TOP3B isoform 1 TOP3B isoform 2 TOP3B isoform 3	MKTVLMVAEKPSLAQSIAKILSRGSLSSHKGLNGACSVHEYTGTFAGQPVRFKMTSVCGH
TOP3B isoform 1 TOP3B isoform 2 TOP3B isoform 3	VMTLDFLGKYNKWDKVDPAELFSQAPTEKKEANPKLNMVKFLQVEGRGCDYIVLWLDCDK
TOP3B isoform 1 TOP3B isoform 2 TOP3B isoform 3	EGENICFEVLDAVLPVMNKAHGGEKTVFRARFSSITDTDICNAMACLGEPDHNEALSVDA
TOP3B isoform 1	RQELDLRIGCAFTRFQTKYFQGKYGDLDSSLISFGPCQTPTLGFCVERHDKIQSFKPETY
TOP3B isoform 2	RQELDLRIGCAFTRFQTKYFQGKYGDLDSSLISFGPCQTPTLGFCVERHDKIQSFKPETY
TOP3B isoform 3	RQELDLRIGCAFTRFQTKYFQGKYGDLDSSLISFGPCQTPTLGFCVERHDKIQSFKPETY
TOP3B isoform 1	WVLQAKVNTDKDRSLLLDWDRVRVFDREIAQMFLNMTKLEKEAQVEATSRKEKAKQRPLA
TOP3B isoform 2	WVLQAKVNTDKDRSLLLDWDRVRVFDREIAQMFLNMTKLEKEAQVEATSRKEKAKQRPLA
TOP3B isoform 3	WVLQAKVNTDKDRSLLLDWDRVRVFDREIAQMFLNMTKLEKEAQVEATSRKEKAKQRPLA
TOP3B isoform 1	LNTVEMLRVASSSLGMGPQHAMQTAERLYTQGYISYPRTETTHYPENFDLKGSLRQQANH
TOP3B isoform 2	LNTVEMLRVASSSLGMGPQHAMQTAERLYTQGYISYPRTETTHYPENFDLKGSLRQQANH
TOP3B isoform 3	LNTVEMLRVASSSLGMGPQHAMQTAERLYTQGYISYPRTETTHYPENFDLKGSLRQQANH
TOP3B isoform 1	PYWADTVKRLLAEGINRPRKGHDAGDHPPITPMKSATEAELGGDAWRLYEYITRHFIATV
TOP3B isoform 2	PYWADTVKRLLAEGINRPRKGHDAGDHPPITPMKSATEAELGGDAWRLYEYITRHFIATV
TOP3B isoform 3	PYWADTVKRLLAEGINRPRKGHDAGDHPPITPMKSATEAELGGDAWRLYEYITRHFIATV
TOP3B isoform 1	SHDCKYLQSTISFRIGPELFTCSGKTVLSPGFTEVMPWQSVPLEESLPTCQRGDAFPVGE
TOP3B isoform 2	SHDCKYLQSTISFRIGPELFTCSGKTVLSPGFTEVMPWQSVPLEESLPTCQRGDAFPVGE
TOP3B isoform 3	SHDCKYLQSTISFRIGPELFTCSGKTVLSPGFTEVMPWQSVPLEESLPTCQRGDAFPVGE
TOP3B isoform 1	VKMLEKQTNPPDYLTEAELITLMEKHGIGTDASIPVHINNICQRNYVTVESGRRLKPTNL
TOP3B isoform 2	VKMLEKQTNPPDYLTEAELITLMEKHGIGTDASIPVHINNICQRNYVTVESGRRLKPTNL
TOP3B isoform 3	VKMLEKQTNPPDYLTEAELITLMEKHGIGTDASIPVHINNICQRNYVTVESGRRLKPTNL
TOP3B isoform 1	GIVLVHGYYKIDAELVLPTIRSAVEKQLNLIAQGKADYRQVLGHTLDVFKRKFHYFVDSI
TOP3B isoform 2	GIVLVHGYYKIDAELVLPTIRSAVEKQLNLIAQGKADYRQVLGHTLDVFKRKFHYFVDSI
TOP3B isoform 3	GIVLVHGYYKIDAELVLPTIRSAVEKQLNLIAQGKADYRQVLGHTLDVFKRKFHYFVDSI
TOP3B isoform 1	AGMDELMEVSFSPLAATGKPLSRCGKCHRFMKYIQAKPSRLHCSHCDETYTLPQNGTIKL
TOP3B isoform 2	AGMDELMEVSFSPLAATGKPLSRCGKCHRFMKYIQAKPSRLHCSHCDETYTLPQNGTIKL
TOP3B isoform 3	AGMDELMEVSFSPLAATGKPLSRCGKCHRFMKYIQAKPSRLHCSHCDETYTLPQNGTIKL
TOP3B isoform 1	YKELRCPLDDFELVLWSSGSRGKSYPLCPYCYNHPPFRDMKKG <mark>MGCNECTHPSCQHSLSM</mark>
TOP3B isoform 2	YKELRCPLDDFELVLWSSGSRGKSYPLCPYCYNHPPFRDMKKG <mark>MGCNECTHPSCQHSLSM</mark>
TOP3B isoform 3	YKELRCPLDDFELVLWSSGSRGKSYPLCPYCYNHPPFRDMKKG <u>ECSHSL</u>
TOP3B isoform 1	LGIGQCVECESGVLVLDPTSGPKWKVACNKCNVVAHCFENAHRVRVSADTCSVCEAALLD
TOP3B isoform 2	LGIGQCVECESGVLVLDPTSGPKWKVACNKCNVVAHCFENAHRVRVSADTCSVCEAALLD
TOP3B isoform 3	LSTGSCSLFSVPTPALHQA-GL
TOP3B isoform 1 TOP3B isoform 2 TOP3B isoform 3	VDFNKAKSPLPGDETQHMGCVFCDPVFQELVELKHAASCHPMHRGGPGRRQGRGRGRARR VDFNKAKSPLPGDETQHMGCVFCDPVFQELVELKHAASCHPMHRGGPGRRQGRGRARR
TOP3B isoform 1	PPGKPNPRRPKDKMSALAAYFV
TOP3B isoform 2	PPGKPNPRRPKDKMSALAAYFV
TOP3B isoform 3	

CLUSTAL O(1.2.4) multiple sequence alignment

Isoform 1 (NP_001269041.1) Isoform 2 (NP_001336777.1) Isoform 3 (NP_001336780.1)

Supplementary Figure 1. Amino acid sequence alignment of three TOP3B isoforms. Clustal Omega alignment was performed using protein sequences of human TOP3B isoforms. The N- and C-terminus of three isoforms are highlighted in blue and red.



Supplementary Figure 2. Expression analysis of TOP3B isoform1 and isoform3 in mouse tissues and human cancer cell lines. (A) The mRNA expression of TOP3B isoform 1 and isoform 3 in mouse tissues were examined by RT-qPCR using isoform-specific primers. (B) The relative expression of isoform 3 compared to isoform 1 was examined in mouse tissues and human cancer cell lines. * 293T is isolated from human embryonic kidneys. (C) Detection of isoform 1 and isoform 3 in several human cancer cell lines. The cells were transfected with control siRNA or TOP3B specific siRNA. Western blot was performed using anti-TOP3B mouse monoclonal antibody. The protein expression of isoform 3 can be detected in all the cell lines tested.



Supplementary Figure 3. Recombinant PRMTs used in the *in vitro* methylation assays are active and TOP3B C-terminal domain is not methylated by PRMT5. (A) *In vitro* methylation was performed on a known arginine methylated protein REF/ALY to confirm that recombinant PRMT1, PRMT3 and PRMT6 are active. (B) Recombinant CARM1 methylates a known substrate PABP1. (C) Myc-tag PRMT5 was purified from 293T cells and incubated with HeLa cell core histone in an *in vitro* methylation reaction. PRMT5 preferentially methylates H4. H3 and H2A are also methylated but to a lesser extent. (D) *In vitro* methylation was performed by incubating Myc-PRMT5 with TOP3B C-terminal domain (708-862). For the same exposure time as histone substrates (one week), no obvious methylation signal was detected.



Supplementary Figure 4. Assess the specificity of MMA and ADMA antibodies. Dot-blot assays were performed to assess the specificity of pan-MMA and pan-ADMA antibodies using synthetic biotinylated Histone H4 peptides (N-terminus 22 amino acids) carrying defined methylation at arginine 3 site: monomethylation (H4R3me1), asymmetrical dimethylation (H4R3me2a), and symmetrical dimethylation (H4R3me2s). The loading of the peptides was examined using anti-Biotin antibody.



Supplementary Figure 5. Inhibition of protein arginine methylation reduces TOP3B stress granule localization. HeLa cells cultured on glass cover slides were untreated or treated with MS023 for 2 days before the cells were treated with 0.5 mM Arsenite for 1 hour. The samples were processed as described in Figure 5 (A), except that anti-TOP3B and anti-PABP1 (marker for stress granules) antibodies were used to examine the endogenous protein localization.



Supplementary Figure 6. Confirmation of Tudor domain functional mutation and GFP-TDRD3 subcellular localization. (A) GST-tag recombinant Tudor domain and methylargininebinding deficient Tudor (E691K) were incubated with HeLa cell lysates. After pull-down, the samples were subjected to western blot detection using ADMA antibody. (B) Both GFP-TDRD3 and GFP-TDRD3 (E691K) were transiently transfected into HeLa cells. The subcellular localization of both proteins was visualized by fluorescence microscope.



Supplementary Figure 7. The interaction between TDRD3 OB-fold and TOP3B catalytic domain is not sufficient to mediate their respective recruitment to SGs. Three TOP3B truncations were expressed as GFP-fusion proteins. After Arsenite treatment, the co-localization of GFP-fusion proteins with TDRD3 (Red) was examined by immunofluorescence. Note that TOP3B (125-625), which is sufficient to interact with TDRD3, doesn't exhibit strong stress granule localization.

Supplementary Table 1 (RT-qPCR primers)

NRAS	CTACAGGGAGCAGATTAAGCG
	TAACTCTTGGCCAGTTCGTG
DDX5	TGATTTGGAGAGAGGTGTGG
	TTCAAAGCCCATATCAAGCA
c-MYC	TTCTCTCCGTCCTCGGATTCTCTG
	TCTTCTTGTTCCTCCTCAGAGTCG
Human TOP3B isoform 1	CGTCCTTGGCACAGTCAATTG
	ATCTTGAAGCGCACTGGCTGG
Human TOP3B isoform 3	GTCCTGTGGTCATCAGGCTCT
	GACTGCAGCTACCTGTGGAC
Mouse Top3b isoform 1	GCAACACCTGCGAGGCTGCC
	GCTGCATGCTTAAGCTCCACCA
Mouse Top3b isoform 3	GAACTGGTCCTGTGGTCCTC
	CTCTCAGCTCACCTGTTACTTG

Supplementary Table 2 (DRIP-qPCR primers)

pFC53 R loop fragment	TTTAGAGCTTGACGGGGAAA
	CAACAGTTGCGTAGCCTGAA
c-MYC	GAGGCTATTCTGCCCATTTG
	GGTGCTTACCTGGTTTTCCA
DDX5	GTGTCATCGGTGTCCTTCCT
	ACTCGAATAACCCGACATGG
NRAS	CGTTTCACTGATGCCAGAAA
	TCCTTCCCATTCTCCCTTCT

Supplementary Table 3 (TOP3B site mutagenesis primers)

R824K	ctgccaccccatgcac AAA ggtggaccagggagaag
	cttctccctggtccaccTTTgtgcatggggtggcag
R829/830K	cgcggtggaccaggg AAAAAG cagggtcgagggcgg
	ccgccctcgaccctgCTTTTTccctggtccaccgcg
R833/835K	gggagaaggcagggt AAAgggAAG ggccgggccaggagg
	cctcctggcccggccCTTcccTTTaccctgccttctccc
R837/839/840K	ggtcgagggcggggc AAGgccAAGAAG ccccctgggaagccc
	gggcttcccagggggCTTCTTggcCTTgccccgccctcgacc
R848/849K	gggaagcccaacccc AAAAAG cccaaggacaagatg
	catcttgtccttgggCTTTTTggggttgggcttccc