

## Supplementary Information

### Analysis of DNAs associated with coconut foliar decay disease implicates a unique single-stranded DNA virus representing a new taxon

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**Figures S1, S2, S3, S4; Tables S1, S2, S3, S4, S5, S6, S7**

Figure S1

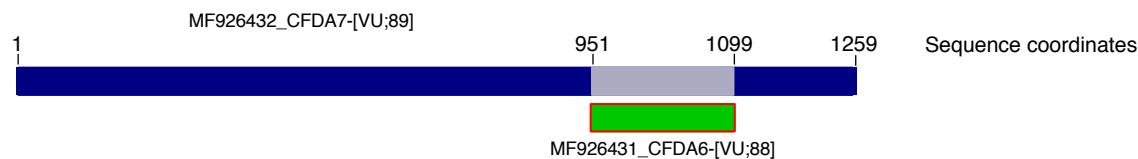
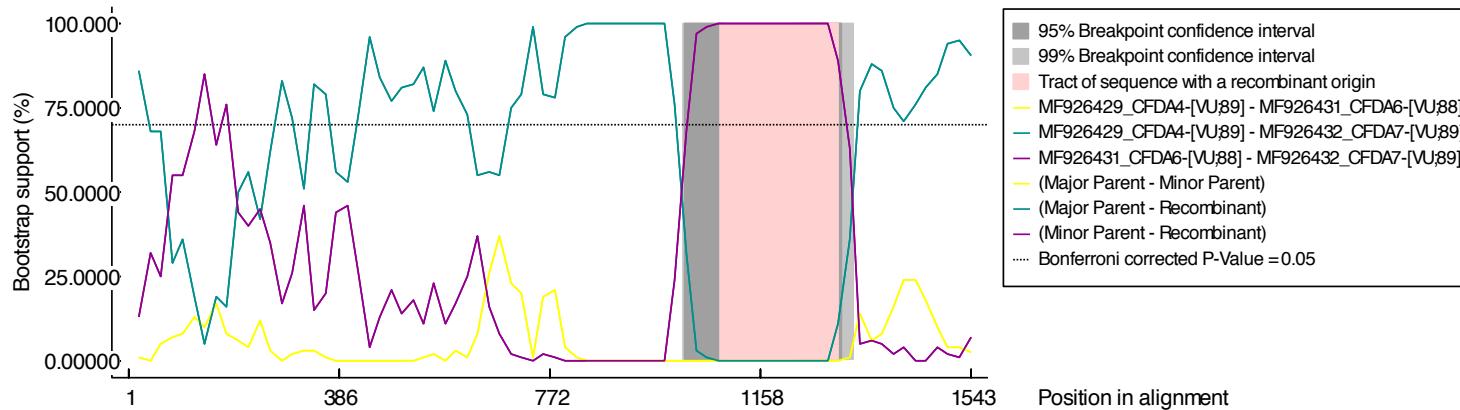
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	Color-coded Domains																				
CFDV-[VU;89]_alphasat_R_Rep	--MAS--SRRWCFTLNFS-AXEP--LSPLSENE-VSYAVWQDEXAPTTQRHLQGYQLKL-KRT-LGGXKLFGG--AHLEKARGTD-EARDYCMKEETRVSFGEPCPSGSHKR---KLMERVIRSPEEMXXEDPSTYRRCAKL--AEEEFXK--EEIWPXXLKPWQLE																				
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CFDV-[VU;89]_alphasat_2_Rep	--MGSS--IRRWCFTLNYSAAAAASLVRRIEISLN-LYYAIVGDEVAPSTGQRHLQGYFIHLK-TGRRRLQGLKTVLGNDRHLEPLTRGSD-EQNRYDCSKE-RVLL--EHGVPTPGVKRP---RLAQRFAEEDPDELRLEDPGYRRCVHG--ASVEWTRAAENPFPFPYHNWQLEV	163																			
CFDV-[VU;89]_alphasat_4_Rep	--MAAR--VRRWCFTLNYSTEEEAAARVERVKSLT-TTYCIVGDEVAPTTQHHLQGYFIHLK-SARCLQGLKTFLQNDKVLHLEAAKGS-EQNRYVCSKE-QIRY--EHGVATRPGSKRR---LEQRFDEDPADLRLLEPGYRRCVVAH--ASWEVYGWALEHPFPHPYHWQLE	162																			
CFDV-[VU;89]_alphasat_8_Rep	--MTQR-CRRWCFTLNYSDEAAAALVRRVQSLSL-TYAIVGKETAPTTQKHLOQFLHLKGGRVLQGLKTVLQNDRHLQAHGS-EQNRYVCSKE-ETIF--EHGIPTRPGVKRR---LVDRFDDPDELRLEDPGYRRCVALR--AAVEWAGWASENPFPFPYHHWQLE	163																			
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CFDV-[VU;89]_alphasat_3_Rep	--MSSR-XRRWCFTLNFSYRSEEAAQVQRIESLD-PTYAIVGDEVAPTTQGHLOQFLHLKGGRVLQGLKTVLQNDRHLQAHGS-EQNRYCSKE-QRFE--EHGATRAGCKR---LLEERAEDPDPLLELLEDPGYRRCVAH--HAVEREWATARDPYPYLHDWQLE	163																			
CFDV-[VU;89]_alphasat_6_Rep	-MAMVS--RRWCFTLNFSYRSEEERTRLLSLFSEE-LHYAIVGDEVAPSTGQRHLQGYLNR-KVRLRALKKY-SDKAHWEIAKGD-EENRVCSCHE-HKFV--ELGSPVVGSNKR---KLAEAIERSPERMRLQEPEIFHRYASAK--KMIOFK--EQYDHPVFDRTWQIKL	159																			
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BBTV_alphaSat_HaiKou2_Rep_AEF97834	--MSS--FKWCFTLNSSAEEAERDFLSRLKEEDD-VYYAVVGDEVAPSQQKHLQGYLSK-KSMKLGLLKKRY-SSKAHWEKARGTD-KENSKYCSKE-TLIL--EIGFPATQGSNKR---KLSEMVSRSPEMRLEQEPEIYHRYLSV--KLKKFK--EEFVHPCLERPWQIQL	157																			
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CBDV_alphaSat_Fr_X7_Rep_YP_008854133	--MAS--FKWCFTLNSSAEEAERDFLALLKEEE-LRYAVVGDEVAPTTQGHLOQFLSL-KLIRLGLLKKR-SPKAHWEIAKGD-EDNAKYCSKE-TLIL--ELGFPASQGSNKR---KLAEEAIERSPERMRLQEPEIFHRYASAK--KMIOFK--EQYDHPVFDTWTQIKL	159																			
BBTV_alphaSat_4_Rep_ACB86656	--MASA--SRWTFTLHYSATERGKFLATLKEEDD-VHYAVVGDETAPNTGRKHLQGYLSK-KRFRISGIKKKY-SSRAHWEKARGSD-YDNKAYCSKE-ALIL--ELGVPCTGSNKR---KLADMVTRSPERMIEQEPEIFHRYASVK--KMKEFK--ERYVYPILDRPWQVQL	157																			
BBTV_alphaSat_TW3S2_Rep_ACJ36782	--MSS--FKWCFTLNSSAEEAERDFLALLKEEDD-VHYAVVGDEVAPTTQGHLOQFLSL-KSIRLGLLKKY-GSRAHWEIAKGD-EDNAKYCSKE-TLIL--ELGTPVPGPSKRR---KLLDRFRE SPEELKMDPDSKYRCLAVE--SIKDAR--NNSEWVHELKEWQNLK	157																			
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SCSV_alphaSat_C2_Rep_AAA68018	--MAS--RRYCFNLNYATEIERETFLSLFSQDE-LNYFVVGDETA-TTGQKHLQGFVFSK-NKIRLGLLKKF-SWRAVEWARTGTD-GENMRCYCSKE-TLIL--ELGIPATPGPSNKR---KYAEMTRSPERMIEQEPEIFHRYDSVN--CMQFK--QEYVPCLERPWQIQL	157																			
FBNSV_alphaSat_1_Rep_AHC72219	--MAC--ANWVFTRFNQGPALP----SLSFDERVQYAWQHERG--THDHQIVGVIQLK-KKARFSTVKEII-GGNPHEVKMGTI-EEASAYVQEKEETRVAvgpwSYDQLLKRGSHT--KTMERYLDEPEEMLKLDPDTALRCKAKK--LKEDYC--SCFSSFKLDRWPQIQL	152																			
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FBNY_alphaSat_C7_EF93_Rep_CAB44025	--MPSI--TRASHWCFTLNFS-GS1PE--IN--WTADQVYSIWQHERV--HDHQYIQLQMKHVT-LKKMKELLPG--AHLEMAKAP--KKAIECYQKKEASIAQWPEIYGTWISGSHKR--KLMERFEDPEEMKLEDPLGPRYRCLSRV--QMCKIRES--CTWNFD-LRPWPQDEL	154																			
MDV_alphaSat_C10_Rep_BAA34048	--MPSIRAIHWCFNLNS-GK1PE--IV--WTADQVYSIWQHERV--HDHQYIQLQMKHVT-LKKMKELLPG--AHLEMARAP--KKAIDCYQKKEATAIDGPWEYGTWISTGSHKR--KLMERFEDPEEMKLEDPLGPRYRCLSRV--QMKTWRK--NSWDYD-LRPWPQDEL	154																			
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PNDV_alphaSat_3_Rep_AHC72280	--MPSQSKTNYWFTLNFK-GEIPI--LS--LDTRVQYALQHEYVS--HHHQLQFQVQMKQST-LQGKMLAPIG--AHFEMVALDKDQSARQYAMKEDSRLEGPWHEYGLYIKKGSHT--KVMERFDSPEEMKVPSLRYRCLSRK--MTEEQRS--STWDYD-LRPWPQDSI	156																			
SCSV_alphaSat_C6_Rep_AAA68022	--MPTQSTSFWFTLNFE-GEIPI--LP--FNEVQYACWQHERV--HDHQYIQLQFKSNTLQRAYKIFNG--LNPHEATRDEVAQKLYMKEKDRSVAvgpwSYEGLVFKRGSHT--KLMERFEDPEEMKLEDPLGPRYRCLSRK--MAEEQKRS--SEEWYD-LRPWPQEEV	157																			
BMLRV_alphaSat_2_Rep_AHC72185	--MPTLQGTFWCFTLNFS-GDVP--LS--FDETQVYAWQHECVS--HDHQYIQLQMKHVT-LKKMKELLPG--AHLEVAKT--PEQASSYAMKESSRVAvgpwYFGE LLKKGSNKR--KLLDRYDRSPEDMELEDPAKARRCRARI--DKEKFVSD--FKVEDD-EQEWRKLV	155																			
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AYVIV_alphaSat_Rep_CDH98093	--MPTIQSFWFCFTIFFTSAPPD--LVPFLFENTHSVYACWQEEESPTTQRHLQGYLQLTKKS-LSAVSKLFGDNLPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQKTA--HEIQISLNLSWQRL	167																			
CYIV_alphaSat_Rep_CBJ19303	--MPTIQSFWFCFTIFFTSAPPD--LVPFLFENTHSVYACWQEEESPTTQRHLQGYLQLTKKS-LSAVSKLFGDNLPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQKTA--HEIQISLNLSWQRL	167																			
VEM_alphaSat_2_Rep_ALK03648	--MPTIQSFWFCFTIFFTSAPPD--LVPFLFENTHSVYACWQEEESPTTQRHLQGYLQLTKKS-LSAVSKLFGDNLPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQKTA--HEIQISLNLSWQRL	167																			
Dragonfly_ass_alphaSat_Rep_AVF91336	--MPTIQSFWFCFTIFFTSAPPD--LVPFLFENTHSVYACWQEEESPTTQRHLQGYLQLTKKS-LSAVSKLFGDNLPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQATV-AEIQISLNLSWQRL	167																			
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CLCuGeV_alphaSat_Rep_CDI44972	--MPSI--SRNWCFTL-FEYTLPL--FLSLPDWAVYLVFQEVEVSPSTRKHIQGYCFTL-KPQLSFLKTKL-QGLNG-AHLEAKGSA--SSNRDYCRKDSRASGPWFGILAEQGSHKR--KTMFERQEDPDEVRLADPKLYRCLATV--TNKRFN--NVELNPLFDRWPQEL	167																			
SCSV_[AU;F]_M_Rep_CAB96405	--MAR-QVICWCFTLNNP--L---APLSLHESMKLVYQTEAGDN-GTIHYQGYVEMK-KRTSLVQMKLLPG--AHLEKRRGSQ-GEARAYAMKEDSRVGEWPWFEGFEKVEFVLEKPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQKTA--HEIQISLNLSWQRL	167																			
FBNV-[E;G;1_93]_M_Rep_NP_619567	--MAR-QVICWCFTLNNP--L---APLSLHESMKLVYQTEQGEA-GNIHFQGYIEMK-KRTSLAGMKLLPG--AHFEKRRGTQ-GEARAYAMKEDSRVGEWPWFEGFEKVEFVLEKPHLEKQKARRTDEARDYCMKEETRVSFGEFGDYPGSPHKR--RQRESVIRSPVRMAEENPSVFRVAKI--AEEEFQKTA--HEIQISLNLSWQRL	167																			
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	190	200	210	220	230	240	250	260	270	280	290	300	310	320	330	340	
CFDV-[VU;89]_alphasat_R_Rep	L	S	A	I	E	E	P	D	R	T	I	W	V	G	P	X	G
CFDV-[VU;89]_alphasat_1_Rep	I	S	A	I	G	E	P	A	D	D	T	I	W	I	C	R	L
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CFDV-[VU;89]_alphasat_4_Rep	C	F	D	V	-	[V	U	;	8	9	_	a	l	p	h	a	s
CFDV-[VU;15]_alphasat_8_Rep	C	F	D	V	-	[V	U	;	1	5	_	a	l	p	h	a	s
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BBTV_alpha <sub>s</sub> AT_Y_Rep_ACJ36781	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV_alpha <sub>s</sub> AT_HaiKou2_Rep_AEF97834	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV_alpha <sub>s</sub> AT_S1_Rep_AAG4003	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CBDV_alpha <sub>s</sub> AT_Tung1_Rep_BAS02103	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CBDV_alpha <sub>s</sub> AT_Fr_X7_Rep_YP_008854133	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV_alpha <sub>s</sub> AT_4_Rep_ACB86656	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV_alpha <sub>s</sub> AT_TW3S2_Rep_ACJ36782	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
MDV_alpha <sub>s</sub> AT_C2_Rep_BAA33981	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNYV_alpha <sub>s</sub> AT_C1_Rep_CAA06791	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
SCSV_alpha <sub>s</sub> AT_C2_Rep_AAA68018	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNSV_alpha <sub>s</sub> AT_1_Rep_AHC72219	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BMLRV_alpha <sub>s</sub> AT_1_Rep_AHC72176	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNYV_alpha <sub>s</sub> AT_C7_EG93_Rep_CAB44025	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
MDV_alpha <sub>s</sub> AT_C10_Rep_BAA34048	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
MDV_alpha <sub>s</sub> AT_C3_Rep_BAA33982	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
PNYDV_alpha <sub>s</sub> AT_3_Rep_AHC72280	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
SCSV_alpha <sub>s</sub> AT_C6_Rep_AAA68022	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BMLRV_alpha <sub>s</sub> AT_2_Rep_AHC72185	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
MDV_alpha <sub>s</sub> AT_C1_Rep_BAA33980	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNSV_alpha <sub>s</sub> AT_2_Rep_AHC72218	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
PNYDV_alpha <sub>s</sub> AT_1_Rep_AHC72279	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
PaLCuV-IN_alpha <sub>s</sub> AT_Rep_AFG29502	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CLCuV_alpha <sub>s</sub> AT_Rep_AIH07616	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
GDS_alpha <sub>s</sub> AT_Rep_ALB26259	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
SilCuV_alpha <sub>s</sub> AT_Rep_CBY89002	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CLCuDaV_alpha <sub>s</sub> AT_Rep_CAD56270	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
ToLCV_alpha <sub>s</sub> AT_Rep_CDH98084	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
TbCSV_alpha <sub>s</sub> AT_Rep_CBK44088	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
AEV-IN_alpha <sub>s</sub> AT_Rep_CEE15328	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
AYVV_alpha <sub>s</sub> AT_Rep_CAB59911	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
TbCSV_alpha <sub>s</sub> AT_Y290_Rep_CAI59924	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
AYVIV_alpha <sub>s</sub> AT_Rep_CDH98093	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CYMV_alpha <sub>s</sub> AT_Rep_CBJ19303	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
VEM_alpha <sub>s</sub> AT_2_Rep_ALK03648	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
Dragonfly_ass_alpha <sub>s</sub> AT_Rep_AFV91336	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
EuVMY_alpha <sub>s</sub> AT_Rep_CBA18108	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CLCuGeV_alpha <sub>s</sub> AT_Rep_CDI44972	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
SCSV-[AU;F]_M_Rep_CAB96405	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNYV-[EG;1_93]_M_Rep_NP_619567	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
MDV-[JP;1]_M_Rep_NP_619769	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBYLV-[ET;231]_M_Rep_CCF74113	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
FBNSV-[ET;Ho1_97]_M_Rep_AdJ00306	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
PNYDV-[DE;15]_M_Rep_ADE87486	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
PYSV-[AT;15]_M_Rep_AHC72281	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV-[Au]_M_Rep_AAB25544	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
BBTV-[TW]_M_Rep_ABH03036	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
CBDV-[IN]_M_Rep_AGG38934	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
ABTV-[MY]_M_Rep_ABP96965	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K
ABTV-[PH]_M_Rep_ABP96960	M	E	V	A	E	P	D	R	T	I	W	V	G	P	X	G	K

B

**Figure S1. Comparison of the master Rep protein sequences of nano- and babuviruses with the Rep protein sequences of selected alphasatellites associated with nanovirids, geminiviruses and CFDV as well as two alphasatellites isolated from insects.**

Alignments were done using ClustalW in MegAlign of DNASTAR. Blue and green boxes A and B indicate the supposed M-Rep signature, i.e. two perfectly conserved sequence signatures characteristic of the M-Rep proteins of nanovirids. Names and accession numbers of the proteins are indicated on the left.



### Figure S2. Recombination analysis

Sequences of CFD alphasatellites were aligned with sequences of ten banana bunchy top and two cardamom bushy dwarf alphasatellites using Muscle (UPGMB clustering, 8 iterations). A recombination analysis by RDP 4<sup>26</sup> identified CFDA7 as a potential recombinant. The recombination event was supported by RDP ( $P$  value  $7.35 \times 10^{-11}$ ), GENECONV ( $P 1.3 \times 10^{-10}$ ), BootScan ( $P 1.09 \times 10^{-12}$ ), MaxChi ( $P 2.77 \times 10^{-9}$ ), Chimaera ( $P 3.51 \times 10^{-9}$ ), SiScan ( $P 3.0 \times 10^{-11}$ ) and 3Seq ( $P 1.54 \times 10^{-5}$ ). Top panel: Bootscan plot as implemented in RDP 4 showing the recombinant region flanked by potential recombination breakpoint regions.

A graphic representation of CFDA7 with the recombinant region (position 951 to 1099) possibly derived from CFDA6 (minor parent) is shown below the plot.

Figure S3

SCSV-[AU;F]\_DNA-R\_AJ290434  
FBNVV-[EG;1-93]\_DNA-R\_NC\_003560  
MDV-[JP;1]\_DNA-R\_NC\_003648  
FBNSV-[ET;Hol-97]\_DNA-R\_GU983866  
PNYDV-[DE;15]\_DNA-R\_GU553134  
PYSV-[AT;15]\_DNA-R\_KC979054  
FBYLV-[ET;231]\_DNA-R\_HE654123  
BMLRV-[AT;3]\_DNA-R\_KC978939  
BMLRV-[AZ;47]\_DNA-R\_KC978949  
BBTV-[AU]\_DNA-R\_S56276  
BBTV-[TW]\_DNA-R\_DQ826390  
CBDV-[IN]\_DNA-R\_JX867550  
ABTV-[MY]\_DNA-R\_EF546813  
ABTV-[PH]\_DNA-R\_EF546807

SCSV-[AU;F]\_DNA-R\_AJ290434  
FBNVV-[EG;1-93]\_DNA-R\_NC\_003560  
MDV-[JP;1]\_DNA-R\_NC\_003648  
FBNSV-[ET;Hol-97]\_DNA-R\_GU983866  
PNYDV-[DE;15]\_DNA-R\_GU553134  
PYSV-[AT;15]\_DNA-R\_KC979054  
FBYLV-[ET;231]\_DNA-R\_HE654123  
BMLRV-[AT;3]\_DNA-R\_KC978939  
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SCSV-[AU;F]\_DNA-R\_AJ290434  
FBNVV-[EG;1-93]\_DNA-R\_NC\_003560  
MDV-[JP;1]\_DNA-R\_NC\_003648  
FBNSV-[ET;Hol-97]\_DNA-R\_GU983866  
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PYSV-[AT;15]\_DNA-R\_KC979054  
FBYLV-[ET;231]\_DNA-R\_HE654123  
BMLRV-[AT;3]\_DNA-R\_KC978939  
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BBTV-[AU]\_DNA-R\_S56276  
BBTV-[TW]\_DNA-R\_DQ826390  
CBDV-[IN]\_DNA-R\_JX867550  
ABTV-[MY]\_DNA-R\_EF546813  
ABTV-[PH]\_DNA-R\_EF546807

A

SCSV-[AU;F]\_DNA-R\_AJ290434  
FBNYV-[EG;1-93]\_DNA-R\_NC\_003560  
MDV-[JP;1]\_DNA-R\_NC\_003648  
FBNSV-[ET;Hol-97]\_DNA-R\_GU983866  
PNYDV-[DE;15]\_DNA-R\_GU553134  
PYSV-[AT;15]\_DNA-R\_KC979054  
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BMLRV-[AT;3]\_DNA-R\_KC978939  
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ABTV-[MY]\_DNA-R\_EF546813  
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PYSV-[AT;15]\_DNA-R\_KC979054  
FBYLV-[ET;231]\_DNA-R\_HE654123  
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MDV-[JP;1]\_DNA-R\_NC\_003648  
FBNSV-[ET;Hol-97]\_DNA-R\_GU983866  
PNYDV-[DE;15]\_DNA-R\_GU553134  
PYSV-[AT;15]\_DNA-R\_KC979054  
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BMLRV-[AT;3]\_DNA-R\_KC978939  
BMLRV-[AZ;47]\_DNA-R\_KC978949  
BBTV-[AU]\_DNA-R\_SS56276  
BBTV-[TW]\_DNA-R\_DQ826390  
CBDV-[IN]\_DNA-R\_JX867550  
ABTV-[MY]\_DNA-R\_EF546813  
ABTV-[PH]\_DNA-R\_EF546807

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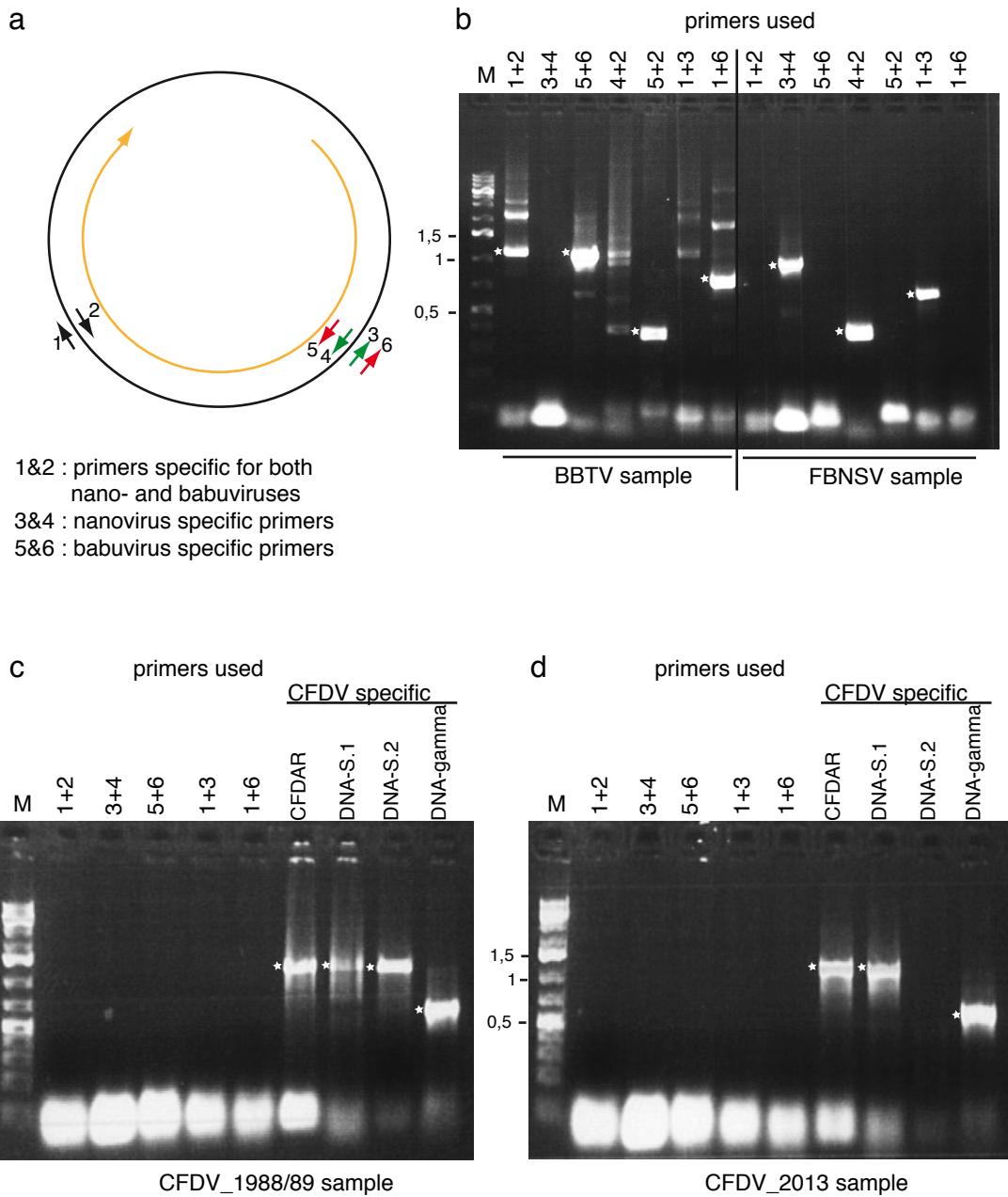
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TATGAACCGTTTAAAGATAGTGAAGATGTGGAGGTATTGTCATGGCTAATTCCCTCCGAAGGAAGGAATCTTCTGAAGATCGATAAAAGCTAGTTGCTGCTGA  
TATGAACCTGTTCTGAAAGTAGTACAGTATGTCATGGCTAATTCCCTCCGAAGGAAGGAATCTTCTGAAGATAGGATAAAAGCTGTTAACTTGTGA

B

**Figure S3. Alignment of coding sequences of nanovirus DNA-R components.**

Alignments were done using ClustalW in MegAlign of DNASTAR. Coloured boxes indicate regions corresponding to the M-Rep amino acid signature regions identified in Fig. S1, boxes A and -B, respectively. These sequences were used to design degenerate oligonucleotide primers (1 to 6, Table S3). Green boxes: fully complementary primers designed for the corresponding region, yellow and blue boxes: overlapping reverse and direct primer, respectively. Names and accession numbers of the respective nano- and babuvirus DNA-R components are indicated on the left

Figure S4



**Figure S4. PCR amplification of nanovirus DNA-R components.**

Degenerate primers (1 to 6, see Table S3) were designed based on the alignments of several babu- and nanovirus DNA-R sequences (see Fig. S3).

a - position and orientation of the primers in relation to nanovirus DNA-R.  
b - control amplifications using total DNA extract of BBTV infected banana leaves (Hawaii-2013 sample, this work) or using the plasmid carrying a dimer of FBNSV DNA-R<sup>30</sup>.

c - and d - PCR amplifications using CFDV samples. PCRs were performed on 1:10 diluted RCA products from a mixture of 23 different virion preparations of the CFDV\_88/89 sample pool (c) or from a mixture of total DNA extracts from eleven different leaves of one symptomatic coconut palm for CFDV\_2013 sample (d). CFDV specific primers were designed based on the cloned CFDV DNAs (the primer pairs used amplify a full-length DNA component as indicated, see Table S3). M- 1 kb plus DNA ladder, the size (K bp) of several bands is indicated on the left of the marker, amplified DNA fragments of expected size are indicated by an asterisk.

**Table S1. CFD samples prepared from symptomatic leaves of coconut palms showing date sampled at VARTC, palm identity, virus preparation date, and characteristics of the buoyant density zone (g/ml) in a Nycodenz® isopycnic density gradient from which the sample was collected**

Sample No (CFD)	Sampling date	Palm ID (Number of palms)	Prep Date	Gradient density	DNA by PAGE	Virions by EM
1		cfd2	(1)	28.02.89	1.23	+
2		cfd2	(1)	28.02.89	1.24	+
<b>3<sup>a,b</sup></b>	11.1988	cfd2 + cfd3	(2)	06.03.89	1.24	+
4		cfd2 + cfd3	(2)	06.03.89	1.24	+
5		cfd3	(1)	04.04.89	1.24	+
6		cfd3	(1)	10.04.89	1.24-1.28	n.t.
<b>7<sup>b</sup></b>	11.1988	cfd4	(1)	18.04.89	1.23	+
8	04.1989	MRD25-21A	(1)	13.06.89	1.24-1.29	+
<b>9<sup>a,b</sup></b>		MRD19 + MRD20	(2)	16.10.89	1.25	+
29		MRD19 + MRD20	(2)	16.10.89	1.26	+
30	09.1989	MRD19 + MRD20	(2)	16.10.89	1.24	+
31		MRD19 + MRD20	(2)	16.10.89	1.28	-
32		MRD19 + MRD20	(2)	16.10.89	1.23	+
10	09.1989	MRD37.14	(1)	30.10.89	1.27	+
<b>11<sup>b</sup></b>		MRD37.14	(1)	30.10.89	1.26	+
21 <sup>c</sup>		MRD caged	(unknown)	16.08.88	1.21-1.29	+
<b>22<sup>b,c</sup></b>	07.1988	MRD caged	(unknown)	23.08.88	1.27-1.29	+
23 <sup>c</sup>		MRD caged	(unknown)	23.08.88	1.23	+
24		TT17-7 + 12-86-14 + 12-86-15	(3)	07.11.88	1.27-1.29	+
25		TT17-7 + 12-86-14 + 12-86-15	(3)	07.11.88	1.21-1.23	+
<b>26<sup>b</sup></b>	09.1988	TT17-7 + 12-86-14 + 12-86-15	(3)	14.11.88	1.23-1.27	+
27		TT17-7 + 12-86-14 + 12-86-15	(3)	14.11.88	1.29	+
28		TT17-7 + 12-86-14 + 12-86-15	(3)	14.11.88	1.23	+

Samples in which DNA or virions were detected by polyacrylamide gel electrophoresis (PAGE) or electron microscopy (EM), respectively, are indicated by (+), or (-) if not detected; n.t. = not tested. MRD, Malayan Red Dwarf; cfd and other identifiers indicate that variety or hybrid source was not recorded

a – CFD samples from which RCA-DNA was used for cloning;

b – CFD samples from which RCA-DNA was pooled for deep sequencing;

c – CFD samples from caged MRD plants inoculated with viruliferous *Myndus taffini*.

**Table S2. Viral DNAs identified in two CFD samples on the basis of similar RFLP patterns and sequencing**

Sample	DNA	Size nt	Enzyme <sup>a</sup>				
			<i>Aat</i> I	<i>Eco</i> RI	<i>Bam</i> HI	<i>Kpn</i> I	<i>Age</i> I
CFD9	DNA-S.1	1286	12 <sup>b</sup>	7			
	DNA-S.2	1263	3	3			
	DNA-gamma	641			22		
	CFDAR	1271			6		
	CFDA2	1277			14		
	CFDA3	1252			3		
	CFDA4	1276			2	6	
CFD3	CFDA7	1259	3				
	CFDA1	1291				3	
	CFDA6	1264					3

a – DNA fragment obtained upon digestion of RCA DNA by a given enzyme was cloned into Litmus28 (for *Aat*I and *Age*I fragments) or pBluescript KSII (+)

b - number of clones showing similar RFLP pattern: insert DNAs of recombinant plasmids were amplified by PCR using M13 direct and reverse primers, and the PCR products were digested by *Hae*III or *Sau*3A enzymes.

**Table S3. Oligonucleotides used**

DNA	Primer name (direction)	Sequence (5'-3')	Tm, C°	Use <sup>a</sup>
CFDV DNA-S.1	CFDV_S1-HindIII-dir	CCCATTAAGCTTAAAGCCCCATTAGCGATGAC	66.9	1
	CFDV_S1-HindIII-rev	GGCTTTAAGCTTAAATGGGCCTTCACGCAATTAC	68.2	1
	CFDV_S1-590_dir	CATTGGCGTTGTGCTCGATAGAG	62.7	2
	CFDV_S1-1160-rev	CTGCTCATCCTATGCCGCCA	63.7	2
CFDV DNA-S.2	CFDV_S2-810-BspEI-dir	TTCCGGAGATACTGCCG	55.2	1, 2
	CFDV_S2-810-BspEI-rev	TATCTCCGGAATACTTCA	49.1	1, 2
	CFDV_S2-590-dir	CATTGGCCTTGGTGTGGACAAAAT	61.0	2
	CFDV_S2-1160-rev	TCTGCTCAAGCTCTGCTCATCC	62.1	2
CFDV DNA-S.1 & DNA-S.2	CFDV-CP1&2-dir	CCGGAGGAATAATGTACACGAAGAAG	63.2	
	CFDV-CP1&2-rev	CCTCCGGCATATAAAGCCGGTCTTC	66.3	
DNA-S.1 related <sup>b</sup>	CFDV_STL1-dir	CGGCCTAGTATTACCCGCCACGCTC	71.1	1
	CFDV_STL1-rev	ACTAGGGCCGCCACGCTTATACAGAGC	69.5	1
DNA-S.2 related <sup>c</sup>	CFDV_STL2-dir	TGCTCCCTACCTCTGCTCATCTG	66.1	1
	CFDV_STL2-rev	CAGAGCAGAGGTAGTGAGC	58.8	1
CFDV DNA- gamma	CFDV-gamma-dir	TGGCGCTGCTGCCGCTTCGCTCTGGAGC	75	1, 2
	CFDV-gamma-rev	GCAGCGCCATTAGCTTCTCCAATTTCACCTC	69.5	1, 2
CFDAR	CFDAR-BamHI-dir	CATAGAGGATCCGAAAAGAAATTGATTCTCG	64.4	1, 2
	CFDAR-BamHI-rev	CTTTCCGGATCCTCTATGTACTGATACAGGAC	66.9	1, 2
CFDA1	CFDA1-dir	CTTGGCTATAATGGGTCCTCC	60.6	2
	CFDA1-rev	CTTCTCCAGCAAGCTACTCACC	62.1	2
CFDA2	CFDA2-392-SstI-dir	TGCTGAGCTCCGCTTGGAAAGAACCTGG	69.5	3
	CFDA2-696-BamHI-rev	CTTCGGATCCTCTATGTACTGGTACAGTAC	66.8	3
	CFDA2-510-dir	ATTGGCAACTTGAGTTGCTCAG	58.4	2
	CFDA2-930-rev	ACAAATGCTAATTGATTATTCAAATATCAA	55.8	2
CFDA3	CFDA3-398-SstI-dir	ATTTGAGCTCGGTTCCCCGGTTGAGTTG	68.1	2, 3
	CFDA3-789-BamHI-rev	TTTCGGATCCTTCATCGATATAGGAAAATA	61.3	2, 3
CFDA4	CFDA4-262-BamHI-dir	GACAGGATCCACCTCGAACAGGCACAC	69.5	2, 3
	CFDA4-563-BamHI-rev	TCGTCAGCTTGTCTTGGAGTTGAGACAGC	68.1	2, 3
CFDA5	CFDA5-dir	AGCAGCCGTGCTCGACGA	60.5	1, 2
	CFDA5-rev	CCGATGAGCAGGGTATG	58.2	1, 2
CFDA6	CFDA6-dir	GTGGGTCCCACTTGGACTTTGTGG	69.5	1
	CFDA6-rev	TGGGACCCACAAGGAGCGTCTGTCGAC	71.0	1
CFDA6	CFDA6-KpnI-1159-dir	TGTGGGTACCACTTGTGGACTTTGTGGTA	68.1	1, 3
	CFDA6-KpnI-1172-rev	AAAGTGGTACCCACAAGGAGCGTCTGTCGA	69.5	1, 2, 3
CFDA6	CFDA6-402-SstI-dir	ATTGGAGCTCGGTTCCCCGGTTCTGTT	68.0	2, 3
	CFDA6-789-BamHI-rev	TTTCGGATCCTTCATCGATATAGGAAAATA	61.3	3
CFDA7	CFDA7-319-SstI-dir	CATGGAGCTCCGACCGCGTCTGGTT	69.5	2, 3
	CFDA7-523-rev	ACCACAGCTGTAAAACCACTTTCATTAAG	62.7	2, 3
CFDA8	CFDA8-210-rev	TTGTACTGTCCGGCACCGGAC	65.8	2
	CFDA8-540-dir	CTTGAATTGCTGTCAAATCGACGGA	63.4	2
CFDA8	CFDA8-200-dir	AAGTCGGTGGCCGGACAGTACAA	66.1	1
	CFDA8-200-rev	CCACCGGACTTCAGATGGAGGAAC	66.1	1
nanovirid DNA-R <sup>d</sup>	1 - b-KNG-dir	AArAATGGnATnA TnC ArAGCGGrAArTA	61.7	1
	2 - b-KNG-rev	TAyTTyCCGCTyTGnAtnATnCCATTyTT	61.7	1
	3 - nanoEYIE-rev	GAyTTrTCrTA nGTGTTACAACACTCmTCdATrTAyTC	67.1	1
	4 - nanoEYIE-dir	GA rTAyA ThGAKGAGTGTGTAACACnTAyGAyAArTC	67.1	1
	5 - babu-EYL-dir	AATACCTTCGAYAGAAGTArAGATACATTATACAG	63.6	1
	6 - babuEYL-rev	rTCGAAGGTATThGGACArTCrTATAAATACTC	64.3	1

a – a pair of primers indicated by the same number was used for PCR amplification of the corresponding DNA for the following purposes: (1) full-length cloning, (2) component-specific detection by PCR, (3) amplification of the fragment for construction of redundant copies of the component;

b – primers designed based on the DNA sequence in the region with potential stem-loop structure (STL) of CFDV DNA-S.1;

c – primers designed on the basis of the DNA sequence of STL region of CFDV\_DNA-S.2;

d – primers designed on the basis of the alignment of the nanovirid DNA-R sequences.

**Table S4. Mapping of primary reads from deep sequencing data of three BBTV samples**

Sample \ DNA	BBTV-Hawaii_2013 (26,758,442 total reads)			BBTV-Nigeria_2013 (25,761,084 total reads)			BBTV-Vietnam_2013 (34,536,988 total reads)		
	Reads	% of reads	Cov. Mean	Reads	% of reads	Cov. Mean	Reads	% of reads	Cov. Mean
DNA-R	1,671,973	6.3	149,484	562,089	2.2	50,015	731,405	2.1	65,039
DNA-S	571,790	2.1	50,314	67,470	0.3	5,952	3,042,114	8.8	271,294
DNA-M	1,306,112	4.9	122,536	264,262	1	24,898	1,302,658	3.8	124,545
DNA-N	7,240,846	27.1	657,758	6,323,995	24.6	565,927	5,100,799	14.8	460,561
DNA-C	5,137,818	19.2	502,970	893,580	3.5	87,631	850,602	2.5	82,171
DNA-U3	8,579,130	32.1	797,873	5,282,003	20.5	487,589	3,922,111	11.4	363,683
BBTA2 <sup>a</sup>							3,161,344	9.2	279,229
total	24,507,669	91.6		13,393,399	52		18,111,033	52.4	

Geneious mapping of total reads against all identified (*de novo* assembled) BBTV DNAs, performed with the settings allowing 10% mismatch per read. Cov. Mean – average coverage.  
a - a variant of the proposed alphasatellite species BBTA2<sup>25</sup>

**Table S5. CFDV mutant spectra**

DNA	Year	Total	Type of base changes															
			A C	A G	A T	C A	C G	C T	G A	G C	G T	T A	T C	T G	transitions	transversions	deletions	insertions
DNA-R	1989														0	0		
	2013	2													0	1	1 (+1)	
DNA-S.1	1989																	
	2013	1													1			
DNA-S.2	2015	0													0	0		
	1989						1			1	1				2	1		
CFDA1	2015	3													0	0		
	1985														0	0		
CFDA1	1988	2					1	1							2	0		
	1989	1							1						1	0		
CFDA1	2013	20	1	1		2	3	3	2	3		1	2		7	11	1 (-3)	1 (+1)

Number and type of changes between the most recent and respective previous sampling dates are shown. For calculation of the nucleotide substitution rates per site per year the deletion of three nucleotides is counted as one event.

**Table S6. Cloning of redundant copies of CFD associated DNAs used for replication assays**

DNA	Primers used for PCR amplification of monomer <sup>a</sup>	Monomer <sup>b</sup>	Primers used for PCR amplification of 0,2 mer <sup>c</sup>	Redundant copy in pBin19
CFDV DNA-S.1	CFDV_S1-HindIII-dir CFDV_S1-HindIII-rev	pBluescriptIIKS(+-)HindIII-S1-5		Dimer first obtained by cloning a monomer into pBluescriptIIKS(+) HindIII site and then transferred into pBin19 as BamHI-KpnI fragment
CFDV DNA-S.2	CFDV_S2-810-BspEI-dir CFDV_S2-810-BspEI-rev	pBluescriptIIKS(+-)BspEI-S2-4		Dimer first obtained by cloning a monomer into pBluescriptIIKS(+) BspEI site and then transferred into pBin19 as BamHI-KpnI fragment
CFDV DNA-gamma		pBluescriptIIKS(+-)KpnI-mini-6		Direct cloning of a monomer into KpnI site of pBin19 with selection of dimer insertion
CFDAR	CFDAR-BamHI-dir CFDAR-BamHI-rev	pBluescriptIIKS(+-)BamHI-AR-1		Direct cloning of a monomer into BamHI site of pBin19 with selection of dimer insertion
CFDA1		Litmus28-AgeI-A1-6 <sup>d</sup>		Dimer first obtained by cloning a monomer into Litmus28 AgeI site and then transferred into pBin19 as BamHI-KpnI fragment
CFDA2		pBluescriptIIKS(+-)BamHI-A2-2	CFDA2-392-SstI-dir CFDA2-696-BamHI-rev	302 bp fragment was PCR amplified using indicated primers, digested by SstI and inserted into pBin19-(SstI-HindIIIBlunt) <sup>e</sup> plasmid. CFDA2 monomer was consequently inserted into BamHI site of this plasmid thus giving rise to CFDA2 1,2 mer.
CFDA3		pBluescriptIIKS(+-)BamHI-A3-24	CFDA3-398-SstI-dir CFDA3-789-BamHI-rev	390 bp fragment was PCR amplified using indicated primers, digested by SstI and inserted into pBin19-(SstI-HindIIIBlunt) plasmid. CFDA3 monomer was subsequently inserted into BamHI site of this plasmid thus giving rise to CFDA3 1,2 mer.
CFDA4		pBluescriptIIKS(+-)BamHI-A4-14	CFDA4-262-BamHI-dir CFDA4-563-BamHI-rev	300 bp fragment was PCR amplified using indicated primers, digested by BamHI and inserted into pBin19-(BamHI-HindIIIBlunt) <sup>f</sup> plasmid. CFDA4 monomer was consequently inserted into BamHI site of this plasmid thus giving rise to CFDA4 1,2 mer.
CFDA6	CFDA6-KpnI-1159-dir CFDA6-KpnI-1172-rev	pBluescriptIIKS(+-)HincII-A6-7	CFDA6-402-SstI-dir CFDA6-789-BamHI-rev	360 bp fragment was PCR amplified using indicated primers, digested by SstI and inserted into pBin19-(SstI-HindIIIBlunt) plasmid. CFDA6 monomer, liberated as KpnI fragment, was ligated, ligation product digested by BamHI and inserted into BamHII site of the plasmid with cloned 360 fragment, thus giving rise to CFDA6 1,2 mer.

CFDA7	Litmus28-AatII-A7-15	CFDA7-319-SstI-dir CFDA7-523-rev	320 bp fragment was PCR amplified using indicated primers, digested by <i>Sst</i> I and inserted into pBin19-( <i>Sst</i> I-HindIIIblunt) plasmid. CFDA7 monomer, liberated as AatI fragment, was ligated, ligation product digested by HindIII and inserted into HindIII site of the plasmid with cloned 320 fragment, thus giving rise to CFDA7 1,2 mer
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a - PCR performed on RCA DNA of sample CFD\_88/89

b - plasmids used to clone a given DNA in the restriction site indicated, numbers at the end identify the recombinant plasmids

c - viral DNA from the plasmid shown under "Monomer" was used for PCR amplification

d - if primers are not indicated in the second column, viral DNA was cloned as monomer after restriction of RCA DNA into the corresponding plasmid, see Table S2 for enzymes used.

e - pBluescriptIIKS(+) plasmid was digested by *Hind*III, the ssDNA ends resulting after digestion were filled using dNTPs and Klenow polymerase. After purification by phenol extraction and ethanol precipitation the plasmid DNA was digested by *Sst*I.

f - pBluescriptIIKS(+) plasmid was digested by *Hind*III, the ssDNA ends resulting after digestion were filled using dNTPs and Klenow polymerase. After purification by phenol extraction and ethanol precipitation the plasmid DNA was digested by *Bam*HI.

**Table S7. GenBank accession numbers of CFD associated DNAs**

DNA \ Year	1985	1988	1989	2013	2015
DNA-S.1			MF926436	MF926437	MF926438
DNA-S.2			MF926439		MF926440
DNA-gamma			MF926441	MF926442	MF926443
CFDAR			MF926434	MF926435	
CFDA1	M29963	MF926423	MF926424	MF926425	
CFDA2			MF926426		
CFDA3			MF926427		
CFDA4			MF926429	MF926444	
CFDA5			MF926430		
CFDA6		MF926431			
CFDA7			MF926432		
CFDA8				MF926433	