromodomain-containing protein 4, bromodomain 2	100	6.1
BRDT(1)	bromodomain testis-specific protein, bromodomain 1	90	5.0
BRDT(2)	bromodomain testis-specific protein, bromodomain 2	97	120
CREBBP	CREB binding protein	100	110
EP300	E1A binding protein p300	100	130
TAF1(2)	TATA box binding protein (TBP)-associated factor 1, bromodomain 2	48	2,200
SMARCA4	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily a, member 4	60	>10,000
CECR2	cat eye syndrome chromosome region, candidate 2	37	NA

Domain ^a	Bromodomain Protein Name	%Inhibition at 1 μ M	K _d (nM)
WDR9(2)	bromodomain and WD repeat domain containing 1, bromodomain 2	36	NA
GCN5L2	GCN5 like 2 or K(lysine) acetyltransferase 2A (KAT2A)	30	NA
BAZ2A	bromodomain adjacent to zinc finger domain, 2A	29	NA
BRD9	bromodomain-containing protein 9	28	NA
PBRM1(5)	polybromo 1, bromodomain 5	25	NA
TRIM24	tripartite motif containing 24 (PHD+bromodomain)	23	NA
BAZ2B	bromodomain adjacent to zinc finger domain, 2B	23	NA
TRIM33	tripartite motif containing 33 (PHD+bromodomain)	15	NA
SMARCA2	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily a, member 2	11	NA
BRPF3	bromodomain and PHD finger containing, 3	10	NA
BRD7	bromodomain containing 7	10	NA
PCAF	P300/CBP-associated factor or K(lysine) acetyltransferase 2B (KAT2B)	6	NA
PBRM1(2)	polybromo 1, bromodomain 2	0	NA
ATAD2A	ATPase family AAA domain containing 2	0	NA
ATAD2B	ATPase family AAA domain containing 2B	0	NA
BRD1	bromodomain-containing protein 1	0	NA
BRPF1	bromodomain and PHD finger containing, 1	0	NA
BPTF	bromodomain PHD finger transcription factor, also known as FALZ	0	NA
TAF1L(2)	TATA box binding protein (TBP)-associated factor 1L, bromodomain 2	0	NA

^a Each assay used a truncated protein that includes a single bromodomain (value in parentheses indicates which bromodomain is used when the full length protein contains two bromodomains; 1 stands for BD1 and 2 stands for BD2). ^b Single concentration (at 1 μ M) primary screening data for all 32 bromodomains are presented as percentage of inhibition. ^c K_d values for 12 bromodomains that exhibited >50% inhibition in the primary screen. Assays that showed <50% inhibition at 1 μ M were not pursued for K_d determination (NA)

Table S5: Profiling PLX51107 in Genscript Leukemia and Lymphoma panel reveals broad activity in various malignant cell lines.

Cell Line	Histologic Type	IC ₅₀ (μ M)	95% CI (μ M)
CCRF-CEM	Leukemia, acute lymphoblastic	0.020	0.015 - 0.027
RPMI 8226	Myeloma, plasmacytoma	0.020	0.015 - 0.027
MV4-11	Leukemia, biphenotypic B myelomonocytic	0.031	0.029 - 0.032
8E5	Leukemia, acute lymphoblastic	0.051	0.032 - 0.081
SC	Monocyte/Macrophage	0.056	0.036 - 0.088
KE-37	Leukemia, T cell	0.067	0.054 - 0.083
HuT 78	Lymphoma	0.067	0.050 - 0.090
Loucy	Leukemia, acute lymphoblastic t(16;20)	0.084	0.067 - 0.11

Cell Line	Histologic Type	IC ₅₀ (μ M)	95% CI (μ M)
	translocation		
BDCM	Leukemia, acute myelogenous	0.084	0.055 - 0.13
RL	Lymphoma, non-Hodgkin's	0.091	0.071 - 0.12
J.RT3-T3.5	Leukemia, acute T cell	0.10	0.081 - 0.13
CA46	Lymphoma, Burkitt's	0.11	0.076 - 0.16
BC-1	Lymphoma, EBV and KSHV positive	0.11	0.068 - 0.19
DOHH2	Lymphoma	0.12	0.096 - 0.14
EOL-1	Lymphoma	0.12	0.092 - 0.16
CEM/C2	Leukemia, acute lymphoblastic	0.14	0.11 - 0.18
K562	Leukemia, myelogenous	0.15	0.10 - 0.23
MOLT-4	Leukemia, acute lymphoblastic	0.16	0.089 - 0.27
P3HR-1	Lymphoma, Burkitt's	0.16	0.13 - 0.21
Kasumi-1	Leukemia, acute myeloblastic	0.18	0.14 - 0.23
MPC-11	Myeloma, Mouse	0.19	0.14 - 0.26
H9	Lymphoma, cutaneous	0.22	0.20 - 0.24
Mo	Leukemia, hairy cell	0.23	0.16 - 0.32
CEM/C3	Leukemia, acute lymphoblastic	0.24	0.18 - 0.31
MJ	Lymphoma, cutaneous T cell, mycosis fungoides	0.24	0.18 - 0.32
JVM-2	Leukemia	0.24	0.15 - 0.40
HH	Lymphoma, cutaneous T cell	0.25	0.15 - 0.39
TF-1	Erythroleukemia	0.26	0.16 - 0.43
KU812	Leukemia, chronic myelogenous	0.27	0.19 - 0.36
AHH-1	Lymphoblastoid	0.30	0.22 - 0.41
GDM-1	Leukemia, myelomonoblastic	0.34	0.23 - 0.48
MOLT-3	Leukemia, acute lymphoblastic	0.39	0.21 - 0.73
KG-1	Leukemia, acute lymphoblastic	0.39	0.22 - 0.70
RPMI 7666	Lymphoblast	0.43	0.25 - 0.74
Daudi	Lymphoma, Burkitt's	0.45	0.37 - 0.54
CESS	Leukemia, myelomonocytic	0.49	0.29 - 0.84
HL-60 clone15	Leukemia, Acute promyelocytic	0.49	0.43 - 0.57
GK-5	B lymphoblast; Epstein-Barr virus (EBV) transformed	0.53	0.30 - 0.91
ARH-77	Leukemia, plasma cell	0.57	0.27 - 1.20

Cell Line	Histologic Type	IC ₅₀ (μ M)	95% CI (μ M)
CEM/C1	Leukemia, acute lymphoblastic	0.58	0.39 - 0.84
HL-60	Leukemia, promyelocytic	0.61	0.54 - 0.68
D1.1	Leukemia, acute T cell, CD4 negative	0.62	0.25 - 1.52
P116	Leukemia, acute T cell	0.62	0.37 - 1.03
SUP-B15	Leukemia, acute lymphoblastic	0.63	0.38 - 1.04
Jurkat	Leukemia, acute T cell	0.67	0.42 - 1.08
KU 812 E	Leukemia, chronic myelogenous	0.73	0.63 - 0.84
CCRF-HSB-2	Leukemia, acute lymphoblastic	0.82	0.52 - 1.28
WIL2-S	B lymphoblast, hereditary spherocytosis, spleen	0.93	0.63 - 1.39
MC116	Lymphoma, undifferentiated	0.95	0.48 - 1.88
J45.01	Leukemia, acute T cell, CD45 deficient	0.99	0.70 - 1.40
P116.cl39	Leukemia, acute T cell	1.88	0.80 - 4.41
Ramos.2G6.4C10	Lymphoma, Burkitt's	1.91	0.96 - 3.83
RS4;11	Leukemia, acute lymphoblastic, t(4;11) translocation	2.13	0.73 - 6.16
CML-T1	Leukemia, T-lymphocyte, chronic myelogenous	~2.5 ^a	NA
SU-DHL-6	Lymphoblast-like, peritoneal effusion	2.86	0.86 - 9.46
I 9.2	Leukemia, acute T cell	2.88	0.94 - 8.77
TALL-1	Lymphoma, T cell	3.30	1.60 - 6.84
SUP-T1	Leukemia, lymphoblastic	3.70	1.60 - 8.56
Raji	Lymphoma, Burkitt's	4.00	2.45 - 6.51
ST486	Lymphoma, Burkitt's	5.19	3.00 - 8.99
U266B1	Myeloma; plasmacytoma	6.59	3.56 - 12.20
U-937	Lymphoma, histiocytic	7.58	5.42 - 10.62
AML-193	Leukemia, Acute monocytic	8.21	4.84 - 13.94
Toledo	Lymphoma, diffuse large cell, non-Hodgkin's B cell	12.06	2.25 - 64.51
NAMALWA	Lymphoblastoid	13.57	9.47 - 19.44
EB-1	Lymphoma, Burkitt's	>20	NA
EB-2	Lymphoma, Burkitt's	>20	NA
GA-10	Lymphoma, Burkitt's	>20	NA
NCI-H929	Myeloma, plasmacytoma	>20	NA

^a Estimated value from incomplete curve.

Table S6A. The top 20 canonical pathways identified by IPA for 1361 differentially expressed genes upon BRD4 inhibition in microarray data of CpG-stimulated B-CLL cells (fold change > 2 and $p < 0.01$).

Ingenuity Canonical Pathways	p-value	Molecules
B Cell Receptor Signaling	6.30957E-12	MAP2K6, RAP2B, RAP2A, POU2F2, NFATC3, PIK3R1, SOS2, PDPK1, PTPRC, PAX5, IKBKB, PIK3CG, CD22, IRS2, RASSF5, GSK3B, MAP2K1, PPP3CA, MAP3K9, MAP3K14, NRAS, CSK, FGFR1, EGR1, MAPK8, IKBKE, MALT1, BCL2L1, CALM1, SYNJ1, FOXO1, SYK, VAV3, IGHG4, MEF2C, MAP2K3, MAP3K3
CD28 Signaling in T Helper Cells	5.24807E-08	FYN, NFATC3, CSK, FGFR1, PIK3R1, CHP1, MAPK8, PDPK1, IKBKE, HLA-DQB1, MALT1, ITPR1, CTLA4, PTPRC, IKBKB, CALM1, CD80, PIK3CG, SYK, HLA-DMB, HLA-DOB, CD86, IRS2, MAP2K1, PPP3CA
T Cell Receptor Signaling	1.34896E-07	FYN, NRAS, NFATC3, CSK, FGFR1, PIK3R1, SOS2, MAPK8, IKBKE, MALT1, CTLA4, PTPRC, TEC, CALM1, IKBKB, TXK, VAV3, RASGRP1, PIK3CG, IRS2, MAP2K1, PPP3CA
PKCθ Signaling in T Lymphocytes	8.31764E-07	FYN, MAP3K14, MAP3K9, NRAS, NFATC3, FGFR1, PIK3R1, SOS2, CHP1, MAPK8, IKBKE, MALT1, HLA-DQB1, IKBKB, CD80, VAV3, PIK3CG, HLA-DMB, CD86, HLA-DOB, IRS2, MAP3K3, PPP3CA
PI3K Signaling in B Lymphocytes	2.0893E-06	CD81, FYN, NRAS, NFATC3, PIK3R1, CHP1, PDPK1, IKBKE, ITPR1, MALT1, PTPRC, BLK, CALM1, IKBKB, CD180, VAV3, PIK3CG, SYK, FOXO3, IRS2, MAP2K1, PPP3CA
Th1 and Th2 Activation Pathway	2.13796E-06	MAP2K6, TNFSF4, ICAM1, NFATC3, CHD4, PIK3R1, HLA-DQB1, NFIL3, NOTCH2, TGFB1, PIK3CG, HLA-DMB, IRS2, STAT1, STAT6, IL10, mir-29, IKZF1, FGFR1, CD80, LTA, IL10RA, S1PR1, CD86, HLA-DOB, JAK3, NOTCH1
CD27 Signaling in Lymphocytes	2.39883E-06	MAP2K6, MAP3K14, MAP3K9, APAF1, MAPK8, IKBKE, IKBKB, BCL2L1, BID, MAP2K3, CD27, MAP2K1, MAP3K3
Regulation of IL-2 Expression in Activated and Anergic T Lymphocytes	4.89779E-06	FYN, IKBKB, CALM1, NRAS, CD80, NFATC3, TGFB1, VAV3, CHP1, SOS2, MAPK8, TOB1, IKBKE, MALT1, MAP2K1, PPP3CA
Th1 Pathway	1.07152E-05	MAP2K6, ICAM1, IL10, mir-29, NFATC3, FGFR1, PIK3R1, HLA-DQB1, NFIL3, NOTCH2, CD80, LTA, PIK3CG, HLA-DMB, IL10RA, CD86, HLA-DOB, IRS2, STAT1, JAK3, NOTCH1
CD40 Signaling	1.34896E-05	MAP2K6, MAP3K14, ICAM1, FGFR1, PIK3R1, MAPK8, IKBKE, IKBKB, LTA, PIK3CG, IRS2, MAP2K3, JAK3, MAP2K1, FCER2
Germ Cell-Sertoli Cell Junction Signaling	2.23872E-05	MAP2K6, TUBA1B, MAP3K14, MAP3K9, NRAS, TUBB4B, FGFR1, PIK3R1, TUBB2A, MAPK8, TUBA4A, PDPK1, TUBA1A, SORBS1, TGFB1, PIK3CG, MAP2K3, IRS2, MTMR2, ACTN4, VCL, MAP3K3, ACTG1, MAP2K1
Death Receptor Signaling	2.63027E-05	MAP3K14, MAPK8, APAF1, ZC3HAV1, PARP8, IKBKE, PARP9, IKBKB, PARP15, TIPARP, BID, SPTAN1, CFLAR, MAP4K4, ACTG1, PARP14
NRF2-mediated Oxidative Stress Response	3.71535E-05	MAP2K6, NRAS, FGFR1, NQO2, PIK3R1, MAPK8, DNAJC3, GCLC, DNAJC1, SOD1, CLPP, FTL, PIK3CG, VCP, CAT, IRS2, MAP2K3, JUND, DNAJB1, GSK3B, FKBP5, MAP2K1, ENC1, ACTG1, DNAJC7
Docosahexaenoic Acid (DHA) Signaling	4.57088E-05	BCL2L1, FOXO1, PIK3CG, PIK3R1, FGFR1, APAF1, BID, PDPK1, IRS2, GSK3B, PNPLA2,

		APP
Granzyme A Signaling	5.01187E-05	SET, ANP32A, HIST1H1C, NME1, HIST1H1E, H1FX, HMGB2
Altered T Cell and B Cell Signaling in Rheumatoid Arthritis	7.76247E-05	MAP3K14, SLAMF1, IL10, LTB, HLA-DQB1, TLR10, CD80, TGFB1, LTA, TLR1, HLA-DMB, TLR6, CD86, HLA-DOB, TNFRSF13B
Role of NFAT in Regulation of the Immune Response	8.51138E-05	FYN, NRAS, NFATC3, FGFR1, PIK3R1, SOS2, CHP1, IKBKE, ITPR1, HLA-DQB1, GNAI2, CALM1, IKBKB, CD80, SYK, PIK3CG, HLA-DMB, CD86, HLA-DOB, MEF2C, IRS2, GSK3B, MAP2K1, PPP3CA
IL-4 Signaling	8.91251E-05	STAT6, NRAS, IL13RA1, NFATC3, FGFR1, PIK3R1, SOS2, HLA-DQB1, SYNJ1, PIK3CG, HLA-DMB, HLA-DOB, IRS2, JAK3, FCER2
Glucocorticoid Receptor Signaling	0.000102329	ICAM1, POU2F2, NFATC3, PIK3R1, HSPA1A/HSPA1B, SOS2, GTF2A1, IKBKB, GTF2B, POLR2A, TGFB1, PIK3CG, FOXO3, IRS2, FKBP5, STAT1, MAP2K1, POLR2L, PPP3CA, MAP3K14, NRAS, IL10, FGFR1, CHP1, MAPK8, IKBKE, BCL2L1, TAF5, SMARCA2, PRKACA, JAK3, HLTF
UVA-Induced MAPK Signaling	0.000109648	NRAS, FGFR1, PIK3R1, MAPK8, ZC3HAV1, PARP8, SMPD1, RPS6KA5, PARP9, BCL2L1, PARP15, TIPARP, PIK3CG, IRS2, STAT1, PARP14

Table S6B. The top 20 upstream regulators predicted by IPA to be inhibited or activated for 1361 differentially expressed genes upon BRD4 inhibition in microarray data of CpG-stimulated B-CLL cells (fold change > 2 and $p < 0.01$).

Upstream Regulator	Molecule Type	Predicted Activation State	Activation z-score	p-value of overlap	Target molecules in dataset
IFNG	cytokine	Inhibited	-5.484	0.0000114	ABCB1, ABCD3, ALOX5AP, APOL6, APP, ARL6IP5, ATP1A1, BCL2L1, BID, BST2, CASP4, CAT, CCL4, CD200, CD80, CD86, CERS6, CFLAR, CIRBP, CLEC2D, CSK, DDIAS, DTX3L, EGR1, EIF2AK2, ERAP2, ERCC8, FCER2, FKBP5, FOXO1, FTL, GART, GBP3, GBP4, GCH1, GLIPR2, GLS, GLUL, GNAI2, HDAC9, HERC6, HLA-DMB, HLA-DOB, HLA-DQB1, HSPA1A/HSPA1B, ICAM1, IER3, IFI16, IFI30, IFIH1, IFIT5, IFITM2, IKBKE, IL10, IL10RA, IRF2, IRF5, IRF7, IRS2, ISG20, ITGAL, ITPR1, JAK3, JUND, KARS, KLF2, KMO, KYNU, LAT2, LPL, LTA, LTB, MAN2A1, MAP2K1, MARCKSL1, MNDA, NCF2, NF1, NMI, NOD2, NOTCH1, OAS2, OAS3, OPN3, P2RY14, PARP9, PECAM1, PLEK, RAD18, RAP2B, RFX5, RTP4, SAMD9, SAMHD1, SELL, SELP, SLAMF1, SLC11A2, SLC12A2, SP110, STAT1, STAT6, TGFB1, TLR1, TLR6, TMEM50B, TRIM21, TXK, WARS, ZFP36

IFNA2	cytokine	Inhibited	-3.899	0.000000866	APOL6, BST2, CCDC92, CD69, CD86, DDX60, EIF2AK2, FAM65B, GBP4, HERC6, HSH2D, IFI16, IFIH1, IFIT5, IFITM2, IL10, IRF5, IRF7, ISG20, LILRB2, LILRB4, MAP2K1, OAS2, OAS3, PARP9, SAMD9, SAMHD1, SP110, STAT1, TNFRSF10D, TRIM21, XAF1, ZC3HAV1
IFNL1	cytokine	Inhibited	-3.628	0.00000059	APOL6, BST2, CD80, DDX60, EIF2AK2, HERC6, IFIH1, IFIT5, IL10, ISG20, OAS2, OAS3, SAMD9, SP110, STAT1, TMEM140, ZC3HAV1
IFNB1	cytokine	Inhibited	-3.486	0.00697	ANXA5, BCL2L1, BST2, BTN3A3, CCL4, CD80, CD86, EIF2AK2, GBP3, GBP4, HMGB2, IFI16, IFIH1, IL10, IRF7, ISG20, NMI, NOD2, NOTCH1, OAS2, RNASEL, STAT1, TNFSF4, TRIM21, VCL, XAF1
TNF	cytokine	Inhibited	-3.412	0.00000104	ABCD2, ACOX1, AIMP1, ALCAM, ALOX5AP, APAF1, APP, ARID5B, ARL6IP5, ARDC3, ATP1A1, ATP2B4, BCKDHB, BCL2L1, BID, BST2, BTN3A3, CALR, CASP4, CAT, CCL4, CD5, CD69, CD80, CD86, CERS6, CFLAR, CNR2, COTL1, CPT1A, CRY1, CTLA4, DAG1, DDIAS, DENND2D, DGAT1, DUSP14, DUSP16, EGLN1, EGR1, EIF2AK2, ELK3, ETV5, ETV6, FCER2, FGFR1, FYN, GADD45A, GADD45B, GBP3, GCH1, GCLC, GKAP1, GLS, GM2A, GNAI2, GPX4, GSK3B, HDAC9, HERC1, HEXB, HIVEP1, HSPA1A/HSPA1B, ICAM1, IER3, IFI16, IFIH1, IFIT5, IKBKE, IL10, IL10RA, IRF5, IRF7, IRS2, ITGAL, ITPR1, JUND, KLF2, KMO, KYNU, L3MBTL3, LDHA, LFNG, LPL, LSS, LTB, MAP2K3, MAP2K6, MAP3K14, MAP4K4, MARCKSL1, MEF2C, MPC1, NCF2, NME1, NOD2, NOTCH1, NR4A3, NRARP, NRROS, OAS2, OAS3, PARP14, PCNA, PDE7B, PDIA4, PECAM1, PIK3CG, PILRB, PLK2, PTPRC, QKI, RANBP9, RAPGEF5, RFTN1, RFX5, RGS2, RND1, SAMD9, SAT1, SCD, SELL, SELP, SIRT1, SKI, SLC11A2, SLC35B2, SLC7A1, SMPD1, SOD1, SORBS1, ST3GAL5, ST6GAL1, ST8SIA4, STAT1, STMN1, TERF2IP, TGFB1, TNFRSF10D, TNFSF9, TP53INP1, TSHZ1, UCP2, VAV3, VCL, VIM, WNT10A, ZFP36
IRF7	transcripti on regulator	Inhibited	-3.297	0.0000139	BCL2L1, CASP4, CD69, CD80, CTLA4, GBP3, GBP4, IFI16, IFIH1, IFITM2, IRF7, ISG20, NMI, OAS2, OAS3, PARP14, PLAC8, RTP4, SAMD9L, SAP30, STAT1, TRIM21, XAF1, ZC3HAV1
BRD4	kinase	Inhibited	-3.202	0.000111	ACSL5, ACSM3, ADAT2, BCL2L1, DCPS, DCTPP1, GTF3C6, KCNA3, MTHFD1L, MTMR2, NME1, RRS1, SFXN4, TTC27, ZNF485

Interferon alpha	group	Inhibited	-3.097	0.000406	ANXA5, APOL3, BCL2L1, BST2, CD69, CD80, CD81, CD86, DAG1, EIF2AK2, GBP3, GLS, ICAM1, IFI16, IFIH1, IFITM2, IFNLR1, IKBKE, IL10, IL10RA, IRF5, IRF7, ISG20, LILRB4, mir-29, MND4, NFIL3, NMI, OAS2, PNPT1, PRKACA, RNASEL, STAT1, TGFB1, TLR1, TRIM21, WARS, ZC3HAV1
EGLN	group	Inhibited	-3.039	0.00254	ALDOC, APAF1, BNIP3, EGLN1, FAM117B, FTL, FYN, GAPDH, GYS1, LDHA, SAP30, TPI1, ZNF337
CD2	transmembrane receptor	Inhibited	-2.975	0.000508	CD48, CD80, CD86, ICAM1, IL10, ITGAL, PTPRC, SELL, STAT1
lfn	group	Inhibited	-2.934	0.000143	CD69, CD80, CD86, EIF2AK2, ICAM1, IFI16, IFIH1, IL10, IL10RA, IRF7, ISG20, OAS2, OAS3, PECAM1, RNASEL, STAT1, TLR1, TRIM21
STAT1	transcription regulator	Inhibited	-2.884	0.085	APOL6, BCL2L1, CASP4, CCL4, CD86, EGR1, EIF2AK2, FOXO1, GBP3, HERC6, ICAM1, IFI16, IFITM2, IL10, IRF5, IRF7, OAS2, PARP9, PECAM1, SAMHD1, SP110, STAT1, WARS
CD40LG	cytokine	Inhibited	-2.812	0.000000694	ALCAM, ALG3, BATF, BCL2L1, BNIP3, CCL4, CD27, CD69, CD80, CD86, CELF2, CFLAR, EGR1, EIF2AK2, GADD45A, GCH1, GPR183, HDDC2, HLA-DQB1, HSPA1A/HSPA1B, ICAM1, IL10, IL10RA, IL13RA1, IL1RAP, IPO8, IRF2, JUND, LTA, LTB, MAP2K3, MARCKSL1, MPHOSPH9, NR4A3, PLEK, RAD50, RGS2, SELL, SELP, SLAMF1, STAT1, TM7SF2, TNFRSF13B, TNFSF4, TNFSF9, TNPO1, ZFP36L1
MGEA5	enzyme	Inhibited	-2.785	0.025	ACTG1, ACTN4, ALDOC, ALOX5AP, BCL2L1, CD48, CD86, CDCA7L, CREB3L2, FGFR1, FLNA, FLNB, G6PD, GADD45A, GLUL, GTF3A, HSDL1, LSS, MAP3K3, NOTCH2, OGDH, PIK3R1, RAP2A, SORBS1, STAT1, TGFB1, TSPAN5, TUBA1A, TUBA4A, VIM
FLT3LG	cytokine	Inhibited	-2.773	0.00861	ADGRE5, CD80, CD86, ICAM1, IL10, PAX5, PTPRC, TCF4
lfnar	group	Inhibited	-2.64	0.00346	CD86, EIF2AK2, IFI16, IFIH1, IRF7, ISG20, OAS2, PNPT1, STAT1, TRIM21, UNC93B1, XAF1
RBL2	other	Inhibited	-2.598	0.11	CASP4, CCNF, CHKA, GSK3B, KPNA2, MAP2K3, MAP3K14, PCNA, STMN1
EBI3	cytokine	Inhibited	-2.596	0.0163	CD80, CD86, HLA-DMB, HLA-DOB, ICAM1, IL10, STAT1
IL12 (complex)	complex	Inhibited	-2.589	0.0248	BCL2L1, CCL4, CD69, CD86, GADD45B, GCNT2, ICAM1, IFIH1, IL10, ISG20, ITGAL, LTA, LTB, PLAC8, SELL, SELP, SET, TGFB1, TXK
IRF3	transcription regulator	Inhibited	-2.534	0.000648	B4GALT5, BTLA, CCL4, CD69, CD86, DDX60, EIF2AK2, IFI16, IFIH1, IL10, IRF5, IRF7, ISG20,

					OAS2, OAS3, PARP14, PLAC8, PNP, SAMD9L, SAP30, STAT1, TNFSF4, VIM
IL5	cytokine	Activated	2.453	0.000000184	ALDOC, ANXA2, BCL2L1, BNIP3, BNIP3L, CASP4, CD69, CYSLTR1, DDX21, EAF2, EGLN1, EGR1, ELL2, FAM65B, GADD45A, GBP4, GLIPR2, HSPH1, ICAM1, IER3, IFI30, IL13RA1, IRF7, LFNG, LTA, LTB, NABP1, NDRG1, NFIL3, P4HA1, PAX5, PTPRC, QSOX1, SAP30, TGFB1, TLR1, TPI1, VIM, ZNF443
FOXO1	transcription regulator	Activated	2.477	0.143	CCNE2, CCNF, CDKN3, HIST1H2BG, LDHA, MAPK8, PCNA, PECAM1, STMN1, VIM
Nr1h	group	Activated	2.517	0.00957	ABCD2, ERCC8, HLA-DOB, IFIH1, IL10, ITGAL, ITPR1, LAT2, LPL, LTB, MARCKSL1, MYLIP, NPC1, SCD, SELP, SP110
EGR2	transcription regulator	Activated	2.534	0.0446	BNIP3L, CHKA, EGR1, FCER2, GOLGB1, GTF2A1, HOMER1, IL10, MAP2, NOTCH1, RALGAPA1, SCD, SLC12A2, SLC16A1
ARNT	transcription regulator	Activated	2.538	0.00063	ALDOC, BNIP3, CD81, G6PD, GAPDH, IL10, IRS2, LDHA, MYO1C, OAZ1, PDPK1, PGM2, S1PR1, SELL, TPI1, TUBA4A, VIM
SMAD4	transcription regulator	Activated	2.597	0.519	CELF2, GADD45A, GADD45B, ICAM1, IER3, IL10, PTPRC, SCD, SERTAD1, SLC25A4, TGFB1, VIM, ZFP36
TRIM24	transcription regulator	Activated	2.685	0.000297	DDX60, EPSTI1, GBP3, GBP4, GLUL, HERC6, HNF1B, IFIH1, IRF7, NMI, PLAC8, RTP4, SAMD9L, SAMHD1, STAT1
CREM	transcription regulator	Activated	2.724	0.0467	CD86, DUSP14, EGR1, GADD45B, IRS2, LSS, MEST, MIDN, MKNK2, NFIL3, NOTCH1, SERTAD1, SLC16A1, TIPARP
ERBB2	kinase	Activated	2.802	0.00104	ABHD5, ALDOC, ANXA2, APLP2, ATP1B3, BAG2, BCKDHB, BCL2L1, BNIP3, CCL4, CCNE2, CDCA7L, CDKN3, CHD4, DAAM1, DAG1, DDX10, DGAT1, DTX1, EGR1, ELK3, ELL2, EPSTI1, ETV5, ETV6, FOXO1, G3BP1, GART, GLS, GSK3B, GTF3A, HIST1H4C, HMGB2, HSPA1A/HSPA1B, IGF2R, IKBKB, LPCAT1, MAN1A1, MAP2K3, MGAT5, MKNK2, MLXIP, MPHOSPH9, NDRG1, NOTCH1, PCNA, PDIA4, PDPK1, PFKFB3, PLAC8, POLE, POLK, POLR1B, POLR2A, POLR2L, QKI, SIRPA, SLC4A7, SORBS1, TUBA1A, VCL, VIM, WSB2
E2F1	transcription regulator	Activated	2.842	1.08E-08	ABC1, ACSL5, APAF1, ATAD2, BCL2L1, BID, BNIP3, CALM1 (includes others), CALR, CCNE2, CCNF, CFLAR, CWC27, EGR1, EIF2AK2, EXOSC9, EZH2, FANCD2, FGFR1, FOXO1, FOXO3, GM2A, HIST1H2AC, HIST1H2AE, HIST1H2BJ, HIST1H4H,

					HIST2H2AA3/HIST2H2AA4, HLTF, HMGB2, HN1, ICAM1, IRS2, KCNA3, LTA4H, MAP3K14, MGEA5, mir-223, MTHFD1, NMI, NUSAP1, PCNA, PIK3R1, PKNOX1, PLK2, PPP1R13B, QKI, RCOR1, RYBP, SERTAD2, STMN1, TGFB1, TNFSF9, TP53INP1, TRIM28, TRMT13, UCP2, VCP, VIM, YARS, ZFP36
XBP1	transcription regulator	Activated	2.852	0.181	APP, CALR, CAT, CRK, DNAJC1, DNAJC3, ERO1B, GOLGB1, ICAM1, MAP1LC3B, P3H1, PDIA4, S1PR1, SOD1, STARD5
MKNK1	kinase	Activated	2.887	0.0112	ANXA2, ANXA5, FLNA, FLNB, IFRD1, KIF1B, MAN2A1, NR4A3, PDLIM5, STXBP1, TTC3, VIM
NFE2L2	transcription regulator	Activated	2.899	0.000941	ABCB4, ACSL5, ACTG1, ARHGEF3, ATP1A1, BCL2L1, BNIP3, CAT, CD86, CLPP, CREG1, DNAJC3, DYNLL1, ESD, ETV6, FKBP5, FOXO3, FTL, G6PD, GCLC, GNAI2, IFRD1, IL10, LPL, MAP1LC3B, MEF2C, NCF2, OAT, PDIA4, PGD, PKIA, PTPRO, RRS1, SAT1, SLC30A4, SOD1, TGFB1, TPI1, VCP
ESRRA	ligand-dependent nuclear receptor	Activated	2.941	0.189	ACOX1, ALDOC, ASAH1, CHKA, FAM102A, GAPDH, IKZF3, LDHA, LPL, MED24, PDP1, SIRT1, TPI1
GH1	growth factor	Activated	3.077	0.387	CAT, EGR1, GADD45A, GCLC, LPL, NRAS, PCNA, SIRT1, SKI, SOD1, TUBA1A, VIM, XRCC1, ZFP36
IL1RN	cytokine	Activated	3.2	0.0174	ERAP2, HDAC9, HERC6, HLA-DQB1, ICAM1, IFIH1, IFIT5, IL10, IRF7, ISG20, OAS2, OAS3, RTP4, SAMD9
PPARG	ligand-dependent nuclear receptor	Activated	3.209	0.0592	ACOX1, ACSL5, APAF1, APP, ASAP1, BCL2L1, CAT, CPT1A, DGAT1, EGLN1, GAPDH, ICAM1, IL10, IRS2, LPL, MANBA, MAP2, MKNK2, MRTO4, NDRG1, PCNA, PECAM1, PFKFB3, RAPGEF1, SAT1, SCD, SLC44A1, SLC9A1, SOD1, SORBS1, TCF4, UCP2, VIM, WIPF1, XYLT1
CD38	enzyme	Activated	3.229	0.0000025	ALDOC, ANXA2, BNIP3, BNIP3L, CD86, EAF2, EGLN1, ELL2, GBP4, GLIPR2, ICAM1, IFI30, LFNG, LTA, NABP1, NDRG1, NFIL3, P4HA1, PAX5, PPP3CA, SAP30, SELL, TPI1, VIM, ZNF443
PPARGC1A	transcription regulator	Activated	3.608	0.276	ACOX1, BNIP3, CALM1 (includes others), CAT, CPT1A, DGAT1, GSK3B, IL10, IRS2, LDHA, LPL, NRF1, PNPLA2, SCD, SOD1, UCP2
FOXO3	transcription regulator	Activated	3.684	0.000000703	APAF1, BNIP3, BNIP3L, CASP4, CAT, CCNE2, CDKN3, CFLAR, CPT1A, EGR1, FOXO1, FOXO3, GADD45A, GADD45B, GCLC, GLUL, IARS, IER3, IFIH1, IL10, IMPDH2, IRS2, LARS, MEF2C, MXD1, PCNA, PPRC1, RTN3, RTN4, SESN1, SLC7A1, SOD1, TGFB1, TP53INP1,

					UCP2, VIM
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Table S7A. The top 20 canonical pathways identified by IPA for 142 genes contained within SEs that are ranked lower in 3 out of 4 CLL patient samples after *ex-vivo* PLX51107 treatment.

Ingenuity Canonical Pathways	p-value	Molecules
Th1 and Th2 Activation Pathway	6.30957E-07	IL10, IKZF1, IL10RA, HLA-DQA1, IL2RA, SPI1, IL24, mir-21, IL2RB
Th2 Pathway	1.31826E-06	IL10, IKZF1, HLA-DQA1, IL2RA, SPI1, IL24, mir-21, IL2RB
T Helper Cell Differentiation	4.46684E-05	IL10, IL10RA, HLA-DQA1, IL2RA, BCL6
Molecular Mechanisms of Cancer	0.00019	ZBTB17, RAC2, RAPGEF1, PMAIP1, TCF4, CCND2, PRKAR1B, RHOH, BCL2
Antigen Presentation Pathway	0.00105	B2M, NLRC5, HLA-DQA1
Leukocyte Extravasation Signaling	0.00575	RAC2, CD44, RASSF5, RHOH, SELPLG
Th1 Pathway	0.00589	IL10, IL10RA, HLA-DQA1, mir-21
Ovarian Cancer Signaling	0.00741	TCF4, CD44, PRKAR1B, BCL2
Allograft Rejection Signaling	0.01023	B2M, IL10, HLA-DQA1
OX40 Signaling Pathway	0.01175	B2M, HLA-DQA1, BCL2
Cytotoxic T Lymphocyte-mediated Apoptosis of Target Cells	0.01230	B2M, BCL2
Death Receptor Signaling	0.01259	PARP15, PARP9, BCL2
Hepatic Fibrosis / Hepatic Stellate Cell Activation	0.01738	IL10, IL10RA, ECE1, BCL2
Protein Kinase A Signaling	0.01995	TCF4, PTPRJ, PTPN1, PRKAR1B, UBASH3B, PDE4B
p53 Signaling	0.02089	PMAIP1, CCND2, BCL2
IL-8 Signaling	0.02188	RAC2, CCND2, RHOH, BCL2
Netrin Signaling	0.02239	RAC2, PRKAR1B
Neuroprotective Role of THOP1 in Alzheimer's Disease	0.02455	PRKAR1B, ECE1
Autoimmune Thyroid Disease Signaling	0.02754	IL10, HLA-DQA1
iCOS-iCOSL Signaling in T Helper Cells	0.02951	HLA-DQA1, IL2RA, IL2RB

Table S7B. The top upstream regulators (activation z-score>2) predicted by IPA for 142 genes contained within SEs that are ranked lower in 3 out of 4 CLL patient samples after *ex-vivo* PLX51107 treatment

Upstream Regulator	Molecule Type	Predicted Activation State	Activation z-score	p-value of overlap	Target molecules in dataset
STAT3	transcription regulator	Inhibited	-2.047	0.0000002	BATF, BCL2, BCL6, BMF, CCND2, HLA-DQA1, IKZF3, IL10, IL2RA, IL2RB, mir-21, PMAIP1, SERPINB1, SERPINB9, TCF4
CD3	complex	Activated	2.097	0.0000003	BATF, BCL2, BCL6, CCND2, CD44, CKS2, DAD1, IL10, IL2RA, IL2RB, PDE4B, PTPRJ, RAC2, RHOH, SELPLG, SRP14
BCL6	transcription regulator	Activated	2.308	0.0000016	BCL2, BCL6, CCND2, CD44, IL10, IL24, IL2RA, IL2RB, LPP
IL21	cytokine	Inhibited	-2.574	0.0000043	BATF, BCL2, BCL6, CCND2, IL10, IL2RA, IL2RB, mir-21
IL4	cytokine	Inhibited	-2.111	0.0000338	BCL11A, BCL2, BCL6, CCND2, CD44, IL10, IL10RA, IL24, IL2RA, IL2RB, mir-21, PDE4B, PIGR, SELPLG, ST6GAL1
CSF2	cytokine	Inhibited	-2.941	0.000222	BCL2, IL10, IL24, IL2RA, mir-21, PDE4B, RHOH, SERPINB9, SPI1, TCF4
STAT5B	transcription regulator	Inhibited	-2.191	0.000275	BCL2, BCL6, CCND2, IL24, IL2RA, IL2RB

CBFB	transcription regulator	Activated	2	0.000675	IKZF3, IL2RA, IL2RB, SLC25A19
Nr1h	group	Activated	2	0.00871	BCL2, CCND2, ECE1, IL10
IL1	group	Inhibited	-2.412	0.0115	BCL2, IL10, IL2RA, IL2RB, mir-21, PIGR
ERBB2	kinase	Inhibited	-2.371	0.021	BCL2, BTG2, CCND2, CDC42EP3, CKS2, ETV6, LPCAT1, mir-21, PTPN1
Creb	group	Inhibited	-2.195	0.024	BCL2, DDX6, IL10, mir-21, PDE4B
MGEA5	enzyme	Inhibited	-2	0.137	BCL2, CD44, PRKAR1B, SERPINB9
NUPR1	transcription regulator	Activated	2	0.286	ETV6, JADE2, PARP9, PTPRJ

Table S7C. The top bio-functions (activation z-score >2) predicted by IPA for 142 genes contained within SEs that are ranked lower in 3 out of 4 CLL patient samples after *ex-vivo* PLX51107 treatment.

Diseases or Functions Annotation	Predicted Activation State	Activation z-score	p-Value	Molecules
leukopoiesis	Decreased	-2.964	7.25E-14	ARID5B, B2M, BATF, BCL11A, BCL2, BCL6, CCND2, CD44, EIF2AK4, ETV6, HLA-DQA1, IGKC, IKZF1, IKZF3, IL10, IL10RA, IL2RA, IL2RB, LAIR1, MAPKAPK2, mir-21, PTPRJ, RAC2, RASSF5, RHOH, SLAMF6, SMARCE1, SPEN, SPI1, TCF4, ZBTB17
hematopoiesis of mononuclear leukocytes	Decreased	-2.576	3.94E-13	ARID5B, B2M, BATF, BCL11A, BCL2, BCL6, CCND2, CD44, EIF2AK4, ETV6, HLA-DQA1, IGKC, IKZF1, IKZF3, IL10, IL10RA, IL2RA, IL2RB, LAIR1, PTPRJ, RASSF5, RHOH, SLAMF6, SMARCE1, SPEN, SPI1, TCF4, ZBTB17
lymphopoiesis	Decreased	-2.744	6.21E-13	ARID5B, B2M, BATF, BCL11A, BCL2, BCL6, CCND2, CD44, EIF2AK4, ETV6, HLA-DQA1, IGKC, IKZF1, IKZF3, IL10, IL10RA, IL2RA, IL2RB, PTPRJ, RASSF5, RHOH, SLAMF6, SMARCE1, SPEN, SPI1, TCF4, ZBTB17
dendropoiesis	Decreased	-2.007	1.05E-09	BATF, BCL11A, BCL6, EIF2AK4, IKZF1, IL10, IL2RA, IL2RB, LAIR1, mir-21, SPI1
development of antigen presenting cells	Decreased	-2.378	1.2E-09	BATF, BCL11A, BCL6, EIF2AK4, IKZF1, IL10, IL2RA, IL2RB, LAIR1, MAPKAPK2, mir-21, SPI1
homeostasis of leukocytes	Decreased	-3.057	2.01E-09	B2M, BATF, BCL11A, BCL2, BCL6, BMF, EIF2AK4, HLA-DQA1, IGKC, IKZF1, IL10, IL10RA, IL2RA, IL2RB, PDE4B, RHOH, SLAMF6, SMARCE1, SPI1, TCF4, ZBTB17
differentiation of T lymphocytes	Decreased	-3.169	2.35E-09	B2M, BATF, BCL11A, BCL2, BCL6, EIF2AK4, HLA-DQA1, IKZF1, IL10, IL2RA, IL2RB, RHOH, SLAMF6, SMARCE1, SPI1, TCF4, ZBTB17
T cell homeostasis	Decreased	-2.784	3.88E-09	B2M, BATF, BCL11A, BCL2, BCL6, BMF, EIF2AK4, HLA-DQA1, IGKC, IKZF1, IL10, IL10RA, IL2RA, IL2RB, RHOH, SLAMF6, SMARCE1, SPI1, TCF4, ZBTB17
development of phagocytes	Decreased	-2.706	7.34E-09	BATF, BCL11A, BCL2, BCL6, EIF2AK4, IKZF1, IL10, IL2RA, IL2RB, LAIR1, MAPKAPK2, mir-21, SPI1
hematopoiesis of phagocytes	Decreased	-2.378	8.56E-09	BATF, BCL11A, BCL2, BCL6, EIF2AK4, IKZF1, IL10, IL2RA, IL2RB, LAIR1, mir-21, SPI1
T cell development	Decreased	-2.598	1.59E-08	B2M, BATF, BCL11A, BCL2, BCL6, EIF2AK4, HLA-DQA1, IGKC, IKZF1, IL10, IL10RA, IL2RA, IL2RB, RHOH, SLAMF6, SMARCE1, SPI1, TCF4, ZBTB17
differentiation of antigen	Decreased	-2.033	1.93E-08	BATF, BCL11A, BCL2, BCL6, EIF2AK4, IKZF1,

presenting cells				IL10, IL2RA, IL2RB, LAIR1, mir-21, SPI1
cell proliferation of T lymphocytes	Increased	3.125	2.59E-08	ADORA2A, B2M, BATF, BCL2, CD44, HLA-DQA1, IKZF1, IKZF3, IL10, IL2RA, IL2RB, LAIR1, PTPRJ, RAC2, RASSF5, RHOH, SERPINB9, SPI1, TIGIT, ZC3H12D
anemia	Increased	2.188	0.00000018	ADORA2A, BCL2, CCND2, CDC42EP3, DAD1, GRHPR, HLA-DQA1, IKZF1, IL10, IL2RA, IL2RB, PTPRJ, RAC2, SLC25A19, SPI1
hypoplasia of lymphoid organ	Increased	2.949	0.00000048	ARID5B, BCL11A, BCL6, IGKC, IKZF1, IL10, IL2RB, RASSF5, RHOH
abdominal neoplasm	Increased	2.126	0.00000212	ADORA2A, APOBEC3D, APOBEC3F, APOBEC3H, ARID5B, B2M, BATF, BCL11A, BCL2, BCL6, BTG2, C1orf186, C1QTNF6, CBX7, CCND2, CD44, CDC42EP3, CHIT1, CKS2, CORO1C, CYTOR, DAD1, DDX6, DTX3L, ECE1, EIF2AK4, EIF2D, ETV6, FAM167A, FAM53B, FAM60A, FGD2, GPR107, GRHPR, HERPUD1, HLA-DQA1, IGHA2, IGKC, IKZF1, IL10, IL10RA, IL24, IL2RA, IL2RB, JAZF1, KLHDC7B, LDLRAD4, LILRA4, LILRA5, LPCAT1, LPP, MAPKAPK2, MCTP2, MIF4GD, mir-21, MSI2, MTSS1, NLRC5, NSMCE1, NUBP1, PARP15, PATL2, PDE4B, PIGR, PLEKHG1, PMAIP1, PPP1R16B, PRKAR1B, PTPN1, PTPRJ, RAD51B, RAPGEF1, RASSF5, RBM17, RHOH, SECISBP2, SELPLG, SERPINB1, SERPINB9, SLAMF6, SLC39A13, SMAP2, SPEN, SPI1, SSTR3, ST6GAL1, SUSD1, TAGAP, TCF4, TCTN1, TIGIT, TRIM69, TVP23A, UBASH3B, USP3, VMP1, VOPP1, XXYLT1, ZBTB17, ZBTB20, ZC3H12D, ZCCHC7, ZMYND8
hypoplasia of organ	Increased	3.216	0.00000247	ARID5B, BCL11A, BCL2, BCL6, CCND2, ECE1, IGKC, IKZF1, IL10, IL2RB, PTPRJ, RASSF5, RHOH, SPEN
morbidity or mortality	Increased	3.531	0.0000037	ADORA2A, ARID5B, B2M, BCL11A, BCL2, BCL6, BMF, CCND2, CD44, CKS2, DAD1, ECE1, EIF2AK4, ETV6, FCMR, GPR107, IKZF3, IL10, IL2RA, IL2RB, MAPKAPK2, mir-21, MTSS1, NUBP1, PDE4B, PTPN1, PTPRJ, RAC2, RAD51B, RAPGEF1, RASSF5, SECISBP2, SERPINB1, SLAMF6, SLC25A19, SPEN, SPI1, TCF4, ZBTB17, ZBTB20
cellular homeostasis	Decreased	-2.981	0.00000421	ADORA2A, B2M, BATF, BCL11A, BCL2, BCL6, BMF, EIF2AK4, HERPUD1, HLA-DQA1, IGKC, IKZF1, IL10, IL10RA, IL24, IL2RA, IL2RB, NUBP1, PDE4B, PMAIP1, PTPRJ, RAC2, RHOH, SLAMF6, SLC39A13, SMARCE1, SPI1, TCF4, UBASH3B, VMP1, ZBTB17
quantity of plasma cells	Increased	2.219	0.0000257	FCMR, IL10, IL2RB, RAC2, RASSF5

Table S8A. The top 20 canonical pathways identified by IPA for 106 genes that are directly regulated by BRD4.

Ingenuity Canonical Pathways	p-value	Molecules
tRNA Charging	0.00047	LARS, GARS, LARS2, IARS
Th2 Pathway	0.00052	STAT6, IKZF1, HLA-DQB1, JAK3, mir-21
IL-4 Signaling	0.00069	STAT6, NRAS, HLA-DQB1, JAK3
Th1 and Th2 Activation Pathway	0.00138	STAT6, IKZF1, HLA-DQB1, JAK3, mir-21

Role of JAK1 and JAK3 in γ c Cytokine Signaling	0.00372	STAT6, NRAS, JAK3
TREM1 Signaling	0.00457	TLR6, TLR1, CCL3
IL-15 Signaling	0.00468	STAT6, NRAS, JAK3
Colorectal Cancer Metastasis Signaling	0.00501	NRAS, GRK3, TLR6, TLR1, JAK3
JAK/Stat Signaling	0.00575	STAT6, NRAS, JAK3
Communication between Innate and Adaptive Immune Cells	0.00724	TLR6, TLR1, CCL3
Altered T Cell and B Cell Signaling in Rheumatoid Arthritis	0.00724	TLR6, TLR1, HLA-DQB1
Oncostatin M Signaling	0.00977	NRAS, JAK3
UVA-Induced MAPK Signaling	0.01047	NRAS, PARP15, PARP9
Primary Immunodeficiency Signaling	0.01862	RFX5, JAK3
Th1 Pathway	0.02188	HLA-DQB1, JAK3, mir-21
Role of Pattern Recognition Receptors in Recognition of Bacteria and Viruses	0.02239	OAS2, TLR6, TLR1
Pyrimidine Ribonucleotides Interconversion	0.02344	NME1, CTPS2
Retinoic acid Mediated Apoptosis Signaling	0.03236	PARP15, PARP9
IL-2 Signaling	0.03388	NRAS, JAK3
Pyrimidine Ribonucleotides De Novo Biosynthesis	0.03467	NME1, CTPS2

Table S8B. The top upstream regulators (activation z-score >2) predicted by IPA for 106 genes that are directly regulated by BRD4.

Upstream Regulator	Molecule Type	Predicted Activation State	Activation z-score	p-value of overlap	Target molecules in dataset
Interferon alpha	group	Inhibited	-2.513	0.000379	CCL3, CCND2, IFI16, LILRB4, mir-21, MX2, OAS2, TLR1
IFN Beta	group	Inhibited	-2.164	0.000398	CCL3, IFI16, MX2, OAS2, XAF1
BCR (complex)	complex	Inhibited	-2.18	0.00044	CCL3, CCND2, CLEC2D, FCMR, KCNN4
STAT1	transcription regulator	Inhibited	-2.433	0.00008	APOL6, CCL3, CCND2, HERC6, IFI16, OAS2, PARP9, SP110
PRL	cytokine	Inhibited	-2.828	0.0000485	DTX3L, EPSTI1, HERC6, MX2, OAS2, SP110, TMEM140, XAF1
VEGFA	growth factor	Inhibited	-2.186	0.0101	BATF, CTLA4, MRPL3, NME1, TLR1
IFNL1	cytokine	Inhibited	-2.449	0.00000125	APOL6, DDX60, HERC6, OAS2, SP110, TMEM140
IL21	cytokine	Inhibited	-2.414	0.000147	BATF, CCND2, HERC6, IFI16, mir-21, OAS2
CSF2	cytokine	Inhibited	-2.771	0.00145	CCL3, CD180, CTLA4, GRK3, HLA-DQB1, mir-21, TIFA, TLR1
TNF	cytokine	Inhibited	-2.761	0.00797	AIMP1, ARID5B, BTN3A3, CCL3, CCND2, CTLA4, DENND2D, ELK3, ETV6, IFI16, mir-21, NCF2, NME1, OAS2, RFX5, TIFA
TGM2	enzyme	Inhibited	-2.607	0.0000901	CCL3, DDX60, NCF2, OAS2, PARP9, SP110, XAF1
IRF7	transcription regulator	Inhibited	-2.2	0.00096	CTLA4, IFI16, MX2, OAS2, XAF1
MSC	transcription regulator	Inhibited	-2	0.000137	DDX60, EPSTI1, PAX5, XAF1
ATF4	transcription regulator	Inhibited	-2.236	0.000767	ARHGAP24, BCAT1, GARS, IARS, LARS
IFNB1	cytokine	Inhibited	-2.401	0.00179	BTN3A3, CCL3, IFI16, mir-21, OAS2, XAF1
IFNG	cytokine	Inhibited	-3.874	0.00000733	APOL6, CCL3, CCND2, CLEC2D, DTX3L, ERCC8, HERC6, HLA-DQB1, IFI16, JAK3, MX2, NCF2, OAS2, PARP9, RFX5, SP110, STAT6, TLR1, TLR6
IFNA2	cytokine	Inhibited	-3.231	1.06E-08	APOL6, CCL3, DDX60, HERC6, IFI16,

					LILRB4, MX2, OAS2, PARP9, SP110, XAF1
MYC	transcription regulator	Inhibited	-2.329	0.0117	ABCE1, BCAT1, CCND2, CD48, DDX21, IFI16, IKZF1, NME1, PAX5, PNO1, TMEM126A
CD3	complex	Activated	3.031	0.000173	AIMP1, BATF, CCL3, CCND2, CTLA4, ELK3, HLA-DQB1, JAK3, KCNN4, NME1, XAF1
NUPR1	transcription regulator	Activated	2.449	0.0288	CTPS2, ETV6, LARS2, MX2, PARP9, RFX5
SOCS1	other	Activated	2	0.00257	CCL3, IFI16, OAS2, STAT6
NKX2-3	transcription regulator	Activated	2	0.0181	DDX60, PARP9, SP110, XAF1
IL1RN	cytokine	Activated	2	0.00339	HERC6, HLA-DQB1, MX2, OAS2

Table S8C. The top bio-functions (activation z-score >2) predicted by IPA for 106 genes that are directly regulated by BRD4.

Diseases or Functions Annotation	Predicted Activation State	Activation z-score	p-Value	Molecules
cell survival	Decreased	-3.891	0.000161	ACSL5, AIMP1, ALKBH8, AMIGO2, CCL3, CD48, CTLA4, ERCC8, ETV6, FANCF, FCMR, JAK3, mir-21, NCF2, NME1, NRAS, PAX5, PFDN1, STAT6, UPF2, UTP15, XRCC4
cell viability	Decreased	-3.75	0.000171	ACSL5, AIMP1, ALKBH8, AMIGO2, CCL3, CD48, CTLA4, ERCC8, ETV6, FANCF, FCMR, JAK3, mir-21, NCF2, NME1, NRAS, PAX5, PFDN1, UPF2, UTP15, XRCC4
migration of cells	Decreased	-2.468	0.00946	AIMP1, ARHGAP24, ARID5B, BATF, BCAT1, CCL3, CD48, CTLA4, DOCK10, ELK3, ETV6, IKZF1, JAK3, KCNN4, mir-21, NCF2, NME1, NRAS, PARP9, PAX5, SH2D3C, STAT6
leukopoiesis	Decreased	-2.439	0.000000196	ARID5B, BATF, CCL3, CCND2, CTLA4, ETV6, HLA-DQB1, IFI16, IKZF1, IKZF3, JAK3, LILRB1, LILRB4, mir-21, PAX5, SLAMF6, STAT6, TLR1, TLR6, XRCC4
Lymphocyte migration	Decreased	-2.333	0.00128	BATF, CCL3, CD48, CTLA4, ETV6, JAK3, KCNN4, SH2D3C
hematopoiesis of mononuclear leukocytes	Decreased	-2.291	8.12E-08	ARID5B, BATF, CCL3, CCND2, CTLA4, ETV6, HLA-DQB1, IFI16, IKZF1, IKZF3, JAK3, LILRB1, LILRB4, PAX5, SLAMF6, STAT6, TLR1, TLR6, XRCC4
cell proliferation of tumor cell lines	Decreased	-2.222	0.00602	ACSL5, CCL3, CCND2, CTLA4, CWC27, DDX21, ETV6, IFI16, IKZF1, JAK3, KCNN4, LILRB1, mir-21, NAT10, NME1, NRAS, PAX5, PPIL1, SH2D3C, SP110
tumorigenesis of tissue	Decreased	-2.204	0.00122	ABCE1, ALKBH8, AMIGO2, ARHGAP24, ARID5B, BATF, BCAT1, BTN3A3, C3orf38, CCND2, CD180, CD48, CLEC2D, CTLA4, CTPS2, CWC27, DDX21, DDX60, DENND2D, DOCK10, DTX3L, ELK3, EPSTI1, ERCC8, ETV6, FANCF, FCRLA, FYCO1, GARS, GPATCH4, GTF3C6, HERC6, HLA-DQB1, IARS, IFI16, IKZF1, JAK3, KCNN4, KIAA0391, LARS, LARS2, LILRA4, LILRB1, LILRB4, LRRC32, MCTP2, METTL3, mir-21, MTX3, MX2, NAT10, NCF2, NME1, NRAS, NUP205, OAS2, PARP15, PFDN1, PIGK, PPIL1, PYROXD1, RFX5, SH2D3C, SLAMF6, SLC43A3, SMYD5, SNX11, SP110, STAT6, TIFA, TIGIT, TLR1, TLR6,

				TMEM126A, TMEM140, TRIT1, TXLNB, UPF2, XAF1, ZC3H12D, ZC3H7A, ZNF484
Bacterial Infections	Increased	2	0.00453	AIMP1, CCL3, FCMR, SLAMF6, SP110, STAT6, TLR1, TLR6
hypoplasia of lymphoid organ	Increased	2.165	0.00118	ARID5B, CTLA4, IKZF1, JAK3, XRCC4