# **Expanded View Figures**

### Figure EV1. Immunofluorescence and electron microscopic analyses of Gtsf1-deficient testes (related to Fig 1).

- A Elevated expression of LINE-1 and IAP in early postnatal periods caused by *Ctsf1* deficiency. Immunostaining of *Ctsf1*<sup>+/-</sup> and *Ctsf1*<sup>-/-</sup> testes at postnatal days 0, 4, and 8 with antibody to Line-1 ORF1 protein (left panel, green) and IAP GAG protein (right panel, green). DNA was stained with DAPI (red). Scale bar, 10 µm.
- B Expression of GTSF1 in early postnatal periods. Immunostaining of Gtsf1<sup>+/-</sup> and Gtsf1<sup>-/-</sup> testes at postnatal days 0, 4, and 8 with antibody to GTSF1 (green). DNA was stained with DAPI (red). Scale bar, 10 μm.
- C Histological analysis of *Gtsf1*-deficiency phenotypes. Immunostaining of *Gtsf1*<sup>+/-</sup> and *Gtsf1*<sup>-/-</sup> E17.5 testes with anti-DNMT3A2 (upper panel, green) and anti-DNMT3L (lower panel, green) antibodies. DNA was stained with DAPI (red). Localization of DNMT3L and DNMT3A/3A2 was unaffected in the prenatal *Gtsf1*<sup>-/-</sup> testes. Scale bar, 10 µm
- D Electron microscopy of Ctsf1<sup>+/-</sup> (upper panel) and Ctsf1<sup>-/-</sup> (lower panel) E17.5 prospermatogonia. Right panels (scale bar, 1 μm) are magnified views of the boxed region in the left panels (scale bar, 5 μm). Arrows indicate intermitochondrial cement.

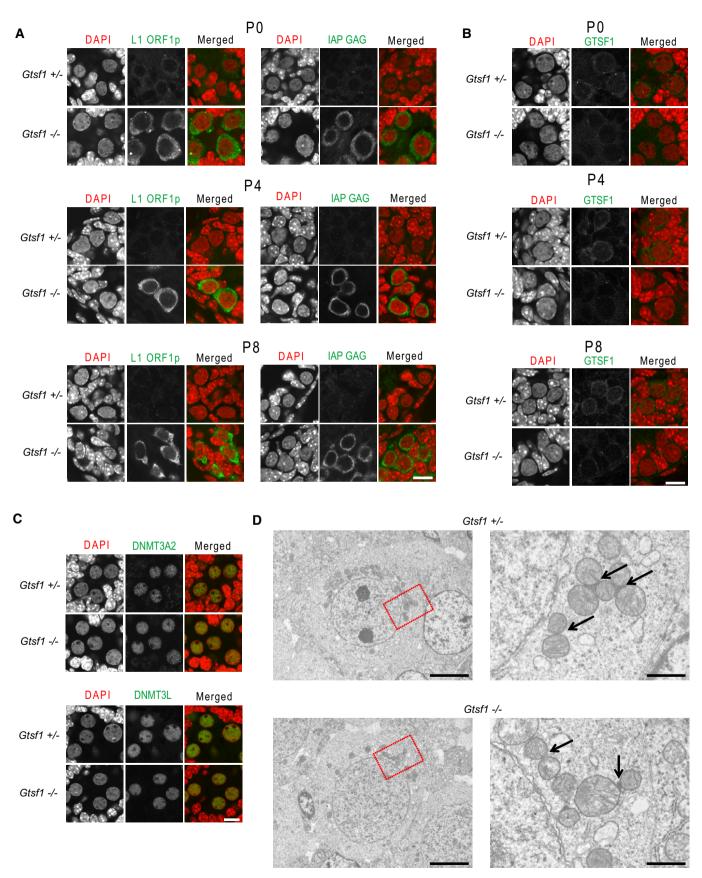
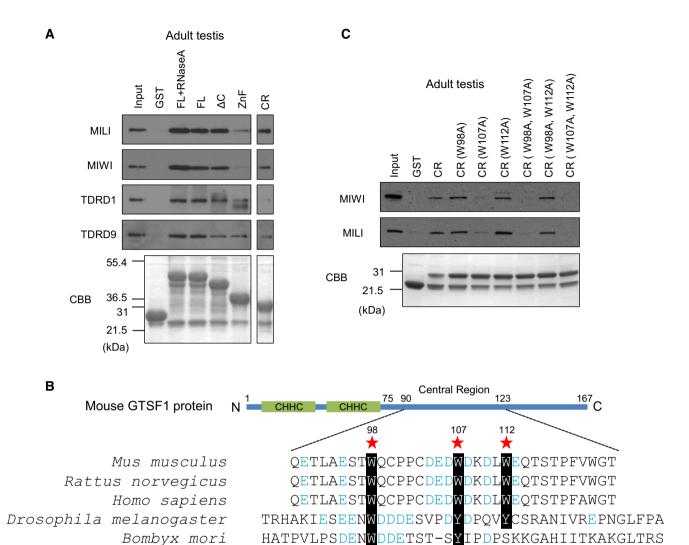


Figure EV1.

### Figure EV2. Analysis of GTSF1-associated proteins (related to Fig 2).

- A GST pull-down analysis of the interaction of GTSF1 with MILI, MIWI, TDRD1, or TDRD9. The GST-fusion proteins bound to glutathione sepharose were incubated with adult testis lysates. In some experiments, testis lysates were pretreated with RNase A prior to incubation with GST-fusion proteins. The proteins bound to GST-fusion proteins were analyzed by SDS-PAGE followed by Western blotting with antibodies to MILI, MIWI, TDRD1, and TDRD9 (upper panels). CBB staining shows the amount of the GST-fusion proteins in each of the reactions (lower panels).
- B Schematic illustration of mouse GTSF1 protein. Amino acid sequences in the central regions of five species, including mouse, are shown. Black-and-white inverted characters represent conserved aromatic amino acid residues in the central regions among the species, which were used for alanine substitutions in pull-down analysis. These aromatic amino acid residues are surrounded by several negatively charged amino acid residues (in blue font).
- C GST pull-down analysis of the interaction of mutated CR proteins with MILI or MIWI complexes. The GST-fusion proteins bound to glutathione sepharose were incubated with adult testis lysates. The proteins bound to GST-fusion proteins were analyzed by SDS-PAGE followed by Western blotting with antibodies to MILI and MIWI (upper panels). CBB staining shows the amount of the GST-fusion proteins in each of the reactions (lower panels).
- D Immunoprecipitation analysis of the binding of GTSF1 to several piRNA pathway components. Myc-tagged GTSF1 was co-expressed with FLAG-tagged MAEL, MILI, MIWI, MIWI2, MVH, or His-tagged TDRD1 in HEK293 cells. Lysates of the transfected cells were immunoprecipitated with an anti-Myc antibody and separated by SDS-PAGE, followed by Western blotting using the indicated antibodies.



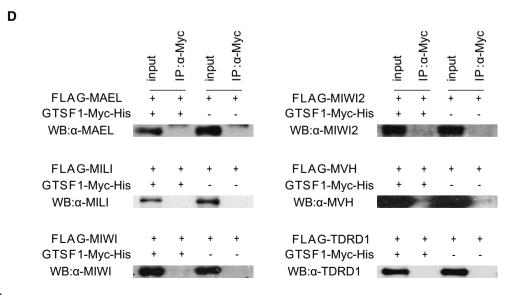
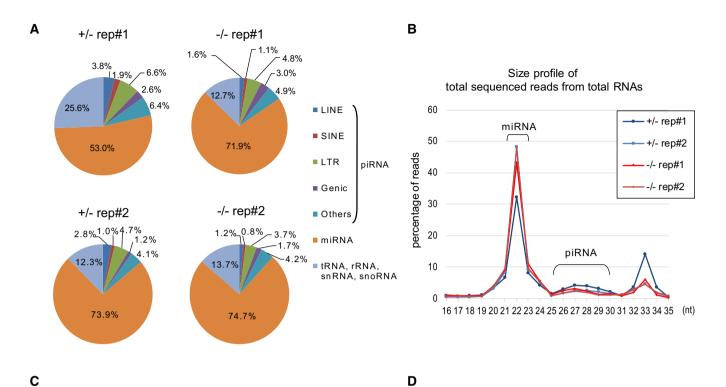


Figure EV2.



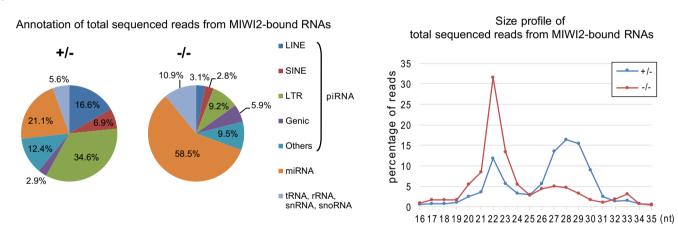


Figure EV3. Analysis of deep-sequencing data obtained from the libraries of total small RNAs and MIWI2-bound piRNAs (related to Fig 3).

- A RNA annotation of total sequenced reads from total RNA libraries prepared from  $Gtsf1^{+/-}$  and  $Gtsf1^{-/-}$  E17.5 testes (n=2).
- B Size profile of total sequenced reads from total RNA libraries prepared from  $Gtsf1^{+/-}$  and  $Gtsf1^{-/-}$  E17.5 testes (n=2). Shown is the percentage of total reads of each length to total reads of 16–35 nt RNAs.
- C RNA annotation of total sequenced reads from MIWI2-bound RNA libraries prepared from  $\mathit{Ctsf1}^{+/-}$  and  $\mathit{Ctsf1}^{-/-}$  E17.5 testes.
- D Size profile of total sequenced reads from MIWI2-bound RNA libraries prepared from Gtsf1<sup>+/-</sup> and Gtsf1<sup>-/-</sup> E17.5 testes. Shown is the percentage of total reads of each length in total reads of 16–35 nt MIWI2-bound RNAs.

EMBO reports e42054 | 2018 © 2018 The Authors

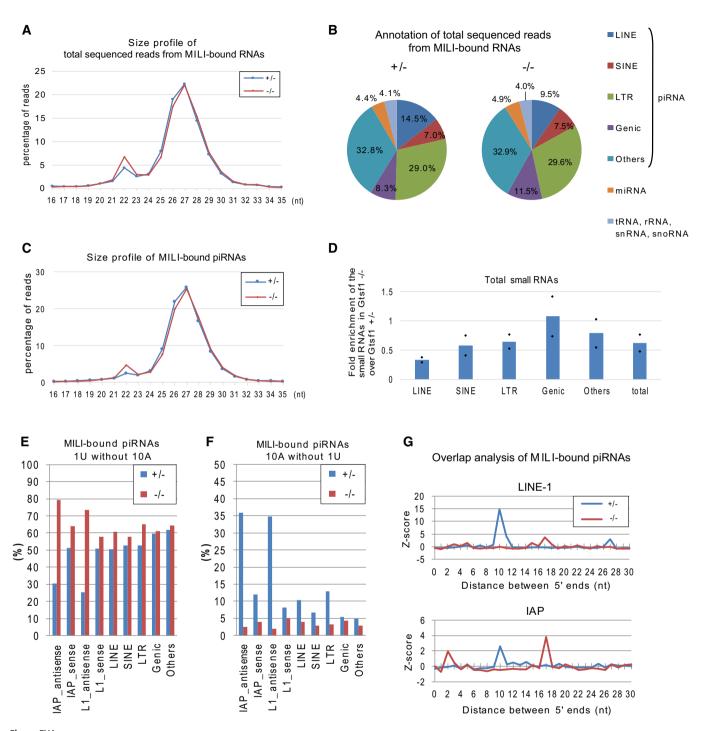


Figure EV4.

### Figure EV4. Analysis of deep-sequencing data obtained from the libraries of MILI-bound piRNAs and total small RNAs (related to Fig 4).

- A Size profile of total sequenced reads from MILI-bound RNA libraries prepared from Gtsf1<sup>+/-</sup> and Gtsf1<sup>-/-</sup> E17.5 testes. Shown is the percentage of MILI-bound small RNA reads of each length in total reads of 16–35 nt MILI-bound RNAs.
- B RNA annotation of total sequenced reads from MILI-bound RNA libraries prepared from  $Gtsf1^{+/-}$  and  $Gtsf1^{-/-}$  E17.5 testes.
- C Size profile of the piRNAs in MILI-bound RNA libraries prepared from Gtsf1+/- and Gtsf1-/- E17.5 testes. Shown is the percentage of MILI-bound piRNA reads of each length in the total reads of 16–35 nt MILI-bound piRNAs.
- D Small RNAs are less abundant in  $Gtsf1^{-/-}$  total small RNA libraries. Shown is the fold enrichment of each annotated piRNA in  $Gtsf1^{-/-}$  over  $Gtsf1^{+/-}$ . Total read number of each annotated piRNA was normalized to that of 22 nt miRNA in the same libraries and plotted relative to its average value in  $Gtsf1^{+/-}$  testes. Bar graphs represent the mean of two biological replicates in each annotated piRNA.
- E The 1U signature for primary processing was increased in transposon-derived piRNAs in the *Ctsf1* mutant. Shown is the percentage of small RNAs containing 1U without 10A in each annotated group of MILI-bound piRNAs from *Ctsf1*<sup>+/-</sup> and *Ctsf1*<sup>-/-</sup> E17.5 testes.
- F The 10A ping-pong signature was lost in transposon-derived piRNAs under Gtsf1 deficiency. Shown is the percentage of small RNAs containing 10A without 1U in each annotated group of MILI-bound piRNAs from Gtsf1<sup>+/-</sup> and Gtsf1<sup>-/-</sup> E17.5 testes.
- G Ping-pong Z-scores [47] for the significance of the distance between the 5' ends of complementary MILI-bound piRNAs for LINE-1 (upper panel) and IAP (lower panel) from the Gtsf1<sup>-/-</sup> and Gtsf1<sup>+/-</sup> E17.5 testes.

#### Figure EV5. Mapping of small RNAs to LINE-1 sequence (related to Figs 3 and 4).

A—H Mapping of small RNAs derived from  $Gtsf1^{+/-}$  (A, C, E, and G) and  $Gtsf1^{-/-}$  (B, D, F, and H) testes to the genomic sequences of LINE-1 (Accession No. M13002), allowing up to three mismatches. The piRNAs for IAP from MILI-bound (A, B), MIWI2-bound (C, D), and two replicates of total (E—H) small RNA libraries were subjected to the analysis. The x-axis shows the position in M13002 sequence. The y-axis shows the normalized read count relative to 22 nt miRNAs in the deep-sequencing data from each library. Sense and antisense reads to retrotransposon transcripts are shown in blue and orange, respectively.

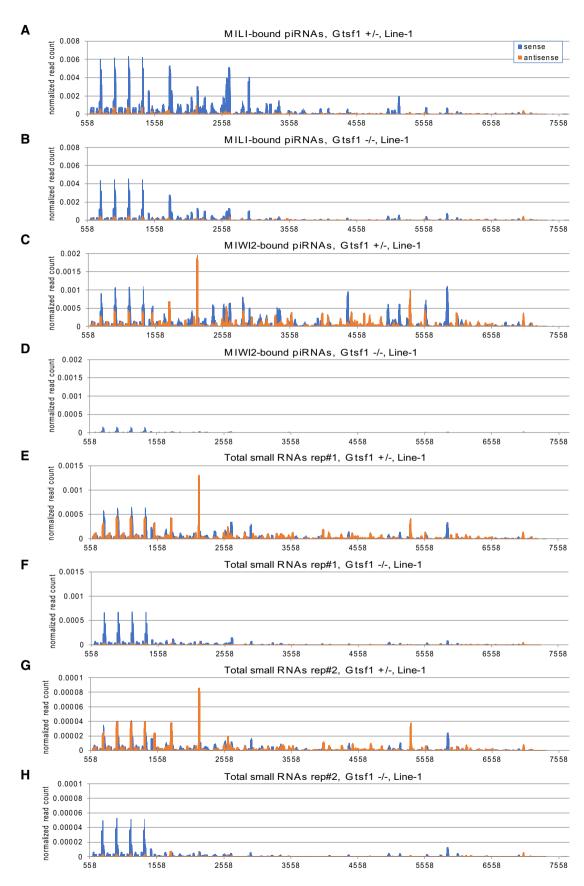


Figure EV5.

## Figure EV6. Mapping of small RNAs to IAP sequence (related to Figs 3 and 4).

A–H Mapping of piRNAs derived from  $Gtsf1^{+/-}$  (A, C, E, and G) and  $Gtsf1^{-/-}$  (B, D, F, and H) testes to the genomic sequences of IAP (Accession No. M17551), allowing up to three mismatches. The piRNAs for IAP from MILI-bound (A, B), MIWI2-bound (C, D), and two replicates of total (E–H) small RNA libraries were subjected to the analysis. The x-axis shows the position in M17551 sequence. The y-axis shows the normalized read count relative to 22 nt miRNAs in the deep-sequencing data from each library. Sense and antisense reads to retrotransposon transcripts are shown in blue and orange, respectively.

EMBO reports e42054 | 2018 © 2018 The Authors

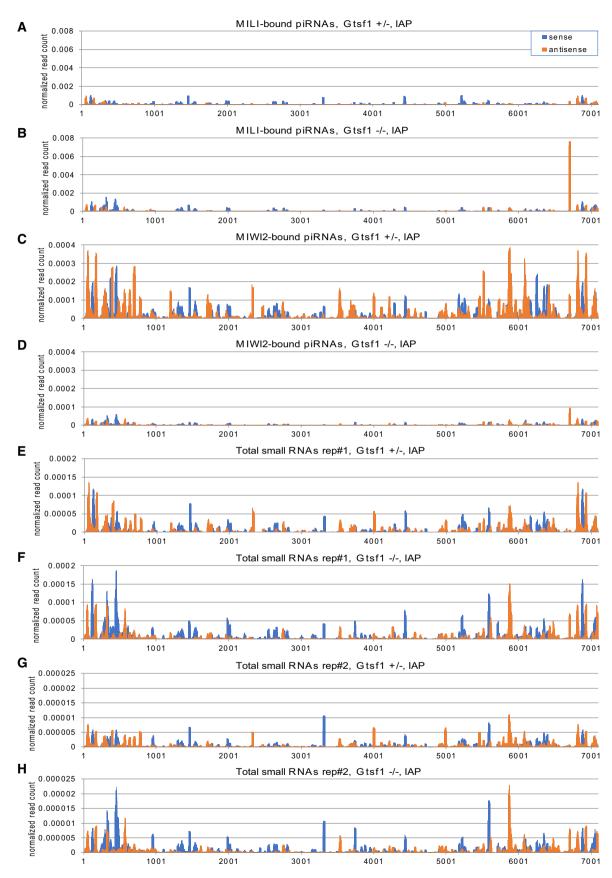


Figure EV6.