

The Diverted Total Synthesis of Carolacton-Inspired Analogs Yields Three Distinct Phenotypes in *Streptococcus mutans* Biofilms

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[^]*authors contributed equally*

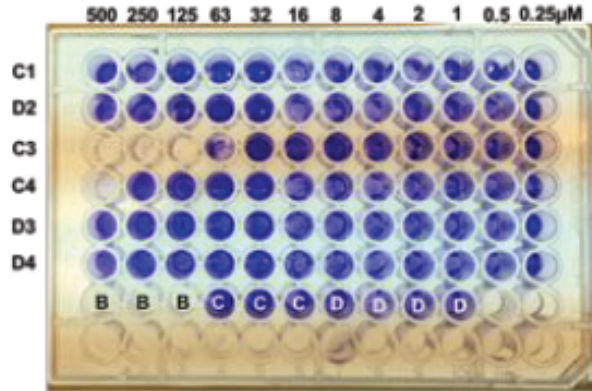
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6. References

| Compound | MIC | MBIC₅₀ | Phenotype |
|-------------------|------------|--------------------------|------------------|
| A1 | >500μM | >500μM | NORMAL |
| A2 | >500μM | >500μM | NORMAL |
| A3 | >500μM | 500μM | INHIBITED |
| A4 | >500μM | >500μM | NORMAL |
| B1 | >500μM | 500μM | INHIBITED |
| B2 | >500μM | >500μM | NORMAL |
| B3 | 500μM | 500μM | INHIBITED |
| B4 | 500μM | 500μM | INHIBITED |
| C1 | >500μM | >500μM | 1 |
| C2 | 500μM | 500μM | INHIBITED |
| C3 | 500μM | 63μM | INHIBITED |
| C4 | 500μM | 500μM | INHIBITED |
| D1 | >500μM | >500μM | NORMAL |
| D2 | >500μM | >500μM | 1 |
| D3 | >500μM | >500μM | NORMAL |
| D4 | >500μM | >500μM | 2 |
| Carolacton | >500μM | >500μM | 2 |
| | | 1 = microcolony | |
| | | 2 = carolacton-like | |

Figure S1. Summary of observed MIC values, MBIC values and phenotypes observed via confocal imaging. Experiments were completed in triplicate.

Full image of crystal violet plate and controls. OD measurements were taken at 595nm.



| | 500uM | 250 | 125 | 63 | 32 | 16 | 8 | 4 | 2 | 1 | 0.5 | 0.25 |
|----------|-------|------|------|------|------|------|------|------|------|------|------|------|
| C3 | 0.09 | 0.13 | 0.16 | 0.31 | 0.87 | 0.83 | 0.56 | 0.44 | 1.64 | 1.22 | 1.11 | 1.03 |
| C4 | 0.17 | 0.32 | 1.55 | 0.85 | 1.69 | 0.92 | 1.35 | 0.93 | 1.18 | 1.13 | 0.94 | 0.89 |
| Controls | 0.05 | 0.07 | 0.10 | 0.59 | 0.61 | 1.00 | 0.60 | 0.48 | 0.58 | 0.69 | | |
| | B | B | B | C | C | C | D | D | D | D | | |

OD values taken directly from plate reader. B= Blank, only media; C= Cells in media; D= 5% DMSO control. Controls were averaged for normalization below.

Blank Average = 0.07 ± 0.02

Cells Average = 0.73 ± 0.23

DMSO Average = 0.59 ± 0.09

| | 500uM | 250 | 125 | 63 | 32 | 16 | 8 | 4 | 2 | 1 | 0.5 | 0.25 |
|----|-------|------|------|------|------|------|------|------|------|------|------|------|
| C3 | 0.03 | 0.11 | 0.17 | 0.46 | 1.55 | 1.47 | 0.94 | 0.72 | 3.03 | 2.23 | 2.01 | 1.86 |
| C4 | 0.20 | 0.48 | 2.86 | 1.51 | 3.14 | 1.64 | 2.47 | 1.66 | 2.14 | 2.06 | 1.69 | 1.58 |

OD values normalized to the Blank and DMSO controls.

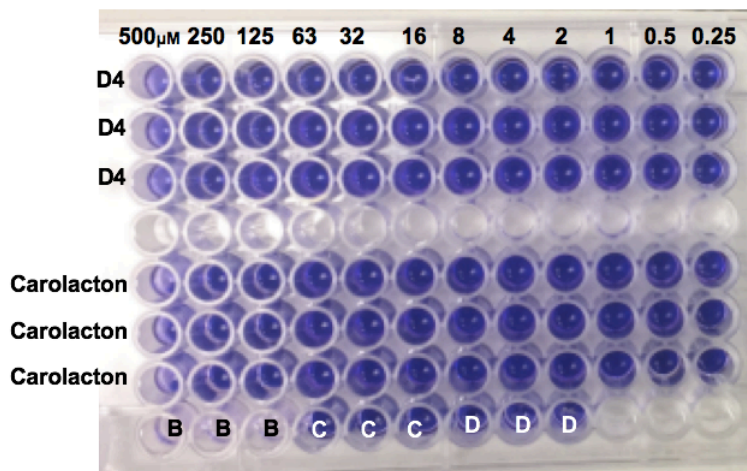


Figure S2. MBIC assay stained with crystal violet and subsequent OD values (normalized to blank and DMSO control)

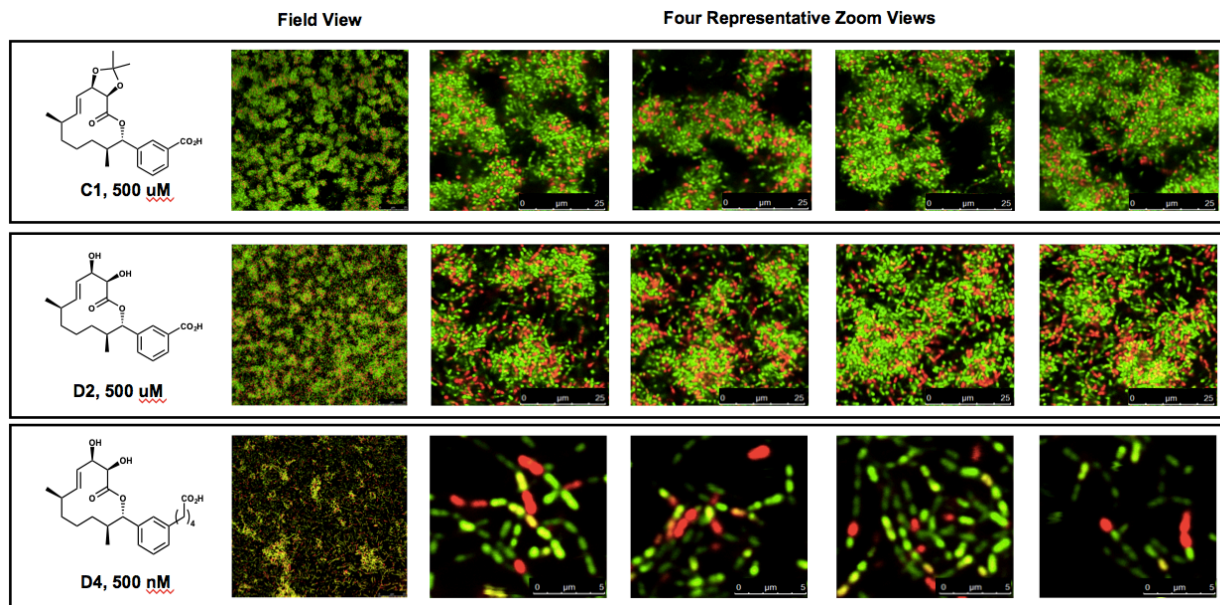


Figure S3. Field and representative zoom views of compounds **C1**, **D2**, and **D4** under confocal microscope.

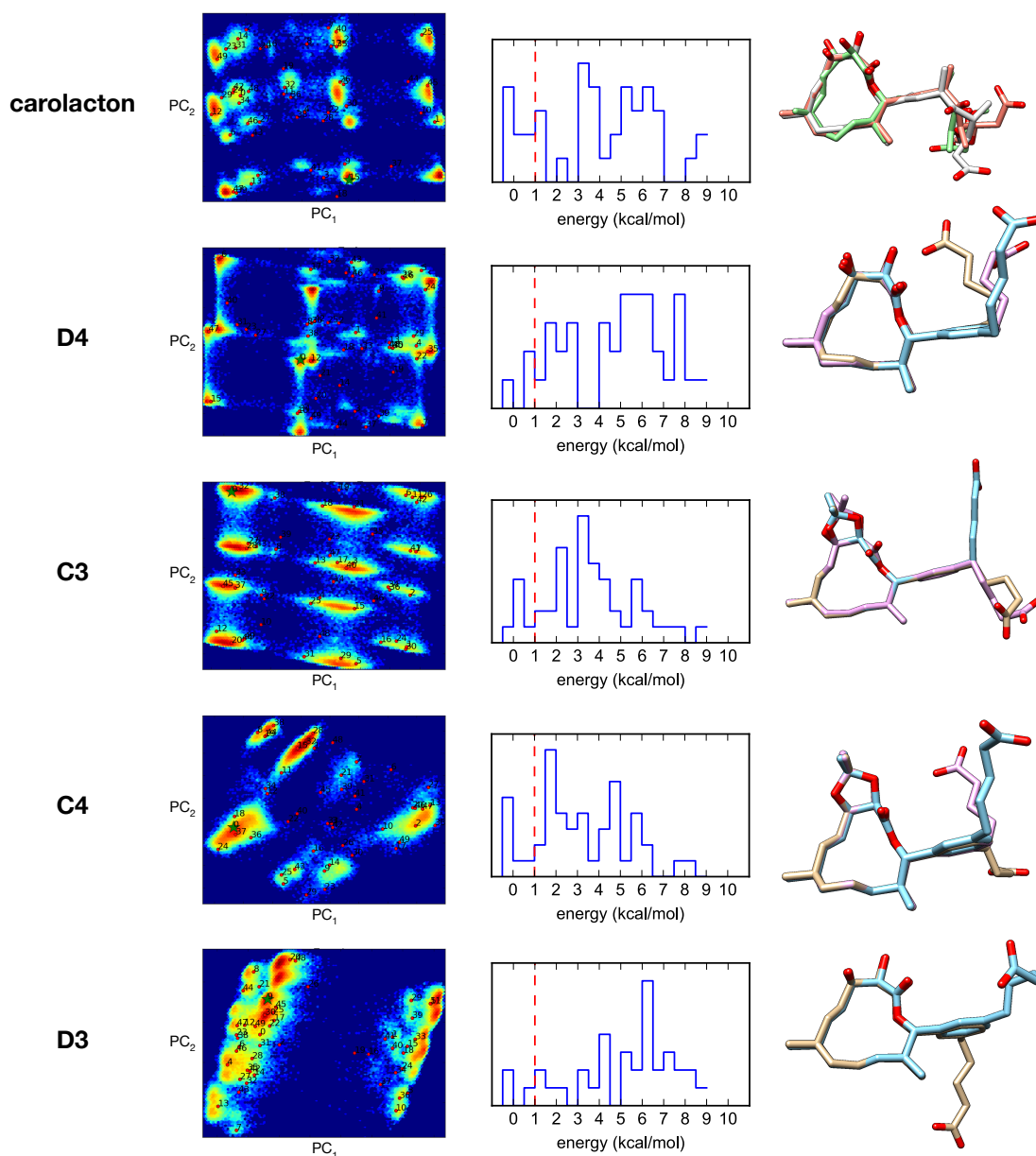


Figure S5. REMD/QM calculations for carolacton, **D4**, **C3**, **C4** and **D3**. Left column: REMD trajectory data projected to the first two principal components (PC_1 and PC_2) of dihedral angle space. Red dots indicate the 50 cluster generators obtained from k -centers clustering, which overlap well with regions of sampled density (shows as a log-scaled color gradient). The dispersion of clusters in the principal component subspace can be mainly attributed to variation in the flexible side chain of the carolacton analogs. Center column: Histograms of DFT energies calculated for the 50 QM-refined conformations. Right column: A superposition of the three lowest-energy conformations (within 0.2, 1.2, 0.7, 0.2 and 0.5 kcal/mol of the calculated ground state, respectively for carolacton, **D4**, **C3**, **C4** and **D3**).

2. Methods

General. NMR spectra were recorded using the following spectrometers: Bruker Advance 500 (500/125 MHz) or Bruker Advance 400 (400/100 MHz). Chemical shifts are quoted in ppm relative to tetramethylsilane and with the indicated solvent as an internal reference. The following abbreviations are used to describe signal multiplicities: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad), dd (doublet of doublets), dt (doublet of triplets), etc. Accurate mass spectra were recorded on an Agilent 6520 Accurate-Mass Q-TOF LC/MS, infrared spectra were obtained using a Thermo Nicolet Nexus 670 FTIR spectrophotometer and specific rotation measurements were made with a 1 dm path length using a Perkin Elmer 341 Polarimeter. Non-aqueous reactions were performed under an atmosphere of argon, in flame-dried glassware, with HPLC-grade solvents dried by passage through activated alumina. Amine bases were freshly distilled from CaH₂ prior to use. Brine refers to a saturated aqueous solution of sodium chloride. Products purified via flash chromatography using Biotage Isolera One Automated column. Reactions monitored via thin-layer chromatography (TLC) using EMD Millipore® TLC silica gel glass plates with KMnO₄ stain.

Materials. *Streptococcus mutans* wild-type strain UA159 was used for all bacterial cultures and was provided by Dr. Bettina Buttaro from Temple University Medical School, Philadelphia, PA. Bacteria were routinely maintained on in Bacto™ Todd- Hewitt (TH) agar plates and liquid cultures were grown in in Bacto™ Todd-Hewitt broth (THB). For growth of biofilms, THB was supplemented with 0.1% sucrose. Incubation was stagnant at 37 °C.

S. mutans MIC assay. Stock solution of carolacton analogs, 1000 μM, were serial diluted in THB media in flat-bottom 96-well microtiter plates. Then 2 μL of overnight cell culture are added to reach final volume of 200 μL. Plates are incubated at 37 °C in 5% CO₂ for 20-24 hours upon which time wells are evaluated visually for bacterial growth. The MIC is determined as the lowest concentration of compound resulting in no bacterial growth visible to the naked eye. Biological triplicates were performed to confirm results.

S. mutans biofilm model. Stock solution of carolacton analogs, 1000 μM, were serial diluted in THB media with 0.1% sucrose (w/v) in glass flat-bottom 96-well microtiter plates. Then 2 μL of overnight cell culture are added to reach final volume of 200 μL. Plates are incubated at 37 °C in 5% CO₂ for 18-20 hours (early stage biofilm) at which time wells are evaluated visually for bacterial growth.

S. mutans MBIC₅₀ assay. Biofilms were prepared with above procedure and evaluated visually and then emptied by inverting carefully, as to not disturb the biofilm. Wells were washed with 200μL of DI H₂O and dried for 6h at 37°C. Once dry, plates were incubated for 10min at room temperature with 50μL of 1% w/v crystal violet (25% ethanol in H₂O). Excess crystal violet was removed by submerging plates in fresh tap water until the run off was colorless. Plates were then inverted and dried at room temperature. Crystal violet was redissolved with 200μL of 95% ethanol, 100μL of which was then transferred to a fresh flat-bottom 96-well plate for absorbance measurements at 595nm. DMSO controls corresponding to each test concentration were performed. Three biological replicates were performed.

Confocal Imaging. Biofilms were grown with method described above. To note, the glass flat-bottom microtiter plates allowed for more efficient imaging compared to method using glass slips. After incubation, media was removed and each well was rinsed twice with PBS to remove planktonic cells. Subsequently, 10 μ L of BacLight LIVE/DEAD stain was added to each well. Images of biofilms were then obtained using the Leica DM IRE2 confocal microscope at Temple University Medical School.

Molecular Modeling. To model the structures of carolacton analogs in aqueous solution, a two-step REMD/QM approach was used, similar to previous work.³⁷ In the first step, replica-exchange molecular dynamics (REMD) was used for exhaustive conformational sampling. Selected structures from populated conformational basins were then further refined using DFT geometry optimization and energy calculation.

3. Biology

3.1 Procedures and General Notes

3.1.1 Bacterial Strains and Culture Conditions

Streptococcus mutans wild-type strain UA159 was obtained from Prof. Bettina Buttaro, Philadelphia, Pennsylvania.

3.1.2 MIC Assay

Stock solution of carolacton analogs, 1000 μM , were serial diluted in THB media in flat-bottom 96-well microtiter plates. Then 2 μL of overnight cell culture are added to reach final volume of 200 μL . Plates are incubated at 37°C in 5% CO_2 for 20-24 hours upon which time wells are evaluated visually for bacterial growth. The MIC is determined as the lowest concentration of compound resulting in no bacterial growth visible to the naked eye. Biological triplicates were performed to confirm results.

3.1.3 *S. mutans* biofilm model.

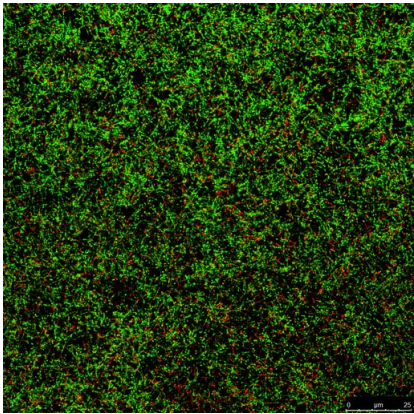
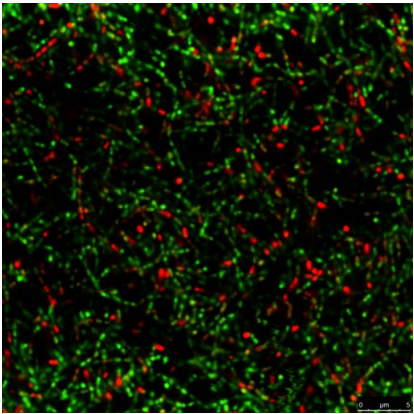
Stock solution of carolacton analogs, 1000 μM , were serial diluted in THB media with 0.1% sucrose (w/v) in glass flat-bottom 96-well microtiter plates. Then 2 μL of overnight cell culture are added to reach final volume of 200 μL . Plates are incubated at 37 °C in 5% CO_2 for 18-20 hours (early stage biofilm) at which time wells are evaluated visually for bacterial growth. After incubation, the media was removed from each desired well and the biofilm was rinsed with phosphate buffered saline (PBS) and then stained with Propidium Iodide/ Syto9.

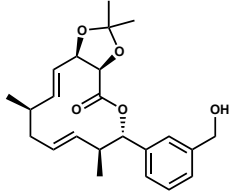
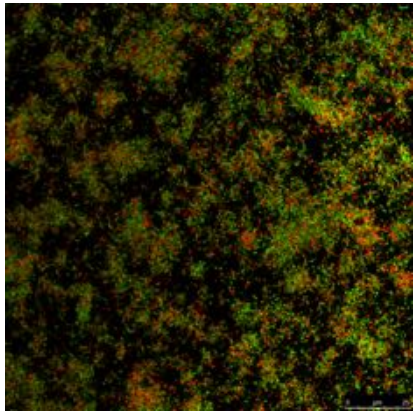
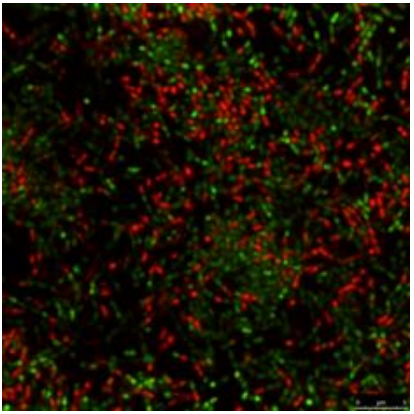
3.1.4 Biofilm Inhibition Assay (MBIC₅₀)

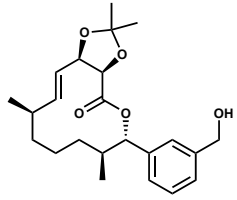
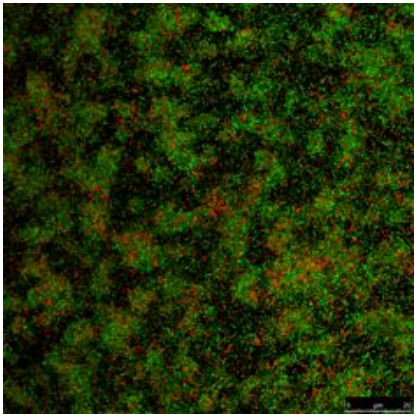
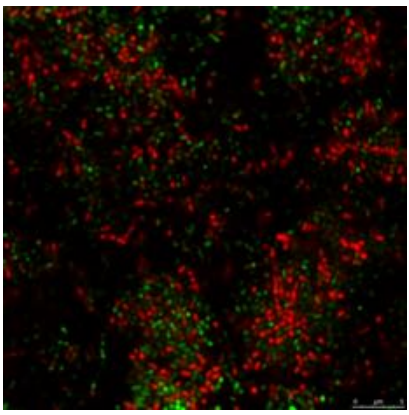
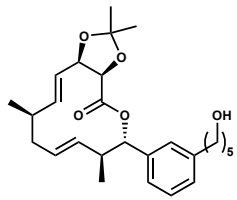
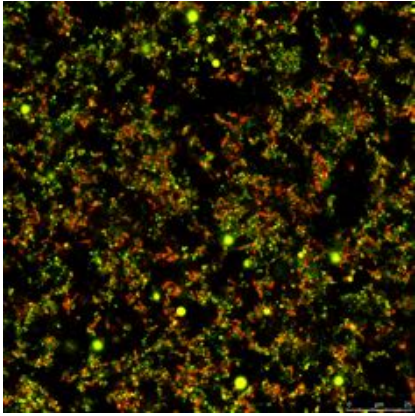
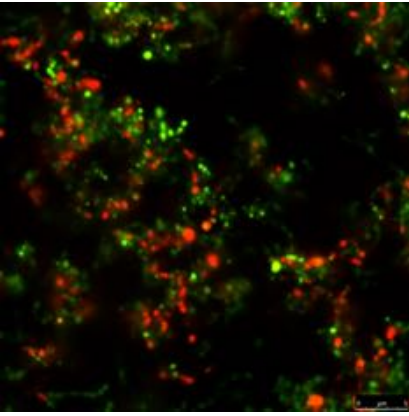
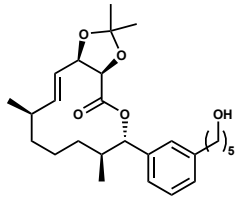
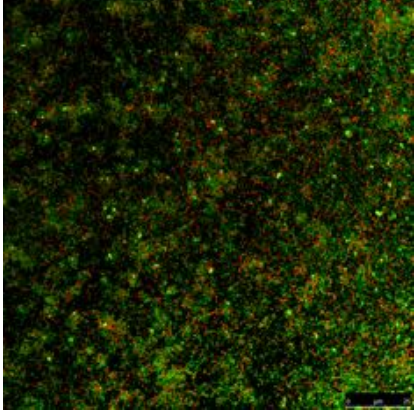
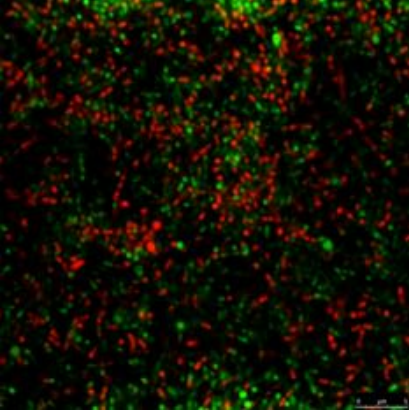
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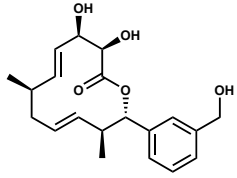
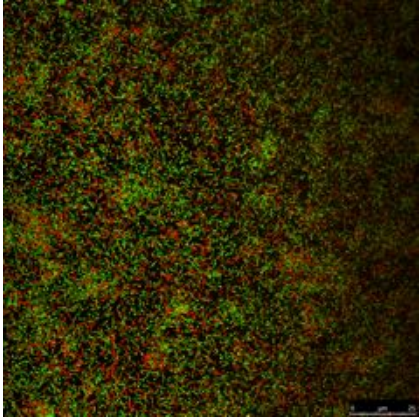
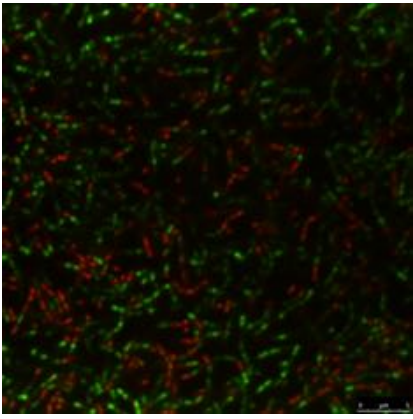
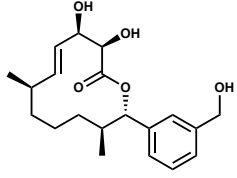
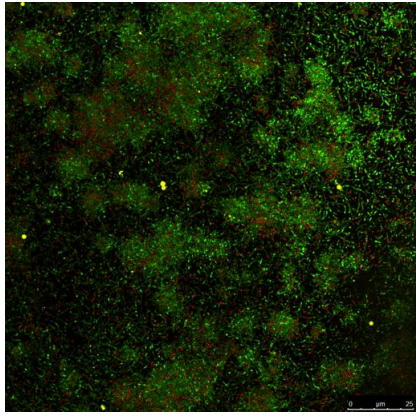
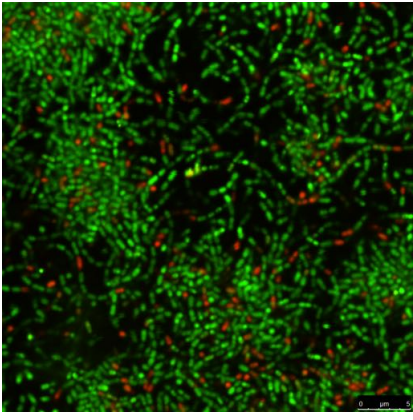
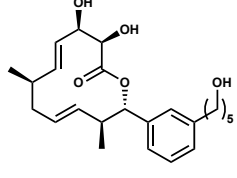
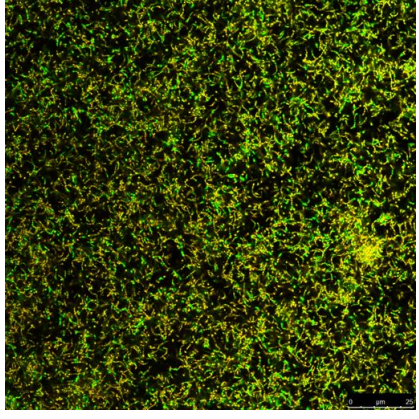
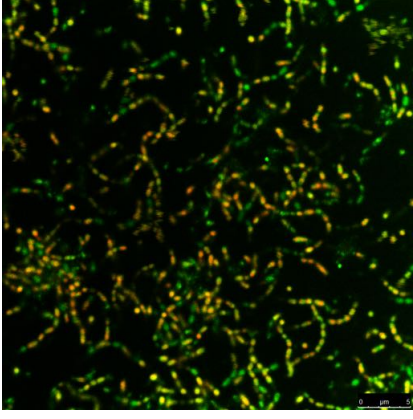
3.2 Biofilm Confocal Images

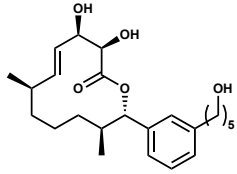
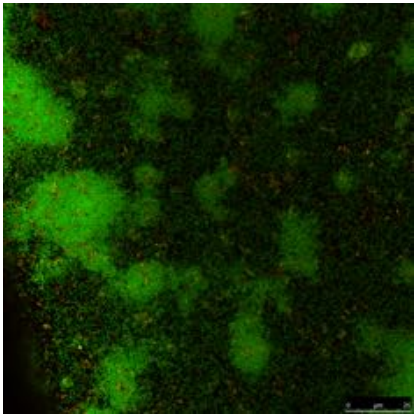
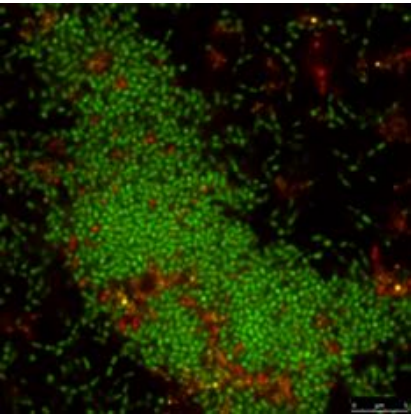
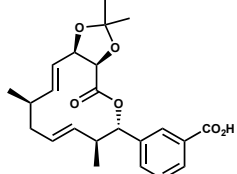
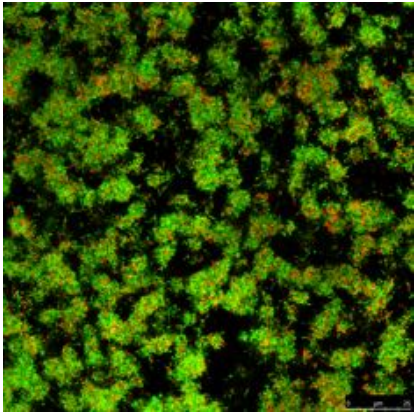
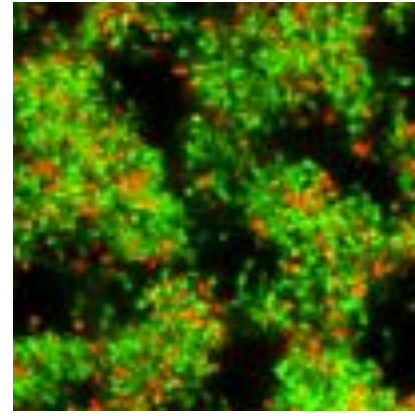
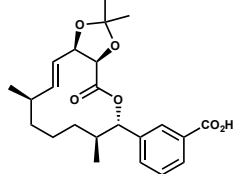
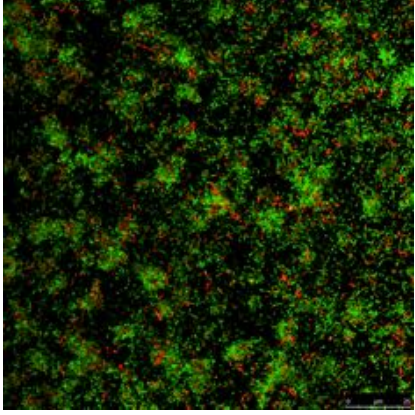
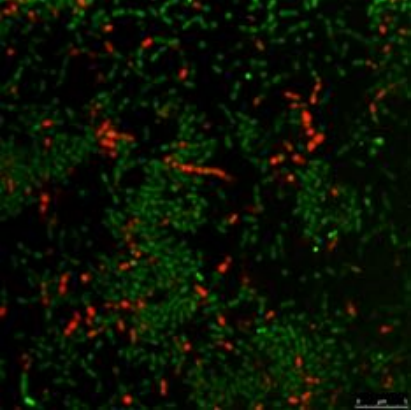
DMSO Control

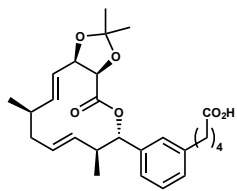


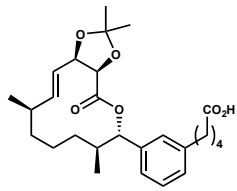


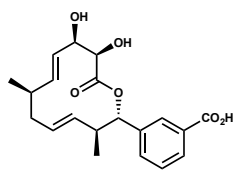
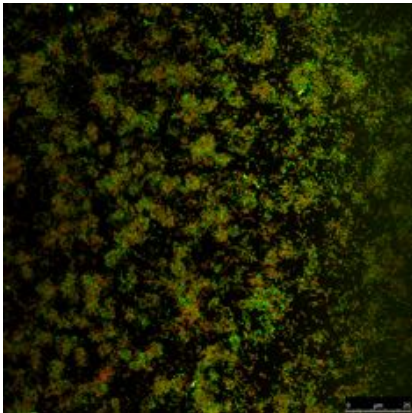
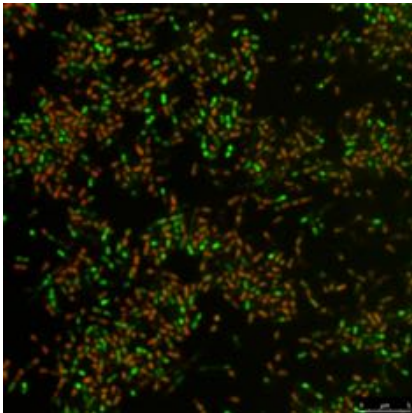
| | Full | Zoom |
|------|---|--|
| DMSO |  |  |

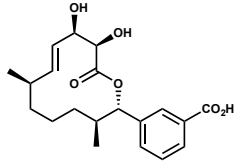
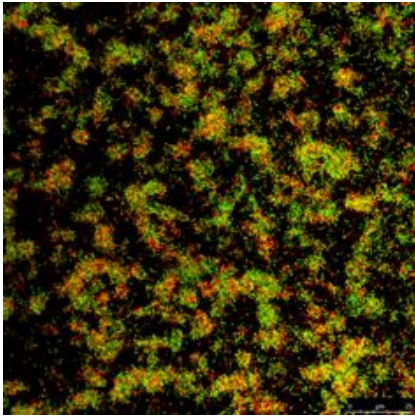
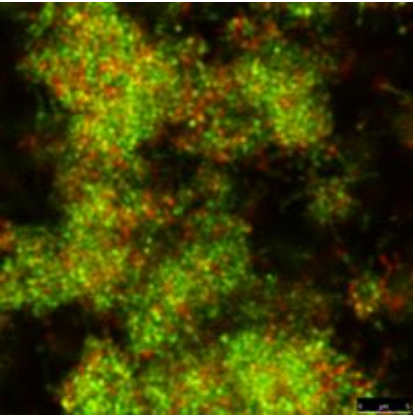
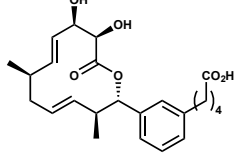
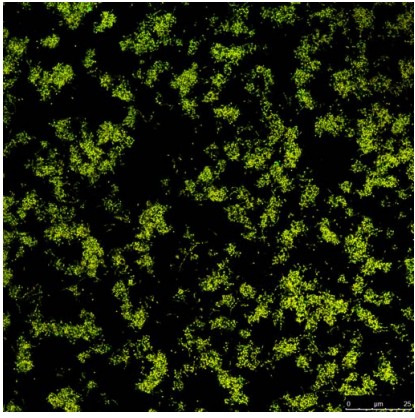
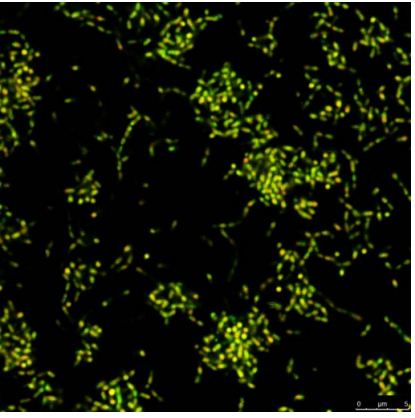
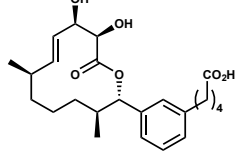
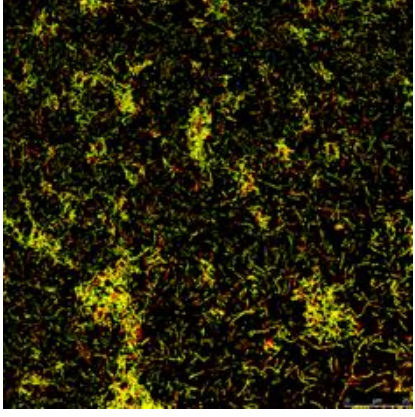
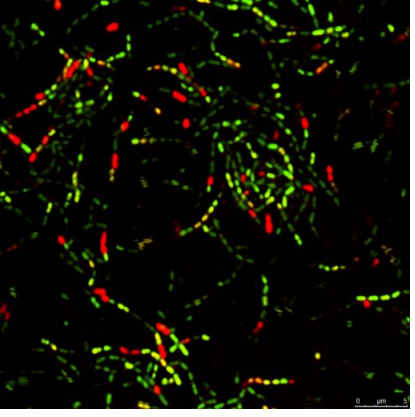
| Compound | Full (500 μ M) | Zoom (500 μ M) |
|---|---|--|
|  A1 |  |  |

| | | |
|--|---|--|
|  <p>A2</p> |  |  |
|  <p>A3</p> |  |  |
|  <p>A4</p> |  |  |

| | | |
|--|---|--|
|  <p>B1</p> |  |  |
|  <p>B2</p> |  |  |
|  <p>B3</p> |  |  |

| | | |
|--|---|--|
|  <p>B4</p> |  |  |
|  <p>C1</p> |  |  |
|  <p>C2</p> |  |  |

| | | |
|--|---|--|
|  <p>C3</p> |  |  |
|  <p>C4</p> |  |  |
|  <p>D1</p> |  |  |

| | | |
|---|---|--|
| <p style="text-align: center;">  D2 </p> |  |  |
| <p style="text-align: center;">  D3 </p> |  |  |
| <p style="text-align: center;">  D4 </p> |  |  |

4. Computational Methods

To model the structures of carolacton analogs in aqueous solution, a two-step REMD/QM approach was used, similar to previous work.² In the first step, replica-exchange molecular dynamics (REMD) was used for exhaustive conformational sampling. Selected structures from populated conformational basins were then further refined using DFT geometry optimization and energy calculation.

REMD simulation. Molecular topologies for carolacton, D4, C3, C4 and D3 were built in UCSF Chimera, and parameterized using the General Amber Force Field (GAFF)³ with partial changes from AM1-BCC.⁴ Molecular simulations were performed using the GROMACS 4.6 dynamics package.⁵ The OBC Generalized Born implicit solvation model was used in all simulations.⁶ Twenty-four replicas were used with exponentially-spaced temperatures ranging from 300 K to 450 K. Swaps between adjacent replicas were attempted every 10 ps, with acceptance ratios around 90%. REMD trajectories were simulated for 1.2 μ s (\times 24 replicas) for carolacton, and 1.6 μ s (\times 24 replicas) for D4, C3, C4 and D3 using stochastic integration (Langevin dynamics) with a 2 fs time step and water-like viscosity. Trajectory snapshots and energies were written every 10 ps.

For each analog, principal component analysis (PCA) of all rotatable dihedral angles (both in the macrolide ring and side chain) was used to project the trajectory data to a low-dimensional subspace, facilitating conformational clustering into 50 conformational basins using the *k*-centers clustering algorithm (Figure S2). The 50 conformations corresponding to each cluster generator were then selected for further refinement using *ab initio* QM.

***Ab initio* QM calculations for carolacton analogs.** DFT calculations were performed using Gaussian09.⁷ Structures were geometry-optimized using HF/6-31+G(d) with the PCM (polarizable continuum solvent model), following by single point energy calculation using B3LYP/6-311+G(2d,p) with PCM.

5. Synthesis

5.1 Instrumentation and General Notes

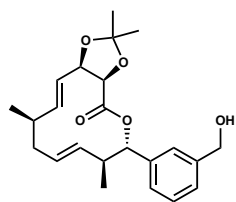
NMR spectra were recorded using the following spectrometers: Bruker Advance 500 (500/125 MHz) or Bruker Advance 400 (400/100 MHz). Chemical shifts are quoted in ppm relative to tetramethylsilane and with the indicated solvent as an internal reference. The following abbreviations are used to describe signal multiplicities: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad), dd (doublet of doublets), dt (doublet of triplets), etc. Accurate mass spectra were recorded on an Agilent 6520 Accurate-Mass Q-TOF LC/MS,

infrared spectra were obtained using a Thermo Nicolet Nexus 670 FTIR spectrophotometer, and specific rotation measurements were made with a 1 dm path length using a Perkin Elmer 341 Polarimeter.

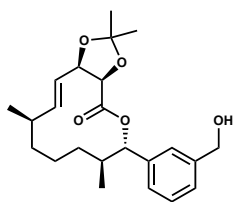
Non-aqueous reactions were performed under an atmosphere of argon, in oven and flame-dried glassware, with HPLC-grade solvents (dichloromethane (DCM), tetrahydrofuran (THF), toluene (PhMe), acetonitrile (ACN)) dried by passage through activated alumina. Amine bases were freshly distilled from CaH₂ prior to use. Brine refers to a saturated aqueous solution of sodium chloride.

All reported yields are based upon purity by ¹HNMR and therefore some yields are based upon samples containing residual amounts of solvent, as

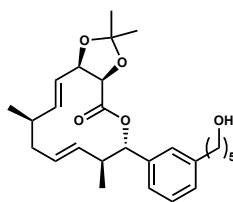
5.2 Experimental procedures



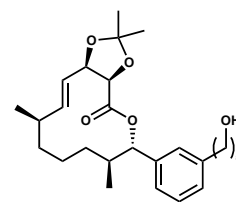
A1



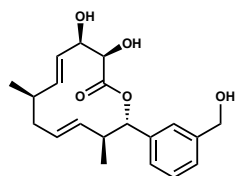
A2



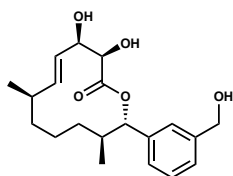
A3



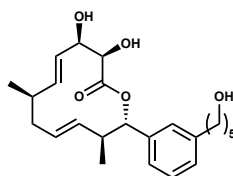
A4



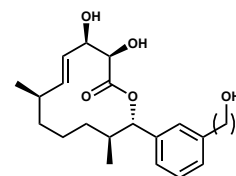
B1



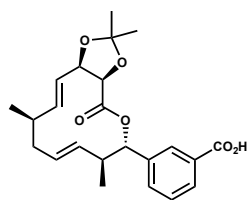
B2



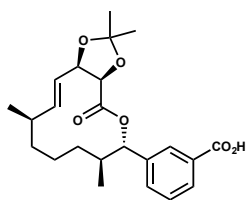
B3



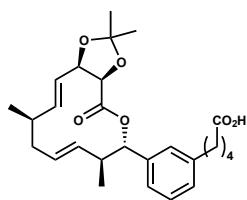
B4



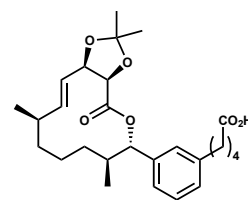
C1



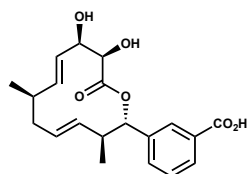
C2



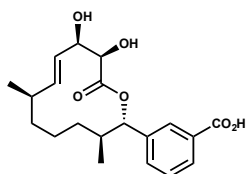
C3



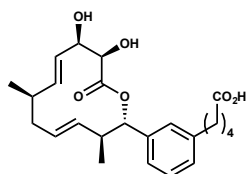
C4



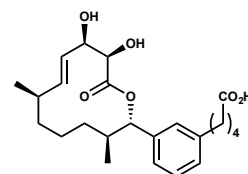
D1



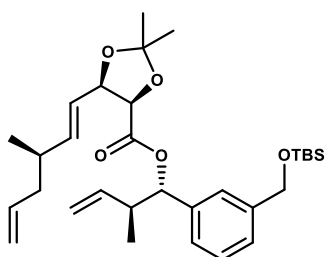
D2



D3



D4



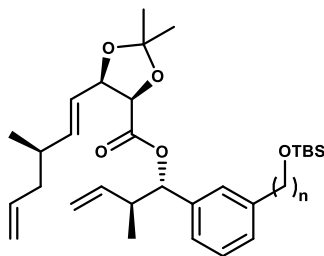
Representative Procedure A: EDC Esterification

(4*R*,5*R*)-(1*S*,2*S*)-1-(3-(((*tert*-Butyldimethylsilyl)oxy)methyl)phenyl)-2-methylbut-3-en-1-yl-2,2-dimethyl-5-((*R*,*E*)-3-methylhexa-1,5-dien-1-yl)-1,3-dioxolane-4-

carboxylate ((-)-9a) A flame dried flask was charged with argon and (-)-**3** (100 mg, 0.416 mmol) and DCM (4.2 mL) and subsequently cooled to 0 °C. Once cool, DMAP (34 mg, 0.027 mmol) and EDCI (106 mg, 0.555 mmol) were added consecutively followed by a solution of (-)-**5a** (85 mg, 0.277 mmol) in DCM (2.8 mL) added via syringe pump. The reaction was stirred 18 hours whereupon it was added to a separatory funnel containing H₂O (15 mL). The organic layer was separated and the aqueous layer was extracted with DCM (3 x 15 mL). The combined organic layers were washed with brine, dried over magnesium sulfate, filtered, and concentrated.

Purification by flash column chromatography (0 to 20% EtOAc in hexanes) afforded the product as a clear oil (130 mg, 89%).

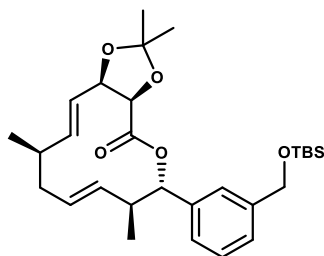
¹H NMR (500 MHz, CDCl₃) δ 7.32 – 7.24 (m, 3H), 7.22 – 7.17 (m, 1H), 5.85 – 5.64 (m, 3H), 5.61 (d, *J* = 7.5 Hz, 1H), 5.19 – 5.10 (m, 1H), 5.10 – 5.03 (m, 2H), 5.01 – 4.92 (m, 2H), 4.79 – 4.71 (m, 3H), 4.61 (d, *J* = 7.0 Hz, 1H), 2.74 – 2.64 (m, 1H), 2.08 – 1.95 (m, 2H), 1.89 – 1.83 (m, 1H), 1.66 (s, 3H), 1.40 (s, 3H), 0.95 (s, 9H), 0.90 (d, *J* = 6.9 Hz, 3H), 0.82 (d, *J* = 6.6 Hz, 3H), 0.10 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.12, 142.70, 141.55, 139.54, 138.49, 136.78, 128.24, 126.06, 125.80, 125.13, 122.13, 116.19, 115.96, 110.95, 80.33, 79.37, 78.34, 64.93, 43.04, 40.72, 35.93, 27.18, 26.08, 25.81, 19.16, 18.53, 16.06, -5.09; **IR** (neat): 3076, 2956, 2928, 2856, 1758, 1733, 1641, 1471, 1462, 1379, 1253, 1217, 1183, 1162, 1082, 1042, 1004, 973, 913, 880, 836, 815, 776, 703, 668, 567, 561 cm⁻¹; [α]_D²⁵ -61.6 (c = 0.92 in CHCl₃); **HRMS** (ES⁺): Found 551.3152 (-1.1 ppm), C₃₁H₄₈O₅SiNa (M+Na⁺) requires 551.3163



(4*R*,5*R*)-(1*S*,2*S*)-1-(3-(5-((*tert*-Butyldimethylsilyl)oxy)pentyl)phenyl)-2-methyl but-3-en-1-yl-2,2-dimethyl-5-((*R,E*)-3-methylhexa-1,5-dien-1-yl)-1,3-dioxolane-4-carboxylate ((-)-**9b**)

Prepared according to Representative Procedure A: (-)-**3** (650 mg, 2.71 mmol), in DCM (27 mL), DMAP (826 mg, 6.76 mmol), EDCI (778 mg, 4.06 mmol), and (-)-**5b** (490 mg, 1.35 mmol) in DCM (14 mL) yielded 759 mg (96%) of the ester. Purified by column chromatography (0 to 20% EtOAc in hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.24 – 7.18 (m, 1H), 7.14 – 7.03 (m, 3H), 5.82 – 5.61 (m, 3H), 5.58 (d, *J* = 7.6 Hz, 1H), 5.13 – 5.06 (m, 1H), 5.06 – 5.00 (m, 2H), 4.99 – 4.91 (m, 2H), 4.77 – 4.70 (m, 1H), 4.59 (d, *J* = 7.0 Hz, 1H), 3.60 (t, *J* = 6.6 Hz, 2H), 2.70 – 2.66 (m, 1H), 2.61 – 2.56 (m, 2H), 2.02 – 1.93 (m, 2H), 1.87 – 1.81 (m, 1H), 1.65 (s, 3H), 1.63 – 1.52 (m, 4H), 1.40 – 1.34 (m, 5H), 0.91 – 0.86 (m, 12H), 0.79 (d, *J* = 6.6 Hz, 3H), 0.04 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.17, 142.73, 142.69, 139.65, 138.52, 136.80, 128.20, 127.65, 124.78, 122.10, 116.18, 115.90, 110.96, 80.35, 79.40, 78.40, 63.35, 43.12, 40.73, 36.06, 35.91, 32.87, 31.41, 27.18, 26.14, 25.83, 25.72, 19.15, 18.53, 16.16, -5.11; **IR** (neat): 2929, 2857, 1735, 1641, 1608, 1461, 1372, 1234, 1163, 1091, 1022, 997, 974, 913, 880, 834, 774, 706, 661 cm⁻¹; **[α]_D²⁵** -47.3 (c = 0.30 in CHCl₃); **HRMS** (ES⁺): Found 602.4218 (-2.3 ppm), C₃₅H₆₀O₅SiN (M+NH₄⁺) requires 602.4241

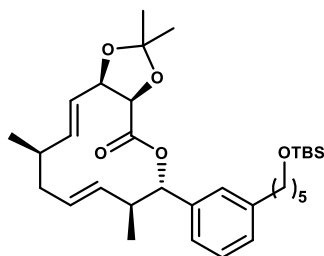


Representative Procedure B: Ring-Closing Metathesis

(3*aR*,6*S*,7*S*,8*E*,11*R*,12*E*,13*aR*)-6-(3-(((*tert*-Butyldimethylsilyl)oxy)methyl)phenyl)-2,2,7,11-tetramethyl-6,7,10,11-tetrahydro-3*aH*-[1,3]dioxolo[4,5-*c*][1]oxacyclo dodecin-4(13*aH*)-one
(-)-6a A flask was charged with (-)-**9a** (80 mg, 0.150 mmol) and DCM (30 mL). Grubbs 2nd generation catalyst (6.0 mg, 0.0076 mmol, 5 mol%) was added and the reaction was stirred at room temperature for 20 hours. The solvent was removed and the crude residue was purified by flash column chromatography (0 to 3% EtOAc in hexanes) to afford the product as a yellowish

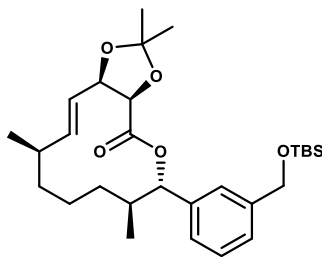
oil (59 mg, 78%). *NOTE: In our experience, degassing solvent prior to use had no noticeable effect on the outcome of the reaction.*

¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.25 (m, 3H), 7.21 – 7.17 (m, 1H), 5.87 – 5.75 (m, 1H), 5.58 (d, *J* = 10.8 Hz, 1H), 5.38 – 5.22 (m, 3H), 4.77 – 4.69 (m, 3H), 4.45 (d, *J* = 6.6 Hz, 1H), 2.66 – 2.49 (m, 1H), 2.39 – 2.19 (m, 2H), 2.16 – 1.98 (m, 1H), 1.66 (s, 3H), 1.37 (s, 3H), 1.08 (d, *J* = 6.7 Hz, 3H), 0.93 (s, 9H), 0.75 (d, *J* = 6.8 Hz, 3H), 0.08 (s, 6H); **¹³C NMR** (101 MHz, CDCl₃) δ 169.85, 141.81, 139.29, 138.85, 134.70, 130.68, 128.43, 126.56, 126.12, 125.54, 123.65, 111.03, 80.72, 79.11, 78.43, 64.91, 43.29, 38.54, 35.90, 26.90, 26.07, 25.99, 20.99, 17.61, -5.07; **IR** (neat): 2955, 2928, 2856, 1796, 1750, 1472, 1461, 1379, 1252, 1222, 1180, 1161, 1081, 1001, 968, 879, 814, 776, 735, 703, 670 cm⁻¹; **[α]_D²⁵** -34.4 (c = 1.00 in CHCl₃); **HRMS** (ES⁺): Found 523.2869 (+1.3 ppm), C₂₉H₄₄O₅SiNa (M+Na⁺) requires 523.2856



(3aR,6S,7S,8E,11R,12E,13aR)-6-(3-(5-((*tert*-Butyldimethylsilyl)oxy) pentyl)phenyl)-2,2,7,11-tetramethyl-6,7,10,11-tetrahydro-3aH-[1,3]dioxolo[4,5-*c*][1]oxacyclo dodecin-4(13aH)-one
(-)-6b Prepared according to Representative Procedure B: **(-)-9b** (750 mg, 1.28 mmol), Grubbs 2nd Generation catalyst (54 mg, 0.064 mmol, 5 mol%), and DCM (128 mL) yielded 710 mg (99%) of the product as a clear oil. Purified by column chromatography (0 to 3% EtOAc in hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.23 – 7.17 (m, 1H), 7.15 – 7.06 (m, 3H), 5.85 – 5.78 (m, 1H), 5.57 (d, *J* = 10.8 Hz, 1H), 5.35 – 5.29 (m, 1H), 5.29 – 5.23 (m, 2H), 4.77 – 4.70 (m, 1H), 4.47 (d, *J* = 6.7 Hz, 1H), 3.60 (t, *J* = 6.6 Hz, 2H), 2.62 – 2.55 (m, 3H), 2.34 – 2.21 (m, 2H), 2.09 – 1.99 (m, 1H), 1.67 (s, 3H), 1.63 – 1.57 (m, 2H), 1.57 – 1.51 (m, 2H), 1.39 – 1.33 (m, 5H), 1.09 (d, *J* = 6.8 Hz, 3H), 0.89 (s, 9H), 0.75 (d, *J* = 6.8 Hz, 3H), 0.04 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.81, 142.99, 139.25, 138.84, 134.79, 130.59, 128.44, 127.93, 125.20, 123.69, 111.00, 80.76, 79.11, 78.42, 63.28, 43.30, 38.53, 35.98, 35.88, 32.79, 31.40, 26.88, 26.10, 25.96, 25.68, 21.00, 18.47, 17.65, -5.15; **IR** (neat): 2954, 2928, 2856, 1751, 1608, 1586, 1459, 1379, 1251, 1222, 1180, 1085, 1002, 968, 880, 834, 813, 775, 705, 662 cm⁻¹; **[α]²⁵_D** -69.4 (c = 1.90 in CHCl₃); **HRMS** (ES⁺): Found 557.3639 (-1.8 ppm), C₃₃H₅₂O₅Si (M+H⁺) requires 557.3657



Representative Procedure C: Hydrogenation

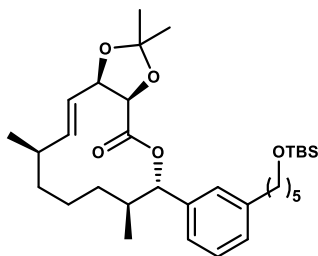
(3aR,6S,7S,11R,13aR,E)-6-(3-(((tert-Butyldimethylsilyl)oxy)methyl)phenyl)-2,2,7,11-

tetramethyl-6,7,8,9,10,11-hexahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-4(13aH)-

one ((-)-7a) To a solution of **(-)-6a** (36 mg, 0.072 mmol) in ethanol (7.2 mL) was added palladium on carbon (10% w/w, 6 mg). The reaction was sparged with hydrogen gas 5 times from a balloon and stirred for 1.5 hours. The reaction was passed through a pad of celite with EtOAc (100 mL) and the solvent was removed. The residue was purified by preparative TLC (3% EtOAc in hexanes, eluted twice, to remove a more non-polar impurity) to afford the product

as a clear oil (25 mg, 69%).

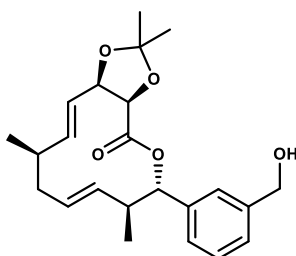
¹H NMR (400 MHz, CDCl₃) δ 7.26 – 7.20 (m, 3H), 7.19 – 7.14 (m, 1H), 5.72 – 5.59 (m, 2H), 5.51 (d, *J* = 11.3 Hz, 1H), 4.87 – 4.82 (m, 1H), 4.70 (s, 2H), 4.52 (d, *J* = 6.6 Hz, 1H), 2.44 – 2.33 (m, 1H), 2.28 – 2.17 (m, 1H), 2.03 – 1.92 (m, 1H), 1.68 (s, 3H), 1.41 – 1.36 (m, 6H), 1.02 (d, *J* = 6.6 Hz, 2H), 0.93 (s, 9H), 0.68 (d, *J* = 7.0 Hz, 3H), 0.08 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.49, 141.63, 139.92, 135.89, 128.32, 126.56, 125.95, 125.46, 122.37, 110.98, 79.40, 78.66, 65.00, 36.26, 35.81, 33.87, 29.44, 26.95, 26.09, 26.03, 21.06, 18.54, 17.99, 15.91, -5.06; **IR** (neat): 2956, 2928, 2856, 2904, 1752, 1462, 1379, 1250, 1224, 1176, 1123, 1081, 1006, 976, 815, 776, 719, 702, 668, 617 cm⁻¹; [α]_D²⁵ -106 (c = 1.88 in CHCl₃); **HRMS** (ES⁺): Found 525.3036 (+2.4 ppm), C₂₉H₄₆O₅SiNa (M+Na⁺) requires 525.3012



(3a*R*,6*S*,7*S*,11*R*,13a*R*,*E*)-6-(3-(5-((*tert*-Butyldimethylsilyl)oxy)pentyl)phenyl)-2,2,7,11-tetramethyl-6,7,8,9,10,11-hexahydro-3a*H*-[1,3]dioxolo[4,5-*c*][1]oxacyclododecin-4(13a*H*)-one ((-)-7b) Prepared according to Representative Procedure C: (-)-6b (20 mg, 0.036 mmol), Pd/C (10% w/w, 2 mg), and EtOH (3.6 mL) yielded 11 mg (55%) of the product as a clear oil. Purified by preparative TLC (3% EtOAc in hexanes, eluted twice).

¹H NMR (500 MHz, CDCl₃) δ 7.20 – 7.15 (m, 1H), 7.11 – 7.08 (m, 2H), 7.07 – 7.03 (m, 1H), 5.71 – 5.60 (m, 2H), 5.49 (d, *J* = 11.5 Hz, 1H), 4.87 – 4.80 (m, 1H), 4.54 (d, *J* = 6.6 Hz, 1H), 3.59 (t, *J* = 6.6 Hz, 1H), 2.58 – 2.54 (m, 2H), 2.42 – 2.36 (m, 1H), 2.28 – 2.17 (m, 1H), 2.01 – 1.94 (m, 1H), 1.68 (s, 3H), 1.63 – 1.50 (m, 6H), 1.39 – 1.37 (m, 3H), 1.37 – 1.32 (m, 3H), 1.08 –

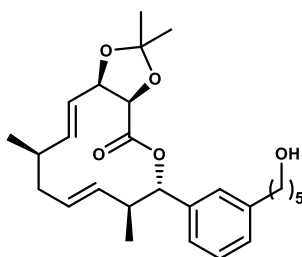
1.04 (m, 2H), 1.02 (d, $J = 6.5$ Hz, 3H), 0.89 (s, 9H), 0.68 (d, $J = 7.0$ Hz, 3H), 0.04 (s, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 169.49, 142.84, 139.98, 135.91, 128.29, 128.24, 127.83, 125.18, 122.37, 110.99, 79.50, 78.69, 63.37, 36.36, 36.02, 35.82, 33.90, 32.85, 31.41, 29.44, 26.96, 26.14, 26.04, 25.73, 21.07, 18.53, 17.97, 15.95, -5.10; IR (neat): 2928, 2856, 2360, 1753, 1608, 1460, 1379, 1250, 1223, 1176, 1085, 1005, 977, 874, 834, 775, 705, 667 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -63.9 ($c = 0.43$ in CHCl_3); HRMS (ES^+): Found 581.3630 (-0.8 ppm), $\text{C}_{33}\text{H}_{54}\text{O}_5\text{SiNa}$ ($\text{M}+\text{Na}^+$) requires 581.3638



Representative Procedure D: TBS removal

(3*aR*,6*S*,7*S*,8*E*,11*R*,12*E*,13*aR*)-6-(3-(Hydroxymethyl)phenyl)-2,2,7,11-tetramethyl-6,7,10,11-tetrahydro-3*aH*-[1,3]dioxolo[4,5-*c*][1]oxacyclododecin-4(13*aH*)-one ((-)-A1) To a solution of (-)-6a (21 mg, 0.0420 mmol) in THF (0.42 mL) was added *tetra*-butylammonium fluoride (1M in THF, 0.13 mL), and the reaction was stirred for 1 hour. The reaction was quenched with saturated aqueous ammonium chloride and diluted in diethyl ether. The organic layer was separated and the aqueous layer was extracted 4 times with diethyl ether. The combined organic layers were washed with water and brine, dried over sodium sulfate, filtered, and concentrated. The crude product was purified by preparative TLC (3:2 hexanes/EtOAc) to afford the product alcohol as a clear oil (15 mg, 94%).

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.31 (m, 1H), 7.32 – 7.26 (m, 1H), 7.28 – 7.22 (m, 1H), 5.82 (dd, *J* = 15.7, 6.6 Hz, 1H), 5.58 (d, *J* = 10.8 Hz, 1H), 5.36 – 5.23 (m, 3H), 4.76 – 4.70 (m, 1H), 4.67 (s, 2H), 4.46 (d, *J* = 6.7 Hz, 1H), 2.69 – 2.52 (m, 1H), 2.36 – 2.20 (m, 2H), 2.11 – 1.99 (m, 1H), 1.65 (s, 3H), 1.37 (s, 3H), 1.08 (d, *J* = 6.5 Hz, 3H), 0.76 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.89, 141.29, 139.45, 139.38, 134.63, 130.77, 128.78, 127.33, 127.02, 126.40, 123.67, 111.10, 80.69, 79.22, 78.43, 65.31, 43.15, 38.49, 35.87, 26.91, 25.99, 20.95, 17.61; **IR** (neat): 3467, 2959, 2929, 2872, 1746, 1455, 1378, 1293, 1251, 1221, 1183, 1161, 1082, 1039, 1000, 969, 910, 880, 785, 729, 705, 673, 647 cm⁻¹; [α]_D²⁵ -76.4 (c = 1.40 in CHCl₃); **HRMS** (ES⁺): Found 387.2169 (+0.3 ppm), C₂₃H₃₀O₅ (M+H⁺) requires 387.2166

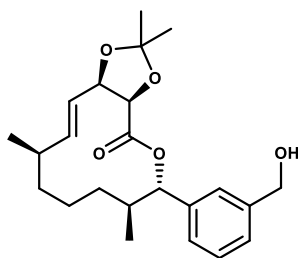


(3aR,6S,7S,8E,11R,12E,13aR)-6-(3-(5-Hydroxypentyl)phenyl)-2,2,7,11-tetramethyl-6,7,10,11-tetrahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-4(13aH)-one((-)-A3)

Prepared according to Representative Procedure D: (-)-**6b** (35 mg, 0.063 mmol), TBAF (1M in THF, 0.63 mL), and THF (0.63 mL) yielded 19 mg (68%) of the product as a clear oil. Purified by preparative TLC (1:1 EtOAc/hexanes).

¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.20 (m, 1H), 7.16 – 7.08 (m, 3H), 5.82 (dd, *J* = 15.6, 6.6 Hz, 1H), 5.56 (d, *J* = 10.8 Hz, 1H), 5.37 – 5.22 (m, 3H), 4.73 (t, *J* = 7.1 Hz, 1H), 4.47 (d, *J* = 6.6 Hz, 1H), 3.63 (t, *J* = 6.6 Hz, 2H), 2.64 – 2.56 (m, 3H), 2.37 – 2.23 (m, 2H), 2.12 – 2.02 (m, 1H), 1.66 (s, 3H), 1.64 – 1.53 (m, 4H), 1.43 – 1.33 (m, 5H), 1.09 (d, *J* = 6.7 Hz, 3H), 0.76 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.88, 142.83, 139.44, 138.90, 134.81, 130.65,

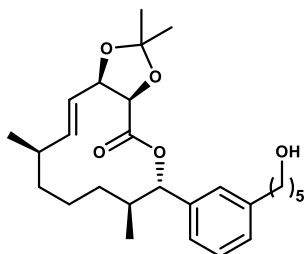
128.50, 128.12, 125.24, 123.70, 111.09, 80.88, 79.21, 78.44, 63.05, 43.20, 38.52, 35.91, 32.73, 31.28, 26.92, 25.99, 25.51, 20.99, 17.65; **IR** (neat): 3313, 2927, 2855, 1748, 1608, 1456, 1379, 1221, 1182, 1083, 969, 880, 786, 753, 705, 667 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -46.2 ($c = 0.58$ in CHCl_3); **HRMS** (ES^+): Found 443.2788 (-0.4 ppm), $\text{C}_{27}\text{H}_{38}\text{O}_5$ ($\text{M}+\text{H}^+$) requires 443.2792



(3aR,6S,7S,11R,13aR,E)-6-(3-(Hydroxymethyl)phenyl)-2,2,7,11-tetramethyl-6,7,8,9,10,11-hexahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-4(13aH)-one ((-)-A2) Prepared according to Representative Procedure D: (-)-7a (28 mg, 0.058 mmol), TBAF (1M in THF, 0.17 mL), and THF (0.56 mL) yielded 19.5 mg (90%) of the product as a clear oil. Purified by preparative TLC (3:2 hexanes/EtOAc).

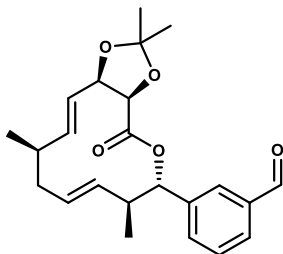
^1H NMR (500 MHz, CDCl_3) δ 7.31 – 7.24 (m, 3H), 7.24 – 7.21 (m, 1H), 5.70 – 5.59 (m, 2H), 5.52 (d, $J = 11.5$ Hz, 1H), 4.87 – 4.82 (m, 1H), 4.66 (s, 2H), 4.54 (d, $J = 6.6$ Hz, 1H), 2.44 – 2.34 (m, 1H), 2.28 – 2.20 (m, 1H), 2.02 – 1.94 (m, 1H), 1.67 (s, 3H), 1.44 – 1.33 (m, 7H), 1.11 – 1.05 (m, 2H), 1.02 (d, $J = 6.6$ Hz, 3H), 0.69 (d, $J = 7.0$ Hz, 3H); **^{13}C NMR** (101 MHz, CDCl_3) δ 169.60, 141.08, 140.42, 135.87, 128.70, 127.19, 126.84, 126.42, 122.32, 111.03, 79.32, 78.63, 65.38, 36.23, 35.81, 33.83, 29.34, 26.94, 26.01, 21.07, 17.91, 15.91; **IR** (neat): 3500, 2931, 2360, 2342, 1748, 1455, 1379, 1223, 1182, 1162, 1123, 1083, 974, 909, 873, 837, 738, 729, 704, 648 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -100 ($c = 0.50$ in CHCl_3); **HRMS** (ES^+): Found 411.2125 (-1.7 ppm), $\text{C}_{23}\text{H}_{32}\text{O}_5\text{Na}$

(M+Na⁺) requires 411.2142



(3*a*R,6*S*,7*S*,11*R*,13*a*R,*E*)-6-(3-(5-Hydroxypentyl)phenyl)-2,2,7,11-tetramethyl-6,7,8,9,10,11-hexahydro-3*a*H-[1,3]dioxolo[4,5-*c*][1]oxacyclododecin-4(13*a*H)-one ((-)-**A4**) Prepared according to representative Procedure D: (-)-**7b** (21 mg, 0.038 mmol), TBAF (1M in THF, 0.11 mL), and THF (0.38 mL) yielded 10.7 mg (64%) of the product as a clear oil. Purified by preparative TLC (3:2 hexanes/EtOAc).

¹H NMR (400 MHz, CDCl₃) δ 7.21 – 7.15 (m, 1H), 7.12 – 7.08 (m, 2H), 7.07 – 7.03 (m, 1H), 5.71 – 5.57 (m, 2H), 5.48 (d, *J* = 11.5 Hz, 1H), 4.85 (dd, *J* = 6.5, 3.2 Hz, 1H), 4.54 (d, *J* = 6.6 Hz, 1H), 3.62 (t, *J* = 6.6 Hz, 2H), 2.58 (t, *J* = 7.6 Hz, 2H), 2.43 – 2.35 (m, 1H), 2.28 – 2.18 (m, 1H), 2.04 – 1.93 (m, 1H), 1.68 (s, 3H), 1.65 – 1.52 (m, 6H), 1.42 – 1.32 (m, 7H), 1.13 – 1.05 (m, 2H), 1.02 (d, *J* = 6.6 Hz, 3H), 0.68 (d, *J* = 7.0 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 169.53, 142.59, 139.95, 135.88, 128.36, 128.27, 128.07, 125.11, 122.34, 111.02, 79.53, 78.64, 63.04, 36.23, 35.88, 35.81, 33.87, 32.73, 31.20, 29.39, 26.94, 26.01, 25.45, 21.07, 17.93, 15.94; **IR** (neat): 3392, 2932, 2859, 2360, 2341, 1748, 1608, 1456, 1380, 1184, 1123, 1083, 976, 874, 836, 785, 752, 705, 667 cm⁻¹; **[α]_D²⁵** -100.6 (c = 0.72 in CHCl₃); **HRMS** (ES⁺): Found 445.2935 (-1.4 ppm), C₂₇H₄₀O₅ (M+H⁺) requires 445.2949

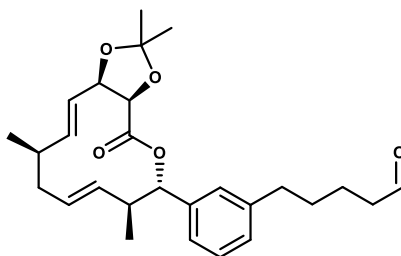


Representative Procedure E: Parikh-Döering Oxidation

3-((3aR,6S,7S,8E,11R,12E,13aR)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,10,11,13a-hexahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)benzaldehyde ((-)-10a) To a solution of (-)-**A1** (10 mg, 0.026 mmol) in DCM (0.26 mL) was added DMSO (0.04 mL, 0.52 mmol) and triethylamine (0.04 mL, 0.26 mmol). The solution was cooled to 0 °C and SO₃-Pyr (33 mg, 0.21 mmol) was added in a single portion. The reaction mixture was removed from the cooling bath and allowed to stir at room temperature for 45 minutes. The reaction was then quenched with saturated aqueous ammonium chloride, diluted in DCM, and poured into a separatory funnel. The organic layer was removed and the aqueous layer was extracted 5x DCM (5 mL). The combined organic layers were washed with brine, dried over sodium sulfate, filtered, and reduced. The residue was purified by preparative TLC (4:1 hexanes/EtOAc) to afford the product as a clear oil (8 mg, 80%).

¹H NMR (500 MHz, CDCl₃) δ 10.01 (s, 1H), 7.88 (s, 1H), 7.85 – 7.78 (m, 1H), 7.64 – 7.55 (m, 1H), 7.50 (t, *J* = 7.6 Hz, 1H), 5.83 (dd, *J* = 15.7, 6.7 Hz, 1H), 5.65 (d, *J* = 10.8 Hz, 1H), 5.36 – 5.21 (m, 3H), 4.78 – 4.72 (m, 1H), 4.48 (d, *J* = 6.6 Hz, 1H), 2.66 – 2.57 (m, 1H), 2.37 – 2.31 (m, 1H), 2.30 – 2.24 (m, 1H), 2.12 – 2.05 (m, 1H), 1.66 (s, 3H), 1.37 (s, 3H), 1.09 (d, *J* = 6.8 Hz, 3H), 0.77 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 192.15, 169.90, 140.35, 139.60, 136.78, 134.20, 134.07, 131.21, 130.07, 129.32, 128.66, 123.52, 111.19, 79.99, 79.23, 78.34,

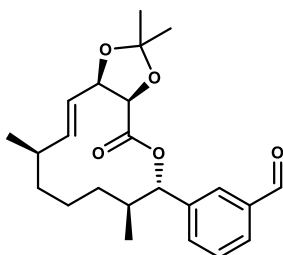
43.20, 38.41, 35.84, 26.88, 25.96, 20.90, 17.45; **IR** (neat): 2958, 2927, 2876, 2851, 2726, 2257, 1821, 1748, 1699, 1604, 1535, 1455, 1379, 1290, 1250, 1220, 1180, 1161, 1040, 1000, 969, 911, 880, 785, 731, 696, 675, 650 cm^{-1} ; $[\alpha]_D^{25}$ -65.3 ($c = 0.30$ in CHCl_3); **HRMS** (ES^+): Found 407.1825 (-0.9 ppm), $\text{C}_{23}\text{H}_{28}\text{O}_5\text{Na}$ ($\text{M}+\text{Na}^+$) requires 407.1834



5-(3-((3*aR*,6*S*,7*S*,8*E*,11*R*,12*E*,13*aR*)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,10,11,13*a*-hexahydro-3*aH*-[1,3]dioxolo[4,5-*c*][1]oxacyclododecin-6-yl)phenyl)pentanal ((-)-**11a**) Prepared according to Representative Procedure E: (-)-**A3** (10 mg, 0.026 mmol), DMSO (0.03 mL, 0.45 mmol), triethylamine (0.03 mL, 0.23 mmol), SO_3 -Pyr (29 mg, 0.18 mmol), and DCM (0.23 mL) yielded 6.2 mg (63%) of the product as a clear oil. Purified by preparative TLC (4:1 hexanes/EtOAc).

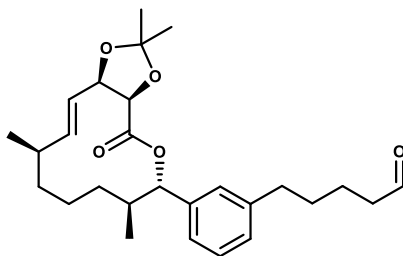
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 9.76 (t, $J = 3.6$ Hz, 1H), 7.22 (t, $J = 7.4$ Hz, 1H), 7.16 – 7.07 (m, 3H), 5.82 (dd, $J = 15.6, 6.6$ Hz, 1H), 5.56 (d, $J = 10.9$ Hz, 1H), 5.36 – 5.23 (m, 3H), 4.76 – 4.70 (m, 1H), 4.47 (d, $J = 6.6$ Hz, 1H), 2.64 – 2.52 (m, 3H), 2.49 – 2.42 (m, 2H), 2.36 – 2.21 (m, 2H), 2.11 – 2.01 (m, 1H), 1.70 – 1.62 (m, 7H), 1.37 (s, 3H), 1.08 (d, $J = 6.7$ Hz, 3H), 0.75 (d, $J = 6.8$ Hz, 3H); **$^{13}\text{C NMR}$** (126 MHz, CDCl_3) δ 202.69, 169.89, 142.25, 139.42, 139.06, 134.77, 130.70, 128.56, 128.47, 125.49, 123.69, 111.08, 80.79, 79.20, 78.46, 43.87, 43.29, 38.54, 35.90, 35.71, 30.97, 26.92, 26.00, 21.86, 21.01, 17.66; **IR** (neat): 3031, 2959, 2024, 2873, 2854, 2720, 2360, 2343, 1748, 1724, 1608, 1559, 1488, 1457, 1379, 1252, 1222, 1183, 1084, 1041, 1000,

970, 880, 786, 706, 667 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -84 ($c = 0.22$ in CHCl_3); **HRMS** (ES^+): Found 463.2487 (+2.7 ppm), $\text{C}_{27}\text{H}_{36}\text{O}_5\text{Na}$ ($\text{M}+\text{Na}^+$) requires 463.2460



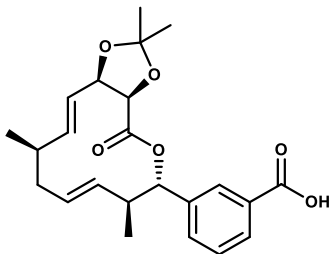
3-((3aR,6S,7S,11R,13aR,E)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,8,9,10,11,13a-octahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)benzaldehyde ((-)-10b) Prepared according to Representative Procedure E: (-)-**A2** (14 mg, 0.036 mmol), DMSO (0.05 mL, 0.72 mmol), triethylamine (0.05 mL, 0.36 mL) SO_3 -Pyr (46 mg, 0.29 mmol), and DCM (0.36 mL) yielded 12.5 mg (89%) of the product as a clear oil. Purified by preparative TLC (4:1 hexanes/EtOAc).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 9.99 (s, 1H), 7.84 – 7.83 (m, 1H), 7.80 – 7.77 (m, 1H), 7.60 – 7.55 (m, 1H), 7.47 (t, $J = 7.6$ Hz, 1H), 5.74 – 5.63 (m, 2H), 5.59 (d, $J = 11.5$ Hz, 1H), 4.86 (dd, $J = 6.5, 3.3$ Hz, 1H), 4.55 (d, $J = 6.6$ Hz, 1H), 2.46 – 2.35 (m, 1H), 2.33 – 2.23 (m, 1H), 2.06 – 1.97 (m, 1H), 1.67 (s, 3H), 1.46 – 1.35 (m, 6H), 1.13 – 1.06 (m, 2H), 1.02 (d, $J = 6.6$ Hz, 3H), 0.69 (d, $J = 7.0$ Hz, 3H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 192.19, 169.65, 141.40, 136.72, 135.92, 134.18, 129.80, 129.23, 128.77, 122.31, 111.13, 78.64, 78.58, 36.33, 35.81, 33.77, 29.27, 26.92, 25.99, 21.05, 17.90, 15.79; **IR** (neat): 2959, 2926, 2852, 2729, 2160, 2031, 1974, 1790, 1749, 1698, 1604, 1455, 1379, 1288, 1224, 1178, 1160, 1124, 1082, 977, 908, 873, 837, 789, 728, 696, 650 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -84.4 ($c = 0.50$ in CHCl_3); **HRMS** (ES^+): Found 409.1999 (+0.8 ppm), $\text{C}_{23}\text{H}_{30}\text{O}_5\text{Na}$ ($\text{M}+\text{Na}^+$) requires 409.1991



5-(3-((3a*R*,6*S*,7*S*,11*R*,13a*R*,*E*)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,8,9,10,11,13a-octahydro-3a*H*-[1,3]dioxolo[4,5-*c*][1]oxacyclododecin-6-yl)phenyl)pentanal ((-)-11b) Prepared according to Representative Procedure E: (-)-A4 (7.0 mg, 0.016 mmol), DMSO (0.02 mL, 0.315 mmol), triethylamine (0.02 mL, 0.157 mmol) SO₃-Pyr (20 mg, 0.126 mmol), and DCM (0.16 mL) yielded 6 mg (86 %) of the product as a clear oil. Purified by preparative TLC (4:1 hexanes/EtOAc) .

¹H NMR (500 MHz, CDCl₃) δ 9.75 (s, 1H), 7.19 (t, *J* = 7.5 Hz, 1H), 7.13 – 7.04 (m, 3H), 5.71 – 5.60 (m, 2H), 5.49 (d, *J* = 11.5 Hz, 1H), 4.86 – 4.84 (m, 1H), 4.54 (d, *J* = 6.6 Hz, 1H), 2.59 (t, *J* = 7.1 Hz, 2H), 2.46 – 2.43 (m, 2H), 2.40 – 2.35 (m, 1H), 2.25 – 2.18 (m, 1H), 2.02 – 1.95 (m, 1H), 1.68 (s, 3H), 1.66 – 1.61 (m, 4H), 1.42 – 1.34 (m, 6H), 1.09 – 1.05 (m, 2H), 1.02 (d, *J* = 6.6 Hz, 3H), 0.68 (d, *J* = 7.0 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 202.74, 169.52, 142.02, 140.15, 135.91, 128.44, 128.22, 127.87, 125.40, 122.36, 111.01, 79.45, 78.67, 43.87, 36.33, 35.82, 35.68, 33.88, 30.92, 29.41, 26.96, 26.03, 21.85, 21.07, 17.95, 15.94; **IR** (neat): 2926, 2855, 1749, 1724, 1608, 1488, 1456, 1379, 1223, 1180, 1124, 1084, 977, 911, 874, 836, 785, 731, 706, 648 cm⁻¹; [α]_D²⁵ -76.6 (c = 0.60 in CHCl₃); **HRMS** (ES⁺): Found 443.2757 (-4.0 ppm), C₂₇H₃₉O₅ (M+H⁺) requires 443.2797

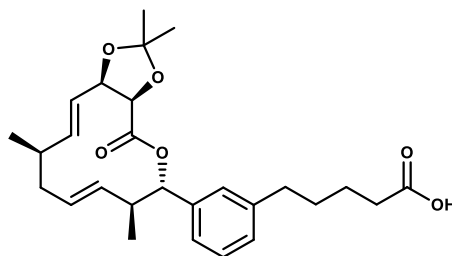


Representative Procedure F: Pinnick Oxidation

3-((3aR,6S,7S,8E,11R,12E,13aR)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,10,11,13a-hexahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)benzoic acid ((-)-C1) To a rapidly stirring solution of **(-)-10a** (8.0 mg, 0.021 mmol) in *tert*-butyl alcohol (0.42 mL) and acetonitrile (0.21 mL) was added 2-methyl-2-butene (neat, 0.11 mL), and the reaction vessel was sparged with argon. The solution was cooled to 0 °C, and a freshly prepared solution of NaH₂PO₄ (12 mg, 0.10 mmol) and NaClO₂ (10 mg, 0.11 mmol) in water (0.62 mL) was added dropwise. The reaction was slowly warmed to room temperature and stirred overnight. The reaction was quenched by addition of a saturated aqueous solution of sodium thiosulfate (1 mL) and diluted in EtOAc. The aqueous layer was separated and acidified to pH 3 with 1N HCl. The acidified aqueous layer was extracted 5x EtOAc (5x 5 mL). The combined organic extracts were washed with brine, dried over sodium sulfate, filtered, and concentrated. The residue was purified by preparative TLC (1% AcOH, 49% hexanes, 50% EtOAc) to afford the product (6 mg, 75%) as an off white residue.

¹H NMR (500 MHz, CDCl₃) δ 8.10 (s, 1H), 8.06 – 8.02 (m, 1H), 7.60 – 7.55 (m, 1H), 7.44 (t, *J* = 7.7 Hz, 1H), 5.87 – 5.79 (m, 1H), 5.65 (d, *J* = 10.8 Hz, 1H), 5.36 – 5.21 (m, 3H), 4.78 – 4.71 (m, 1H), 4.50 (d, *J* = 6.6 Hz, 1H), 2.70 – 2.57 (m, 1H), 2.35 – 2.24 (m, 2H), 2.11 – 2.02 (m, 1H), 1.66 (s, 3H), 1.38 (s, 3H), 1.09 (d, *J* = 6.8 Hz, 3H), 0.77 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 170.96, 169.90, 139.76, 139.54, 134.21, 133.65, 131.14, 130.31, 129.67, 129.40,

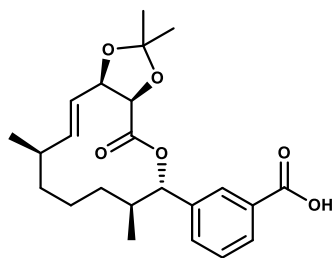
128.81, 123.56, 111.17, 80.09, 79.21, 78.36, 43.28, 38.46, 35.87, 26.89, 25.97, 20.94, 17.50; **IR** (neat): 2957, 2925, 2853, 2360, 1749, 1727, 1693, 1554, 1486, 1455, 1379, 1252, 1222, 1177, 1083, 1039, 970, 913, 880, 814, 784, 758, 696, 668 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -169 ($c = 0.40$ in CHCl_3); **HRMS** (ES^+): Found 423.1777 (-0.1 ppm), $\text{C}_{23}\text{H}_{28}\text{O}_6\text{Na}$ ($\text{M}+\text{Na}^+$) requires 423.1778



5-(3-((3aR,6S,7S,8E,11R,12E,13aR)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,10,11,13a-hexahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)phenyl)pentanoic acid ((-)-C3) Prepared according to Representative Procedure F: (-)-**11a** (5.0 mg, 0.011 mmol), *tert*-butyl alcohol (0.23 mL), 2-methyl-2 butene (0.06 mL), acetonitrile (0.11 mL), NaH_2PO_4 (7.0 mg, 0.057 mmol), NaClO_2 (6.0 mg, 0.062 mmol), and H_2O (0.34 mL) yielded 3.3 mg (65%) of the product as an off-white residue. Purified by preparative TLC (1% AcOH, 49% hexanes, 50% EtOAc).

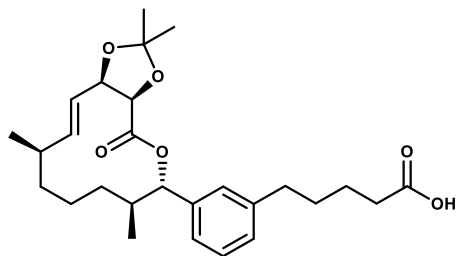
^1H NMR (500 MHz, CDCl_3) δ 7.22 (t, $J = 7.5$ Hz, 1H), 7.16 – 7.12 (m, 2H), 7.10 – 7.08 (m, 1H), 5.85 – 5.79 (m, 1H), 5.56 (d, $J = 10.8$ Hz, 1H), 5.35 – 5.24 (m, 3H), 4.76 – 4.72 (m, 1H), 4.48 (d, $J = 6.7$ Hz, 1H), 2.65 – 2.55 (m, 3H), 2.39 – 2.34 (m, 2H), 2.34 – 2.30 (m, 1H), 2.28 – 2.24 (m, 1H), 2.10 – 2.02 (m, 1H), 1.69 – 1.63 (m, 7H), 1.38 (s, 3H), 1.09 (d, $J = 6.8$ Hz, 3H), 0.76 (d, $J = 6.8$ Hz, 3H); **^{13}C NMR** (101 MHz, CDCl_3) δ 177.74, 169.87, 142.32, 139.48, 138.99, 134.78, 130.68, 128.55, 128.52, 125.43, 123.65, 111.12, 80.82, 79.21, 78.40, 43.16, 38.51, 35.88, 35.54, 33.63, 30.76, 26.91, 26.00, 24.37, 21.01, 17.65; **IR** (neat): 2925, 2854, 1747, 1707, 1608, 1488, 1455, 1412, 1378, 1221, 1182, 1162, 1084, 1042, 1000, 969, 880, 786, 755, 705, 667 cm^{-1} ;

$[\alpha]_{\text{D}}^{25}$ -67.2 ($c = 0.33$ in CHCl_3); **HRMS** (ES^+): Found 457.2576 (-0.9 ppm), $\text{C}_{27}\text{H}_{36}\text{O}_6$ ($\text{M}+\text{H}^+$) requires 457.2585



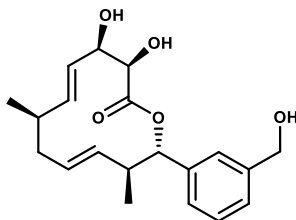
3-((3aR,6S,7S,11R,13aR,E)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,8,9,10,11,13a-octahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)benzoic acid ((-)-C2) Prepared according to Representative Procedure F: **(-)-10b** (5.0 mg, 0.013 mmol), *tert*-butyl alcohol (0.26 mL), 2-methyl-2 butene (0.07 mL), acetonitrile (0.13 mL), NaH_2PO_4 (8.0 mg, 0.065 mmol), NaClO_2 (6.0 mg, 0.071 mmol), and H_2O (0.39 mL) yielded 5.0 mg (96%) of the product as an off-white residue. Purified by preparative TLC (1% AcOH, 49% hexanes, 50% EtOAc).

^1H NMR (500 MHz, CDCl_3) δ 8.06 (s, 1H), 8.02 – 7.98 (m, 1H), 7.57 – 7.53 (m, 1H), 7.40 (t, $J = 7.7$ Hz, 1H), 5.72 – 5.62 (m, 2H), 5.58 (d, $J = 11.5$ Hz, 1H), 4.87 (dd, $J = 6.6, 3.6$ Hz, 1H), 4.57 (d, $J = 6.5$ Hz, 1H), 2.46 – 2.35 (m, 1H), 2.31 – 2.23 (m, 1H), 2.05 – 1.98 (m, 1H), 1.68 (s, 3H), 1.42 – 1.36 (m, 6H), 1.11 – 1.07 (m, 2H), 1.03 (d, $J = 6.6$ Hz, 3H), 0.70 (d, $J = 7.0$ Hz, 3H); **^{13}C NMR** (126 MHz, CDCl_3) δ 170.82, 169.60, 140.81, 135.91, 133.64, 130.10, 129.55, 129.30, 128.68, 122.34, 111.09, 78.75, 78.59, 36.38, 35.81, 33.80, 29.31, 26.94, 26.00, 21.06, 17.92, 15.81; **IR** (neat): 2927, 2853, 2553, 1781, 1749, 1681, 1608, 1589, 1454, 1413, 1380, 1283, 1228, 1175, 1123, 1084, 979, 940, 893, 873, 825, 782, 758, 723, 695, 677 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -130.5 ($c = 0.36$ in CHCl_3); **HRMS** (ES^+): Found 369.1669 (-0.3 ppm), $\text{C}_{20}\text{H}_{26}\text{O}_5\text{Na}$ ($\text{M}+\text{Na}^+$) requires 369.1672



5-(3-((3aR,6S,7S,11R,13aR,E)-2,2,7,11-Tetramethyl-4-oxo-4,6,7,8,9,10,11,13a-octahydro-3aH-[1,3]dioxolo[4,5-c][1]oxacyclododecin-6-yl)phenyl)pentanoic acid ((-)-C4) Prepared according to Representative Procedure F: **(-)-11b** (6.0 mg, 0.014 mmol), *tert*-butyl alcohol (0.27 mL), 2-methyl-2 butene (0.07 mL), acetonitrile (0.14 mL), NaH₂PO₄ (8.1 mg, 0.068 mmol), NaClO₂ (6.7 mg, 0.075 mmol), and H₂O (0.41 mL) yielded 5.7 mg (92%) of the product as an off-white residue. Purified by preparative TLC (1% AcOH, 49% hexanes, 50% EtOAc).

¹H NMR (500 MHz, CDCl₃) δ 7.21 – 7.16 (m, 1H), 7.13 – 7.09 (m, 2H), 7.07 – 7.03 (m, 1H), 5.70 – 5.59 (m, 2H), 5.49 (d, J = 11.5 Hz, 1H), 4.85 (ddd, J = 6.5, 3.9, 0.9 Hz, 1H), 4.55 (d, J = 6.5 Hz, 1H), 2.64 – 2.57 (m, 2H), 2.41 – 2.32 (m, 3H), 2.26 – 2.19 (m, 1H), 2.02 – 1.94 (m, 1H), 1.68 (s, 3H), 1.66 – 1.60 (m, 4H), 1.41 – 1.36 (m, 6H), 1.09 – 1.05 (m, 2H), 1.02 (d, J = 6.6 Hz, 3H), 0.68 (d, J = 7.0 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 169.51, 142.11, 140.09, 135.91, 128.38, 127.90, 125.33, 122.32, 111.08, 79.50, 78.62, 76.91, 36.22, 35.83, 35.51, 33.89, 33.62, 32.08, 30.68, 29.40, 26.94, 26.03, 24.36, 22.85, 21.09, 17.95, 15.94, 14.28; **IR** (neat): 2921, 2852, 1750, 1708, 1607, 1554, 1456, 1377, 1284, 1224, 1178, 1124, 1085, 977, 874, 836, 787, 720, 706 cm⁻¹; **[α]_D²⁵** -48.1 (c = 0.54 in CHCl₃); **HRMS** (ES⁺): Found 481.2583 (+ 1.7 ppm), C₂₇H₃₈O₆Na (M+Na⁺) requires 481.2566



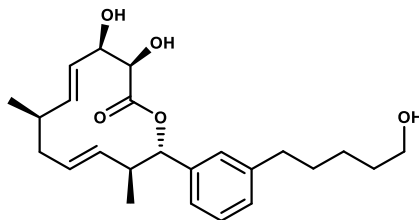
Representative Procedure G: Global Deprotection

(3*R*,4*R*,5*E*,7*R*,9*E*,11*S*,12*S*)-3,4-Dihydroxy-12-(3-(hydroxymethyl)phenyl)-7,11-

dimethyloxacyclododeca-5,9-dien-2-one ((-)-B1) To a solution of (-)-6a (16 mg, 0.032 mmol) in MeOH (1.4 mL) and THF (0.32 mL) was added 1N HCl (1.4 mL), and the reaction vessel was sparged with argon. The reaction was stirred for 22.5 hours. Water was added, and the mixture was diluted in EtOAc. The organic layer was separated and the aqueous layer was extracted 5 times with EtOAc (5x 5 mL). The combined organic fractions were washed with brine, dried over sodium sulfate, filtered, and concentrated. Purification by preparative TLC to remove less polar impurities (5% MeOH in DCM) afforded the product (9 mg, 82%) as an off-white residue.

¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.19 (m, 4H), 5.42 – 5.26 (m, 4H), 5.08 – 4.96 (m, 1H), 4.68 (s, 2H), 4.48 – 4.40 (m, 1H), 4.11 – 4.04 (m, 1H), 2.67 – 2.53 (m, 1H), 2.27 – 2.18 (m, 2H), 1.81 – 1.68 (m, 1H), 1.08 (d, *J* = 6.6 Hz, 3H), 0.71 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 172.01, 141.37, 139.31, 135.26, 134.12, 131.42, 128.81, 127.27, 127.10, 126.49, 126.12, 81.54, 74.04, 73.78, 65.26, 43.51, 40.92, 38.36, 21.41, 17.69; **IR** (neat): 3398, 2956, 2924, 2870, 2848, 2246, 2160, 2029, 1977, 1725, 1616, 1555, 1491, 1453, 1374, 1235, 1195, 1111, 1081, 1018, 966, 907, 840, 791, 705, 673, 646, 591 cm⁻¹; **[α]_D²⁵** -62.4 (c = 0.74 in CHCl₃);

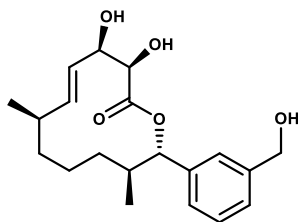
HRMS (ES⁺): Found 369.1669 (-0.3 ppm), C₂₀H₂₆O₅Na (M+Na⁺) requires 369.1672



(3*R*,4*R*,5*E*,7*R*,9*E*,11*S*,12*S*)-3,4-Dihydroxy-12-(3-(5-hydroxypentyl)phenyl)-7,11-

dimethyloxacyclododeca-5,9-dien-2-one ((-)-B3**)** Prepared according to Representative Procedure G: (-)-**6b** (19 mg, 0.034 mmol), MeOH (1.7 mL), THF (0.37 mL), and 1N HCl (1.7 mL) yielded 8 mg (57 %) of the product as an off-white residue. Purified by preparative TLC (5% MeOH in DCM).

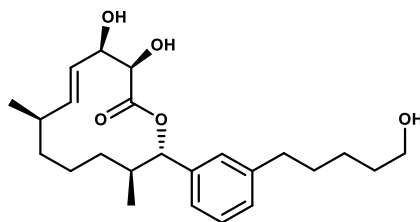
¹H NMR (500 MHz, CDCl₃) δ 7.26 – 7.22 (m, 1H), 7.15 – 7.09 (m, 3H), 5.41 – 5.35 (m, 2H), 5.35 – 5.29 (m, 2H), 5.07 – 4.97 (m, 1H), 4.45 (d, *J* = 2.8 Hz, 1H), 4.09 (d, *J* = 2.8 Hz, 1H), 3.61 (t, *J* = 6.5 Hz, 2H), 2.67 – 2.54 (m, 4H), 2.25 – 2.18 (m, 2H), 1.80 – 1.70 (m, 1H), 1.69 – 1.52 (m, 6H), 1.41 – 1.32 (m, 2H), 1.08 (d, *J* = 6.5 Hz, 3H), 0.72 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 171.96, 142.83, 138.81, 135.22, 134.31, 131.26, 128.62, 128.53, 128.05, 126.56, 125.17, 81.67, 74.11, 73.80, 62.99, 43.42, 40.93, 38.36, 35.82, 32.62, 31.10, 25.29, 21.42, 17.69; **IR** (neat): 3409, 2925, 2854, 1729, 1608, 1589, 1488, 1453, 1373, 1235, 1192, 1111, 1080, 1018, 965, 887, 839, 794, 706, 647 cm⁻¹; **[α]_D²⁵** -36.2 (c = 0.64 in CHCl₃); **HRMS** (ES⁺): Found 403.2481 (+0.2 ppm), C₂₄H₃₄O₅ (M+H⁺) requires 403.2479



(3*R*,4*R*,7*R*,11*S*,12*S*,*E*)-3,4-Dihydroxy-12-(3-(hydroxymethyl)phenyl)-7,11-

dimethyloxacyclododec-5-en-2-one ((-)-B2**)** Prepared according to Representative Procedure G: (-)-**7a** (14 mg, 0.028 mmol), MeOH (1.4 mL), THF (0.30 mL), and 1N HCl (1.4 mL) yielded 5 mg (50 %) of the product as an off-white residue. Purified by preparative TLC (5% MeOH in DCM).

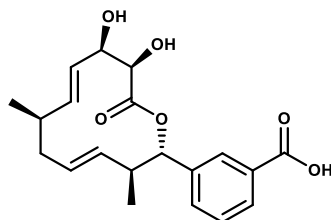
¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.26 (m, 3H), 7.24 – 7.21 (m, 1H), 5.61 (dd, *J* = 15.4, 2.3 Hz, 1H), 5.55 – 5.45 (m, 1H), 5.29 (d, *J* = 11.4 Hz, 1H), 4.68 (s, 2H), 4.53 – 4.46 (m, 1H), 4.19 (dd, *J* = 8.8, 3.7 Hz, 1H), 3.25 (d, *J* = 8.8 Hz, 1H), 2.47 – 2.35 (m, 2H), 2.29 – 2.19 (m, 1H), 2.02 – 1.94 (m, 1H), 1.79 (s, 1H), 1.48 – 1.28 (m, 3H), 1.13 – 1.07 (m, 2H), 1.01 (d, *J* = 6.5 Hz, 3H), 0.67 (d, *J* = 7.0 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 171.98, 141.23, 140.28, 134.54, 128.73, 127.08, 126.93, 126.00, 125.57, 80.86, 73.78, 73.02, 65.32, 36.77, 35.84, 34.60, 28.76, 21.86, 18.96, 15.66; **IR** (neat): 3441, 3413, 3240, 2965, 2936, 2857, 2360, 2342, 1727, 1451, 1373, 1353, 1307, 1247, 1203, 1125, 1096, 1051, 1016, 982, 908, 890, 864, 837, 799, 784, 718, 668, 654 cm⁻¹; **[α]_D²⁵** -49.8 (c = 0.90 in CHCl₃); **HRMS** (ES⁺): Found 371.1874 (+4.0 ppm), C₂₀H₂₈O₅Na (M+Na⁺) requires 371.1834



(3R,4R,7R,11S,12S,E)-3,4-Dihydroxy-12-(3-(5-hydroxypentyl)phenyl)-7,11-

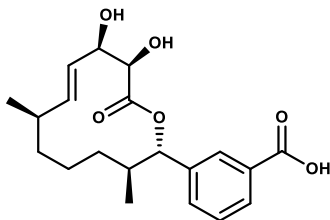
dimethyloxacyclododec-5-en-2-one ((-)-B4) Prepared according to Representative Procedure G: (-)-**7b** (11 mg, 0.020 mmol), MeOH (1.0 mL), THF (0.22 mL), and 1N HCl (1.0 mL) yielded 5 mg (63 %) of the product as an off-white residue. Purified by preparative TLC (5% MeOH in DCM).

¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.17 (m, 1H), 7.14 – 7.04 (m, 3H), 5.66 – 5.44 (m, 2H), 5.26 (d, *J* = 11.4 Hz, 1H), 4.50 (s, 1H), 4.19 (d, *J* = 3.6 Hz, 1H), 3.60 (t, *J* = 6.5 Hz, 2H), 2.61 (t, *J* = 7.4 Hz, 2H), 2.54 – 2.46 (m, 1H), 2.42 – 2.33 (m, 1H), 2.27 – 2.17 (m, 1H), 2.01 – 1.93 (m, 1H), 1.67 – 1.51 (m, 7H), 1.38 – 1.31 (m, 4H), 1.11 – 1.04 (m, 2H), 1.01 (d, *J* = 6.5 Hz, 3H), 0.68 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 171.94, 142.63, 139.77, 134.50, 128.45, 127.97, 125.59, 124.99, 81.00, 73.81, 73.01, 63.02, 36.65, 35.78, 34.63, 32.62, 30.98, 28.77, 25.18, 21.87, 18.92, 15.66, 1.17; IR (neat): 3423, 2930, 1732, 1553, 1450, 1373, 1201, 1125, 1055, 1013, 979, 844, 780, 710 cm⁻¹; [α]_D²⁵ -52.9 (c = 0.17 in CHCl₃); HRMS (ES⁺): Found 427.2445 (-1.0 ppm), C₂₄H₃₆O₅Na (M+Na⁺) requires 427.2455



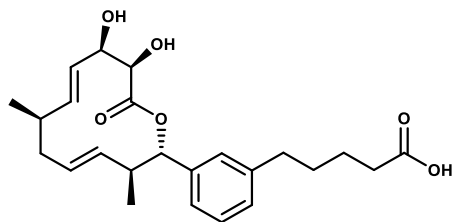
3-((2*S*,3*S*,4*E*,7*R*,8*E*,10*R*,11*R*)-10,11-Dihydroxy-3,7-dimethyl-12-oxooxacyclododeca-4,8-dien-2-yl)benzoic acid ((-)-D1) To a solution of (-)-C1 (6.0 mg, 0.015 mmol) in H₂O (0.08 mL) at room temperature was added TFA (0.08 mL), and the reaction vessel was sparged with argon. The reaction was allowed to stir for 48 hours. The solvent was removed and the residue was co-evaporated with ethanol twice to remove residual water. If necessary, nonpolar impurities may be removed by taking up the product in acetonitrile, washing 5 x 1mL pentane, and reducing the acetonitrile fraction. In this manner, 4.4 mg (79%) of the product was obtained as an off-white residue.

¹H NMR (500 MHz, CDCl₃) δ 8.08 (s, 1H), 8.04 (d, *J* = 7.7 Hz, 1H), 7.58 (d, *J* = 7.5 Hz, 1H), 7.46 (t, *J* = 7.5 Hz, 1H), 5.45 – 5.31 (m, 4H), 5.03 (dd, *J* = 14.9, 9.6 Hz, 1H), 4.49 (s, 1H), 4.16 – 4.11 (m, 1H), 2.68 – 2.57 (m, 1H), 2.24 (d, *J* = 12.6 Hz, 2H), 1.77 (q, *J* = 12.2 Hz, 1H), 1.09 (d, *J* = 6.4 Hz, 3H), 0.72 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 172.11, 170.57, 139.68, 135.35, 133.69, 133.46, 131.81, 130.41, 129.78, 129.32, 128.89, 126.45, 81.00, 74.10, 73.82, 43.48, 40.91, 38.36, 21.40, 17.55; IR (neat): 3409, 2957, 2926, 2871, 1721, 692, 1609, 1591, 1453, 1411, 1375, 1192, 1109, 1080, 1019, 967, 909, 861, 839, 800, 757, 730, 697, 667, 656 cm⁻¹; [α]_D²⁵ -139 (c = 0.50 in CHCl₃); HRMS (ES⁺): Found 383.1503 (+3.2 ppm), C₂₀H₂₄O₆Na (M+Na⁺) requires 383.1471



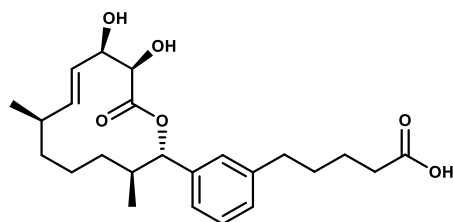
3-((2S,3S,7R,10R,E)-10,11-dihydroxy-3,7-dimethyl-12-oxooxacyclododec-8-en-2-yl)benzoic acid ((-)-D2) To a solution of (-)-C2 (11.8 mg, 0.029 mmol) in H₂O (1.45 mL) at room temperature was added TFA (1.45 mL), and the reaction vessel was sparged with argon. The reaction was allowed to stir for 48 hours. The solvent was removed and the residue was co-evaporated with ethanol twice to remove residual water. Crude product was purified via preparative TLC (1:1 hexanes in ethyl acetate with 1% acetic acid) to yield 6.1 mg (58%). If necessary, nonpolar impurities may be removed by taking up the product in acetonitrile, washing 5 x 1mL pentane, and reducing the acetonitrile fraction.

¹H NMR (500 MHz, CDCl₃) δ 8.04 (t, *J* = 24.3 Hz, 1H), 7.56 (d, *J* = 25.4 Hz, 1H), 7.43 (s, 1H), 5.61 (d, *J* = 15.2 Hz, 1H), 5.50 (s, 1H), 5.33 (d, *J* = 10.7 Hz, 1H), 4.52 (s, 1H), 4.23 (s, 1H), 3.34 (s, 1H), 2.38 (s, 1H), 2.24 (s, 1H), 2.00 (s, 1H), 1.29 (d, *J* = 46.8 Hz, 3H), 1.09 (s, 1H), 1.00 (s, 1H), 0.92 (s, 1H), 0.72 (s, 1H), 0.66 (s, 1H); **¹³C NMR** (126 MHz, CDCl₃) δ 171.88, 169.23, 140.53, 134.38, 133.14, 130.08, 128.99, 128.78, 128.65, 125.41, 80.17, 77.60, 73.65, 72.86, 36.68, 35.70, 34.41, 28.48, 21.75, 15.40 ; **IR** (neat): 3335, 2955, 2922, 2853, 1719, 1454, 1377, 1260, 1195, 1084, 1018, 978, 952 cm⁻¹; **[α]_D²⁵** -33 (c = 0.0023 in CHCl₃); **HRMS** (ES⁺): Found 385.1629 (+ 0.2 ppm), C₂₀H₂₅O₆Na (M+Na⁺) requires 385.1627



5-(3-((2*S*,3*S*,4*E*,7*R*,8*E*,10*R*,11*R*)-10,11-dihydroxy-3,7-dimethyl-12-oxooxacyclododeca-4,8-dien-2-yl)phenyl)pentanoic acid ((-)-D3) To a solution of **C3** (35 mg, 0.077 mmol) in H₂O (0.40 mL) at room temperature was added TFA (0.40 mL), and the reaction vessel was sparged with argon. The reaction was allowed to stir for 48 hours. The solvent was removed and the residue was co-evaporated with ethanol twice to remove residual water. The crude product was purified by preparatory TLC (1:1 hexanes in ethyl acetate with 1% acetic acid). If necessary, nonpolar impurities may be removed by taking up the product in acetonitrile, washing 5 x 1mL pentane, and reducing the acetonitrile fraction. In this manner, 18.8 mg (59%) of the product was obtained as an off-white residue.

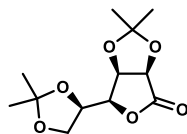
¹H NMR (500 MHz, CDCl₃) δ 7.26 – 7.22 (m, 1H), 7.16 – 7.12 (m, 2H), 7.10 (d, *J* = 7.6 Hz, 1H), 5.39 (d, *J* = 4.6 Hz, 2H), 5.34 – 5.29 (m, 2H), 5.06 – 4.99 (m, 1H), 4.46 (s, 1H), 4.13 (s, 1H), 2.70 – 2.50 (m, 3H), 2.33 (t, *J* = 6.9 Hz, 2H), 2.22 (d, *J* = 13.6 Hz, 2H), 1.76 (dd, *J* = 24.2, 12.1 Hz, 2H), 1.66 (ddd, *J* = 23.0, 15.9, 8.3 Hz, 5H), 1.27 (d, *J* = 15.7 Hz, 6H), 1.08 (d, *J* = 6.5 Hz, 3H), 0.72 (d, *J* = 6.8 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 177.33, 172.22, 142.62, 139.09, 135.50, 134.47, 131.54, 128.99, 128.73, 128.03, 126.70, 125.63, 81.87, 74.23, 73.95, 43.59, 41.15, 38.58, 35.68, 33.78, 30.76, 30.06, 24.43, 21.64, 17.90; **IR** (neat): 3434, 2923, 2851, 1707, 1453, 1410, 1193, 1081, 966 cm⁻¹; **[α]_D²⁵** -28 (c = 0.0028 in CHCl₃; **HRMS** (ES⁺): Found 439.2102 (- 0.5 ppm), C₂₄H₃₂O₆Na (M+Na⁺) requires 439.2097



5-(3-((2S,3S,7R,10R,11R,E)-10,11-dihydroxy-3,7-dimethyl-12-oxooxacyclododec-8-en-2-yl)phenyl)penanoic acid ((-)-D4) To a solution of **C4** (5 mg, 0.011 mmol) in H₂O (0.60 mL) at room temperature was added TFA (0.60 mL), and the reaction vessel was sparged with argon. The reaction was allowed to stir for 48 hours. The solvent was removed and the residue was co-evaporated with ethanol twice to remove residual water. The crude product was purified by preparatory TLC (1:1 hexanes in ethyl acetate with 1% acetic acid). If necessary, nonpolar impurities may be removed by taking up the product in acetonitrile, washing 5 x 1mL pentane, and reducing the acetonitrile fraction. In this manner, 4.6 mg (59%) of the product was obtained as an off-white residue.

¹H NMR (400 MHz, CDCl₃) δ 7.14 (t, *J* = 7.5 Hz, 1H), 7.02 (dd, *J* = 12.4, 7.7 Hz, 3H), 5.57 – 5.38 (m, 2H), 5.19 (d, *J* = 11.4 Hz, 1H), 4.45 (dt, *J* = 4.1, 2.1 Hz, 1H), 4.17 (d, *J* = 3.6 Hz, 1H), 2.63 – 2.47 (m, 2H), 2.31 (ddd, *J* = 16.5, 11.6, 4.6 Hz, 1H), 2.24 (t, *J* = 7.0 Hz, 2H), 2.19 – 2.09 (m, 1H), 1.93 (td, *J* = 14.5, 9.4 Hz, 1H), 1.57 (ddd, *J* = 23.2, 15.5, 8.6 Hz, 4H), 1.40 – 1.14 (m, 7H), 1.02 (d, *J* = 9.0 Hz, 2H), 0.94 (d, *J* = 6.6 Hz, 3H), 0.61 (d, *J* = 7.0 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 177.12, 172.22, 142.46, 140.12, 134.71, 128.83, 128.57, 127.77, 125.79, 125.51, 99.99, 99.86, 81.19, 73.95, 73.14, 36.86, 36.02, 35.62, 34.84, 33.77, 30.62, 28.92, 24.35, 22.11, 19.07, 15.86; **IR** (neat): 3392, 2923, 2852, 2361, 2341, 1735, 1456, 1376, 1241, 1199, 1088 cm⁻¹

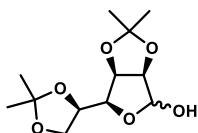
¹; $[\alpha]_{\text{D}}^{25}$ -410 (c = 0.002 in CHCl₃ ; **HRMS** (ES⁺): Found 419.2408 (- 2.6 ppm), C₂₄H₃₅O₆ (M+H⁺) requires 419.2434



(3aR,6S,6aR)-6-((R)-2,2-Dimethyl-1,3-dioxolan-4-yl)-2,2-dimethyldihydrofuro

[3,4-*d*][1,3]dioxol-4(3aH)-one (12) A round bottom flask was charged with **D-gulonic acid lactone** (40.0 g, 224 mmol), acetone (320 mL), *p*-toluenesulfonic acid monohydrate (2.21 g, 11.6 mmol), and 2,2-dimethoxypropane (67.0 mL, 326 mmol). The mixture was allowed to stir 40 hours, becoming clear as the reaction proceeds. A second portion of 2,2-dimethoxypropane (30 mL, 244 mmol) is added and the reaction was allowed to stir a further 20 hours. Upon completion, sodium bicarbonate (3.00 g, 35.7 mmol) was added to the mixture and allowed to stir for 15 minutes. The newly formed suspension was concentrated and the clumpy solid slurry was dissolved in dichloromethane (200 mL). The solution was added to a separatory funnel and water (200 mL) was added, forming two layers. The aqueous layer was extracted with dichloromethane (2x 200 mL). The combined organic layers were then dried over MgSO₄, filtered, and reduced to give a white solid, which was then recrystallized in hot ethyl acetate to afford the bis-protected product as white crystals. The mother liquor was concentrated and dissolved in acetone (200 mL) and another portion of *p*-toluenesulfonic acid monohydrate (2.21 g, 11.6 mmol) is added. The mixture was allowed to stir a further 48 hours, after which the same workup procedure was followed, yielding white crystals (41.69 g, 72%). Experimental data matched that previously described.^{1,6,7}

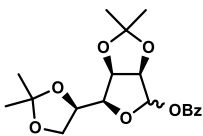
¹H NMR (400 MHz, CDCl₃) δ 4.84 (d, *J* = 5.6 Hz, 1H), 4.74 (dd, *J* = 5.6, 3.5 Hz, 1H), 4.50 – 4.36 (m, 2H), 4.26 – 4.17 (m, 1H), 3.86 – 3.76 (m, 1H), 1.48 (s, 3H), 1.47 (s, 3H), 1.40 (s, 3H), 1.38 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 173.38, 114.39, 110.28, 80.89, 75.97, 75.74, 75.14, 65.05, 26.57, 25.66, 25.14



(3aR,6S,6aR)-6-((R)-2,2-Dimethyl-1,3-dioxolan-4-yl)-2,2-dimethyltetrahydrofuro[3,4-*d*]

[1,3]dioxol-4-ol (13) A flame dried round bottom flask was charged with **12** (4.65 g, 18.0 mmol) and dry dichloromethane (90 mL). The stirring solution was cooled to -78°C and neat DIBAL-H (3.8 mL, 21.6 mmol) was added via syringe. The reaction was maintained at that temperature for 3 hours, then it was added to a stirring solution of 50% saturated Rochelle's salt (90 mL) at 0°C. The thick white slurry was warmed to room temperature and stirred for 3 hours. The reaction was then poured into a separatory funnel and the organic layer was separated. The aqueous layer was extracted with dichloromethane (3x 90 mL). The organic fractions were combined, washed with brine, dried over sodium sulfate, filtered, and concentrated. The lactol was obtained as a white solid (4.51 g, 96%) and used without further purification. If necessary, the product may be recrystallized from hot ethanol. Experimental data matched that previously described.^{1,6,7}

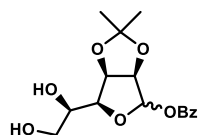
¹H NMR (500 MHz, CDCl₃) δ 5.44 (d, *J* = 2.5 Hz, 1H), 4.68 (dd, *J* = 5.9, 3.8 Hz, 1H), 4.61 (d, *J* = 5.9 Hz, 1H), 4.34 (dt, *J* = 8.5, 6.9 Hz, 1H), 4.19 (dd, *J* = 8.5, 6.5 Hz, 1H), 4.10 (dd, *J* = 8.5, 3.8 Hz, 1H), 3.71 (dd, *J* = 8.4, 7.1 Hz, 1H), 3.40 (d, *J* = 2.5 Hz, 1H), 1.43 – 1.41 (m, 6H), 1.36 (s, 3H), 1.27 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 112.80, 109.78, 101.24, 85.72, 81.97, 79.85, 75.53, 66.00, 26.72, 25.96, 25.42, 24.73



(3aR,6S,6aR)-6-((R)-2,2-Dimethyl-1,3-dioxolan-4-yl)-2,2-dimethyltetrahydro-

furo[3,4-*d*][1,3]dioxol-4-yl benzoate (14) A flame dried round bottom flask was charged with **13** (41.9 g, 160 mmol) and dry dichloromethane (1000 mL). The solution was cooled to -78°C , whereupon triethylamine (56 mL, 400 mmol) was added, followed by DMAP (2.0 g, 16 mmol), and freshly distilled benzoyl chloride (28.0 mL, 241 mmol). The reaction was allowed to stir overnight while warming to room temperature. Upon completion, the reaction was added to an equal volume of water at 0°C and the layers were separated. The aqueous layer was extracted with dichloromethane (2x 300 mL), and the organic layer was washed with 0.5M HCl (3 x 300 mL), H_2O (2 x 300 mL), NaHCO_3 (3 x 300 mL), and brine (400 mL). The organic fractions were then combined and dried over MgSO_4 , filtered, and reduced, yielding a white solid (58.35 g, >99%). Experimental data matched that previously described.^{1,6,7}

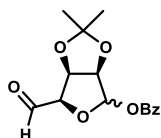
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.04 – 7.97 (m, 2H), 7.61 – 7.53 (m, 1H), 7.47 – 7.39 (m, 2H), 6.50 (s, 1H), 4.90 – 4.80 (m, 2H), 4.43 (q, $J = 7.2$ Hz, 1H), 4.29 – 4.20 (m, 2H), 3.76 (t, $J = 7.8$ Hz, 1H), 1.51 (s, 3H), 1.46 (s, 3H), 1.39 (s, 3H), 1.32 (s, 3H); **$^{13}\text{C NMR}$** (101 MHz, CDCl_3) δ 164.95, 133.54, 129.94, 129.70, 128.56, 113.70, 110.11, 101.87, 85.55, 84.91, 79.66, 75.66, 66.07, 26.90, 26.14, 25.48, 24.93



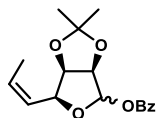
(3*aR*,6*S*,6*aR*)-6-((*R*)-1,2-Dihydroxyethyl)-2,2-dimethyltetrahydrofuro[3,4-*d*]

[1,3]dioxol-4-yl benzoate (15) A flame dried round bottom flask was charged with **14** (10.0 g, 27.4 mmol), H₂O (17 mL), and acetic acid (150 mL). The slurry was heated to 35°C and allowed to stir overnight. The now clear solution was reduced, co-evaporated with toluene (40 mL), reduced, and recrystallized from hot ethanol, yielding white crystals (8.57 g, 96%). Experimental data matched that previously described.^{1,6,7}

¹H NMR (500 MHz, CDCl₃) δ 8.08 – 7.94 (m, 2H), 7.64 – 7.55 (m, 1H), 7.52 – 7.41 (m, 2H), 6.43 (s, 1H), 4.95 – 4.88 (m, 2H), 4.30 – 4.24 (m, 1H), 4.19 – 4.09 (m, 1H), 3.89 – 3.69 (m, 2H), 2.84 (s, 1H), 2.21 (s, 1H), 1.52 (s, 3H), 1.35 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 165.18, 133.68, 129.93, 129.49, 128.60, 113.56, 101.08, 85.74, 82.56, 79.56, 70.86, 63.15, 26.13, 24.79



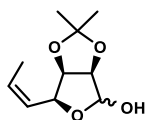
(3*aR*,6*R*,6*aR*)-6-Formyl-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl benzoate (16) A round bottom flask was charged with **15** (3.07 g, 9.47 mmol), 1,4-dioxane (100 mL), and water (100 mL). Sodium metaperiodate (4.05 g, 18.9 mmol) was added and the reaction was allowed to stir for ~90 minutes. The mixture was then filtered over celite, rinsed with ethyl acetate (3 x 100 mL), and added to a separatory funnel. The aqueous layer was extracted with ethyl acetate (3 x 100 mL) and the organic layers were combined, washed with brine (200 mL), dried over MgSO₄, filtered, and reduced. The resulting viscous oil was then co-eluted with toluene and once again reduced, yielding the intermediate aldehyde as a cloudy oil which was used for the next reaction without further purification.



(3aR,6S,6aR)-2,2-Dimethyl-6-((Z)-prop-1-en-1-yl)tetrahydrofuro[3,4-d][1,3]dioxol-4-yl

benzoate (1) A flame dried round bottom flask was charged with freshly prepared ethyltriphenylphosphonium iodide⁸ (4.59 g, 11.0 mmol) and tetrahydrofuran (82 mL), forming an insoluble white suspension. A solution of 1.0 M KHMDS (10.9 mL) was added, immediately forming an orange solution. The ylide solution was allowed to stir 90 minutes, and was then cooled to -78 °C, whereupon a solution of **16** (assuming quantitative yield) in THF (60 mL) was added to the ylide. The reaction was then allowed to stir overnight, warming to room temperature. Upon completion, the reaction was quenched with saturated ammonium chloride (100 mL). The layers were separated and the aqueous layer was extracted with ethyl acetate (3 x 150 mL). The organic layers were combined and washed with brine (200 mL), dried over MgSO₄, filtered, and reduced. Purification via column chromatography (7:3 hexanes/ EtOAc) afforded the product olefin (1.96 g, 70% two steps) as a white crystalline solid. Experimental data matched that previously described.¹

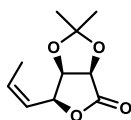
¹H NMR (400 MHz, CDCl₃) δ 8.06 – 8.01 (m, 2H), 7.62 – 7.56 (m, 1H), 7.48 – 7.43 (m, 2H), 6.41 (s, 1H), 5.88 – 5.79 (m, 1H), 5.70 – 5.62 (m, 1H), 5.00 (dd, *J* = 8.2, 3.7 Hz, 1H), 4.90 (d, *J* = 5.7 Hz, 1H), 4.82 (dd, *J* = 5.8, 3.7 Hz, 1H), 1.74 (dd, *J* = 7.0, 1.7 Hz, 3H), 1.54 (s, 3H), 1.36 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 165.22, 157.74, 133.49, 129.87, 128.56, 123.94, 113.25, 101.53, 99.77, 85.65, 81.06, 77.88, 26.26, 25.07, 14.03



(3aR,6S,6aR)-2,2-Dimethyl-6-((Z)-prop-1-en-1-yl)tetrahydrofuro[3,4-d]

[1,3]dioxol-4-ol (8) A flame dried round bottom flask was charged with **1** (4.77 g, 15.6 mmol), tetrahydrofuran (44 mL), and methanol (44 mL). K₂CO₃ (2.59 g, 18.8 mmol) was then added to the flask and the reaction was allowed to stir 3 h, at which point the starting material was completely consumed by TLC (if starting material remained, the reaction was allowed to continue stirring until which time TLC shows complete conversion to product). The mixture was then filtered, reduced, and purified by column chromatography (3:1-1:2 hexanes/ EtOAc) to afford the lactol (3.11 g, 99%) as a clear oil. Experimental data matched that previously described.¹

¹H NMR (400 MHz, CDCl₃) δ 5.87 – 5.77 (m, 1H), 5.68 – 5.59 (m, 1H), 5.41 (d, *J* = 2.2 Hz, 1H), 5.00 (dd, *J* = 8.6, 3.7 Hz, 1H), 4.71 (dd, *J* = 5.8, 3.7 Hz, 1H), 4.65 (d, *J* = 5.8 Hz, 1H), 2.42 (d, *J* = 2.4 Hz, 1H), 1.75 (dd, *J* = 7.0, 1.8 Hz, 3H), 1.48 (s, 3H), 1.33 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 129.84, 124.40, 112.66, 101.14, 85.99, 81.44, 75.58, 26.24, 24.96, 13.92

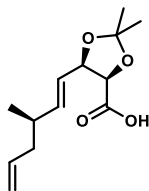


(3aR,6S,6aR)-2,2-Dimethyl-6-((Z)-prop-1-en-1-yl)dihydrofuro[3,4-d]

[1,3]dioxol-4(3aH)-one (2) A flame dried round bottom flask was charged with oxalyl chloride (0.11 mL, 1.3 mmol) and dry dichloromethane (6.5 mL). The stirring solution was cooled to -78 °C and DMSO (0.37 mL, 5.2 mmol) was added. The solution was maintained at this temperature for 1 hour and 20 minutes at which time a solution of **8** (126 mg, 0.63 mmol) in dry dichloromethane (1.7 mL + 2x 0.4 mL washes) was added via cannula at -78 °C. The reaction was maintained at this temperature for 2 hours and 30 minutes and then triethylamine (0.50 mL,

3.6 mmol) was added. The reaction was allowed to stir in the presence of the cooling bath for 2 hours, then at room temperature for an additional 1 hour. The reaction was then poured into a separatory funnel and washed with 0.5M HCl (2x 10 mL), saturated sodium bicarbonate (2x 10 mL), and brine. The organic fraction was dried over sodium sulfate, filtered and concentrated. Elution through a short plug of silica gel with dichloromethane (~50 mL) and concentration of the eluate afforded the product lactone as a white solid (117 mg, 94%), which may be used without further purification. Experimental data matched that previously described.¹

¹H NMR (500 MHz, CDCl₃) δ 6.00 – 5.92 (m, 1H), 5.72 – 5.65 (m, 1H), 5.28 (ddd, *J* = 8.8, 3.6, 1.2 Hz, 1H), 4.83 (d, *J* = 5.3 Hz, 1H), 4.77 (dd, *J* = 5.3, 3.6 Hz, 1H), 1.79 (dd, *J* = 7.1, 1.8 Hz, 3H), 1.50 (s, 3H), 1.40 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 174.26, 132.57, 122.31, 114.15, 77.85, 76.35, 74.64, 65.65, 26.91, 25.97, 13.87

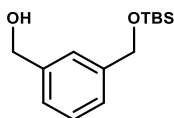


(4*R*,5*R*)-2,2-Dimethyl-5-((*R*,*E*)-3-methylhexa-1,5-dien-1-yl)-1,3-dioxolane-4-carboxylic acid

(3) A flame dried round bottom flask (previously rinsed in a base bath and oven dried) was charged with copper (I) cyanide (1.13 g, 12.61 mmol) and dry THF (630 mL). The slurry was cooled to -10 °C in an ice-salt water bath and freshly prepared allylmagnesium bromide⁹ (0.61M, 16.6 mL, 10.13 mmol) was added very slowly. The solution was stirred at -10 °C for 30 minutes prior to dropwise addition of a solution of **2** (1.00 g, 5.04 mmol) in THF (10 mL). The reaction was maintained at this temperature for ~ 10 minutes, then allowed to warm to room temperature and stirred an additional 3 hours at which point the starting material has been consumed (by

TLC). The reaction was quenched by addition of a 9:1 solution of saturated aqueous ammonium chloride and 1-M ammonium hydroxide (630 mL total volume). The solution was stirred ~ 1 hour until it reached a deep blue color. The reaction was then poured into a separatory funnel and the aqueous layer was separated and acidified to ~ pH 2 with 1-M HCl. The acidified aqueous layer was extracted with diethyl ether (5x 50 mL). The combined organic layer was washed with saturated aqueous ammonium chloride (2x 50mL) and brine, dried over sodium sulfate, filtered, and concentrated. Purification via column chromatography (9:1 DCM/MeOH) provided the acid as a pale yellow oil (1.07 g, 88%). Experimental data matched that previously described¹

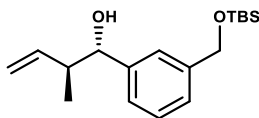
¹H NMR (500 MHz, CDCl₃) δ 5.84 (dd, *J* = 15.4, 7.1 Hz, 1H), 5.78 – 5.68 (m, 1H), 5.40 – 5.33 (m, 1H), 5.03 – 4.97 (m, 2H), 4.84 (t, *J* = 7.6 Hz, 1H), 4.64 (d, *J* = 7.5 Hz, 1H), 2.31 – 2.24 (m, 1H), 2.15 – 2.09 (m, 1H), 2.06 – 1.98 (m, 1H), 1.63 (s, 3H), 1.42 (s, 3H), 0.98 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 173.78, 142.58, 136.64, 122.04, 116.33, 111.27, 78.80, 76.91, 40.85, 36.10, 27.05, 25.41, 19.53



(3-(((*tert*-Butyldimethylsilyloxy)methyl)phenyl)methanol (17) Prepared according to known procedures:³ A flask was charged with sodium hydride (60% w/w, 583 mg, 14.6 mmol) and THF (32 mL). The slurry was cooled to 0°C and a solution of 1,3-benzenedimethanol (5.87 g, 42.5 mmol) in THF (9 ml) was added slowly *via* cannula. The mixture is stirred at this temperature for 15 minutes before addition of *tert*-butylammonium iodide (123 mg, 0.332 mmol) and portion-wise addition of *tert*-butyldimethylsilyl chloride (1.0 g, 6.6 mmol). The reaction was allowed to warm to room temperature slowly and stirred 20 hours. The reaction was quenched by addition

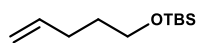
of water. The organic layer was separated and the aqueous layer was extracted 4 times with DCM. The combined organic layers were washed with brine, dried over sodium sulfate, filtered, and concentrated. The crude product was passed through a short column of silica gel (4:1 hexanes/EtOAc). The eluent was concentrated to afford a clear oil (1.13 g, 68%), which was used directly in the next step.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36 – 7.31 (m, 2H), 7.27 (s, 2H), 4.75 (s, 2H), 4.69 (d, $J = 4.4$ Hz, 2H), 0.95 (s, 9H), 0.11 (s, 6H)



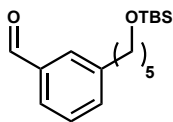
(1*S*,2*S*)-1-(3-(((*tert*-Butyldimethylsilyl)oxy)methyl)phenyl)-2-methylbut-3-en-1-ol ((-)-5a) A flask was charged with a stir bar and powdered 4Å mol sieves (250 mg) and was then flame dried. A solution of *E*-crotylboronate in toluene (prepared *via* known procedure,⁴ 1.29M, 8.5 mL) was added, followed by additional toluene (19 mL). The solution was cooled to -78°C. **4a** (922 mg, 3.68 mmol) was then added as a solution in toluene (9 mL + 2 mL wash) slowly *via* syringe pump over ~20 minutes. The reaction was stirred for 3 hours at this temperature. NaOH was then added (2M (aq), 14 mL) and the reaction was transferred to a 0 °C cooling bath and stirred for 20 minutes. The reaction was then filtered through a pad of celite. The organic layer was separated and the aqueous layer was extracted 4 times with diethyl ether. The combined organic layers are washed with water and brine, dried over sodium sulfate, filtered, and concentrated. The crude product was purified by flash column chromatography (4:1 hexanes/EtOAc) to afford the product alcohol as a clear oil (961 mg, 85%). The product was thus obtained as a single diastereomer as determined by $^1\text{H NMR}$.

¹H NMR (500 MHz, CDCl₃) δ 7.36 – 7.18 (m, 4H), 5.88 – 5.75 (m, 1H), 5.25 – 5.14 (m, 2H), 4.75 (s, 2H), 4.36 (d, *J* = 7.8 Hz, 1H), 2.52 – 2.42 (m, 1H), 2.12 (s, 1H), 0.94 (s, 9H), 0.87 (d, *J* = 6.8 Hz, 3H), 0.10 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 142.50, 141.61, 140.84, 128.30, 125.57, 124.67, 116.91, 78.05, 65.10, 46.39, 26.09, 18.56, 16.68, -5.07; **IR** (neat): 3436, 3078, 3031, 2956, 2928, 2856, 1710, 1639, 1607, 1488, 1472, 1462, 1387, 1360, 1254, 1155, 1097, 1032, 1005, 938, 910, 836, 774, 707, 680, 662 cm⁻¹; [α]_D²⁵ -39 (c = 1.00 in CHCl₃); **HRMS** (ES⁺): Found 329.1896 (-1.7 ppm), C₁₈H₃₀O₂SiNa (M+Na⁺) requires 329.1913



***tert*-Butyldimethyl(pent-4-en-1-yloxy)silane (18)** Prepared according to known procedures.⁵ A flask was charged with 4-penten-1-ol (2.35 mL, 23.2 mmol) and DMF (23 mL). The solution was cooled to 0°C, followed by addition of *tert*-butyldimethylsilyl chloride (4.55 g, 30.2 mmol), imidazole (2.06 g, 30.2 mmol), and DMAP (142 mg, 1.16 mmol). The reaction was allowed to slowly warm to room temperature and was stirred for 19 hours. The reaction was quenched by addition of brine and the organic layer was separated. The aqueous layer was extracted 5 times with EtOAc. The combined organic layers were washed with a saturated aqueous ammonium chloride, sodium bicarbonate, water, and brine, dried over sodium sulfate, filtered, and concentrated. The crude product was purified by flash column chromatography (5% EtOAc in hexanes) to afford the product alkene as a clear oil (3.68 g, 79%). *NOTE: The product is a volatile compound. Care should be taken when removing solvent. Refrain from leaving under vacuum for extended periods.* Experimental data matched that previously described.⁵

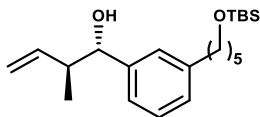
¹H NMR (500 MHz, CDCl₃) δ 5.88 – 5.78 (m, 1H), 5.07 – 4.92 (m, 2H), 3.62 (t, *J* = 6.5 Hz, 2H), 2.13 – 2.07 (m, 2H), 1.65 – 1.58 (m, 2H), 0.89 (s, 9H), 0.05 (s, 6H)



3-(5-((*tert*-Butyldimethylsilyloxy)pentyl) benzaldehyde (4b) A 2-neck flask fitted with a reflux condenser and septum was charged with **18** (1.25 g, 6.24 mmol) and THF (2.8 mL). The solution was cooled to 0 °C followed by slow addition of 9-BBN (0.5M in THF, 12.5 mL). The reaction was allowed to warm to room temperature in the presence of the cooling bath, and was then stirred a further 19 hours at room temperature. DMF (16 mL), PdCl₂(dppf)-CH₂Cl₂ (69 mg, 0.094 mmol), *meta*-bromobenzaldehyde (0.36 mL, 3.12 mmol) and potassium carbonate (1.55 g, 11.2 mmol) were added. The mixture was heated to 50°C and stirred a further 24 hours. The reaction was then cooled to room temperature and poured into a separatory funnel containing water (50 mL). Toluene was added (it may be necessary to add additional water for layers to separate) and the organic layer was separated. The aqueous layer was extracted 5 times with toluene (4x 50 mL), and the combined organic layers were washed 4 times with water (4x 25 mL) and one time with brine, dried over sodium sulfate, filtered, and concentrated to afford a viscous brown oil as the crude product. Purification by flash column chromatography (0 to 10% EtOAc in hexanes) afforded the product aldehyde as a clear oil (821 mg, 86%).

¹H NMR (500 MHz, CDCl₃) δ 10.00 (s, 1H), 7.72 – 7.61 (m, 2H), 7.54 – 7.40 (m, 2H), 3.60 (t, *J* = 6.5 Hz, 2H), 2.72 – 2.68 (m, 2H), 1.70 – 1.62 (m, 2H), 1.55 – 1.50 (m, 2H), 1.41 – 1.33 (m, 2H), 0.88 (s, 9H), 0.03 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 192.74, 143.91, 136.66, 134.86, 129.52, 129.05, 127.65, 63.17, 35.76, 32.76, 31.21, 26.09, 25.57, 18.49, -5.14; **IR** (neat): 2928, 2856, 2726, 1702, 1603, 1588, 1471, 1462, 1386, 1360, 1301, 1250, 1142, 1096, 1005, 938, 910,

833, 774, 692, 651 cm^{-1} ; **HRMS** (ES^+): Found 307.2117 (+2.4 ppm), $\text{C}_{18}\text{H}_{30}\text{O}_2\text{Si}$ ($\text{M}+\text{H}^+$) requires 307.2093



(1*S*,2*S*)-1-(3-(5-((*tert*-Butyldimethylsilyloxy)pentyl)phenyl)-2-methylbut-3-en-1-ol ((-)-**5b**)

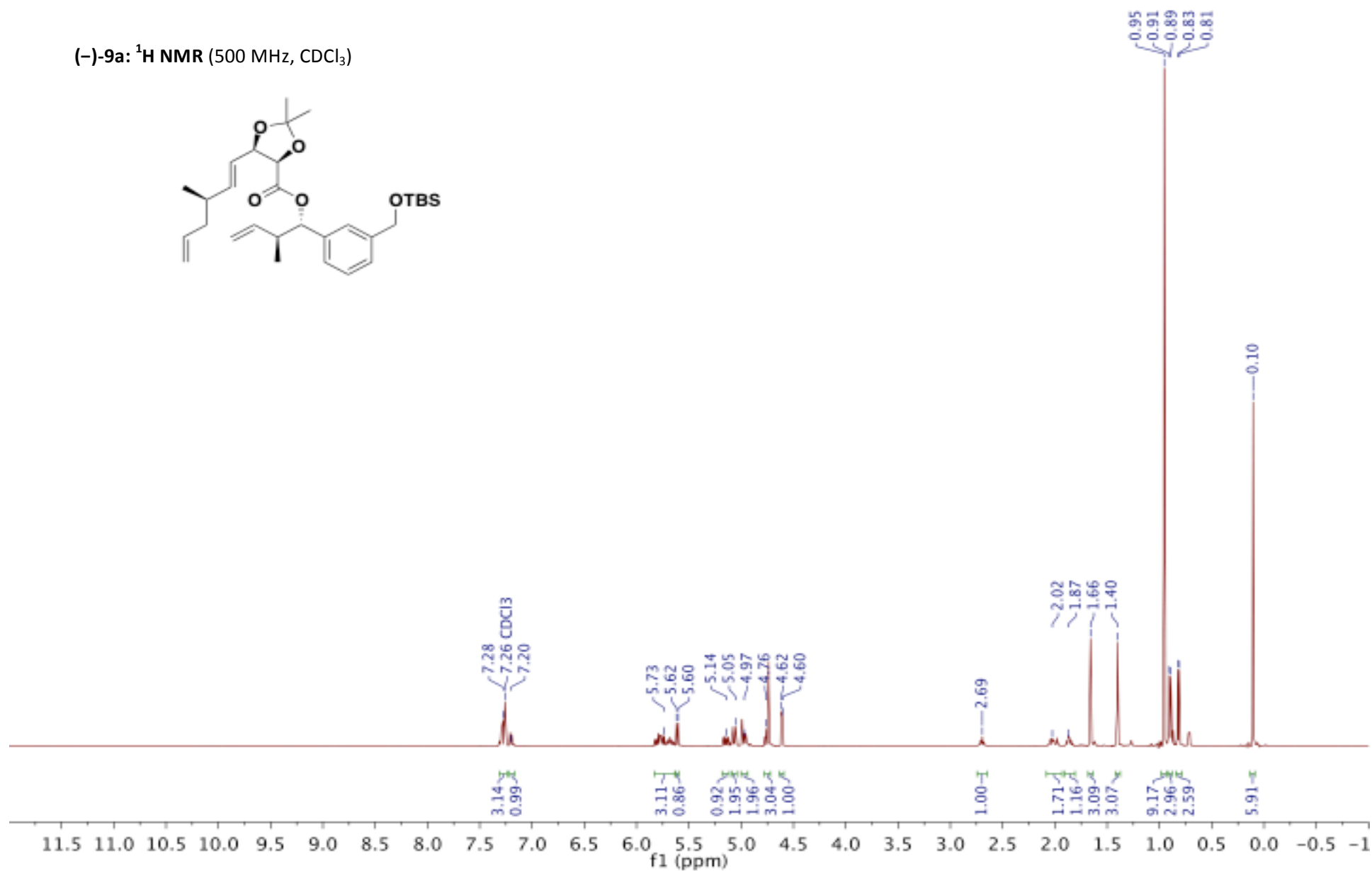
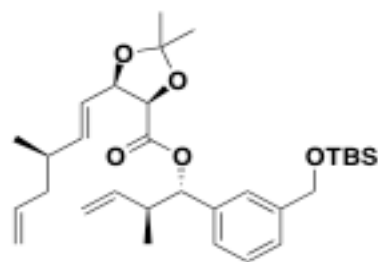
Prepared according to the procedure for (-)-**5a**: **4a** (821 mg, 2.68 mmol), *E*-crotylboronate (6.2 mL, 8.0 mmol), toluene (20 mL), and NaOH (2M, 10mL) yielded 810mg (83%) of the product as a clear oil. Purified by column chromatography (10% EtOAc in hexanes).

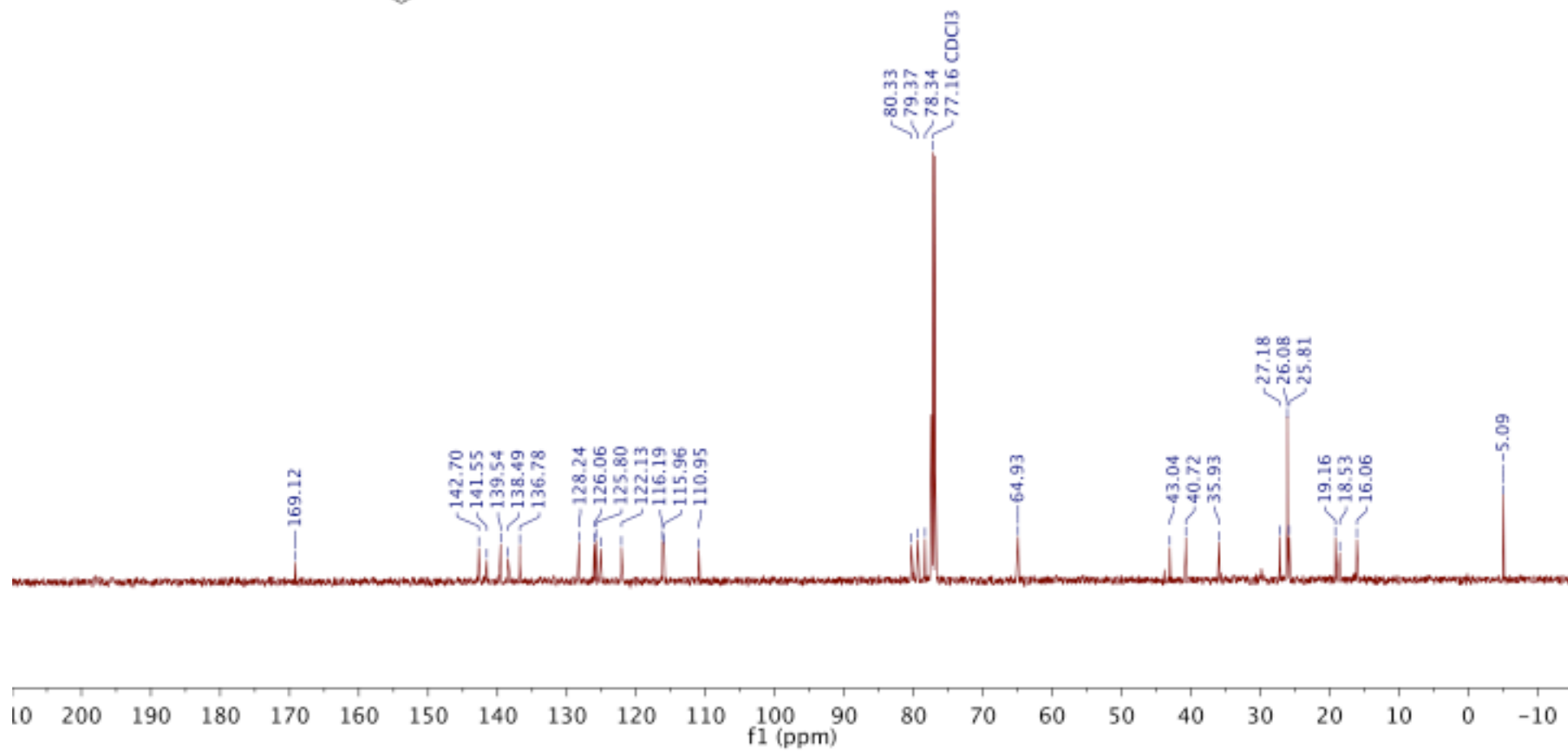
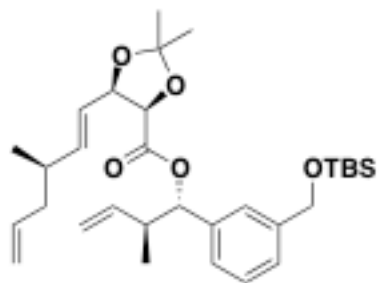
^1H NMR (500 MHz, CDCl_3) δ 7.26 – 7.22 (m, 1H), 7.15 – 7.12 (m, 2H), 7.11 – 7.08 (m, 1H), 5.86 – 5.76 (m, 1H), 5.23 – 5.15 (m, 2H), 4.33 (dd, $J = 8.0, 2.0$ Hz, 1H), 3.60 (t, $J = 6.6$ Hz, 2H), 2.66 – 2.57 (m, 2H), 2.53 – 2.42 (m, 1H), 1.67 – 1.59 (m, 2H), 1.57 – 1.51 (m, 2H), 1.40 – 1.33 (m, 2H), 0.89 (s, 9H), 0.86 (d, $J = 6.8$ Hz, 3H), 0.04 (s, 6H); **^{13}C NMR** (126 MHz, CDCl_3) δ 142.84, 142.48, 140.92, 128.24, 127.86, 126.98, 124.31, 116.79, 78.07, 63.31, 46.37, 36.09, 32.83, 31.47, 25.66, 18.49, 16.71, -5.14; **IR** (neat): 3438, 3080, 3022, 2928, 2856, 1639, 1607, 1471, 1462, 1387, 1361, 1254, 1155, 1098, 1005, 911, 833, 813, 774, 707, 679, 661 cm^{-1} ; $[\alpha]_{\text{D}}^{25}$ -24.1 ($c = 4.20$ in CHCl_3); **HRMS** (ES^+): Found 385.2544 (+0.5 ppm), $\text{C}_{22}\text{H}_{38}\text{O}_2\text{SiNa}$ ($\text{M}+\text{Na}^+$) requires 385.2539

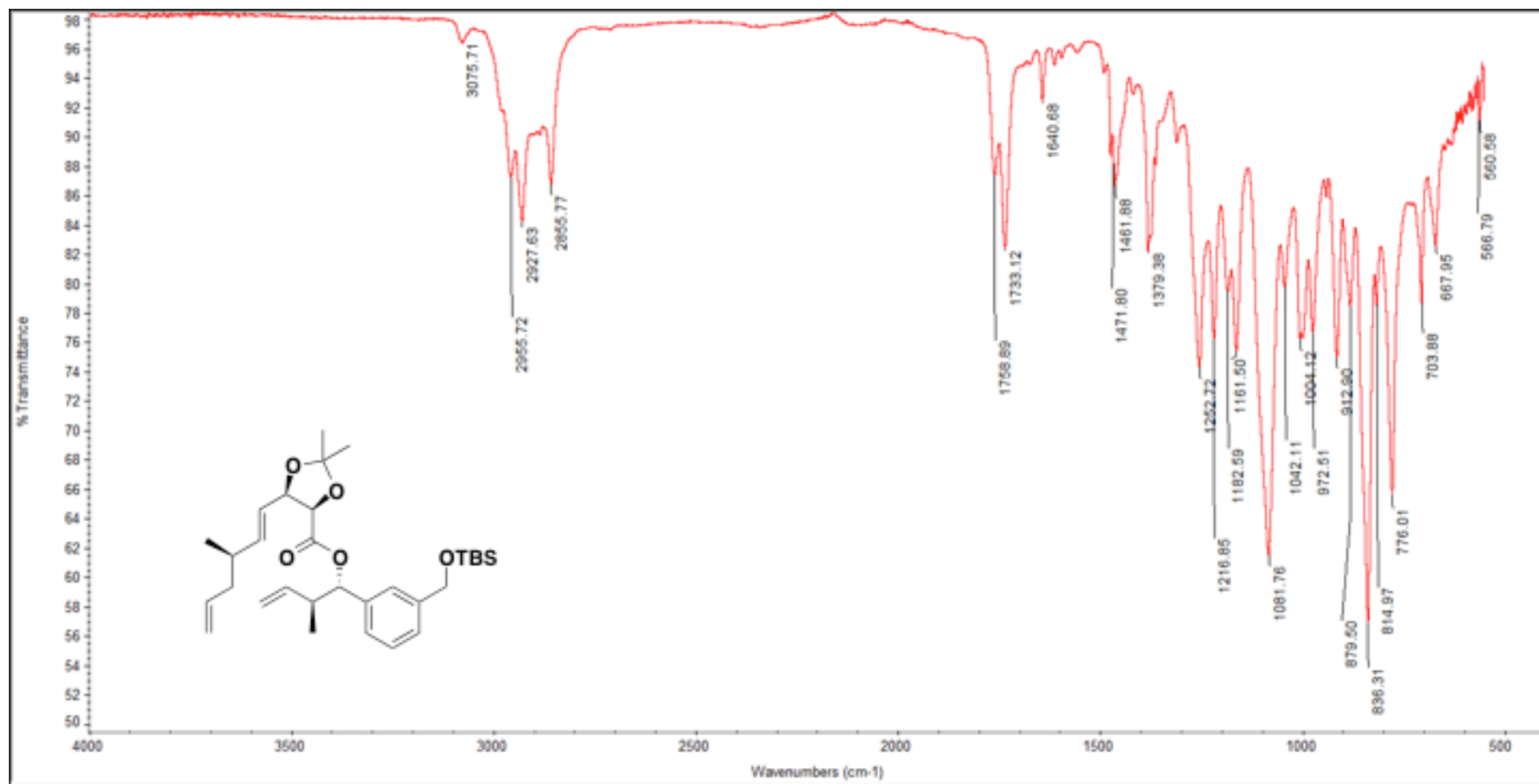
The synthetic route to acid **3** was greatly improved upon compared to lab's previous efforts.¹ In addition to reducing the time it took to accomplish this route, we were also able to make it safer by reducing the use of toxic chemicals. Most importantly, we achieved a 36% overall yield in the route to **3**, a 50% increase from past efforts.

5.3 Experimental Spectra

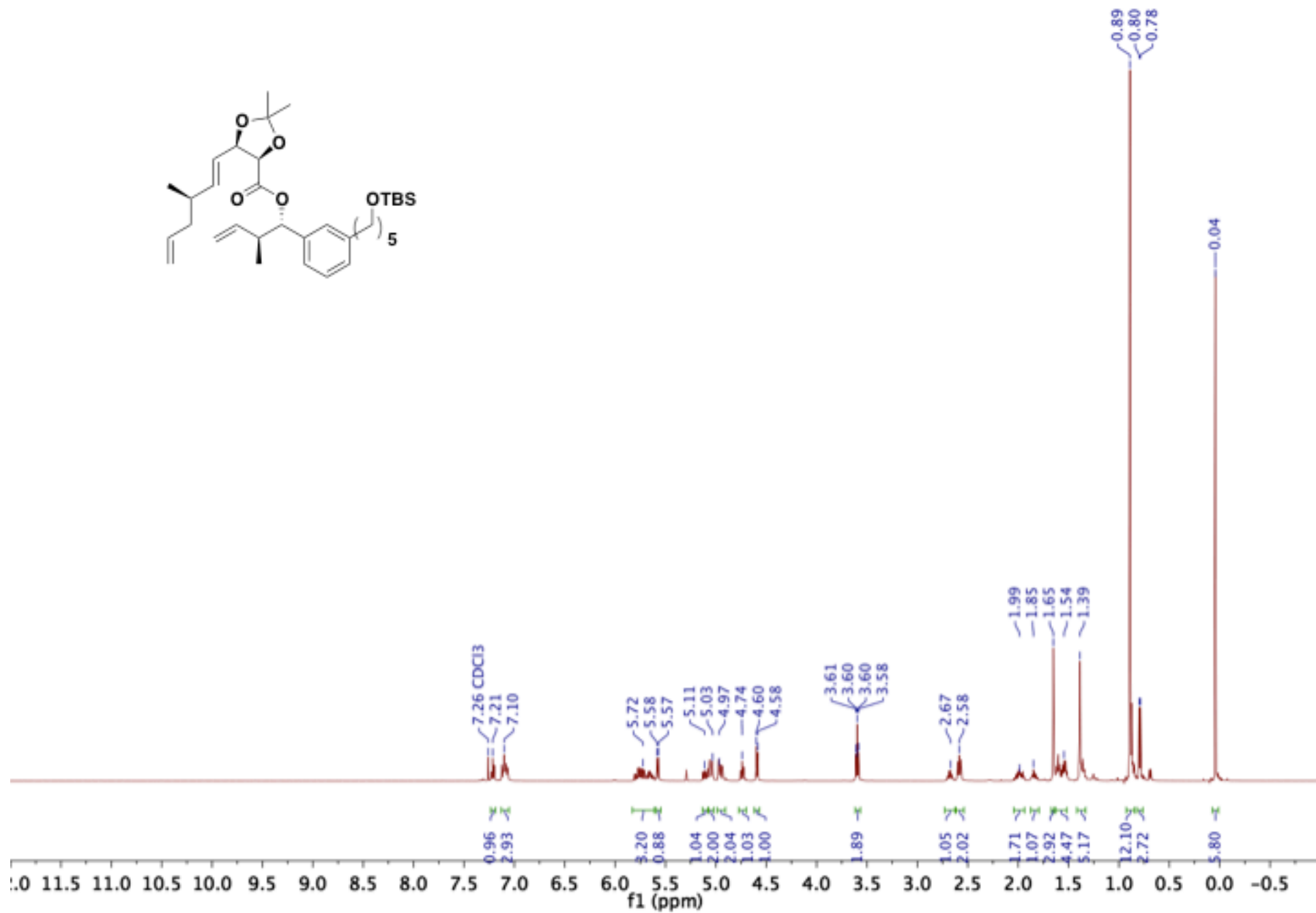
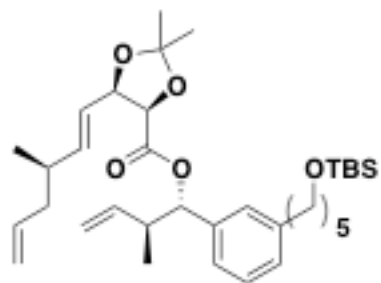
(-)-9a: ^1H NMR (500 MHz, CDCl_3)



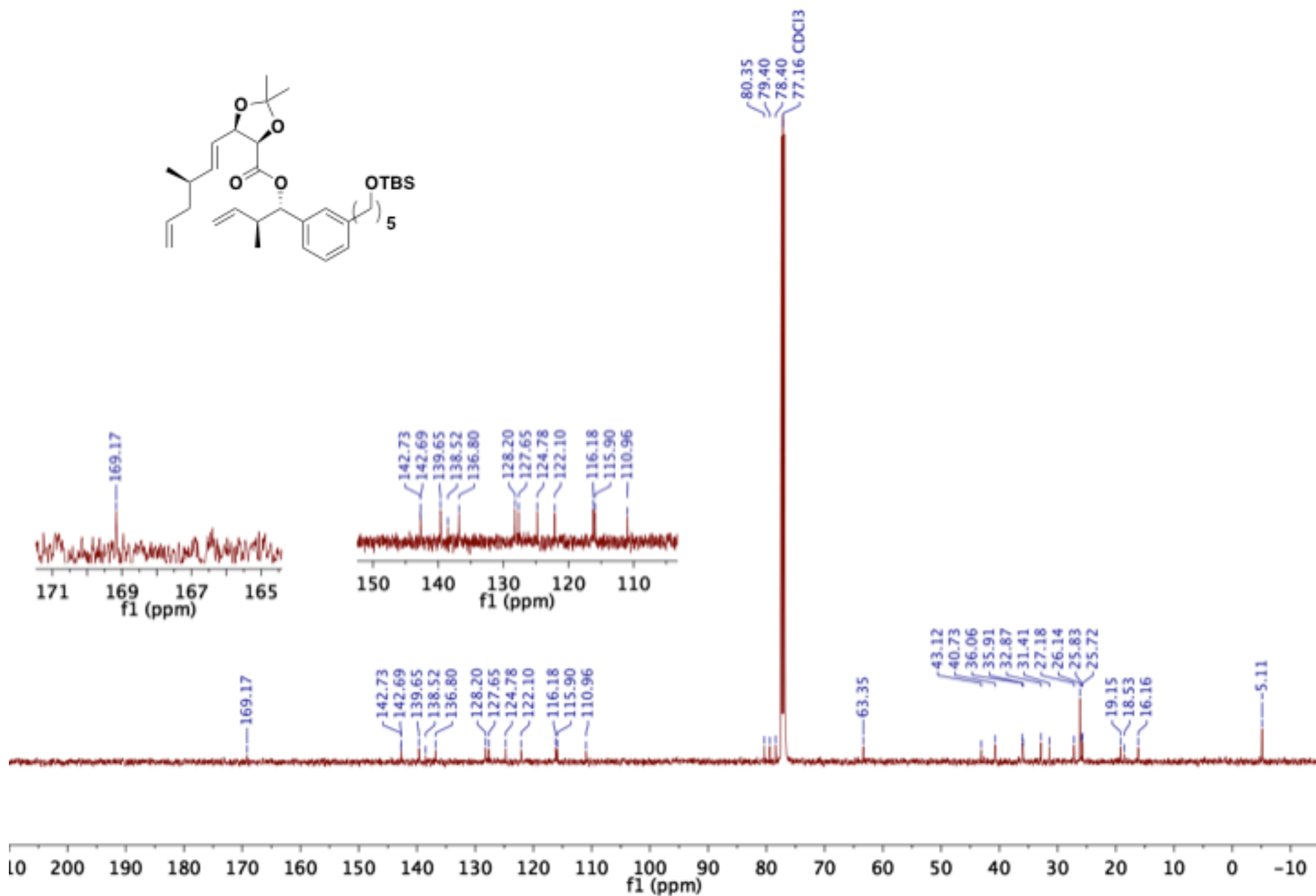
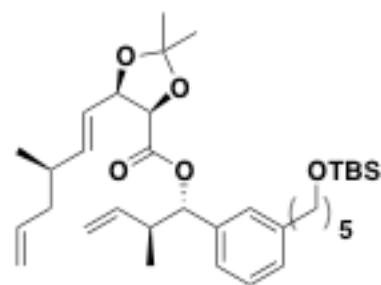


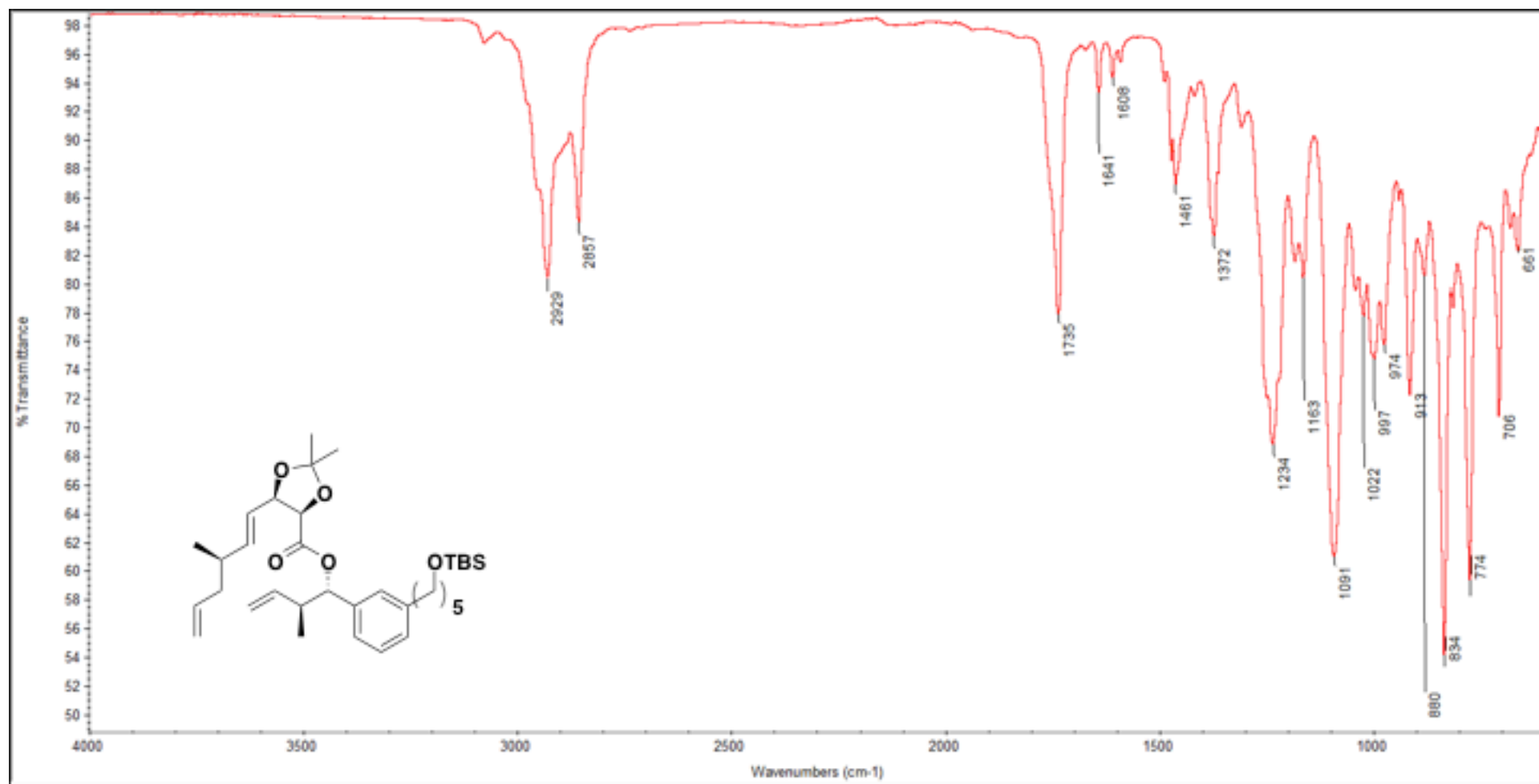


(-)-9b: ^1H NMR (500 MHz, CDCl_3)

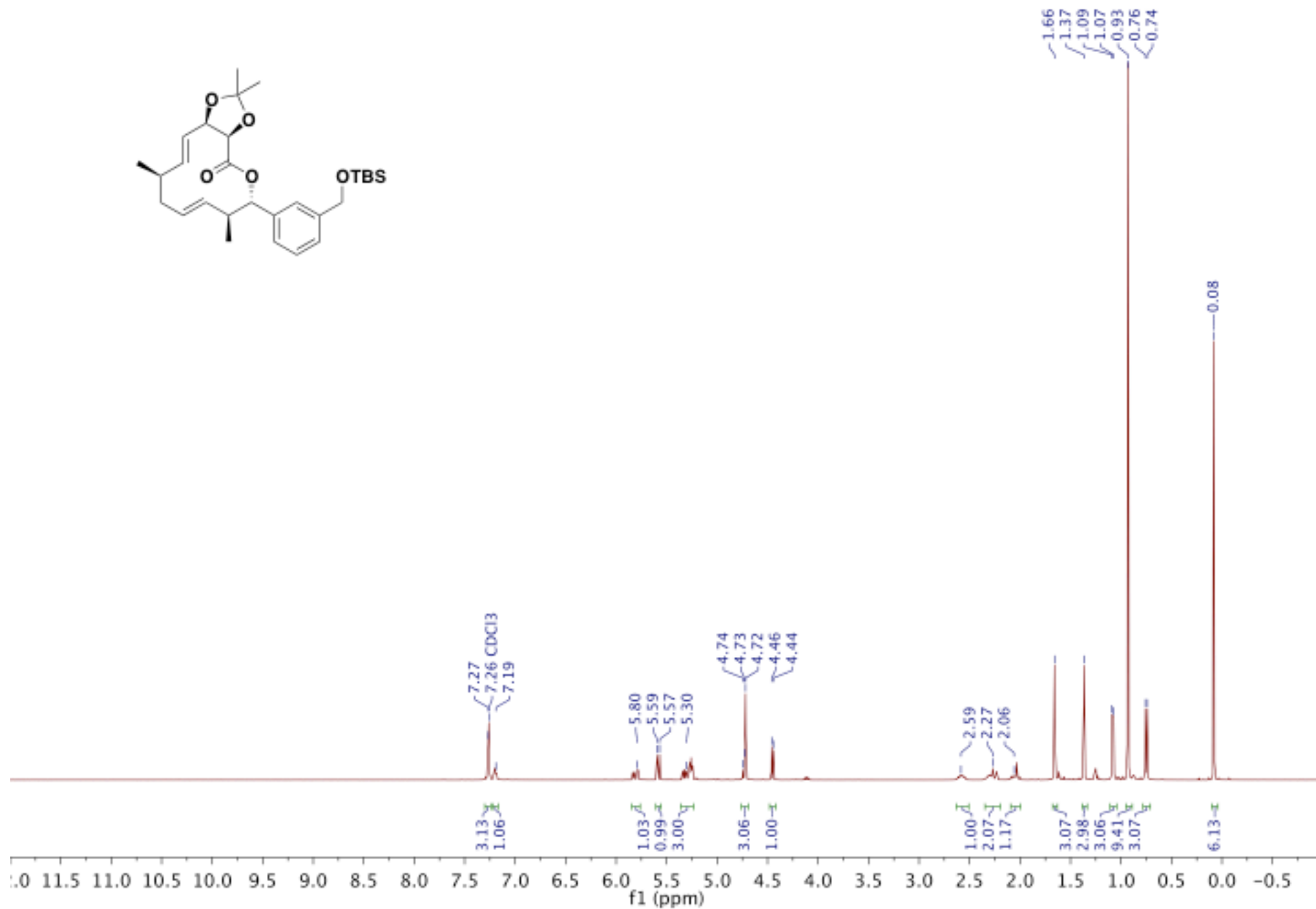
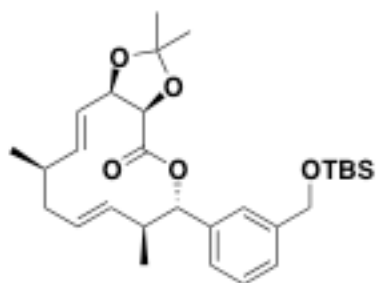


(-)-9b: ^{13}C NMR (126 MHz, CDCl_3)

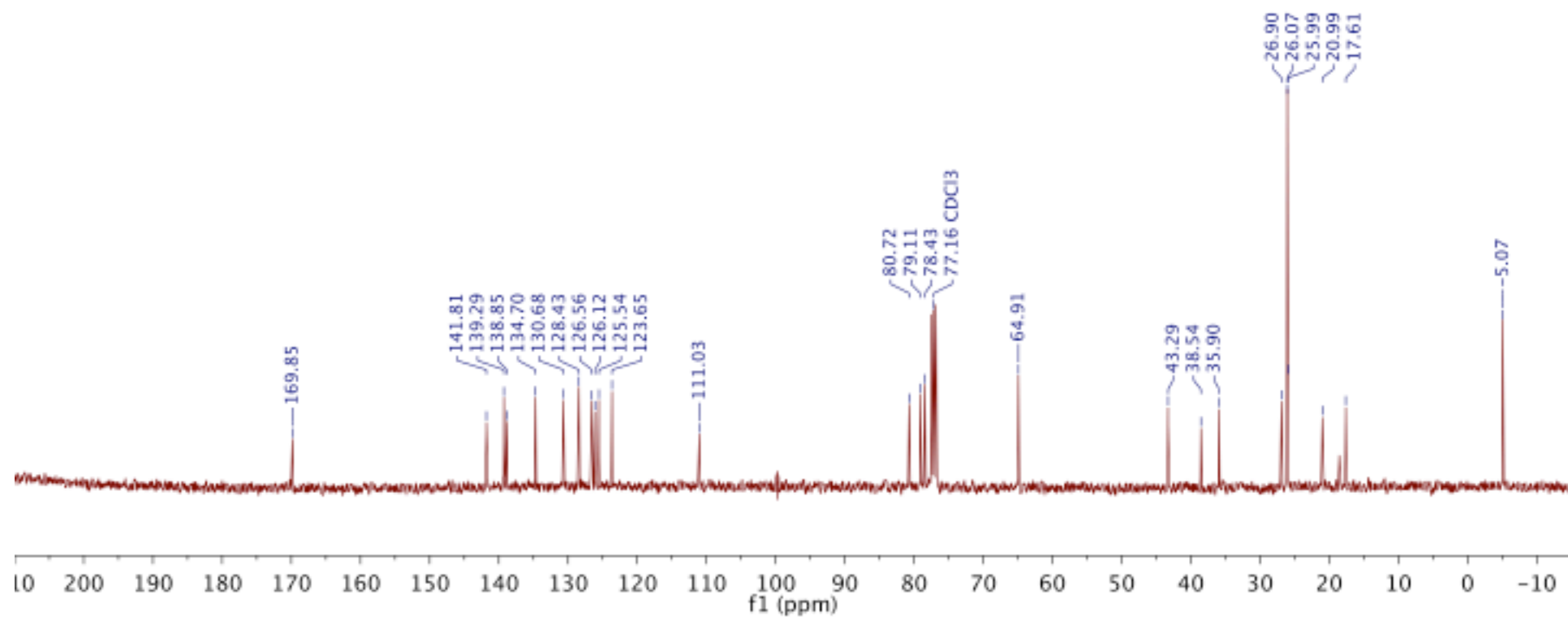
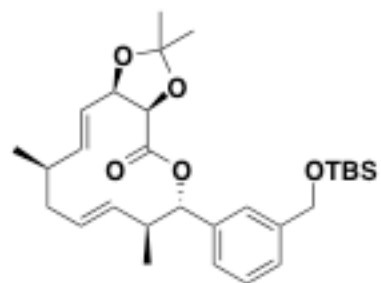


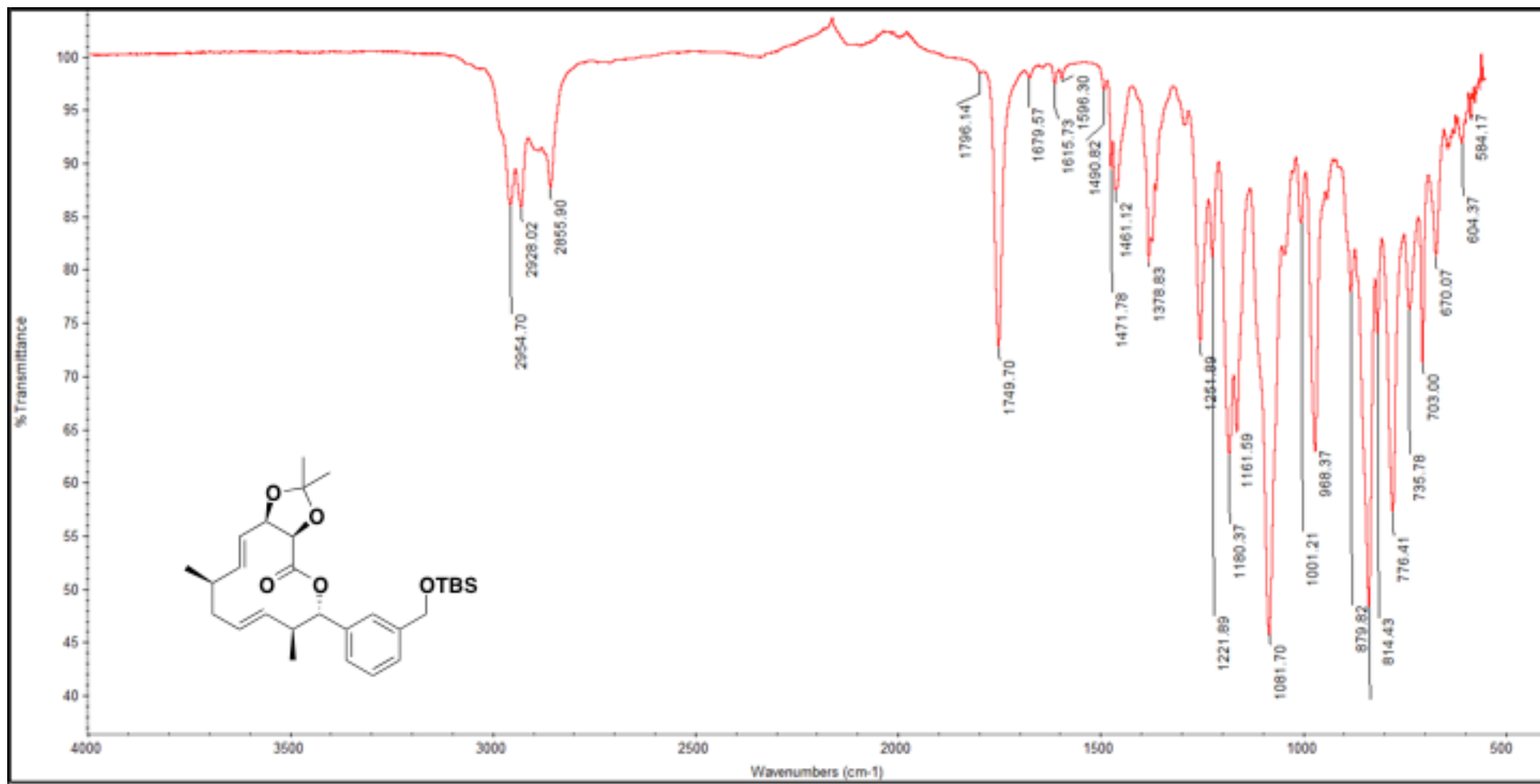


(-)-6a: ¹H NMR (500 MHz, CDCl₃)

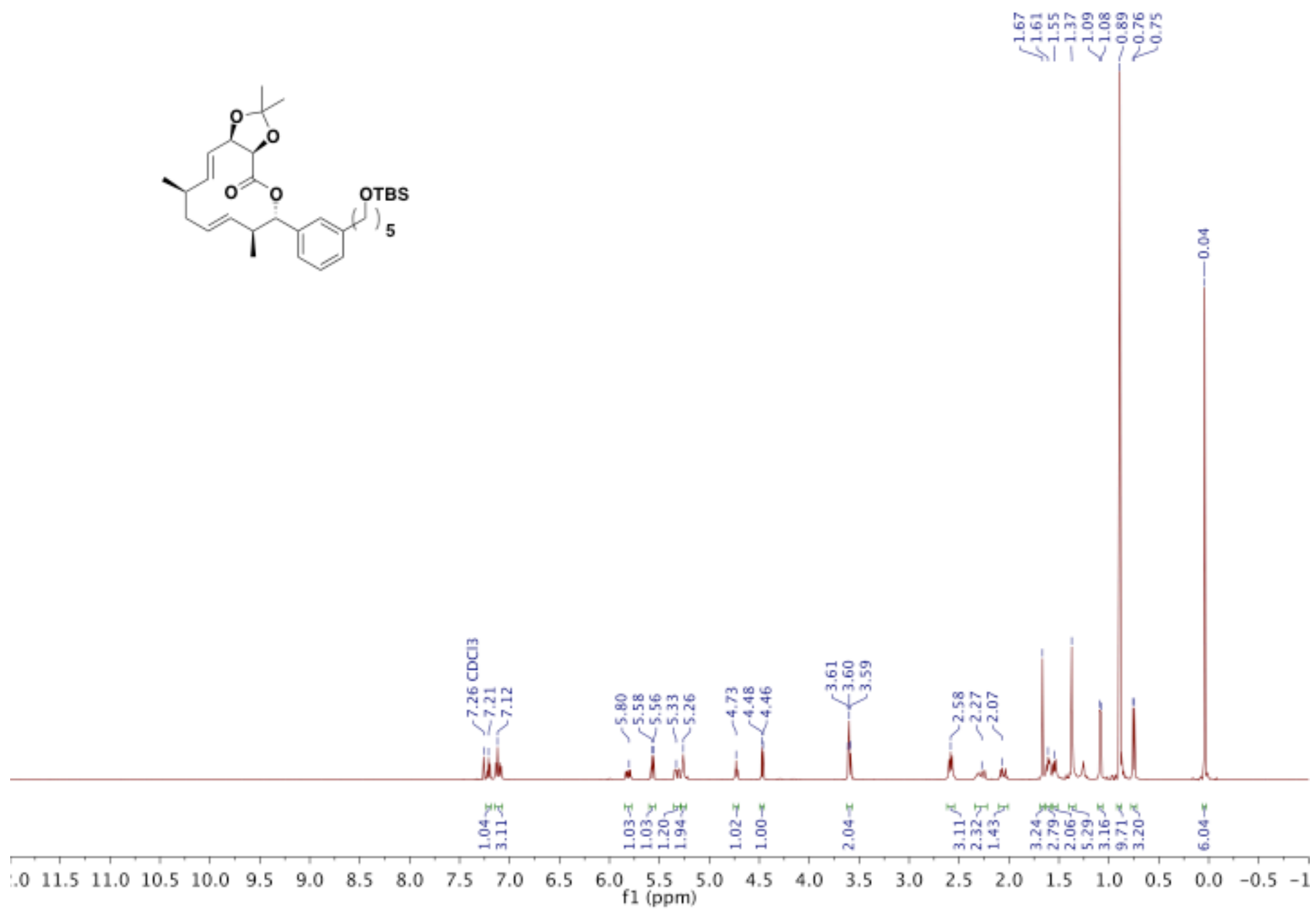
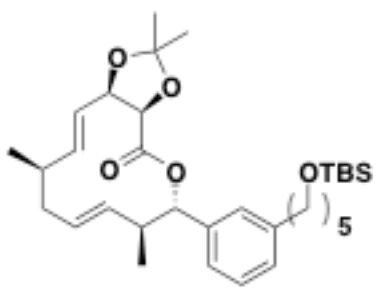


(-)-6a: ^{13}C NMR (126 MHz, CDCl_3)

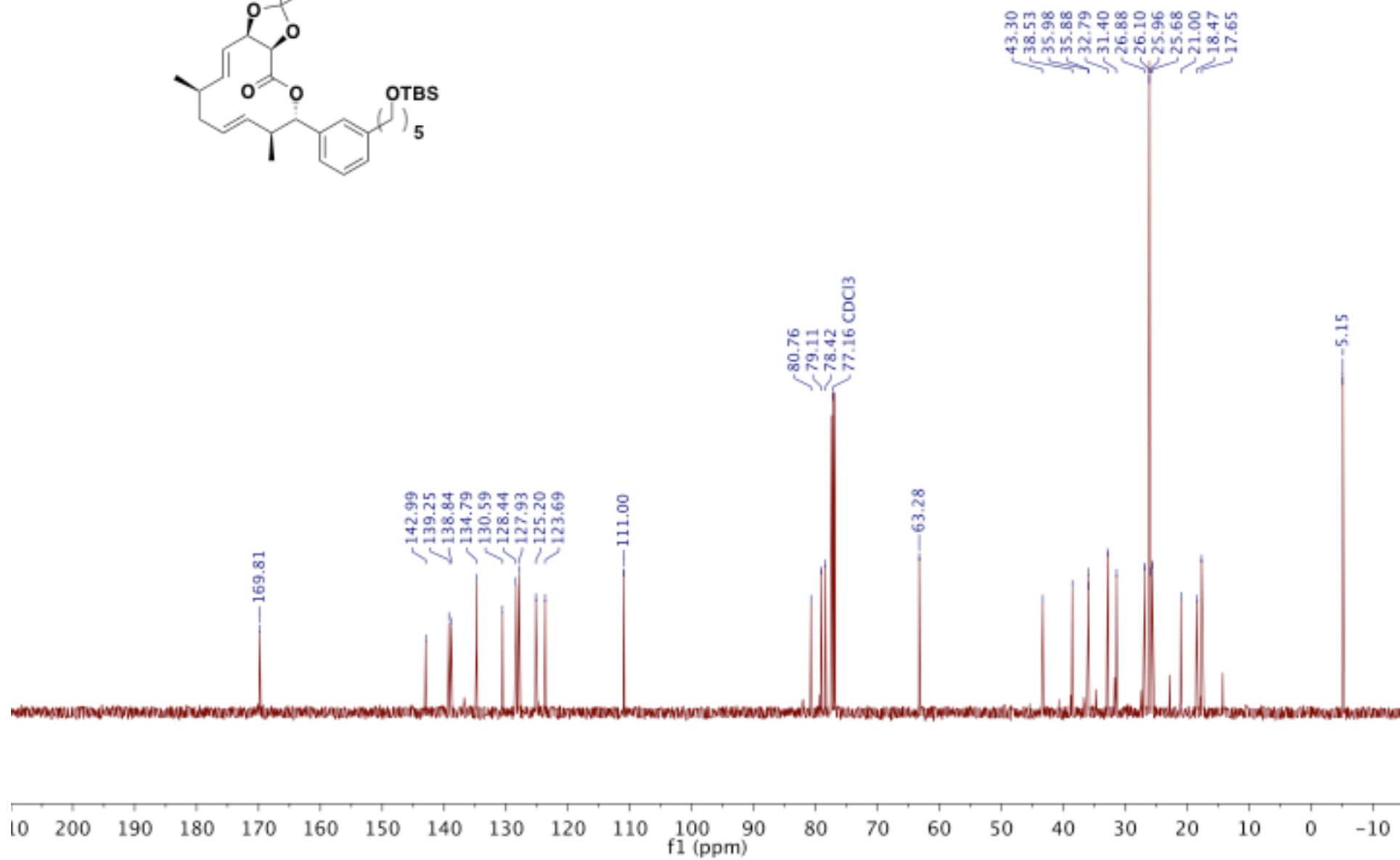
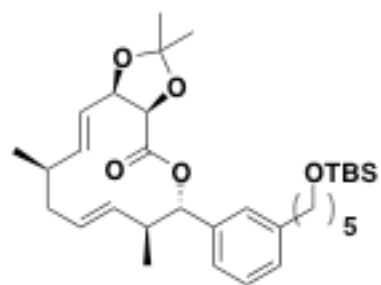


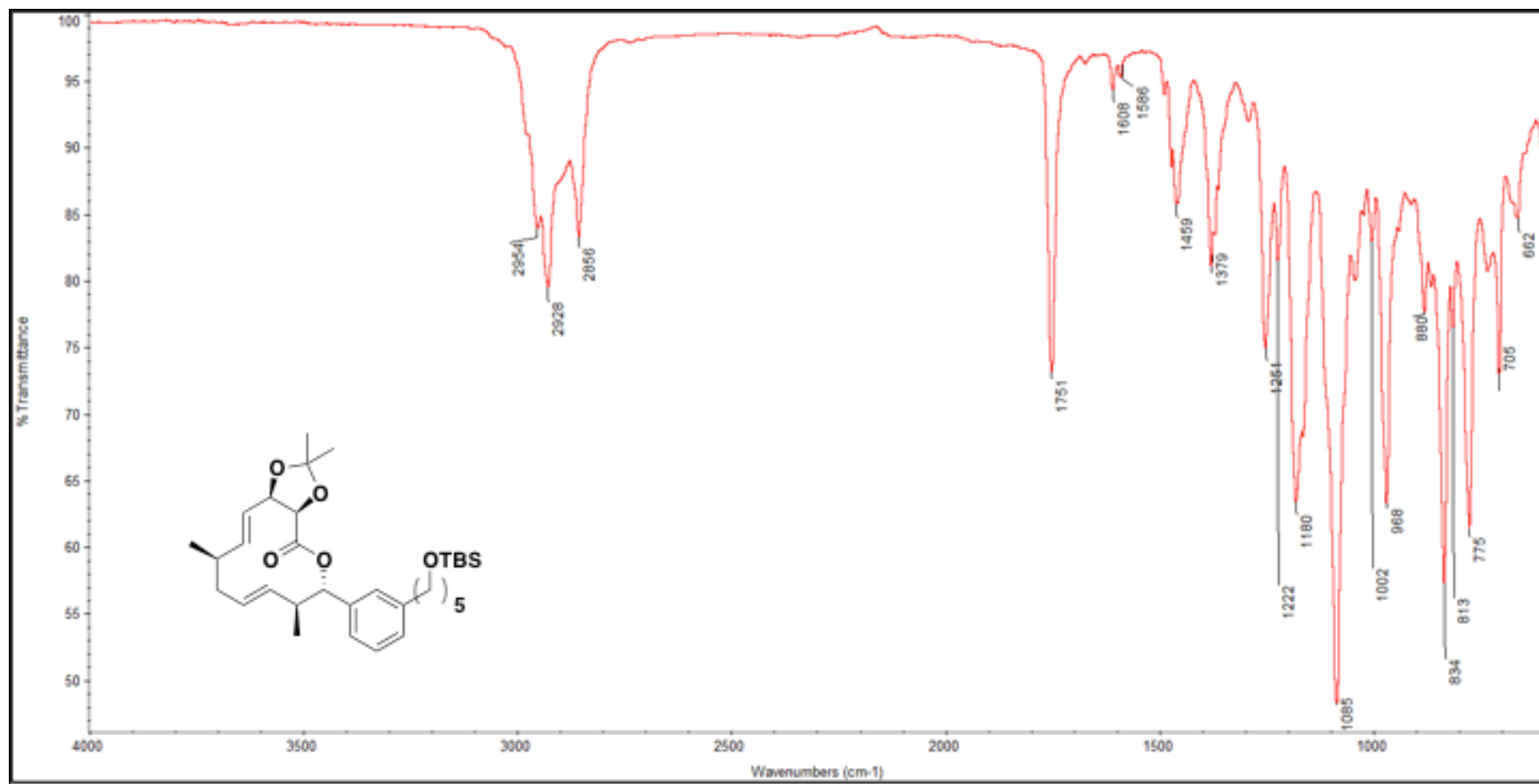


(-)-6b: ¹H NMR (500 MHz, CDCl₃)

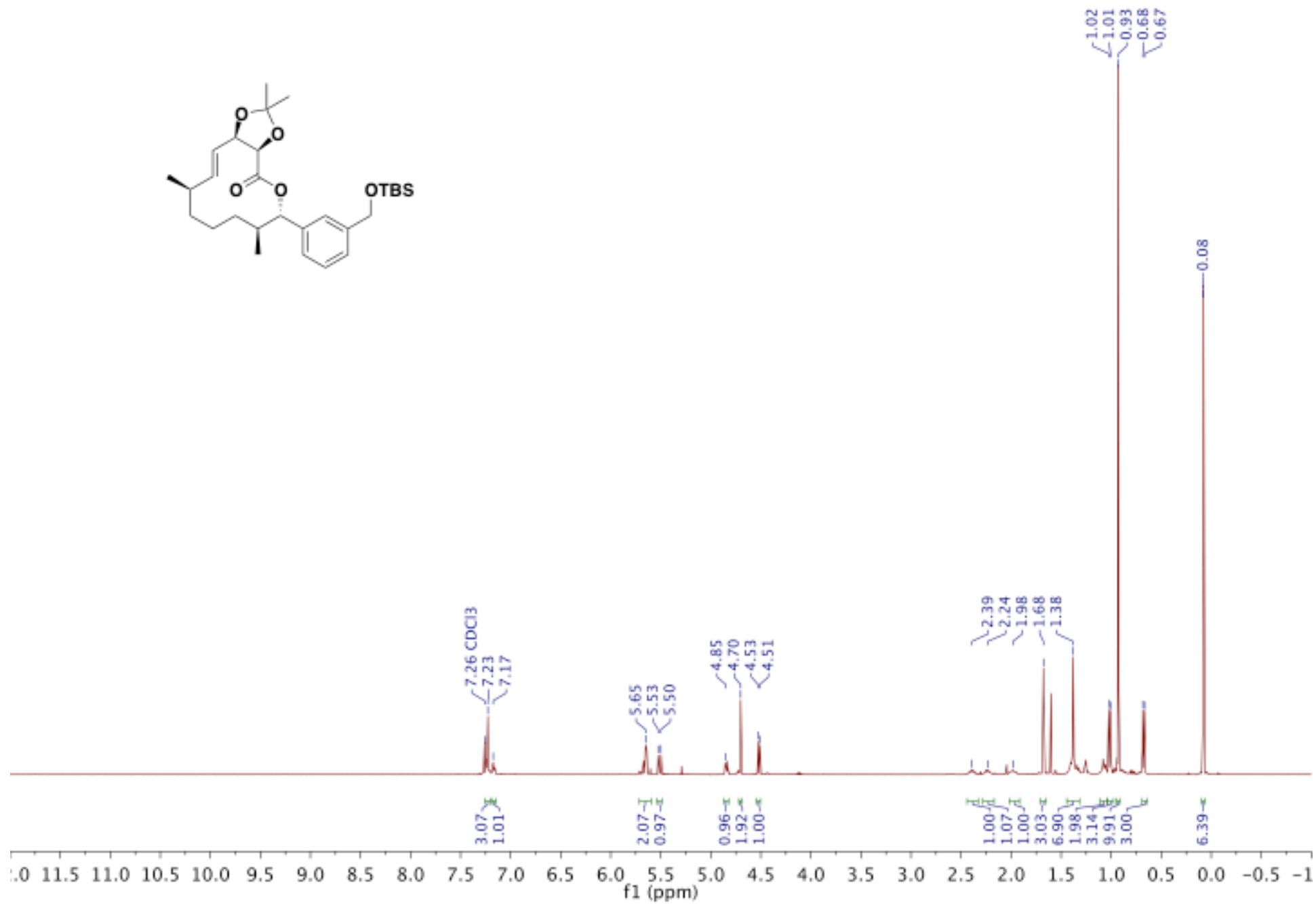
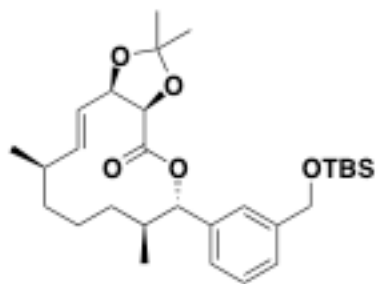


(-)-6b: ^{13}C NMR (126 MHz, CDCl_3)

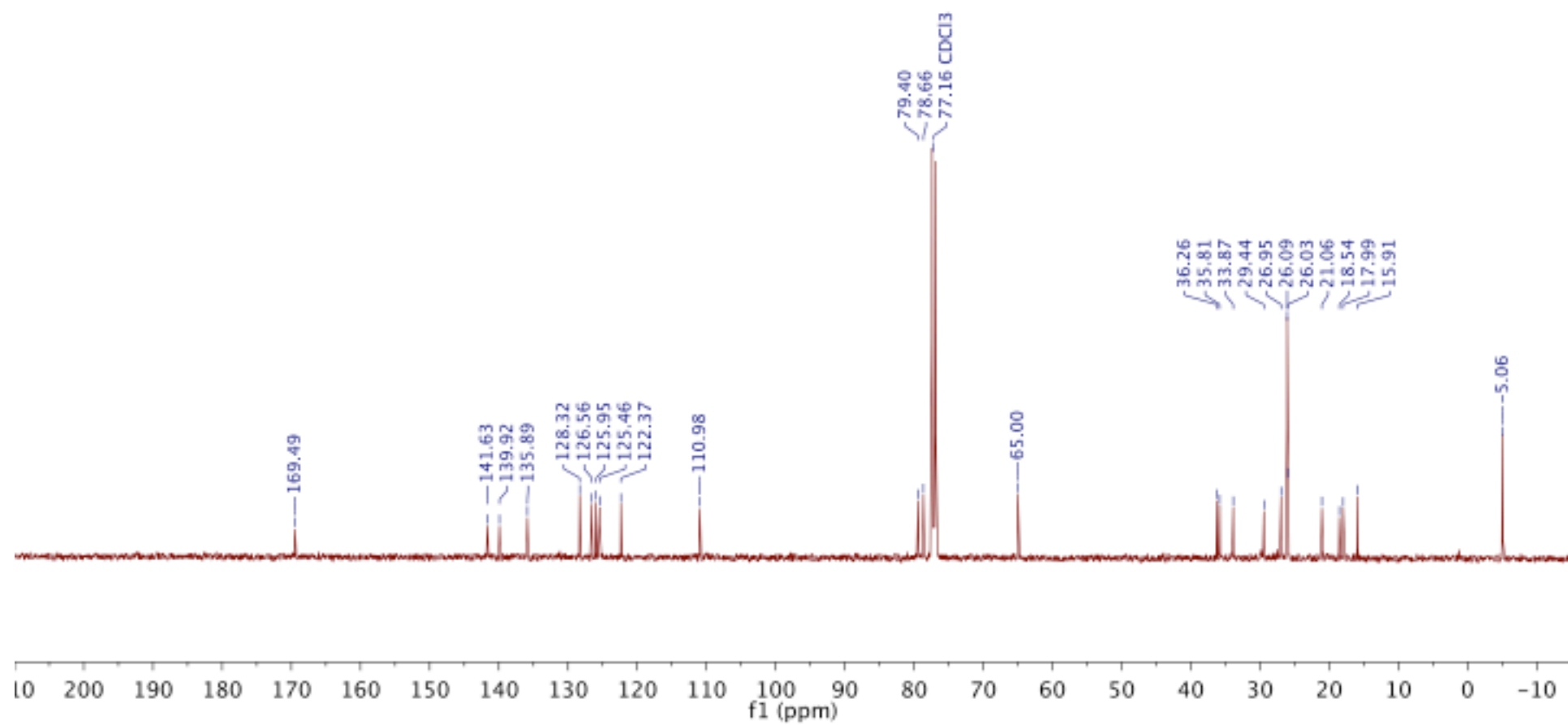
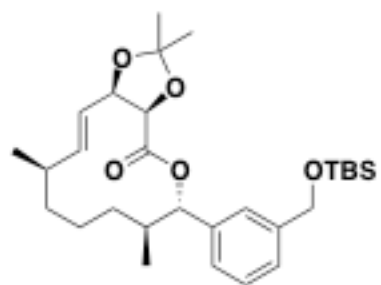


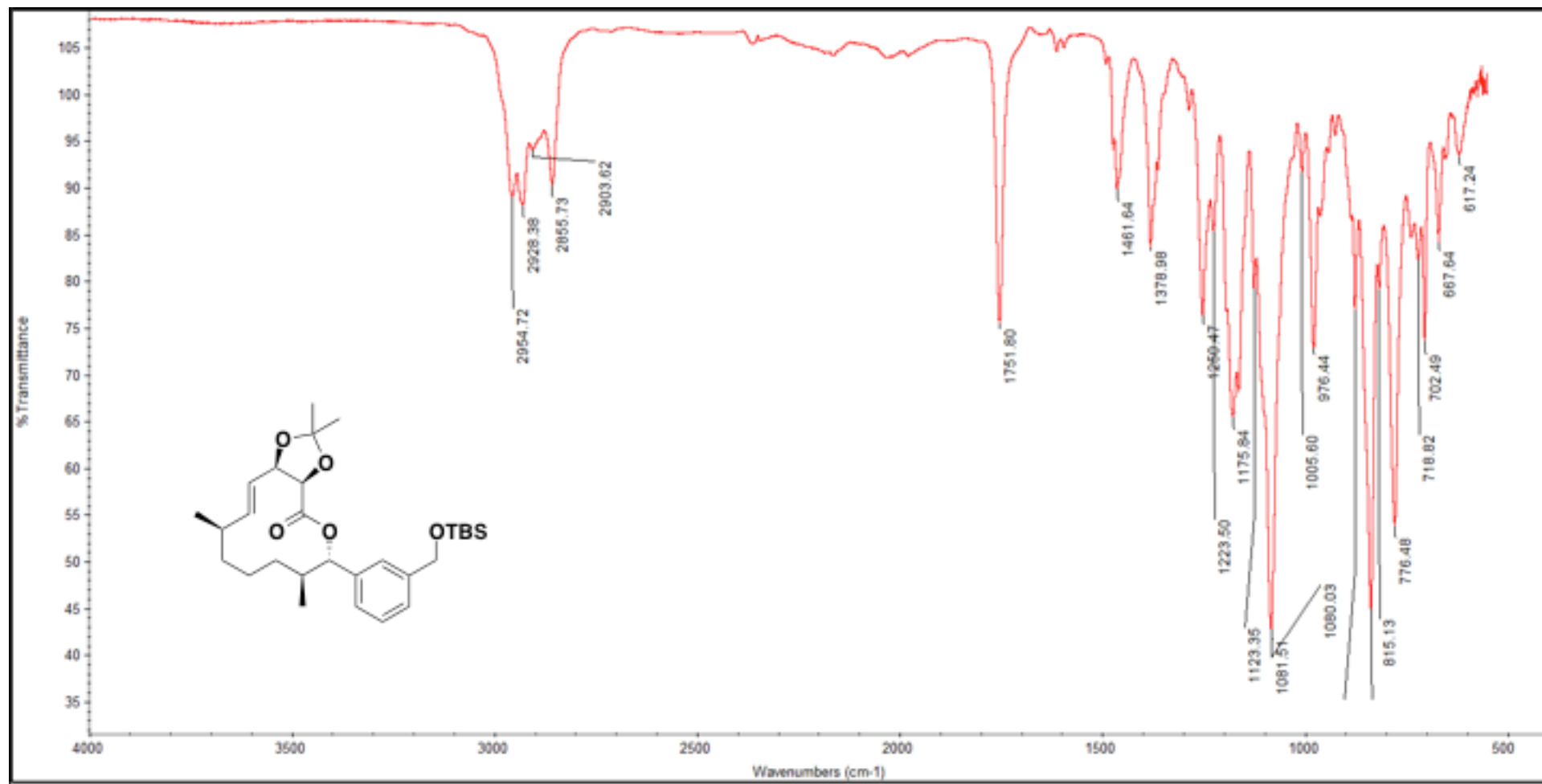


(-)-7a: ¹H NMR (500 MHz, CDCl₃)

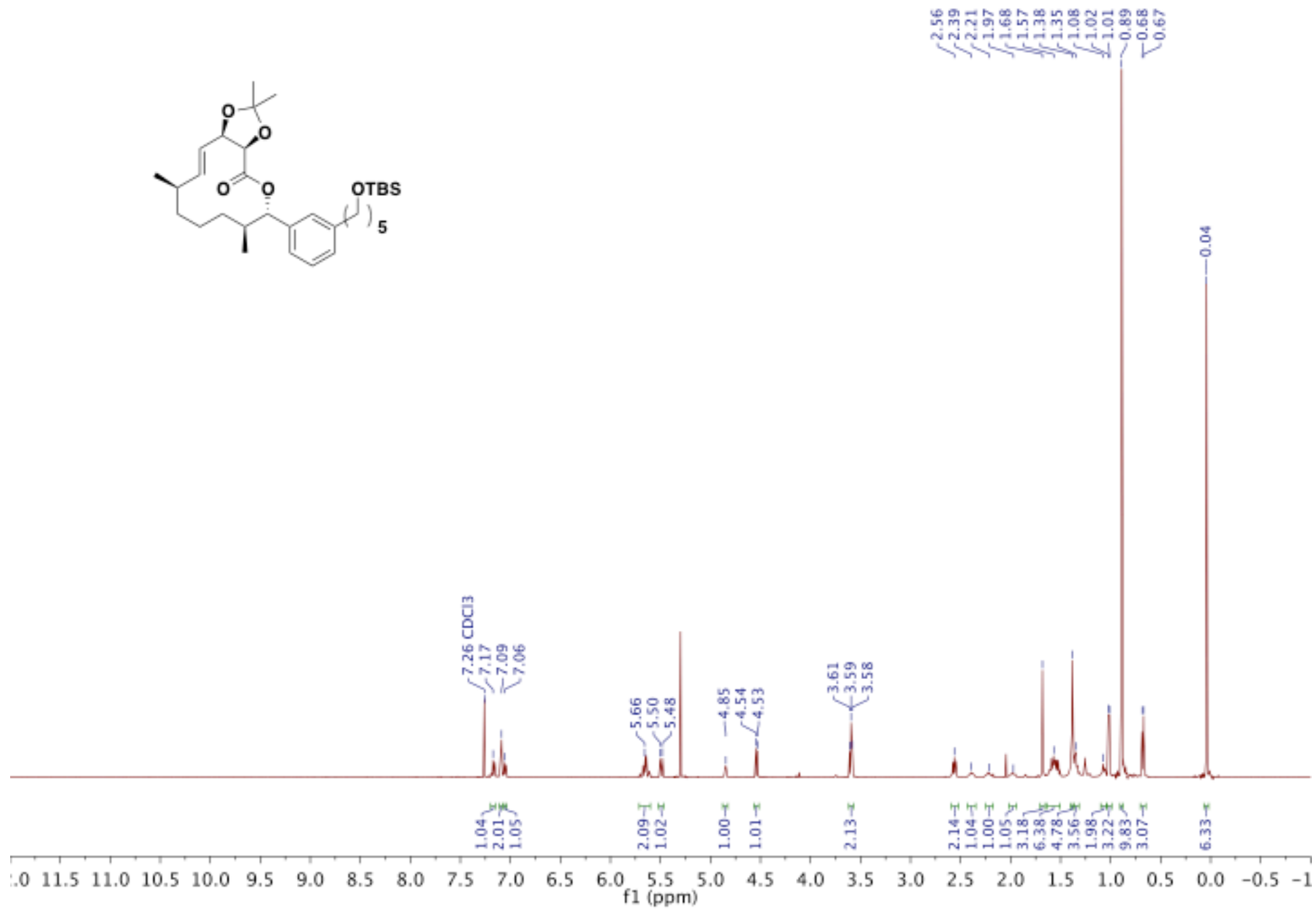
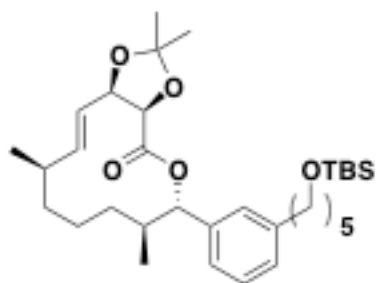


(-)-7a: ^{13}C NMR (126 MHz, CDCl_3)

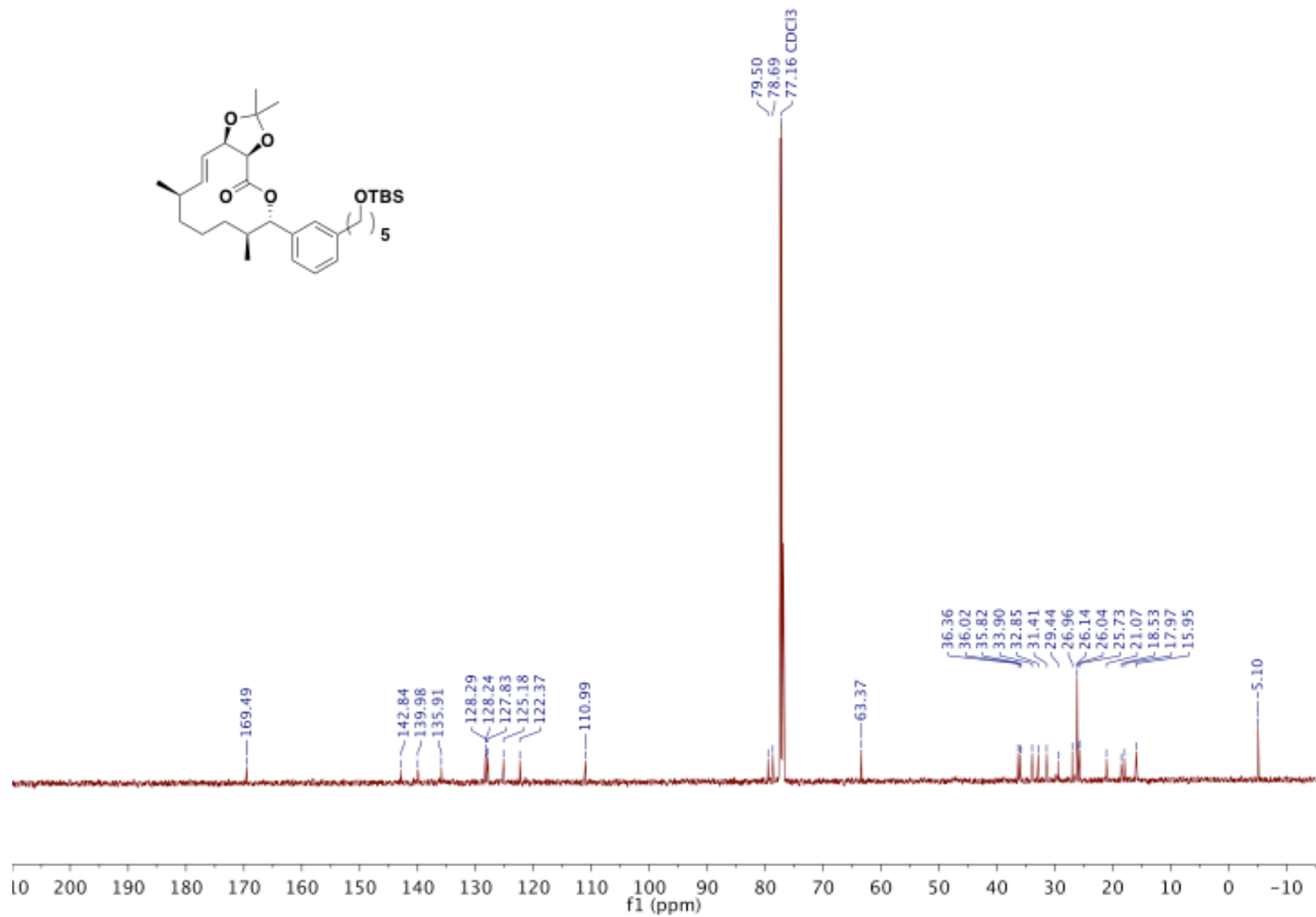
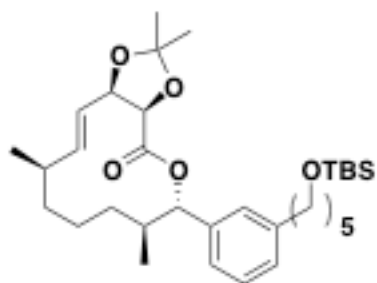


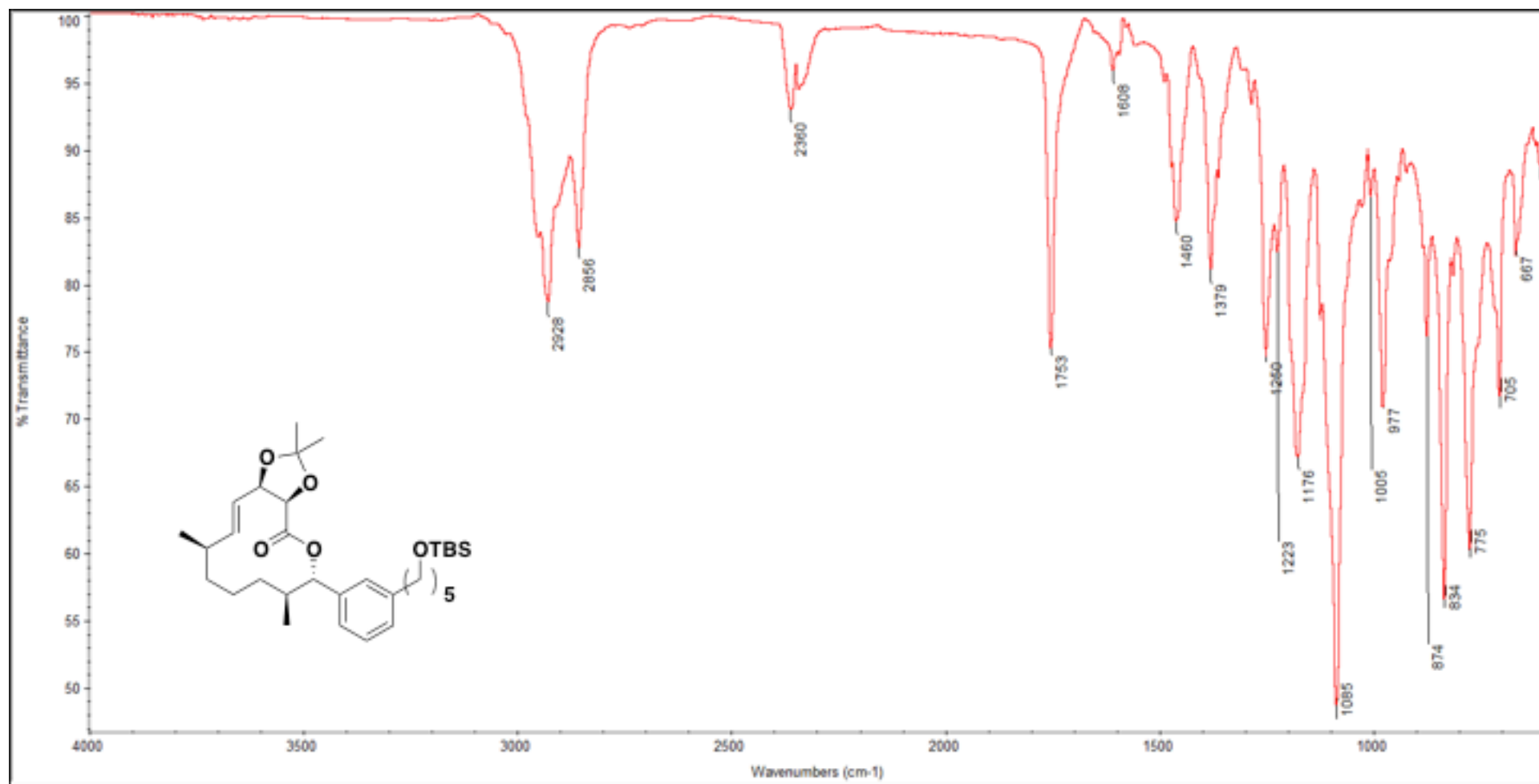


(-)-7b: ^1H NMR (500 MHz, CDCl_3)

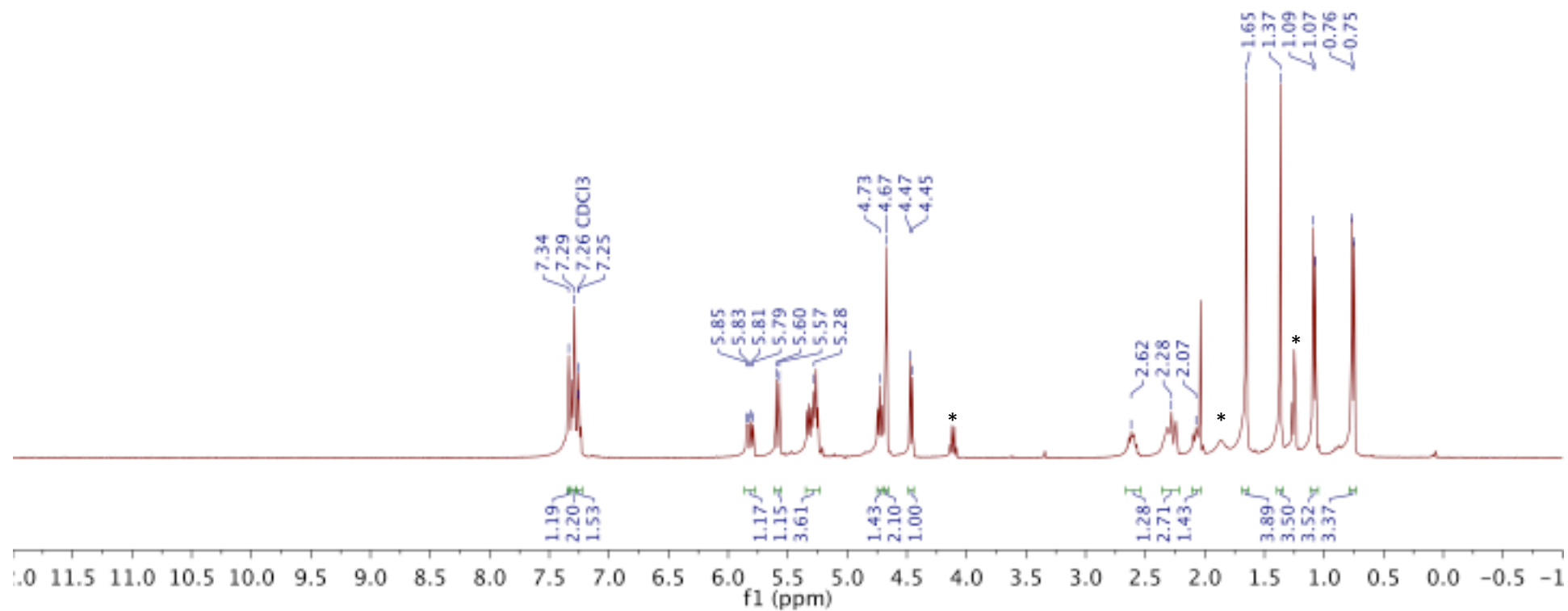
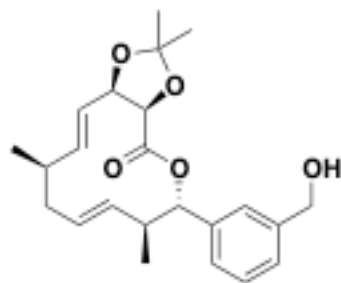


(-)-7b: ^{13}C NMR (126 MHz, CDCl_3)



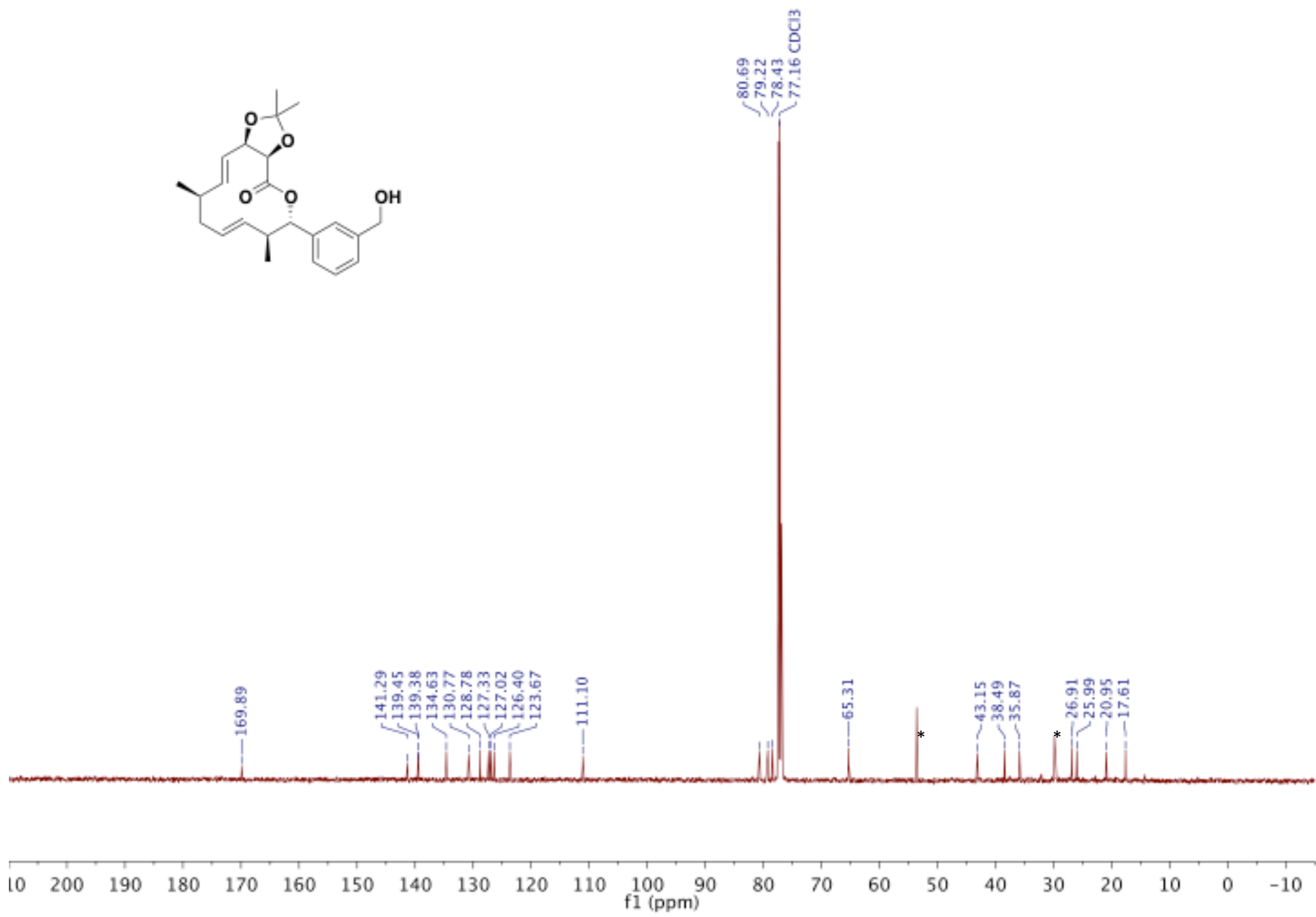
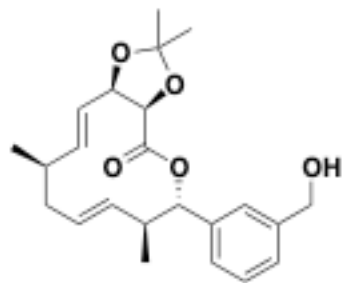


(-)-A1: ¹H NMR (500 MHz, CDCl₃)

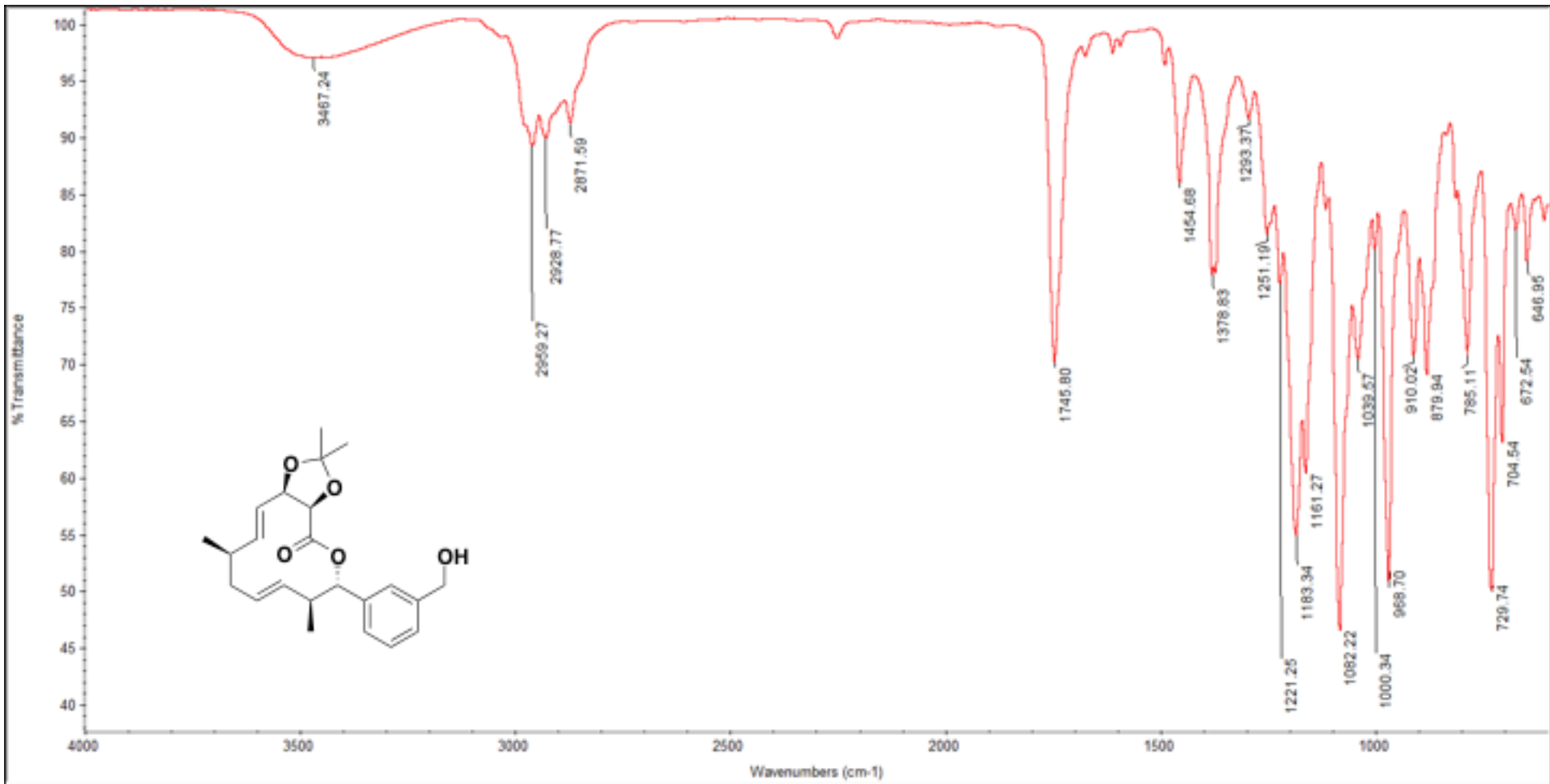


*Impurities present in spectrum

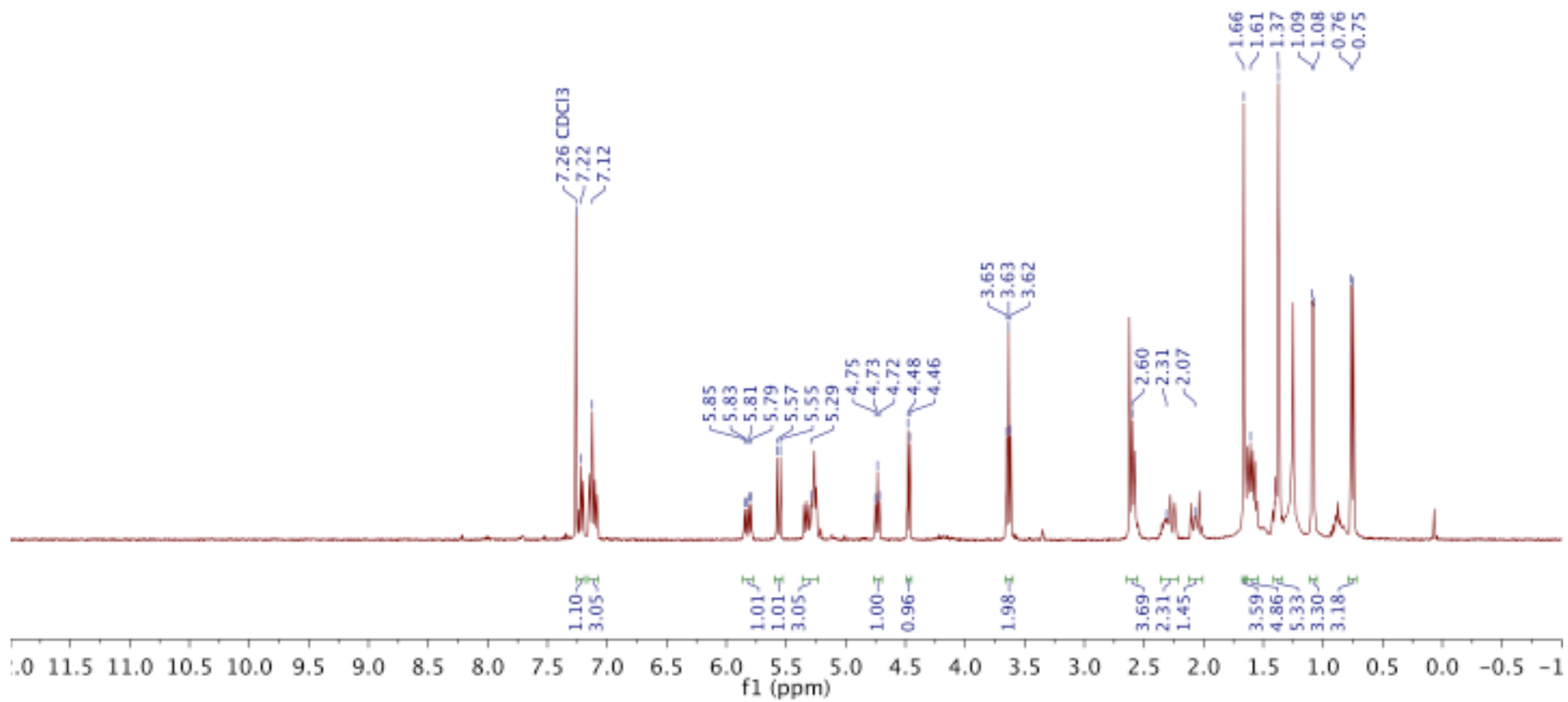
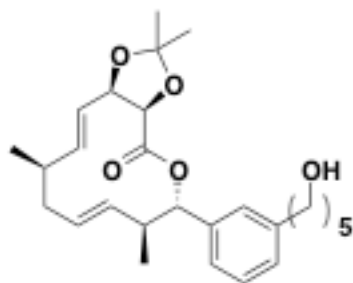
(-)-A1: ¹³C NMR (126 MHz, CDCl₃)



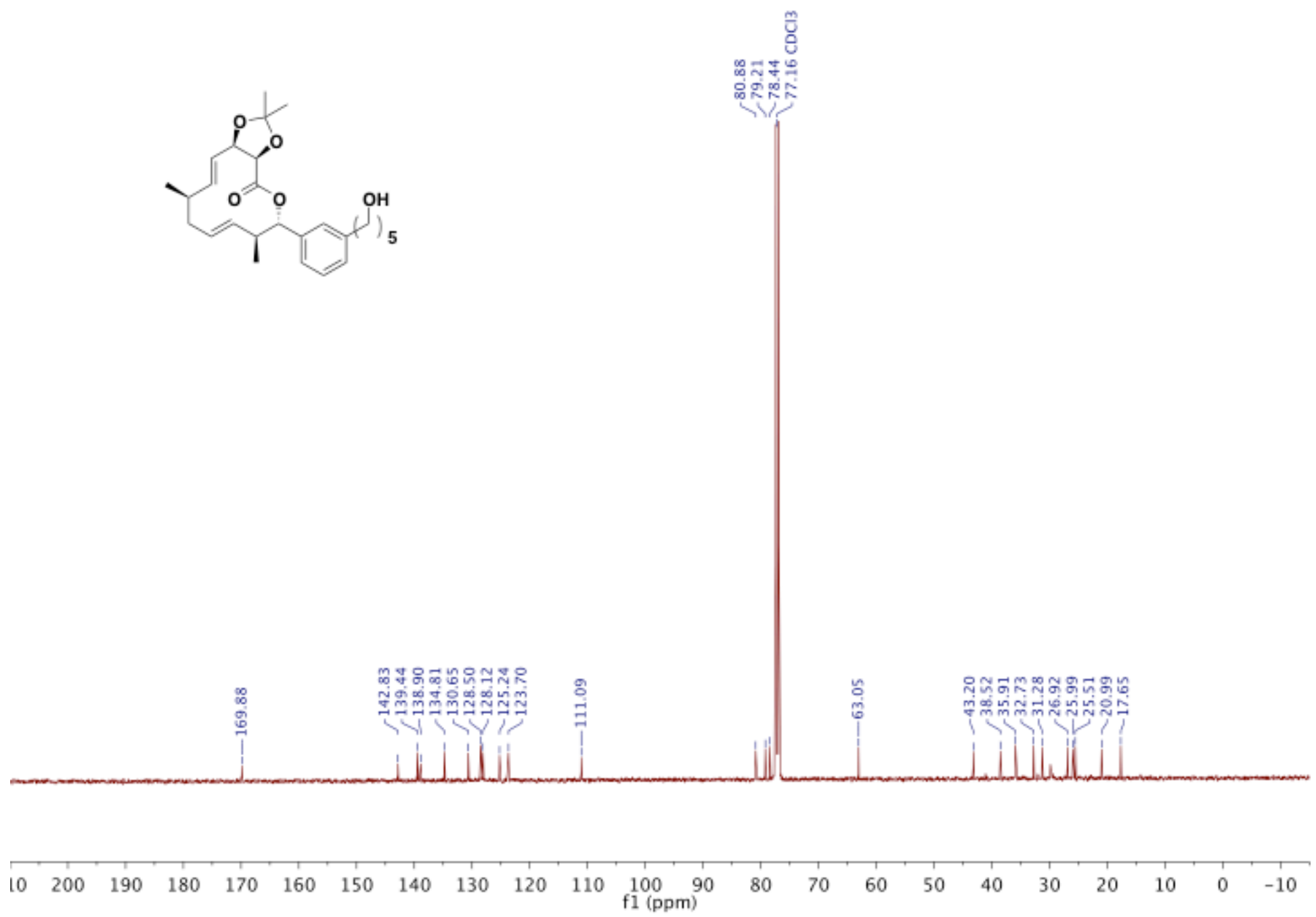
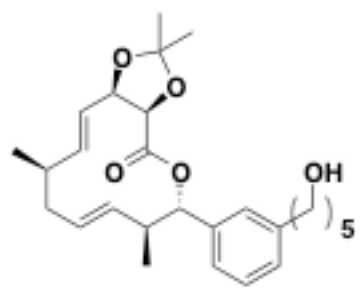
*Impurities present in spectrum

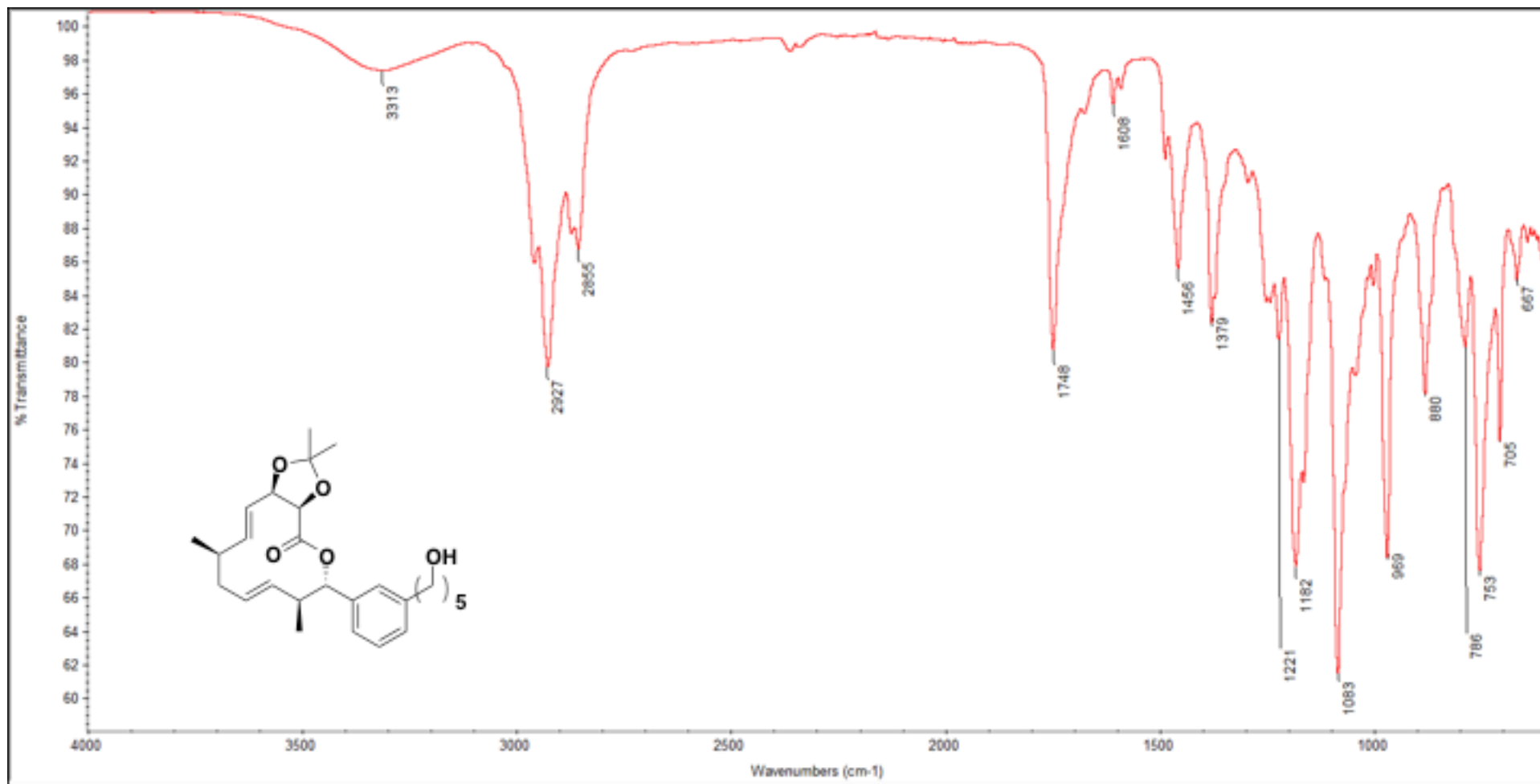


(-)-A3: ^1H NMR (500 MHz, CDCl_3)

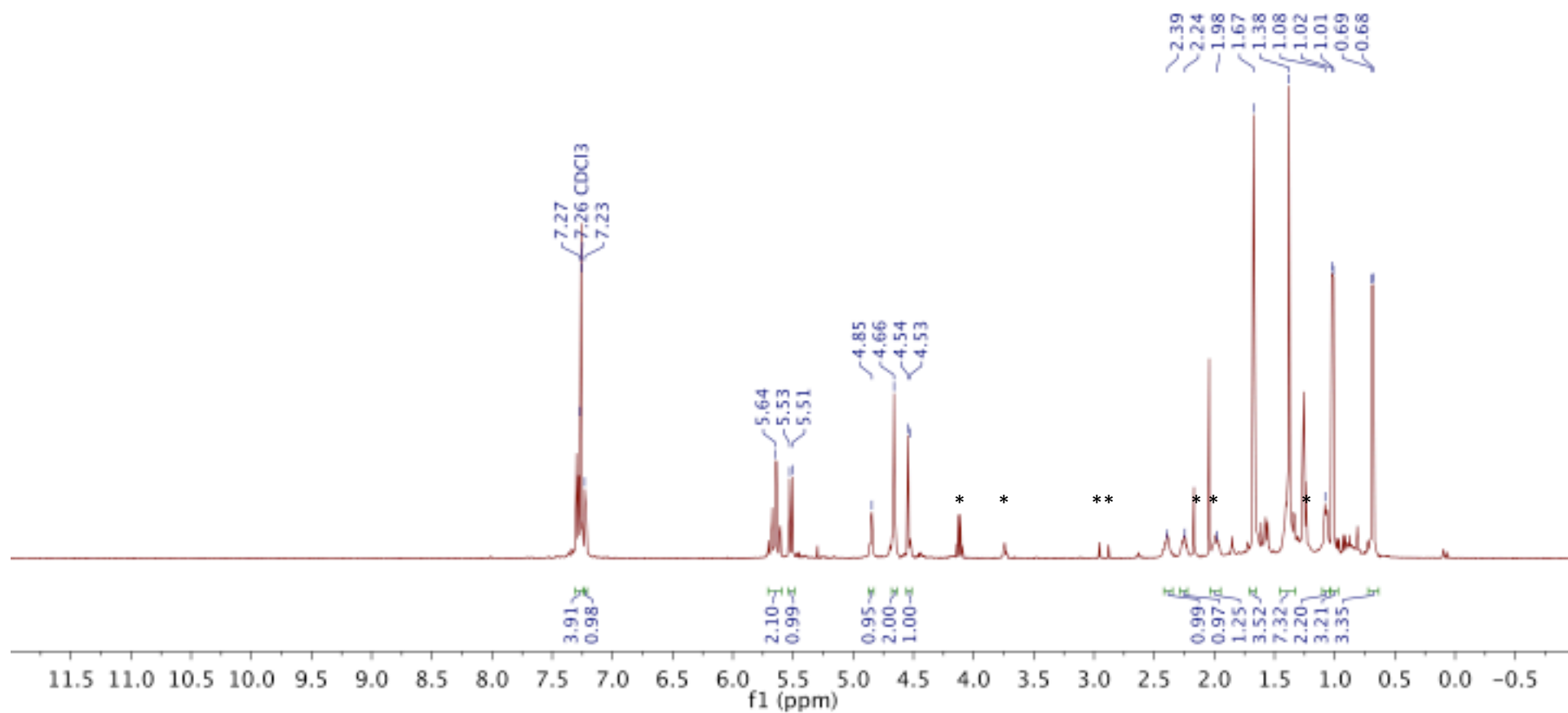
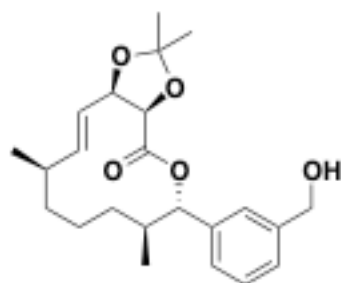


(-)-A3: ^{13}C NMR (126 MHz, CDCl_3)



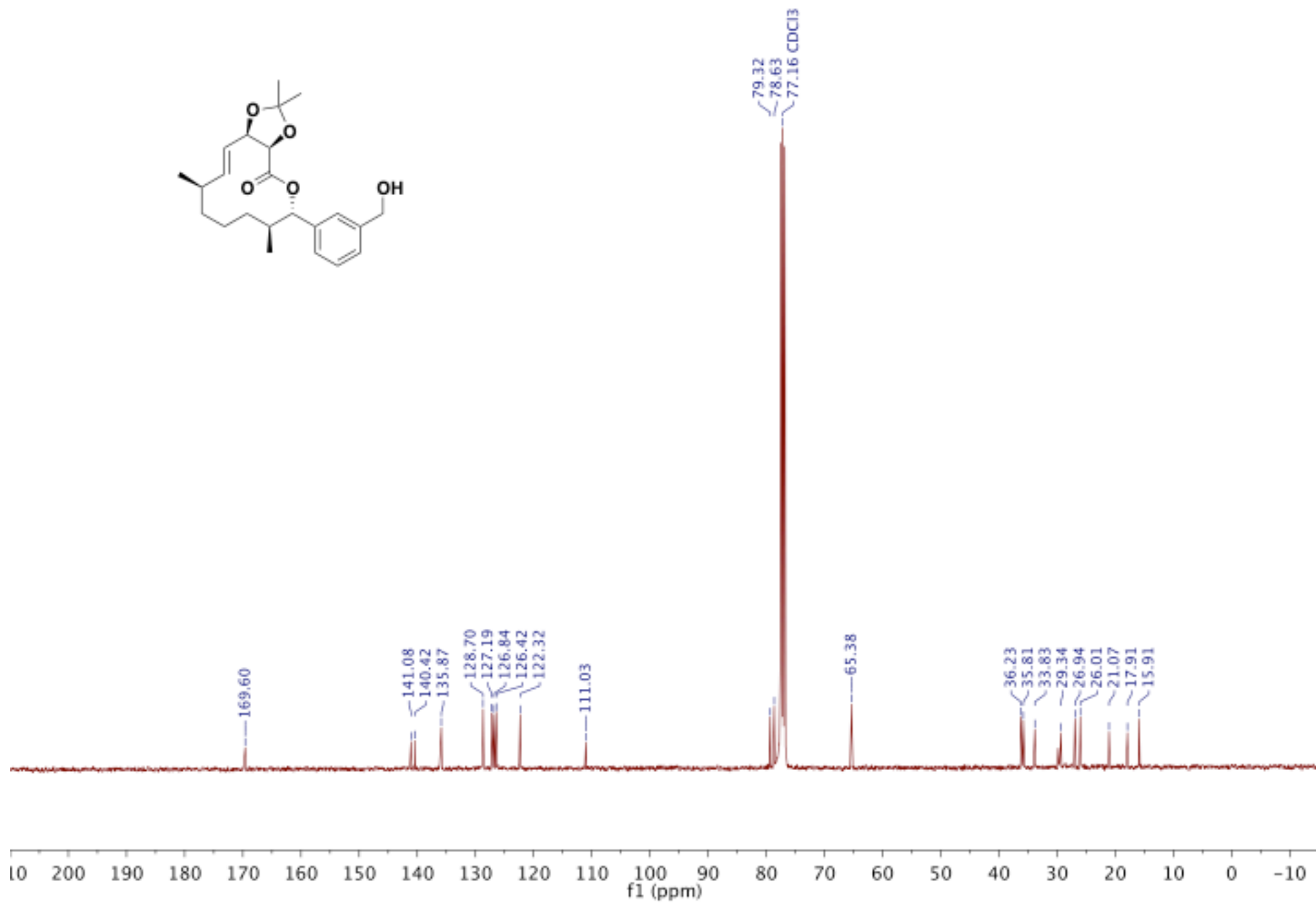
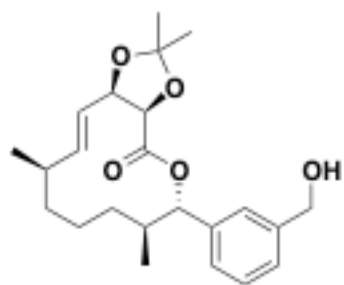


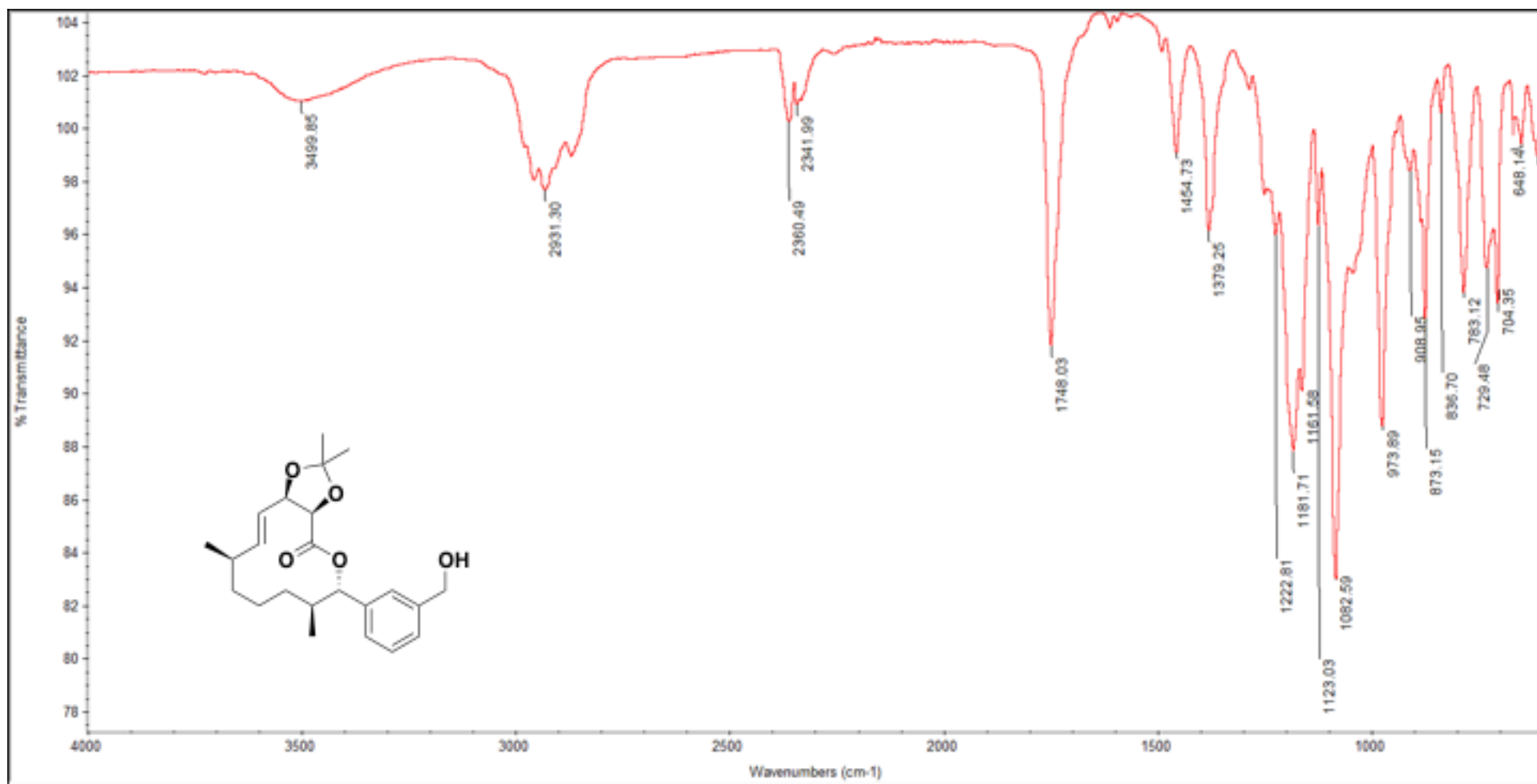
(-)-A2: ^1H NMR (500 MHz, CDCl_3)



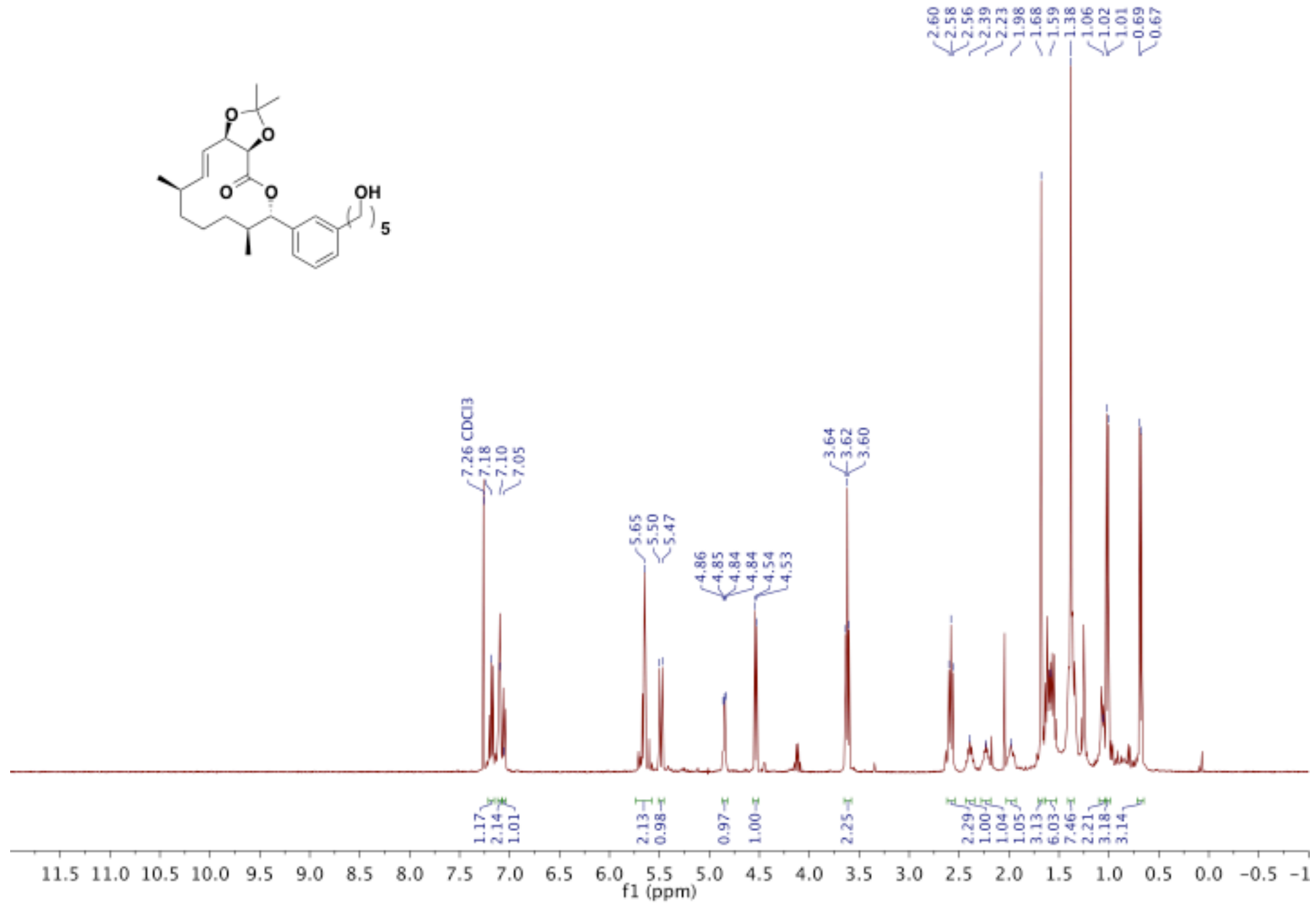
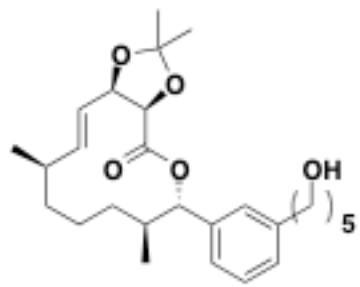
*Impurities present in spectrum

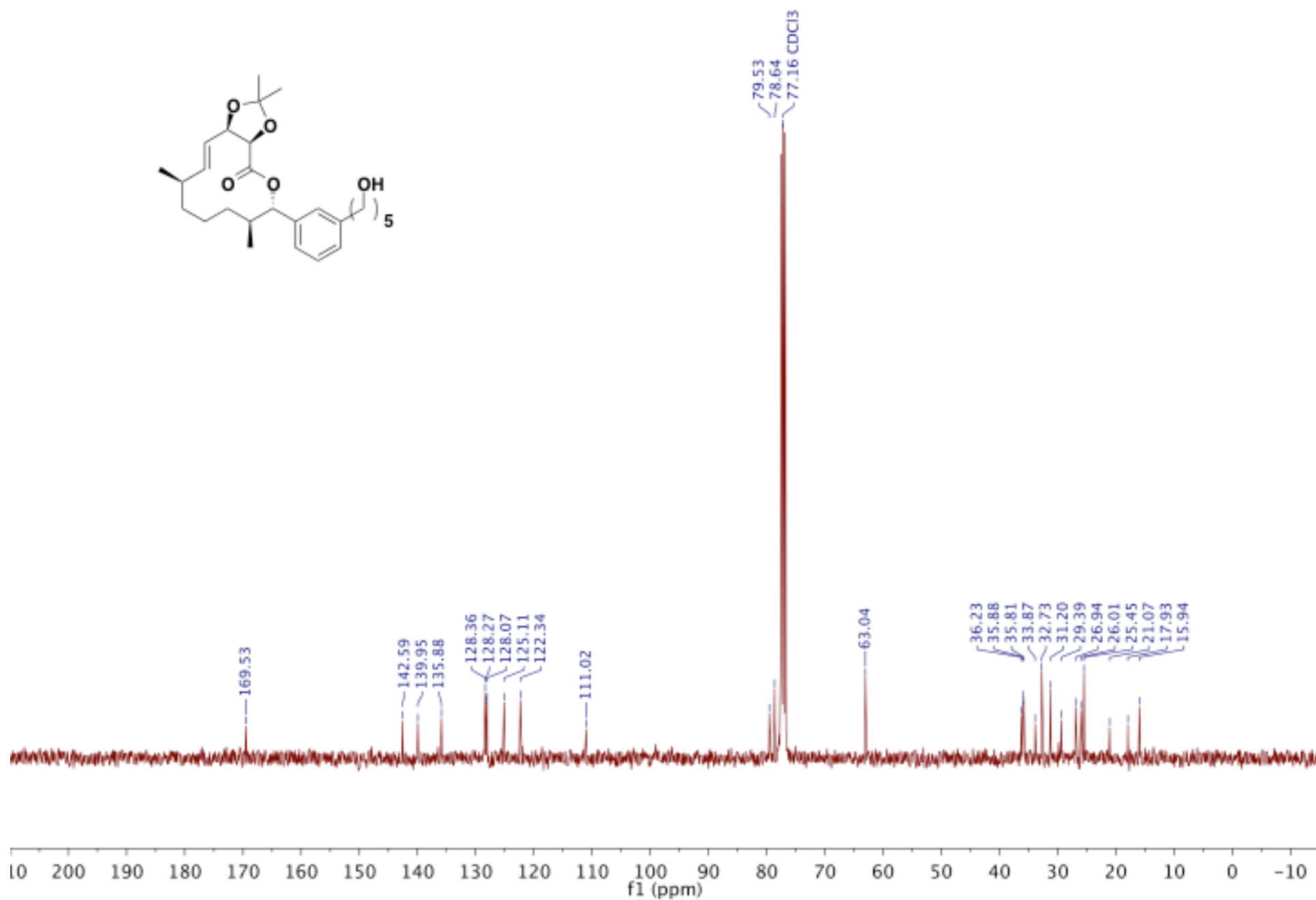
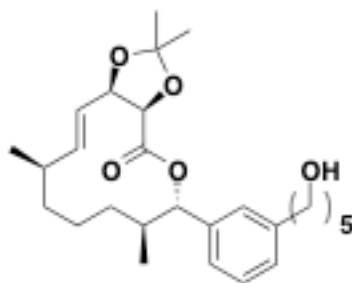
(-)-A2: ^{13}C NMR (126 MHz, CDCl_3)



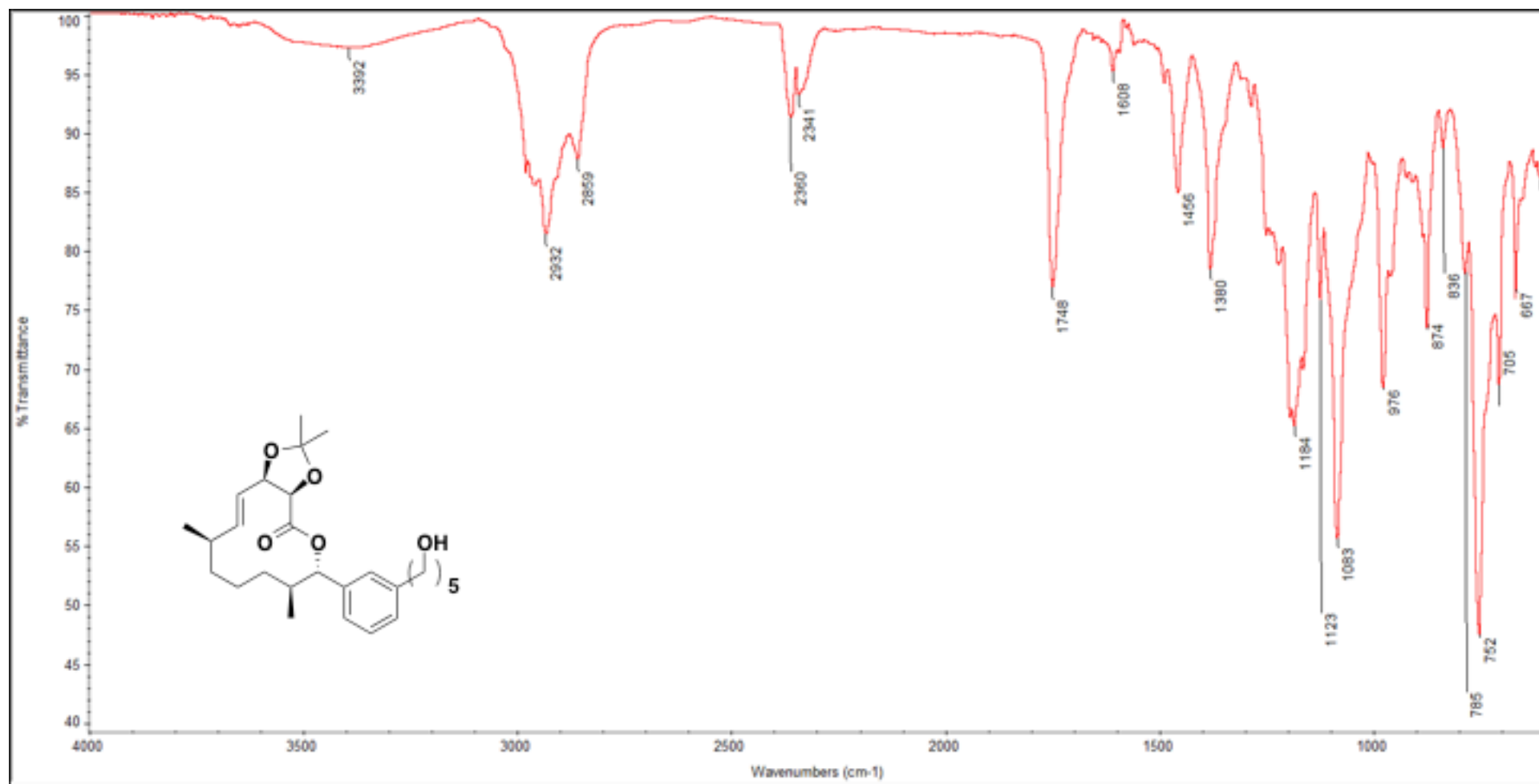


(-)-A4: ¹H NMR (500 MHz, CDCl₃)

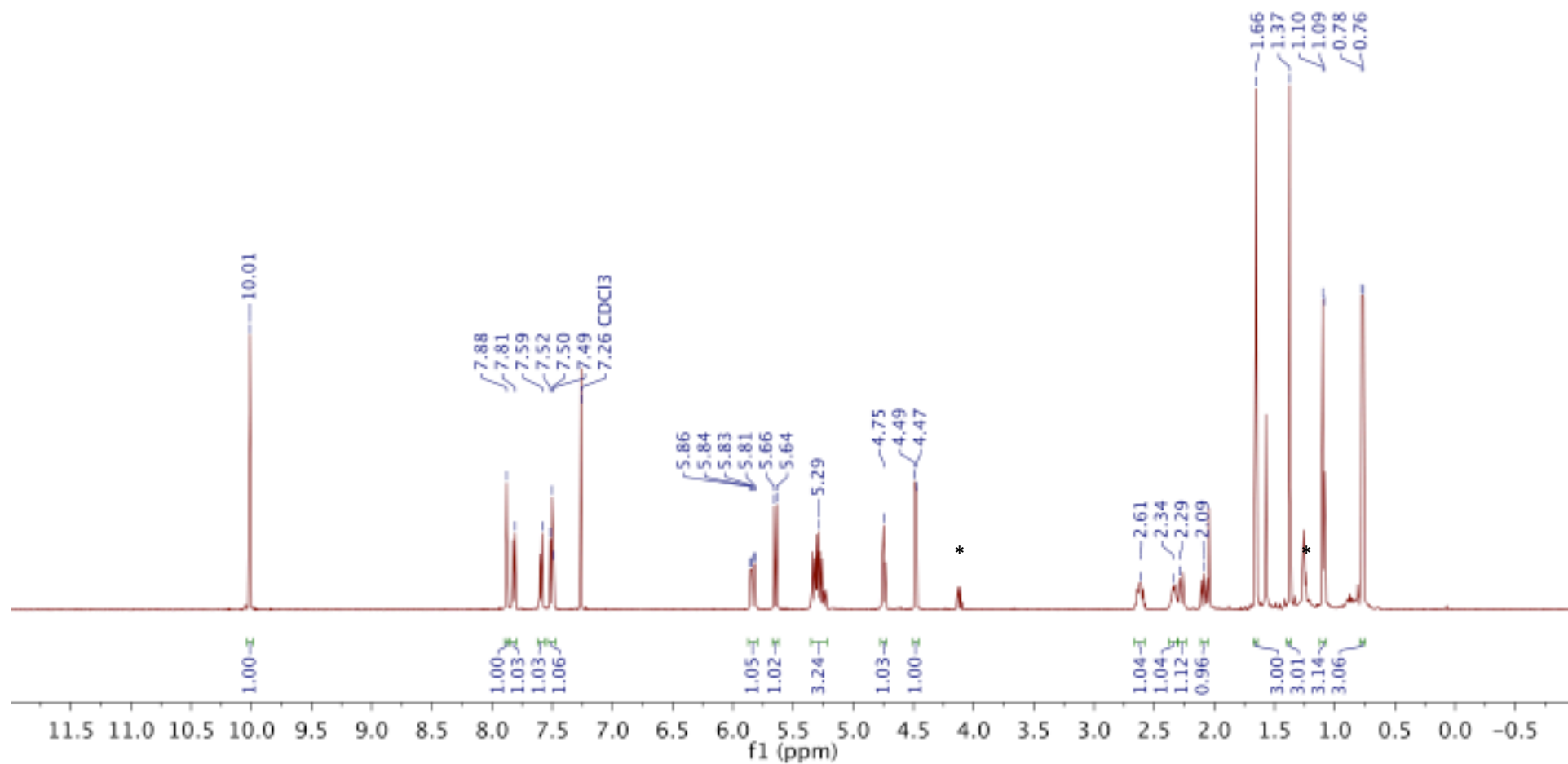
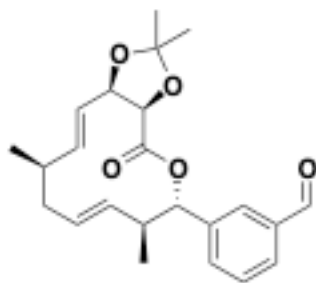


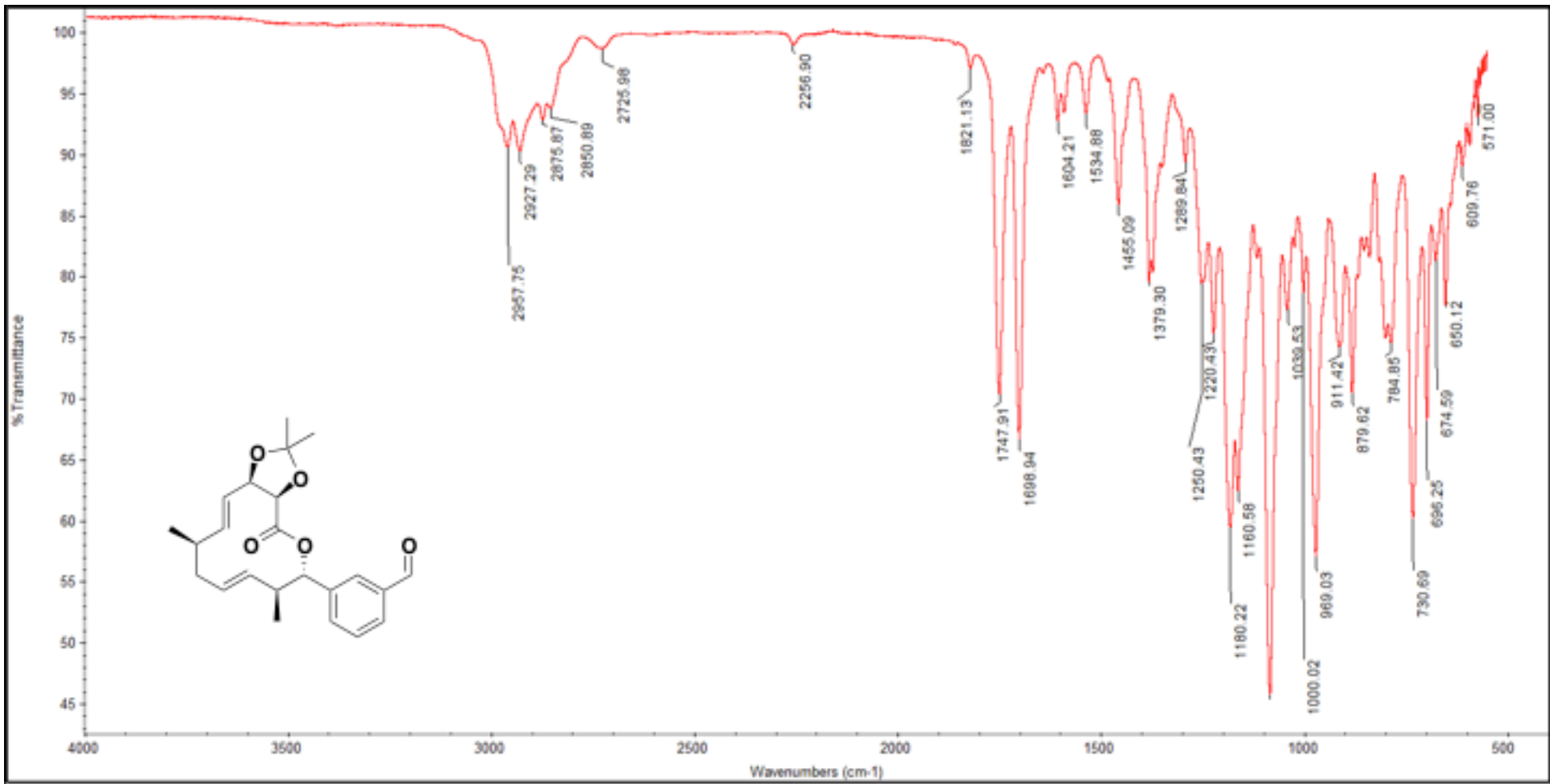


(-)-A4: IR

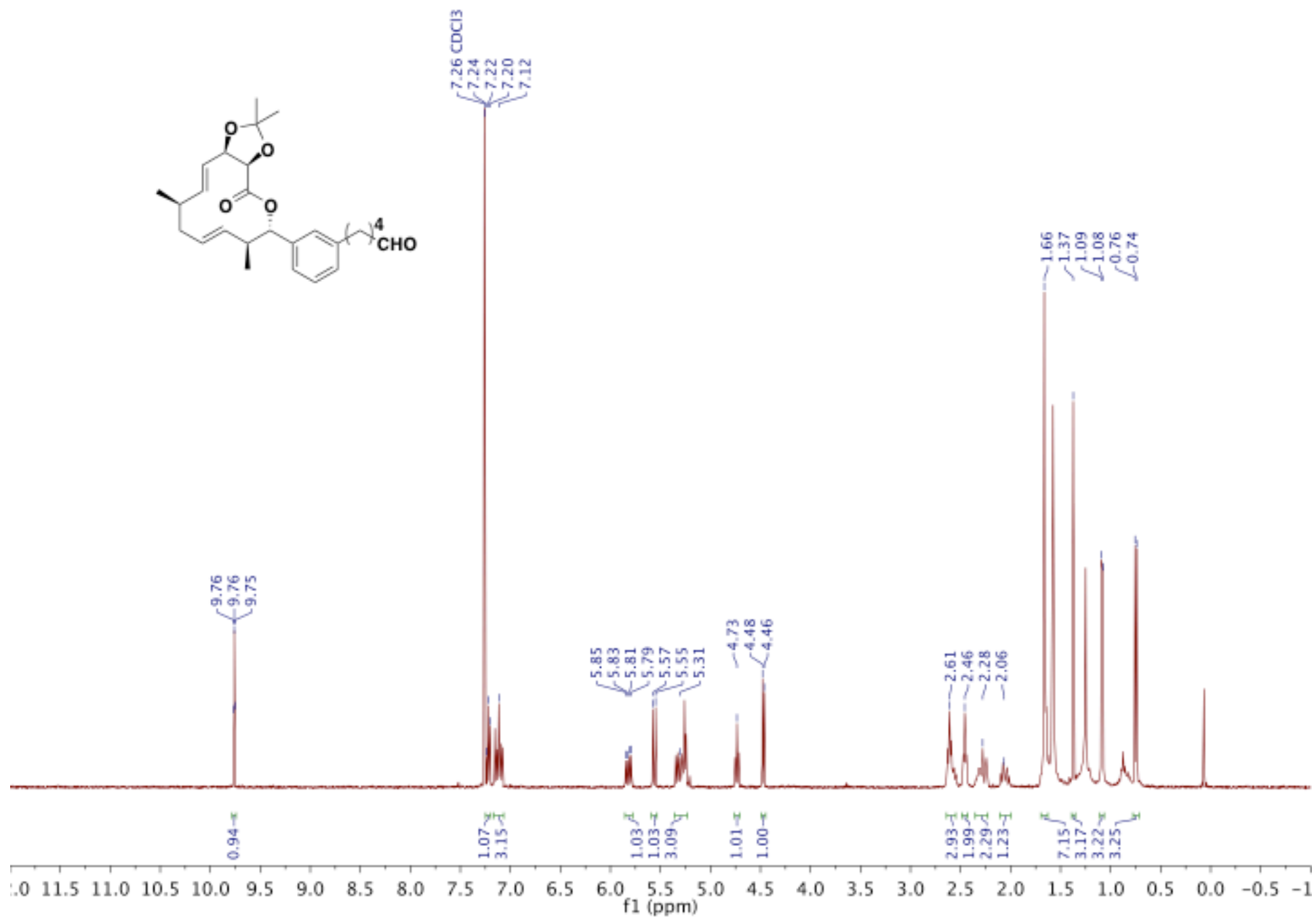


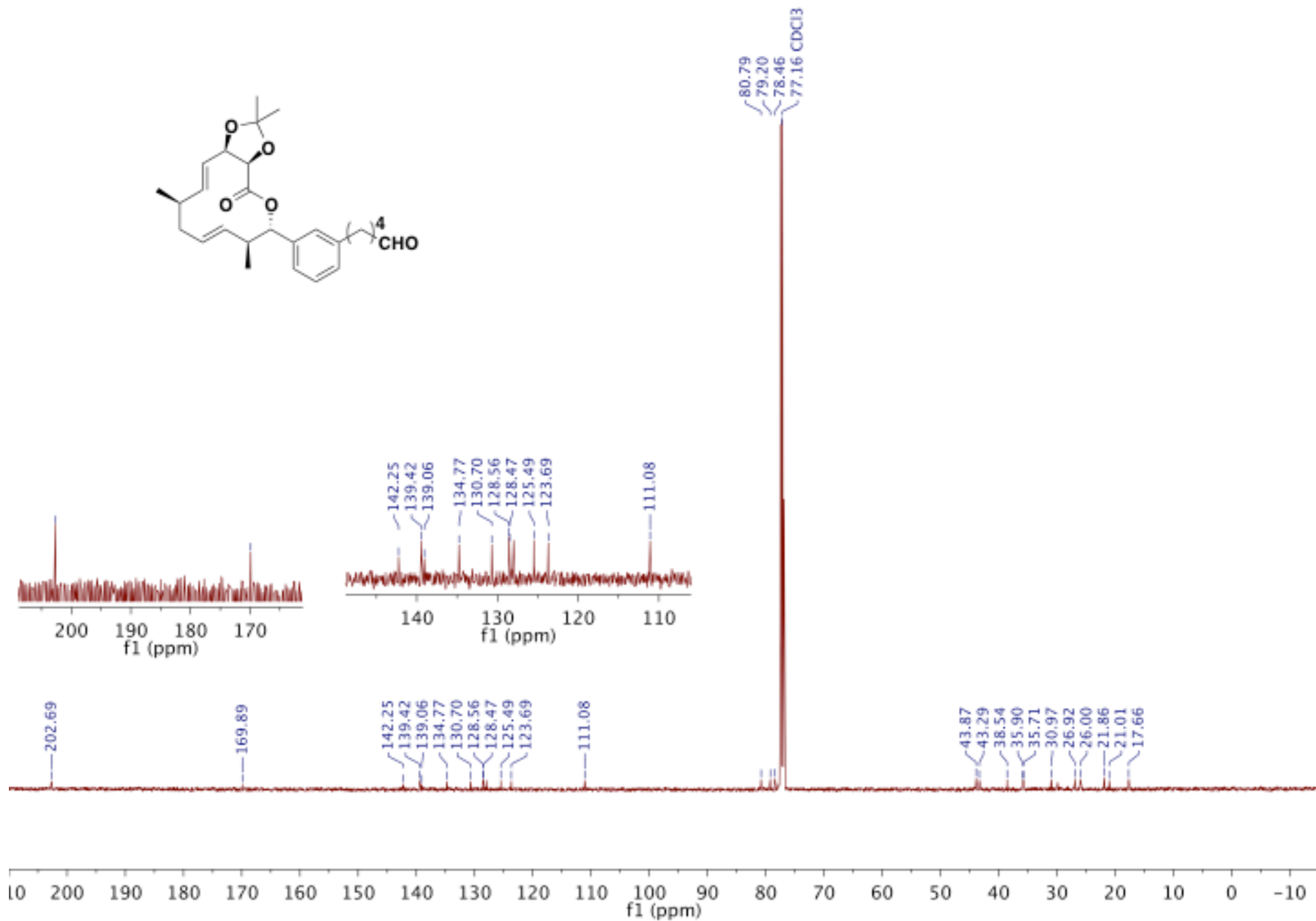
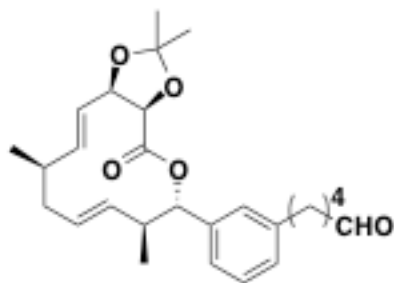
(-)-10a: ¹H NMR (500 MHz, CDCl₃)

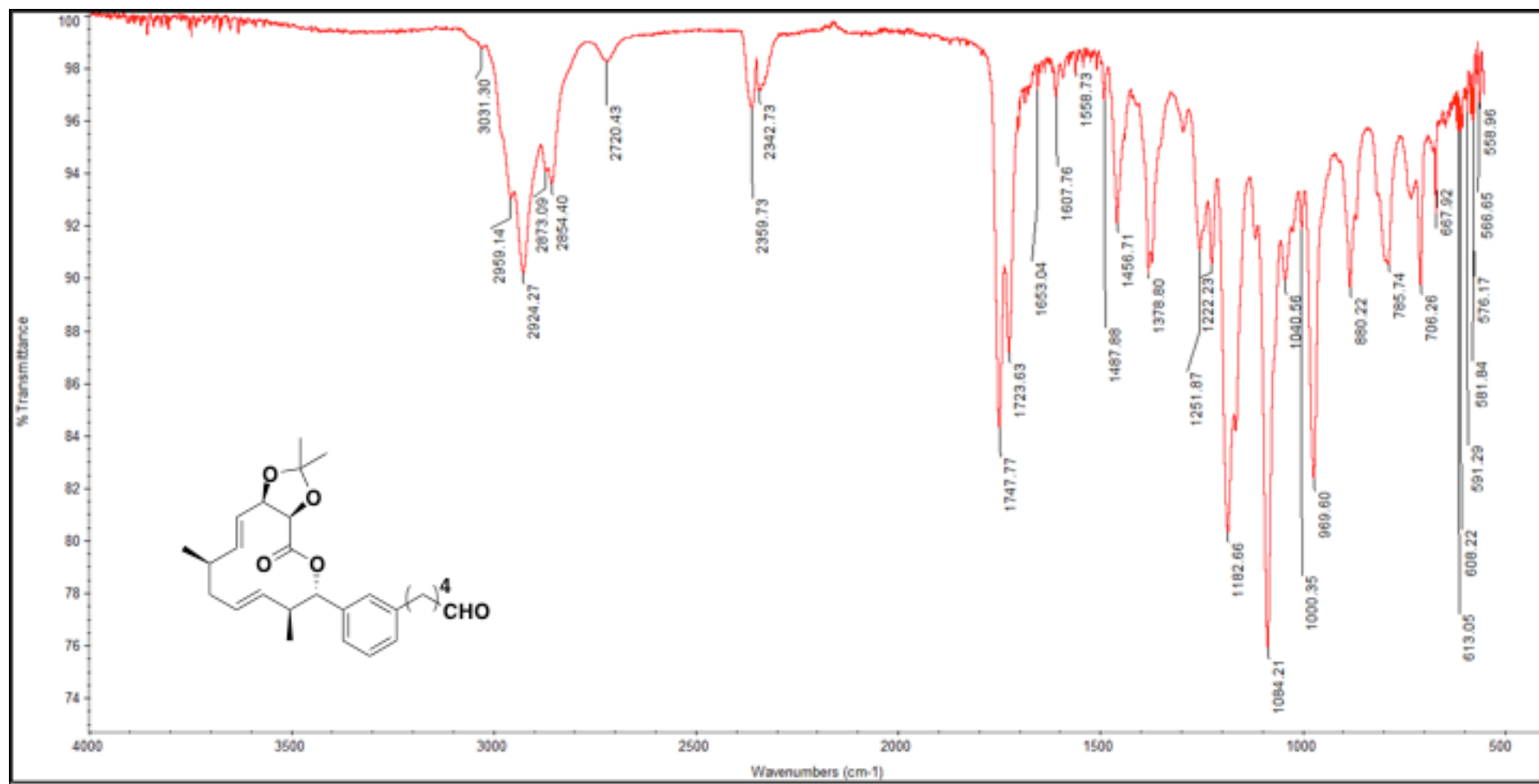




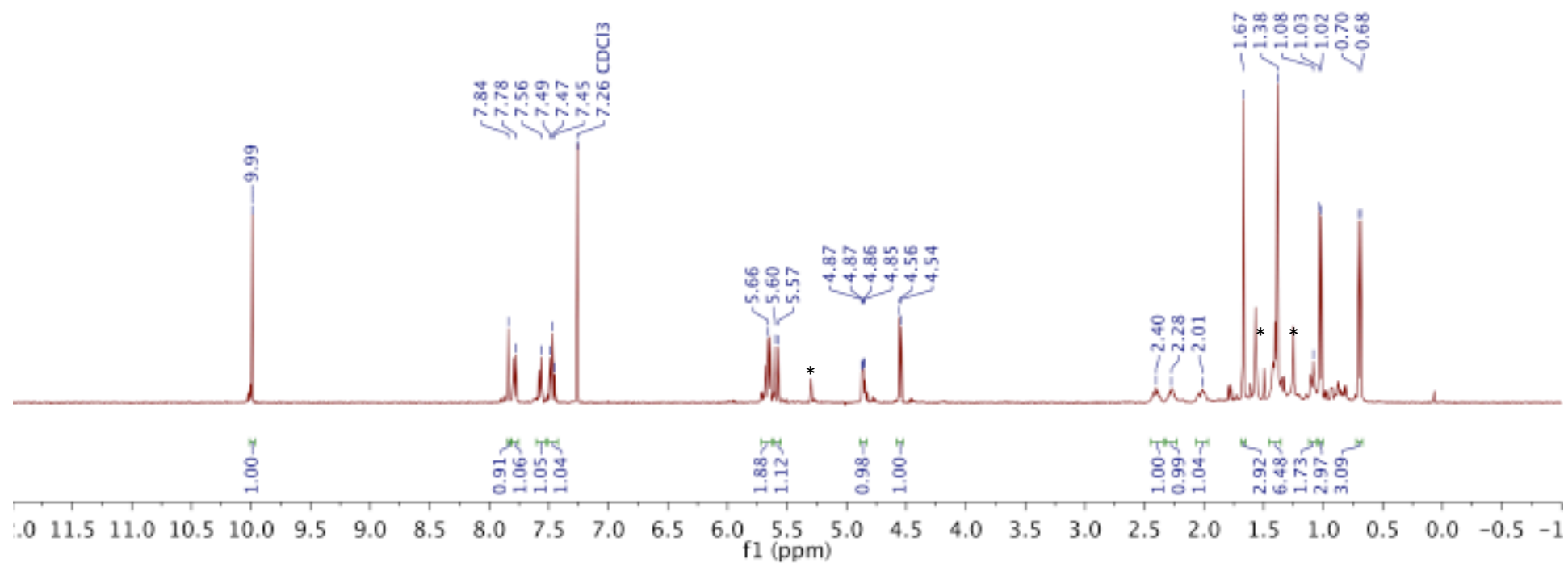
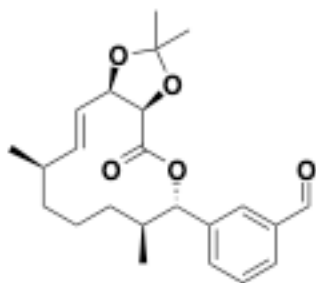
(-)-11a: ¹H NMR (500 MHz, CDCl₃)







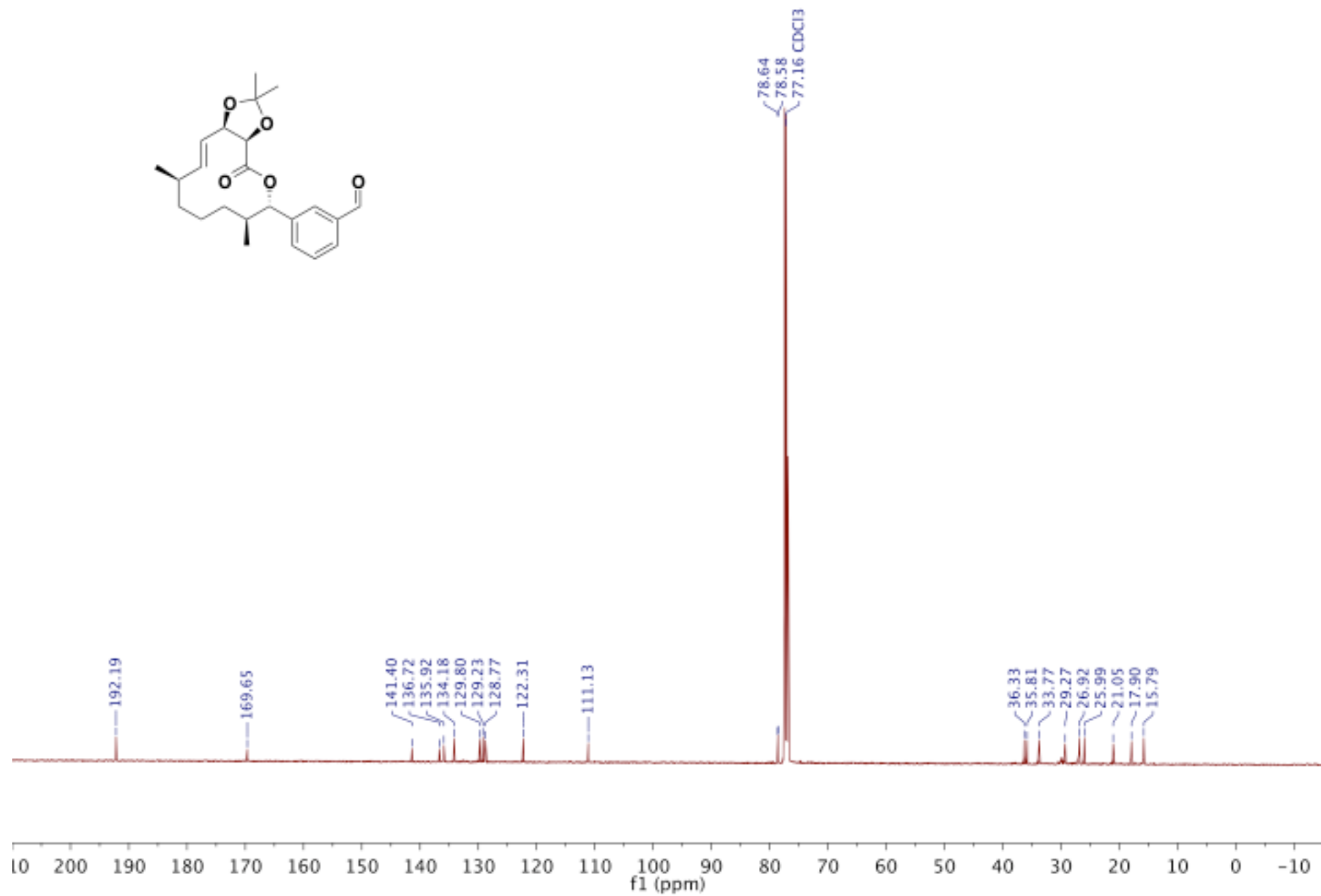
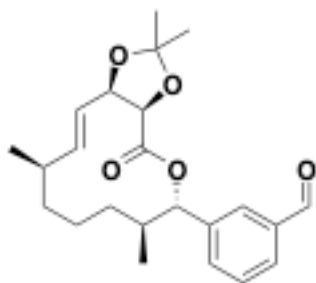
(-)-10b: ¹H NMR (500 MHz, CDCl₃)

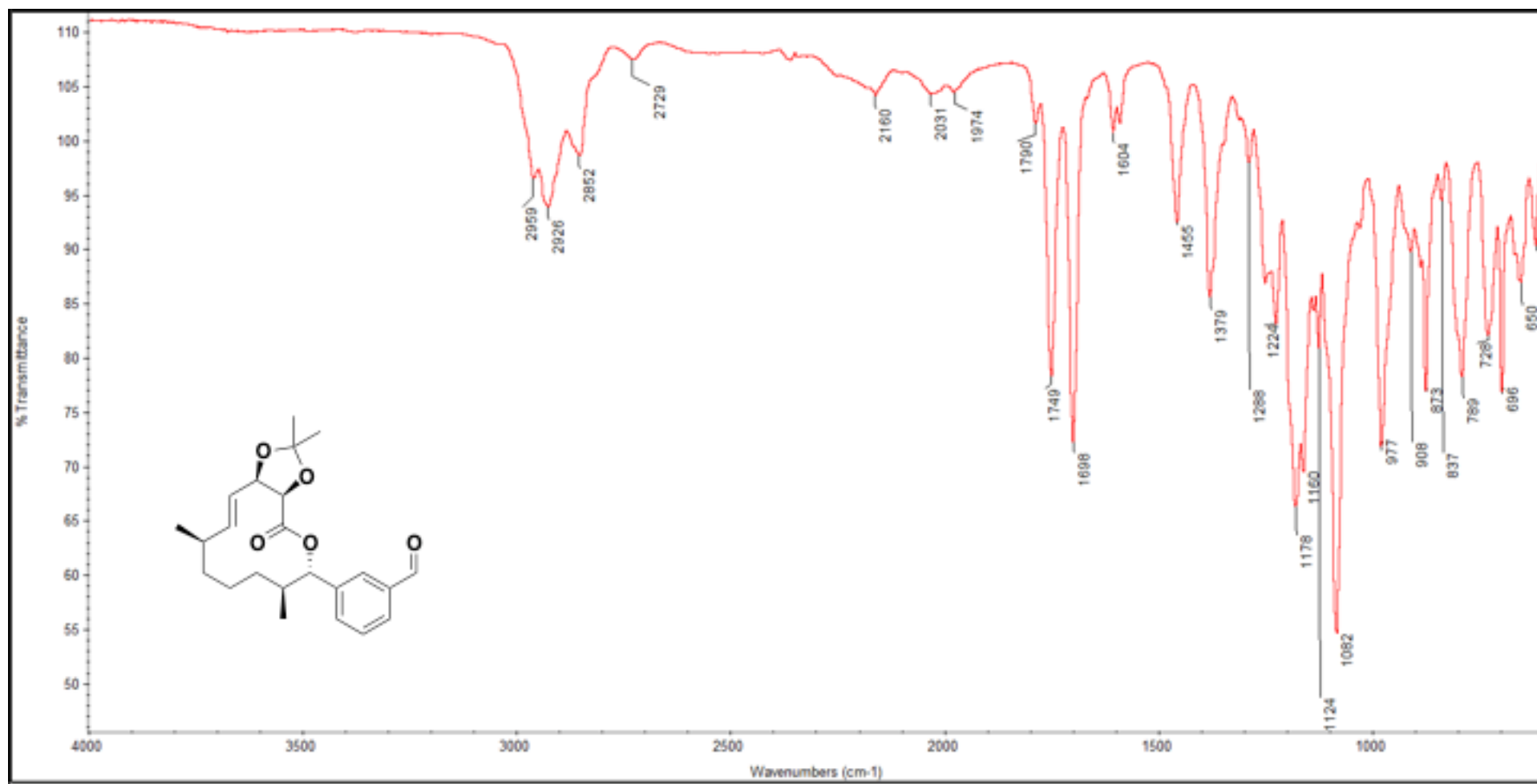


S92

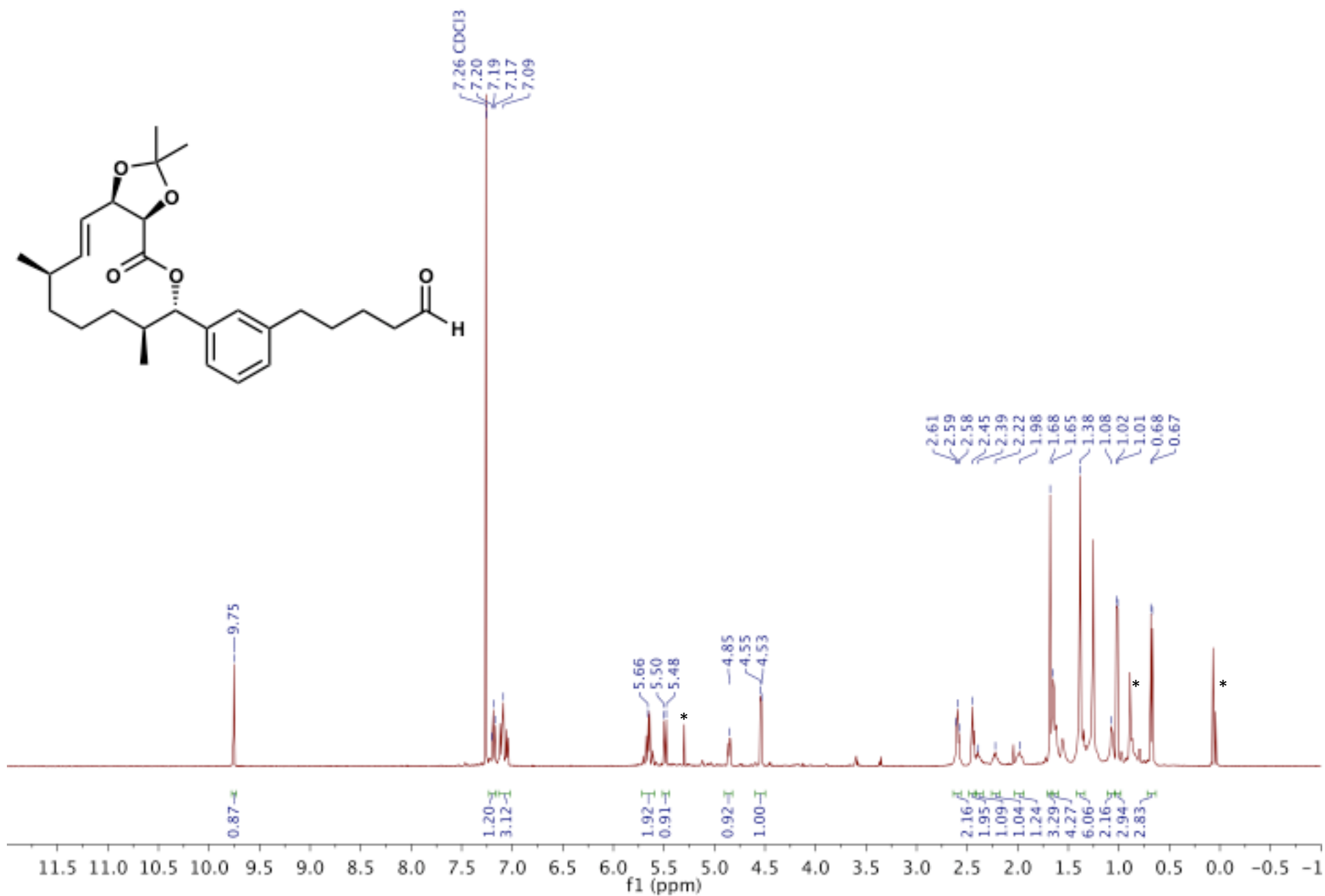
*Impurities present in spectrum

(-)-10b: ^{13}C NMR (126 MHz, CDCl_3)



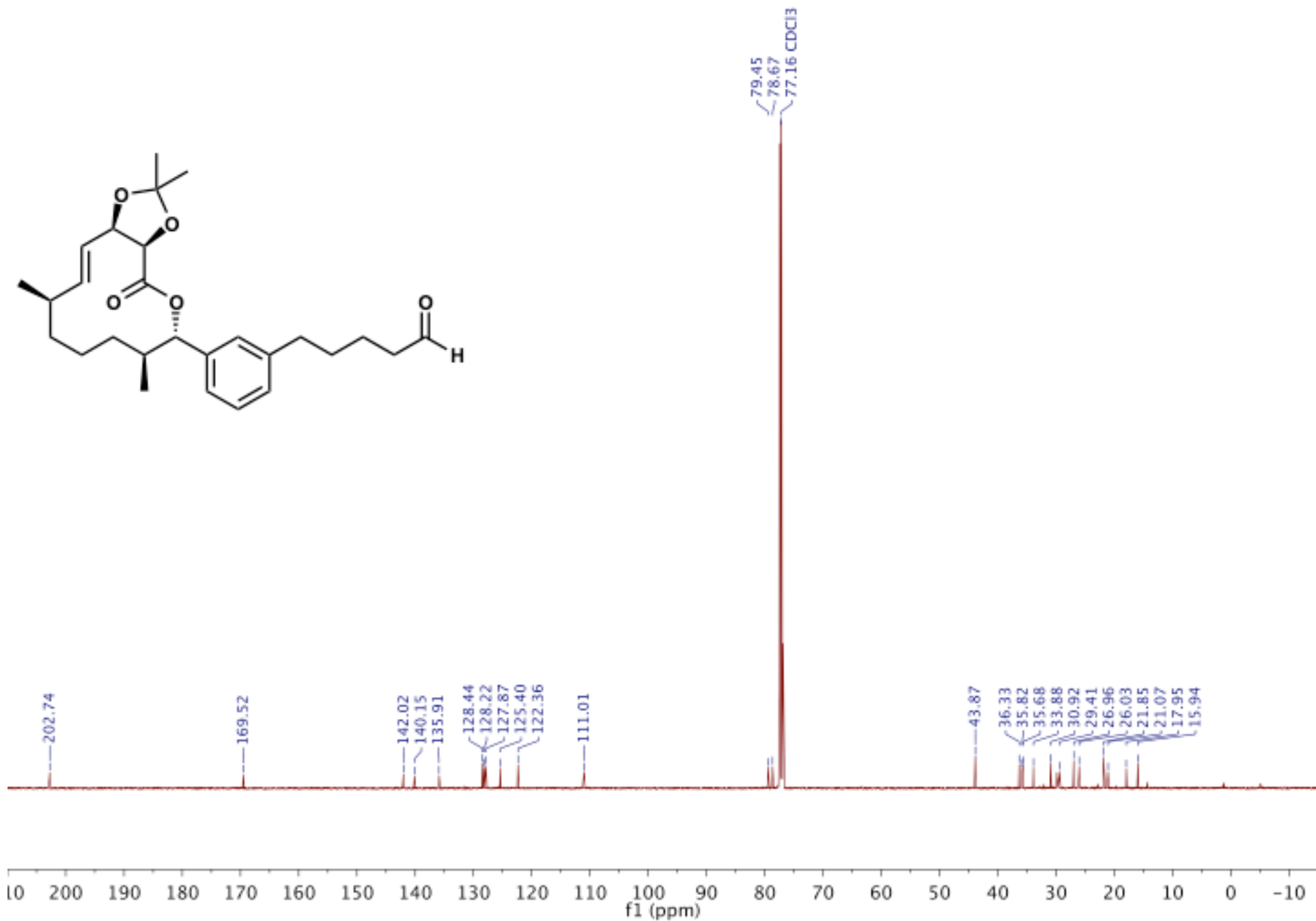


(-)-11b: ¹H NMR (500 MHz, CDCl₃)

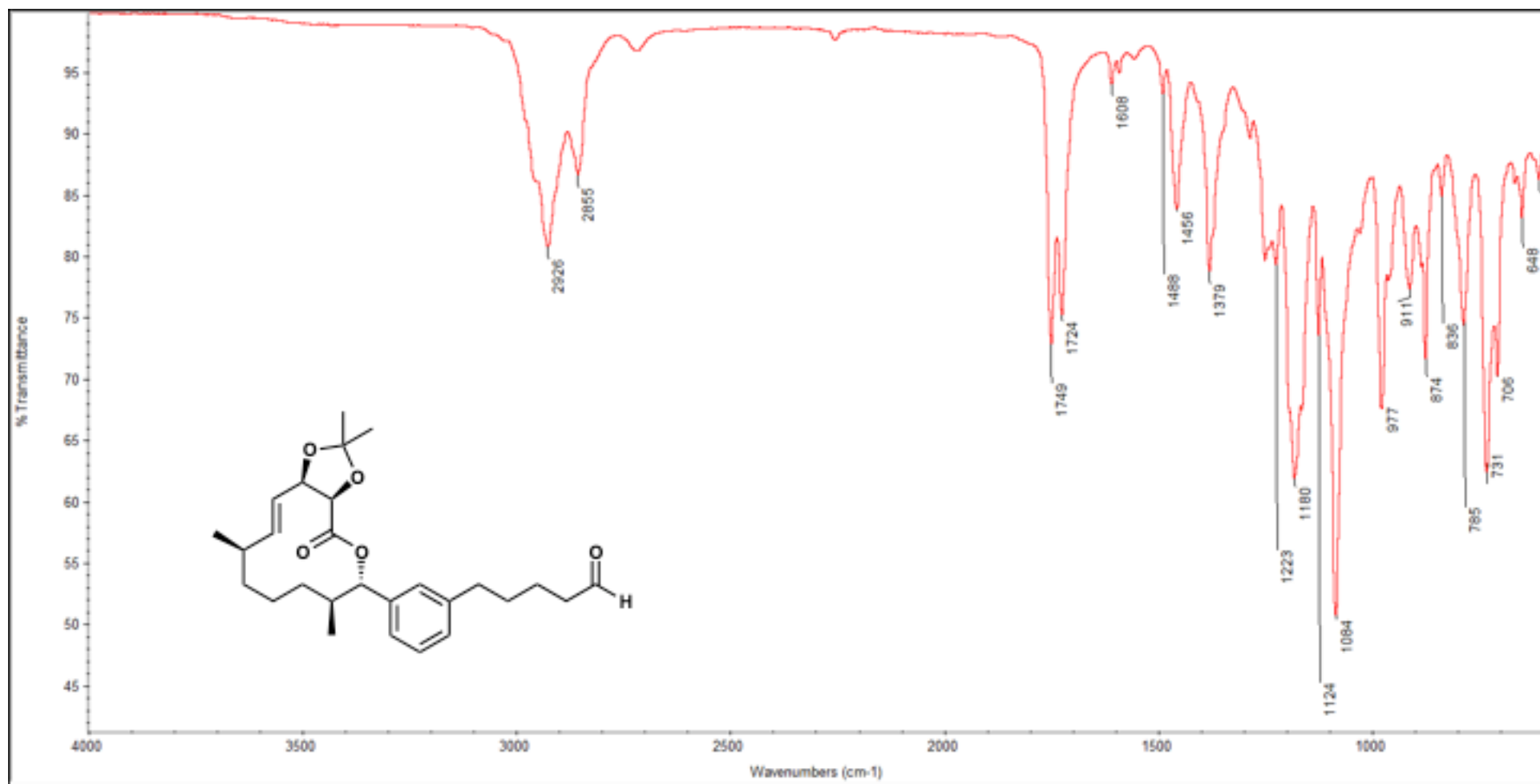


*Impurities present in spectrum

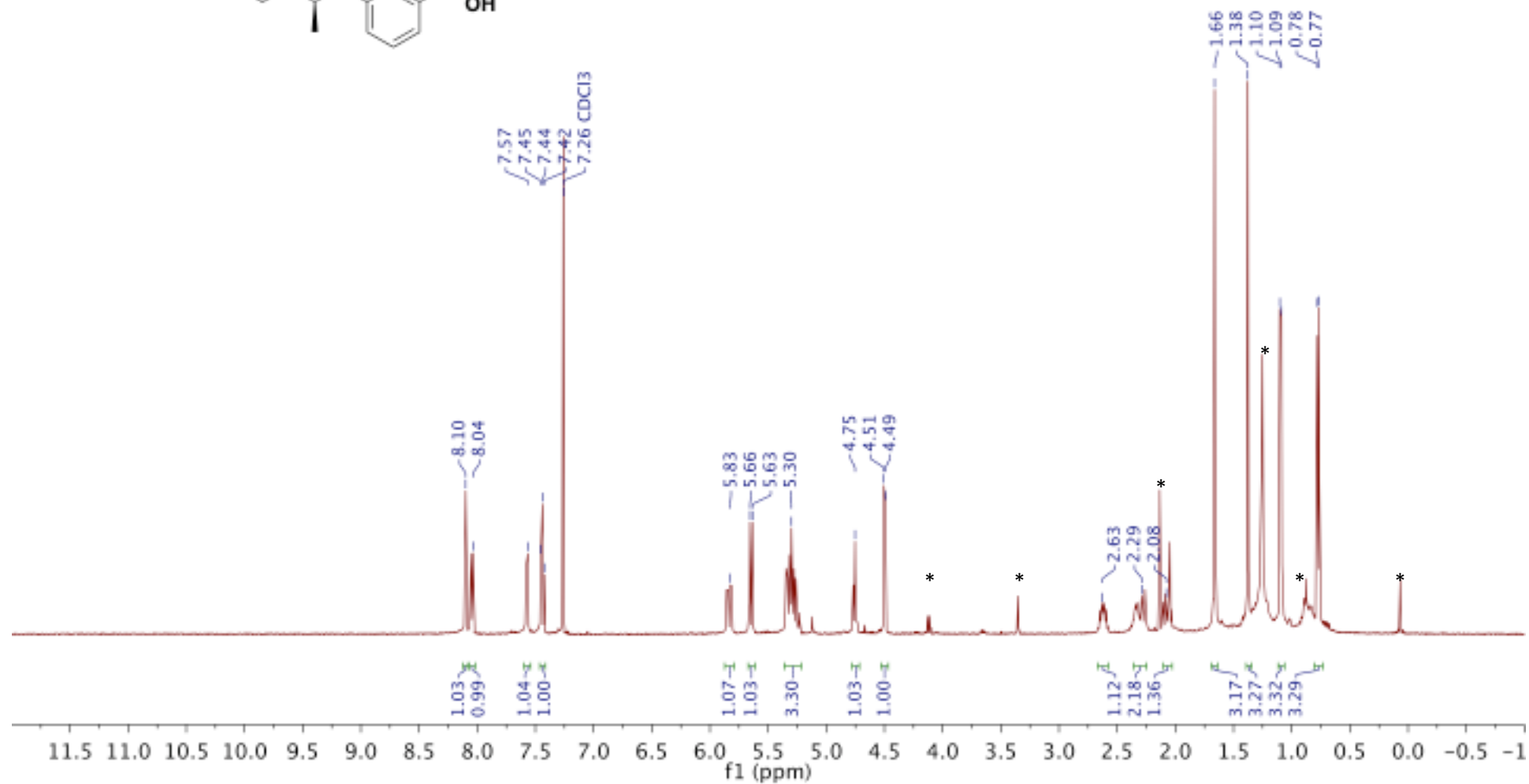
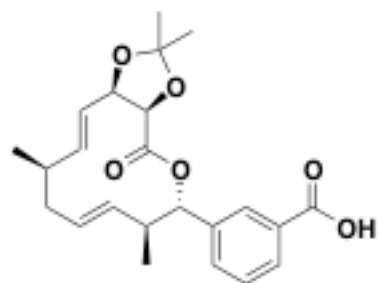
(-)-11b: ^{13}C NMR (126 MHz, CDCl_3)



(-)-11b: IR



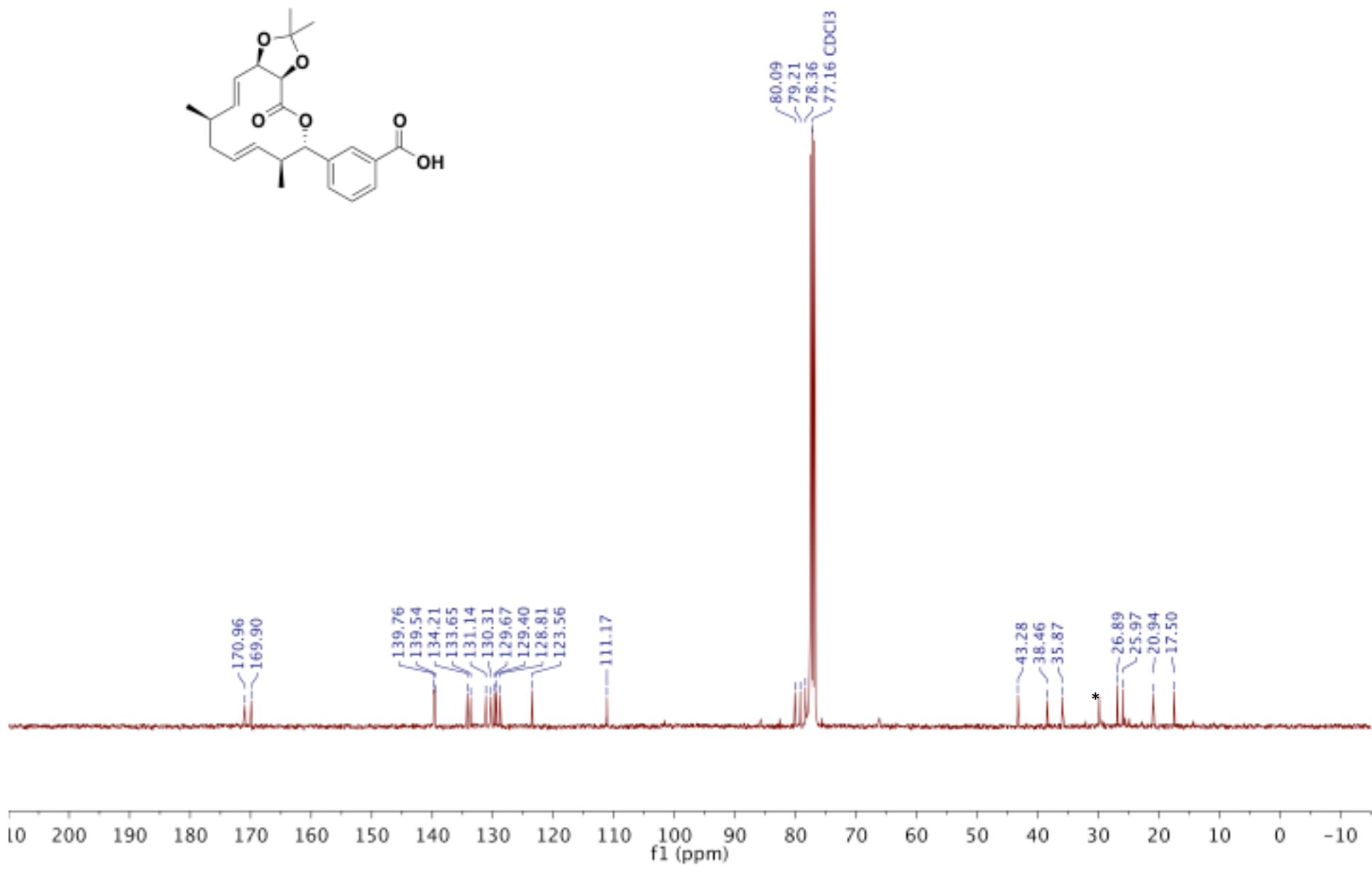
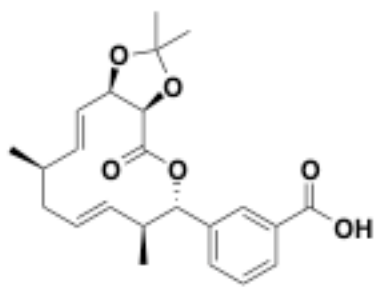
(-)-C1: ^1H NMR (500 MHz, CDCl_3)



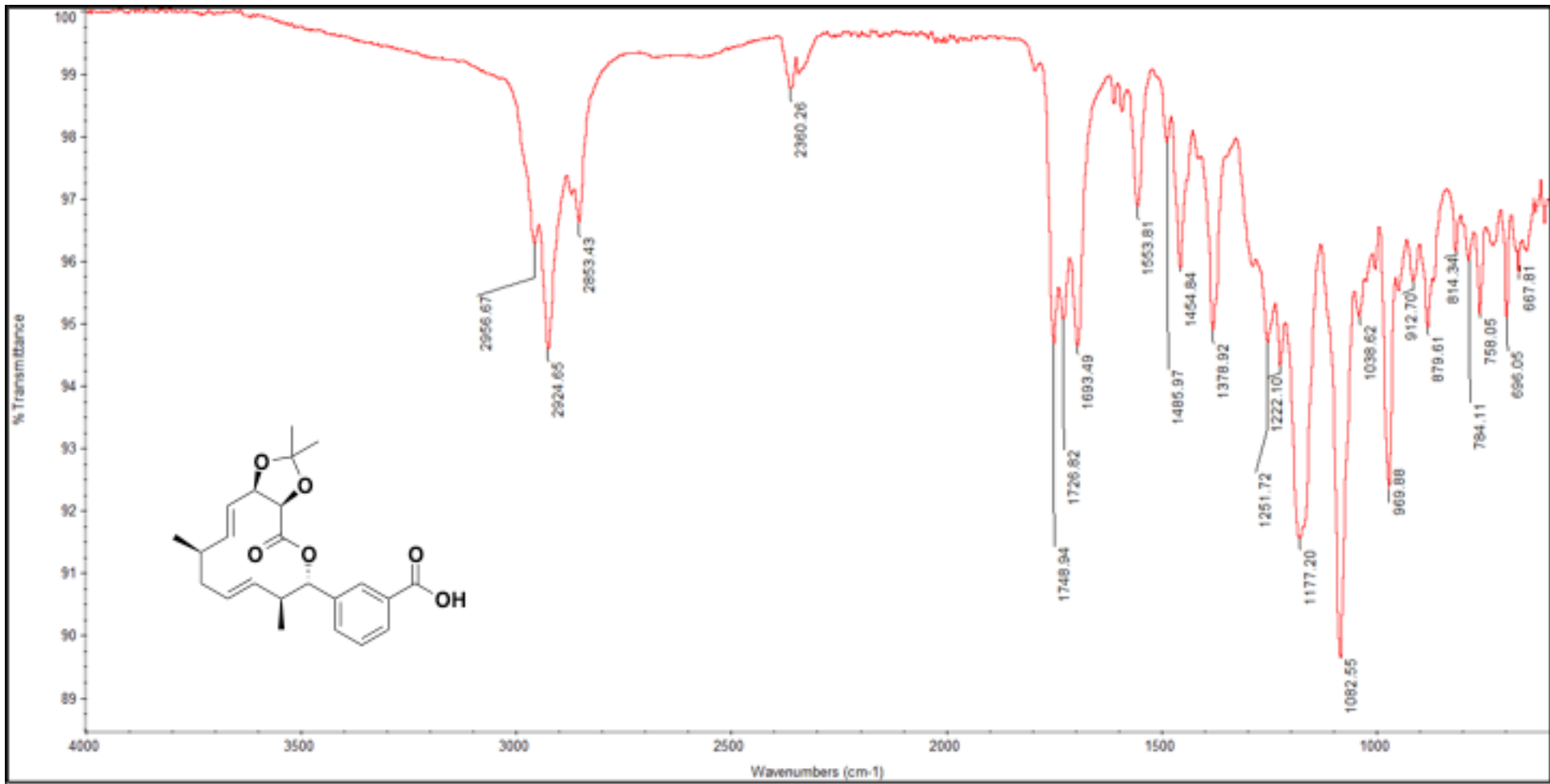
S98

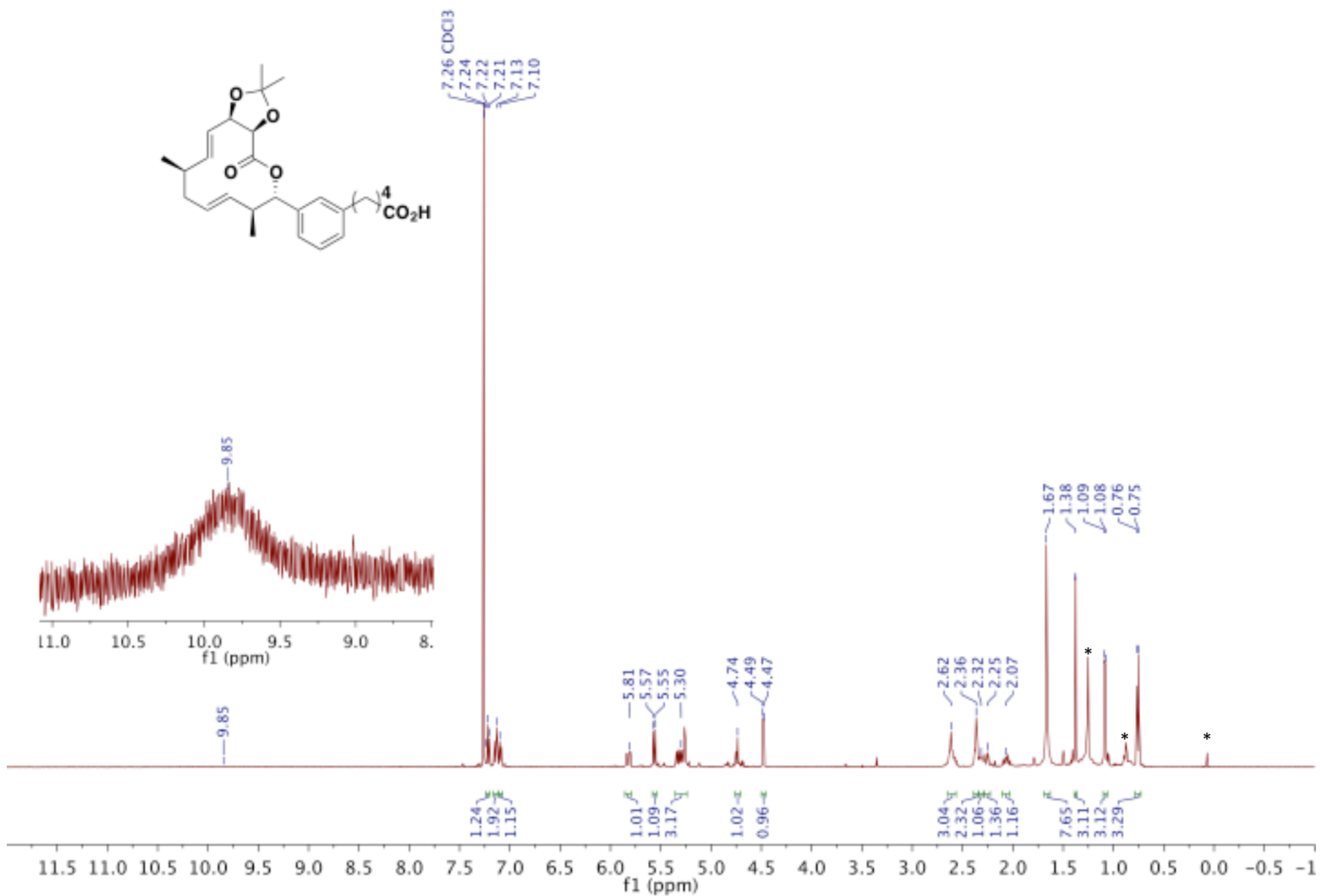
*Impurities present in spectrum

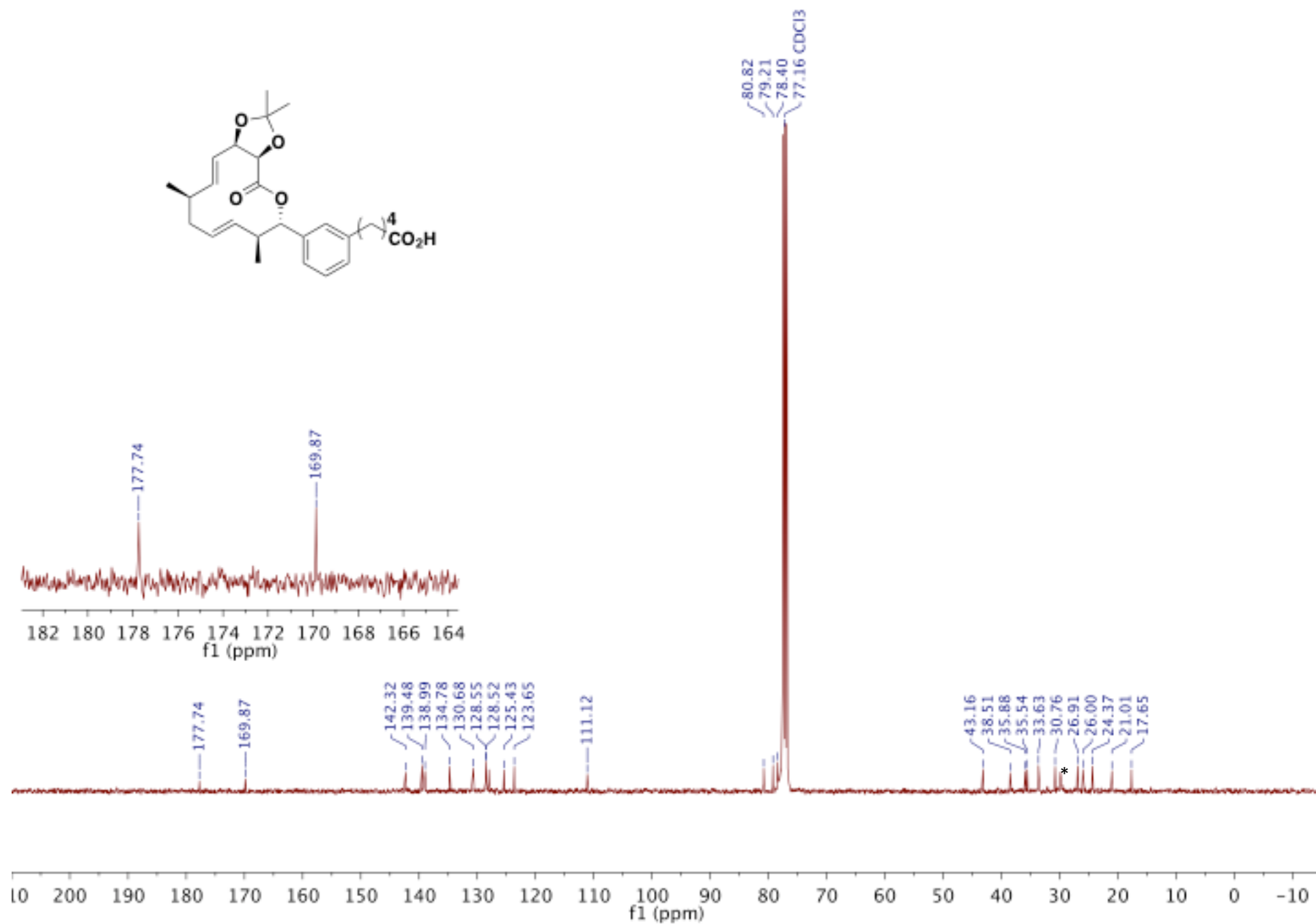
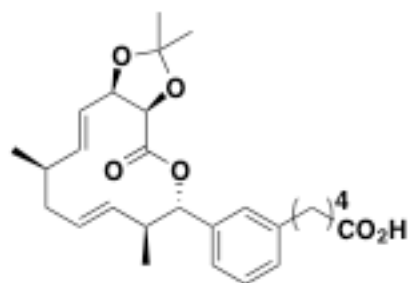
(-)-C1: ^{13}C NMR (126 MHz, CDCl_3)



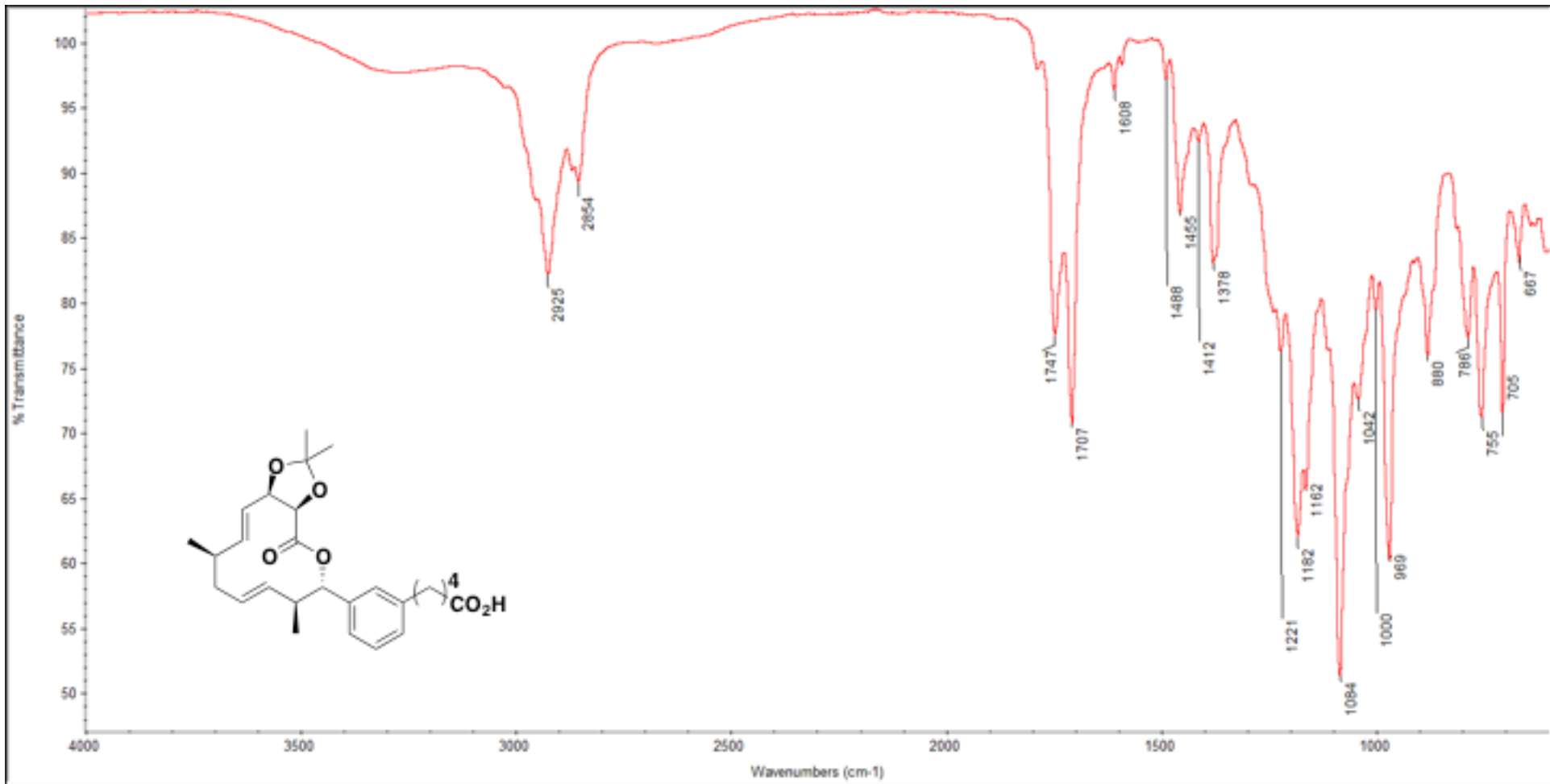
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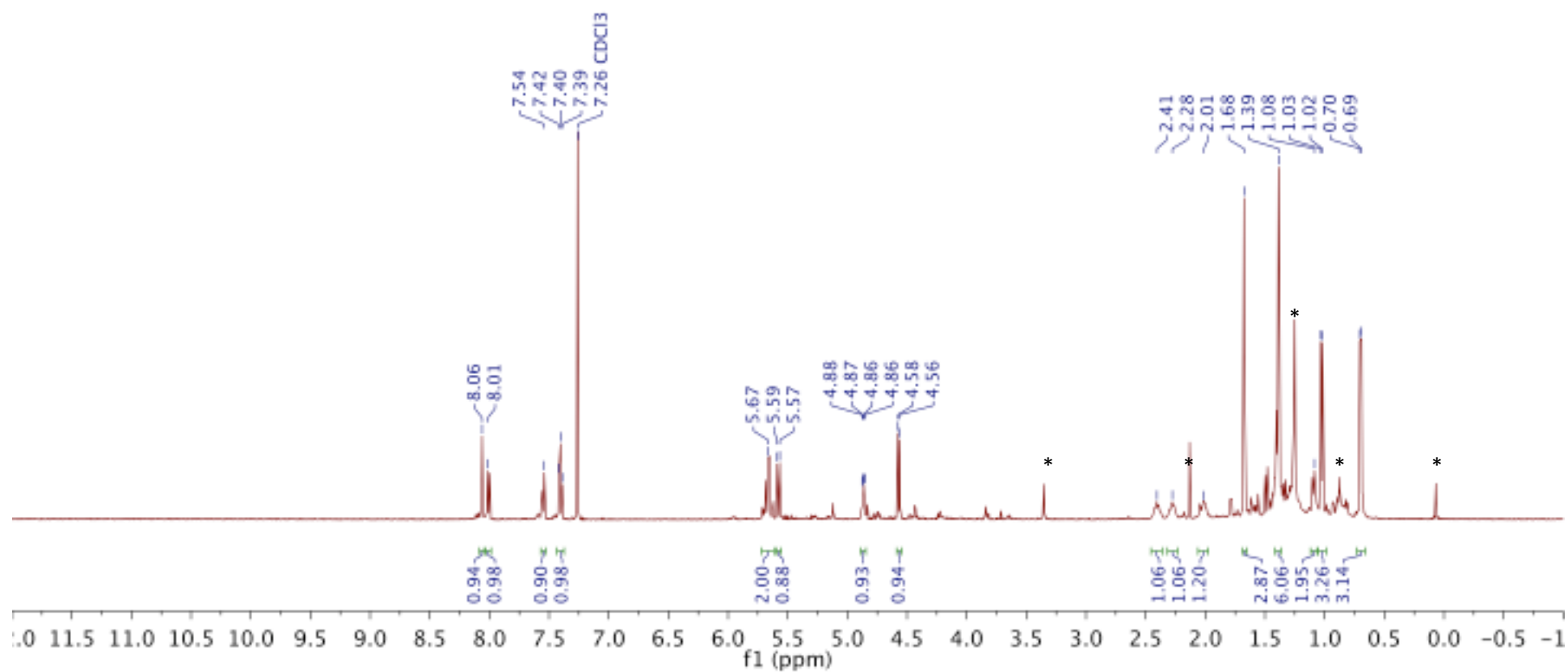
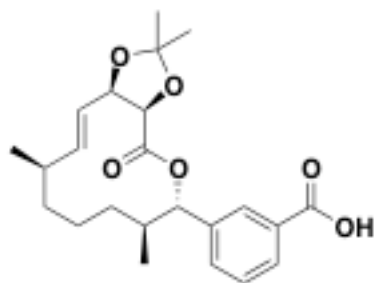




*Impurity present in spectrum

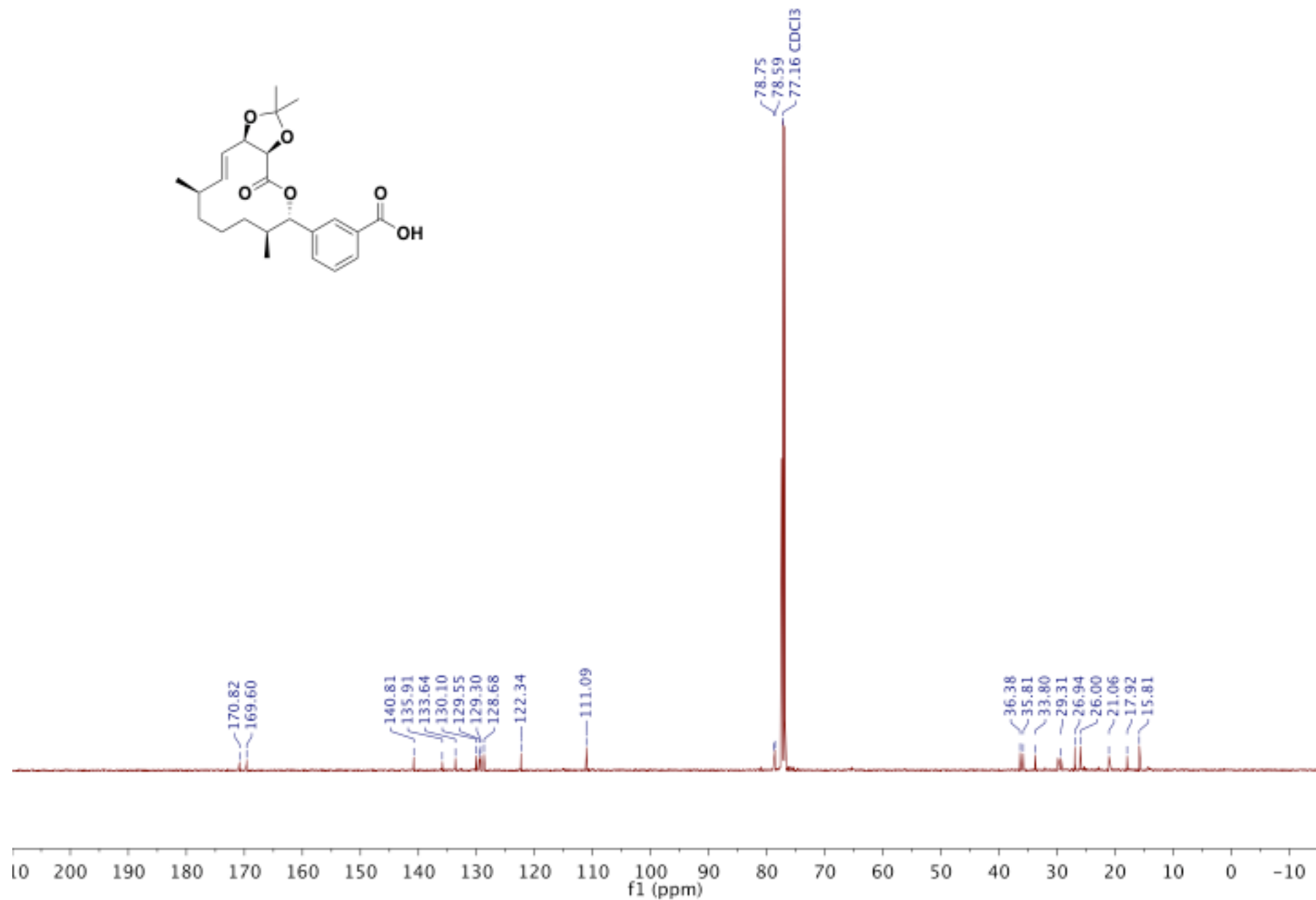
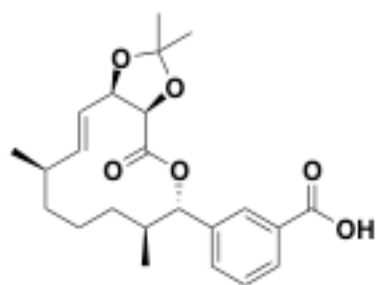


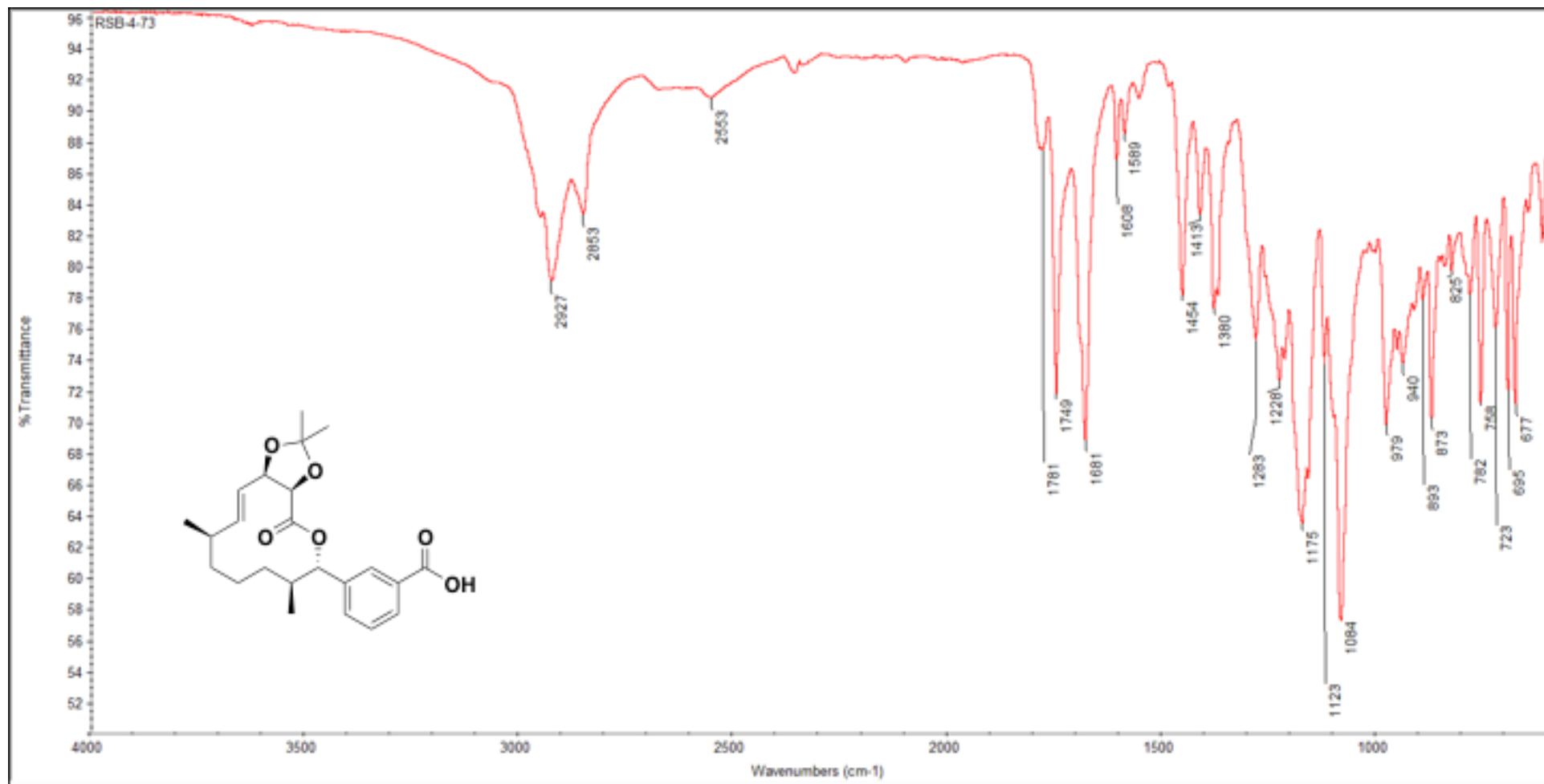
(-)-C2: ¹H NMR (500 MHz, CDCl₃)



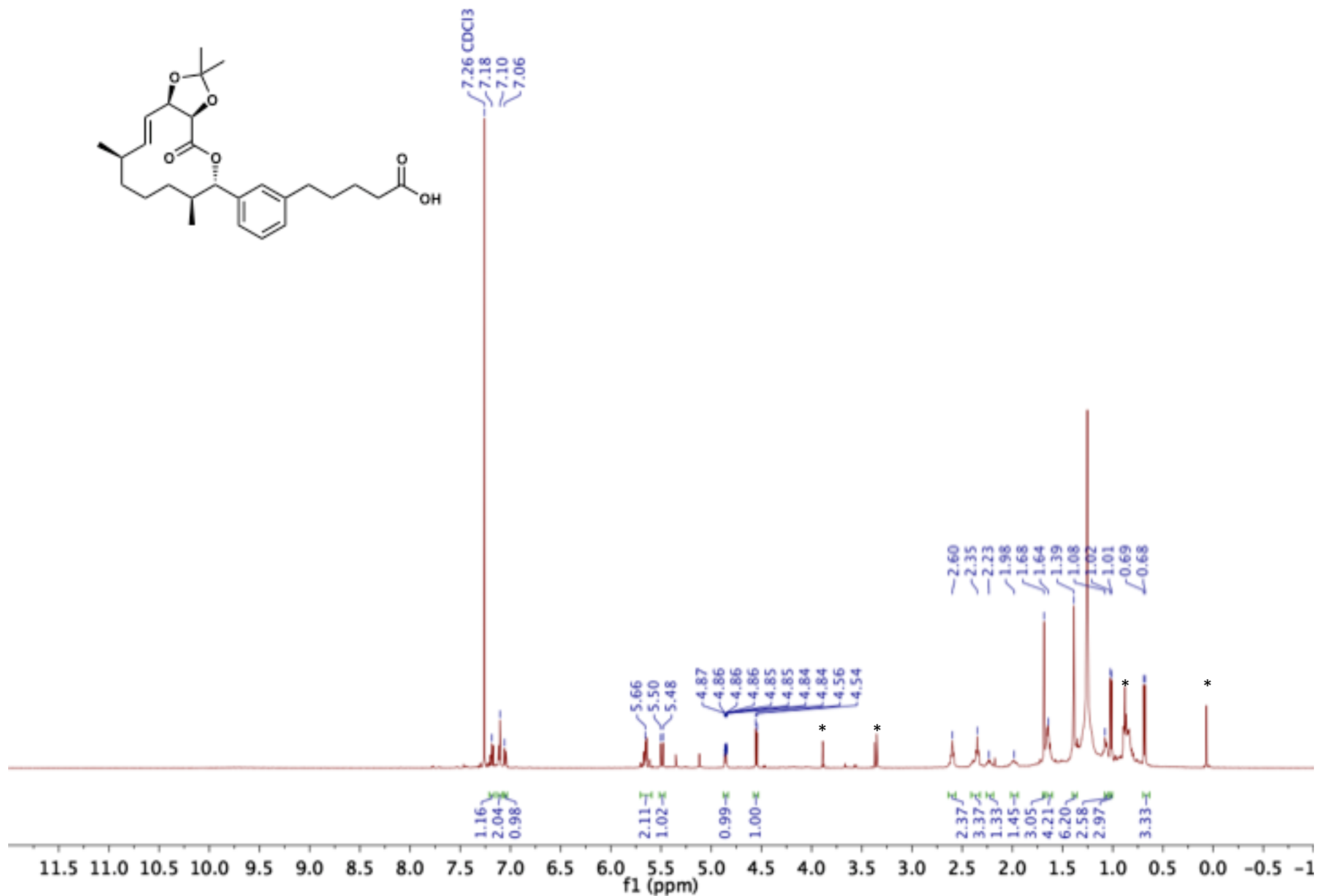
*Impurities present in spectrum

(-)-C2: ^{13}C NMR (126 MHz, CDCl_3)



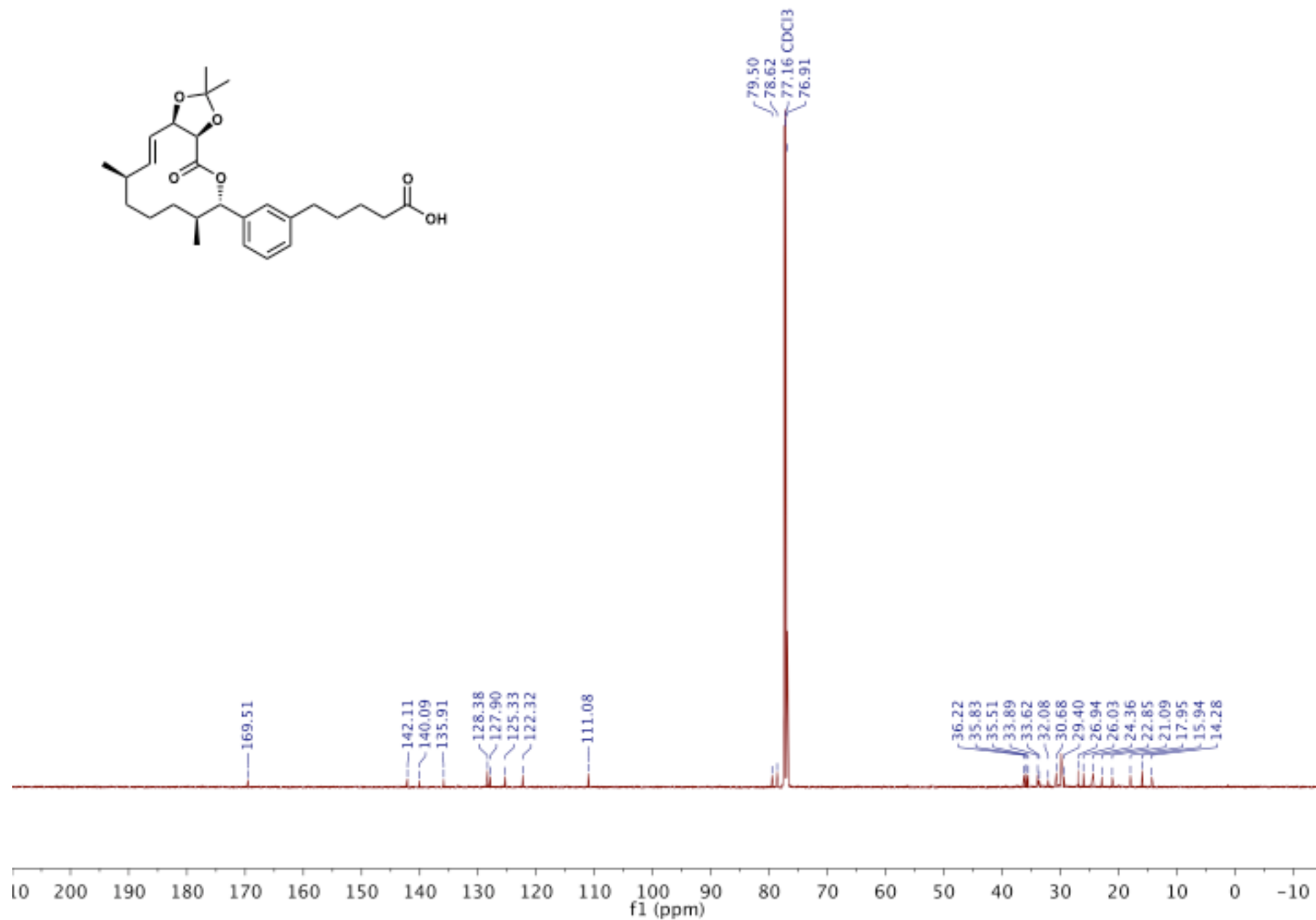
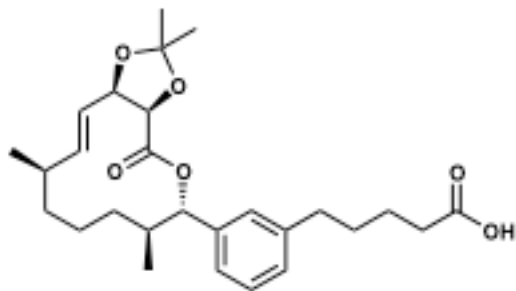


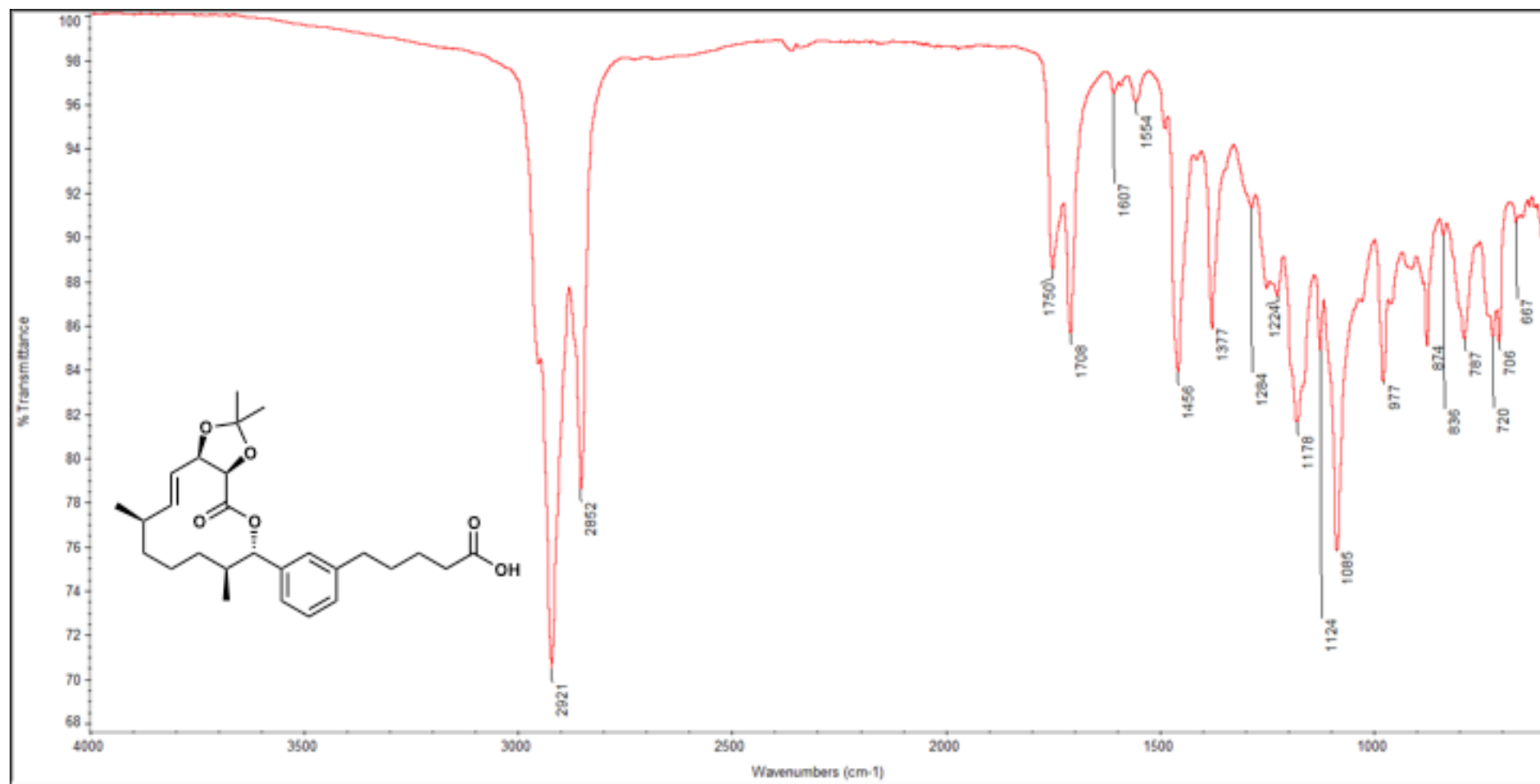
(-)-C4: ¹H NMR (500 MHz, CDCl₃)



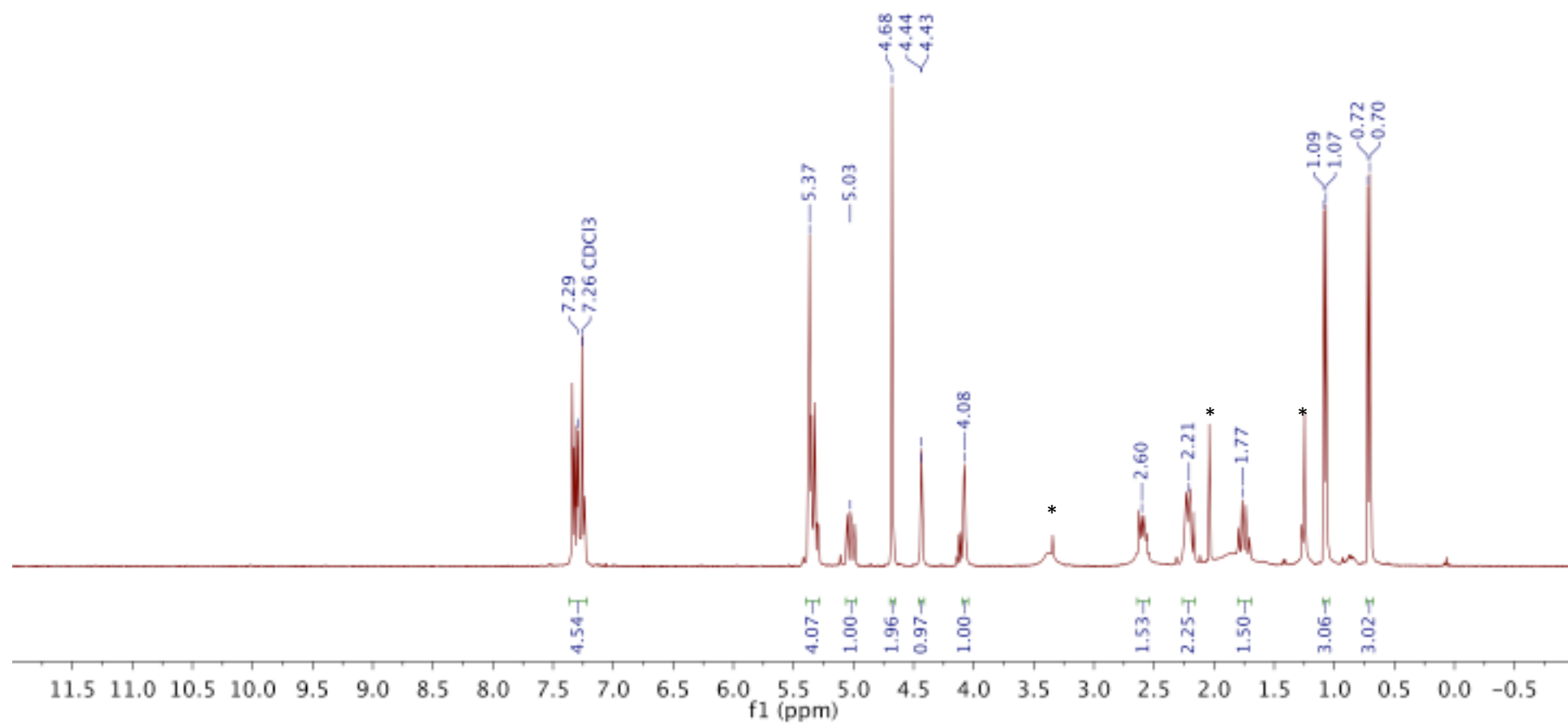
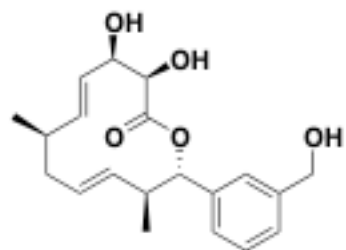
*Impurities present in spectrum

(-)-C4: ^{13}C NMR (126 MHz, CDCl_3)



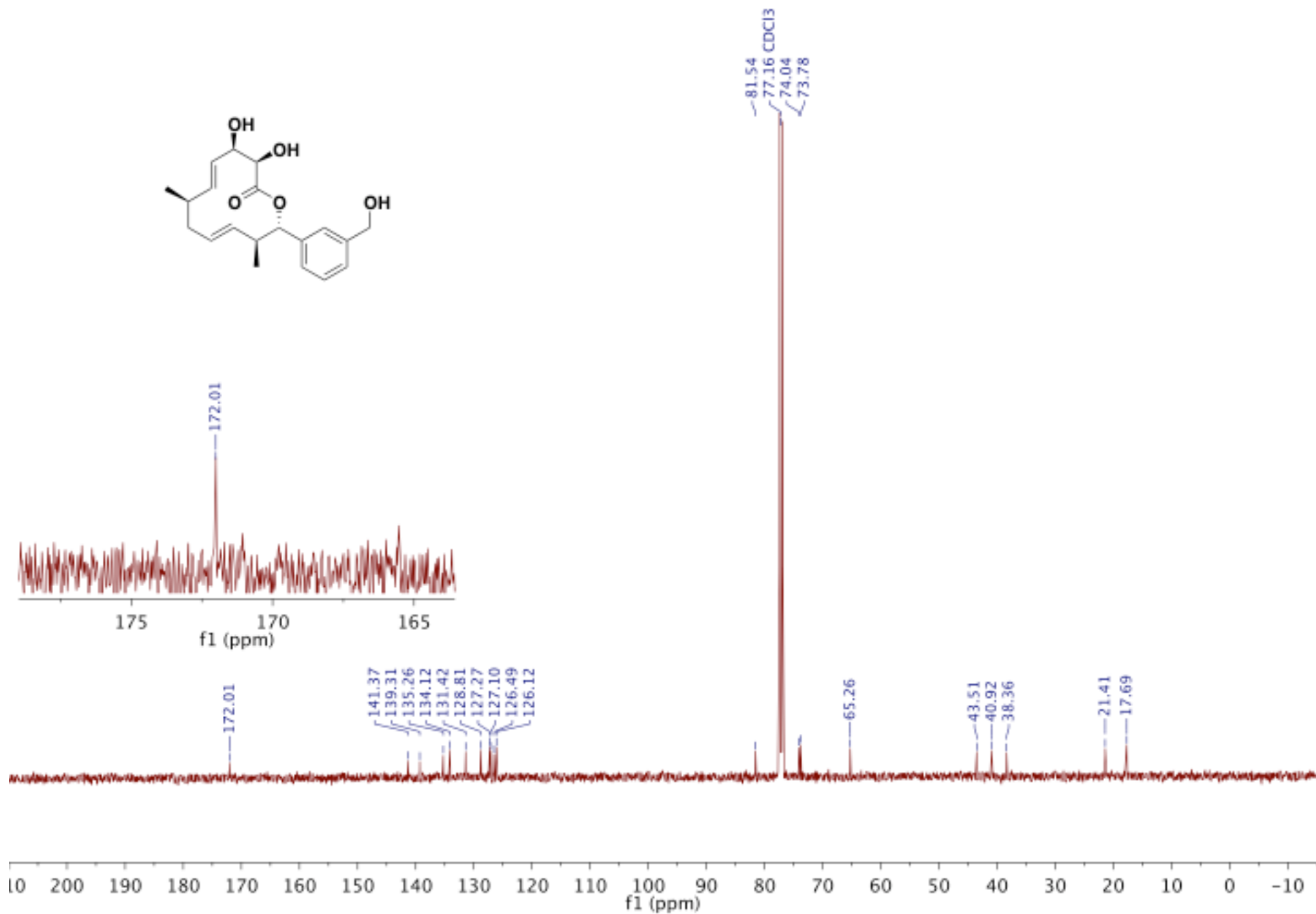
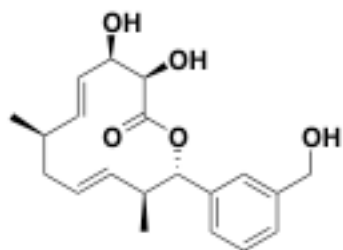


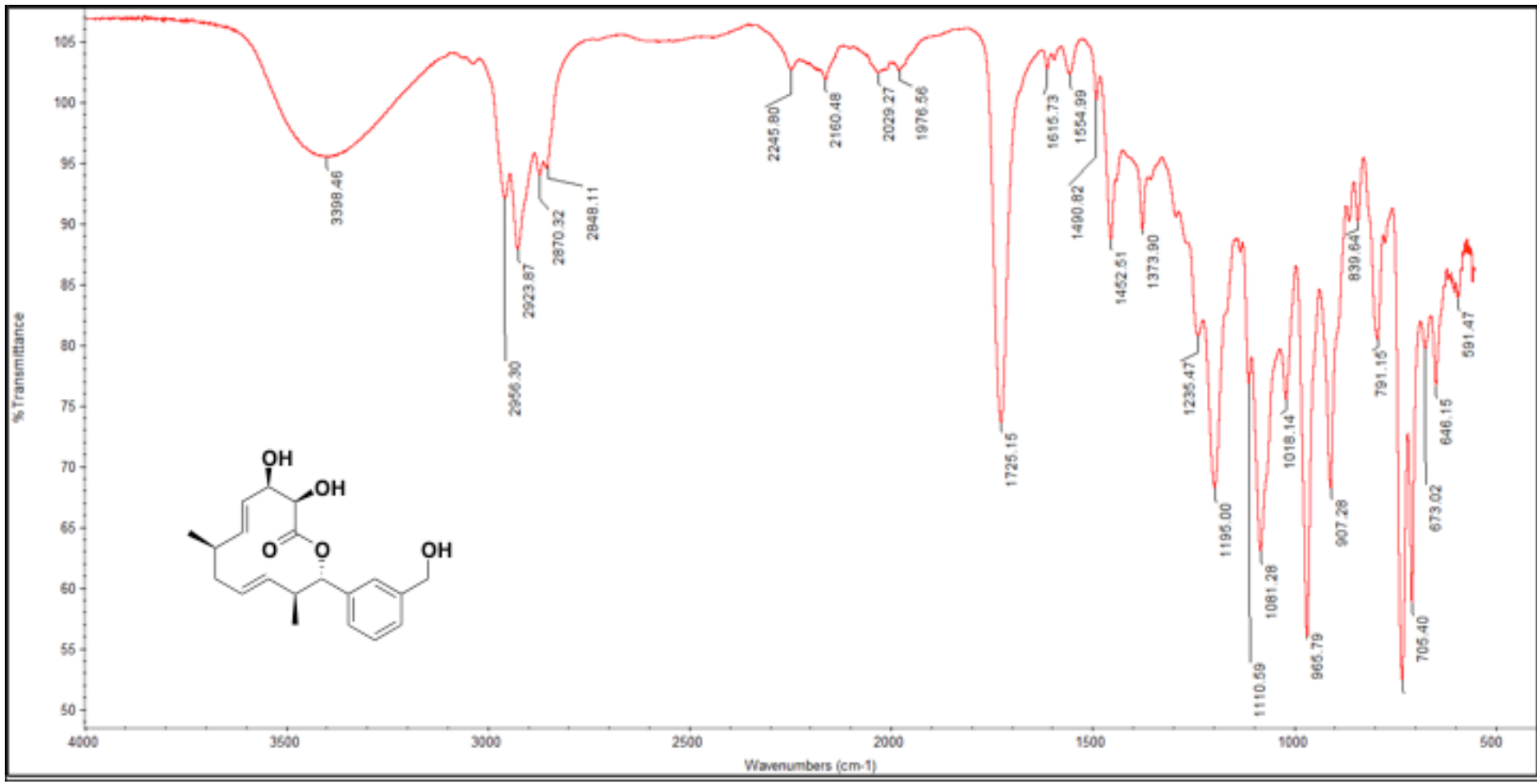
(-)-B1: ^1H NMR (500 MHz, CDCl_3)

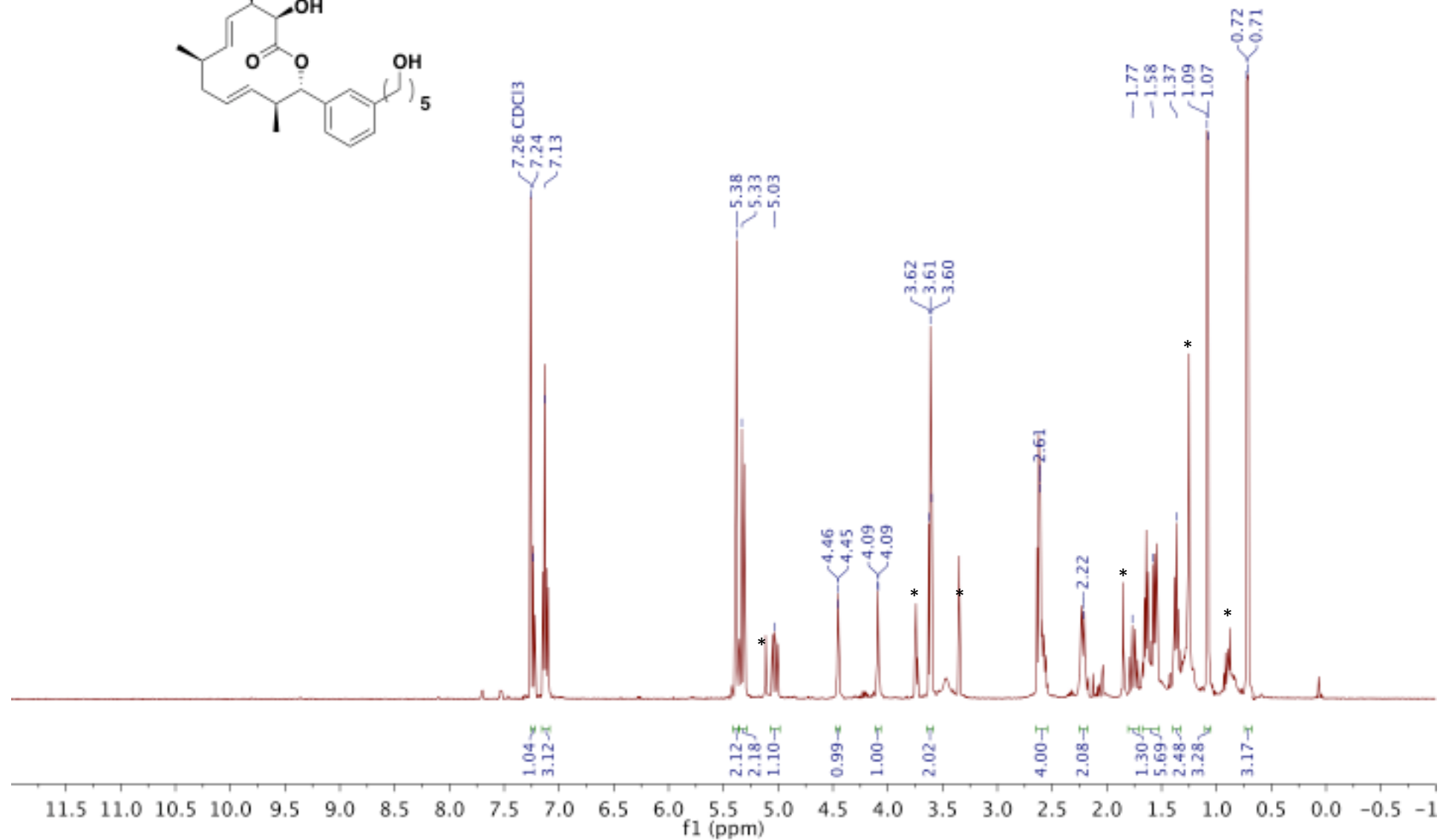
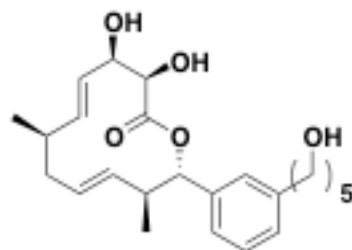


*Impurities present in spectrum

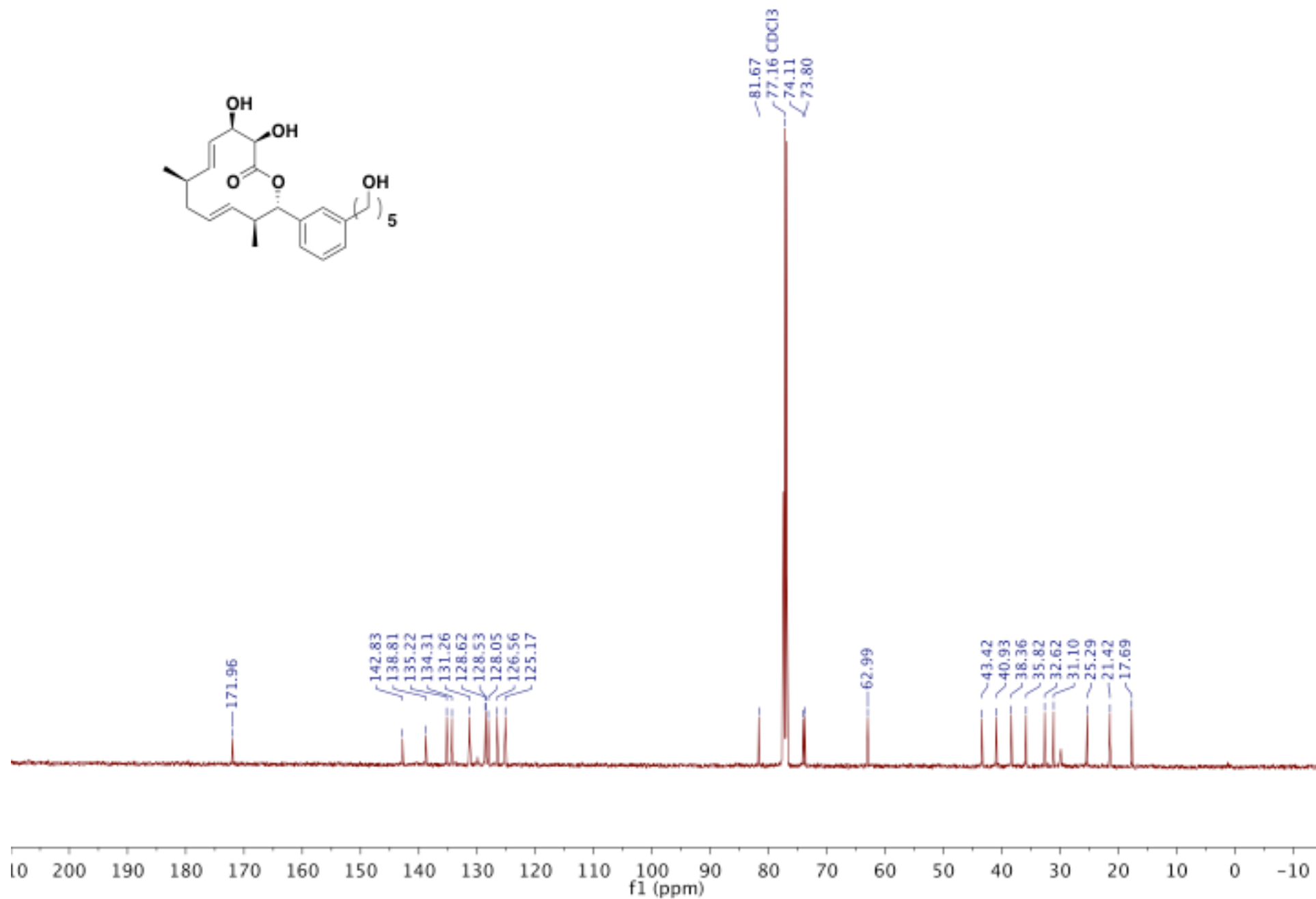
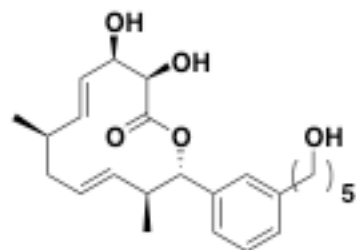
(-)-B1: ^{13}C NMR (126 MHz, CDCl_3)

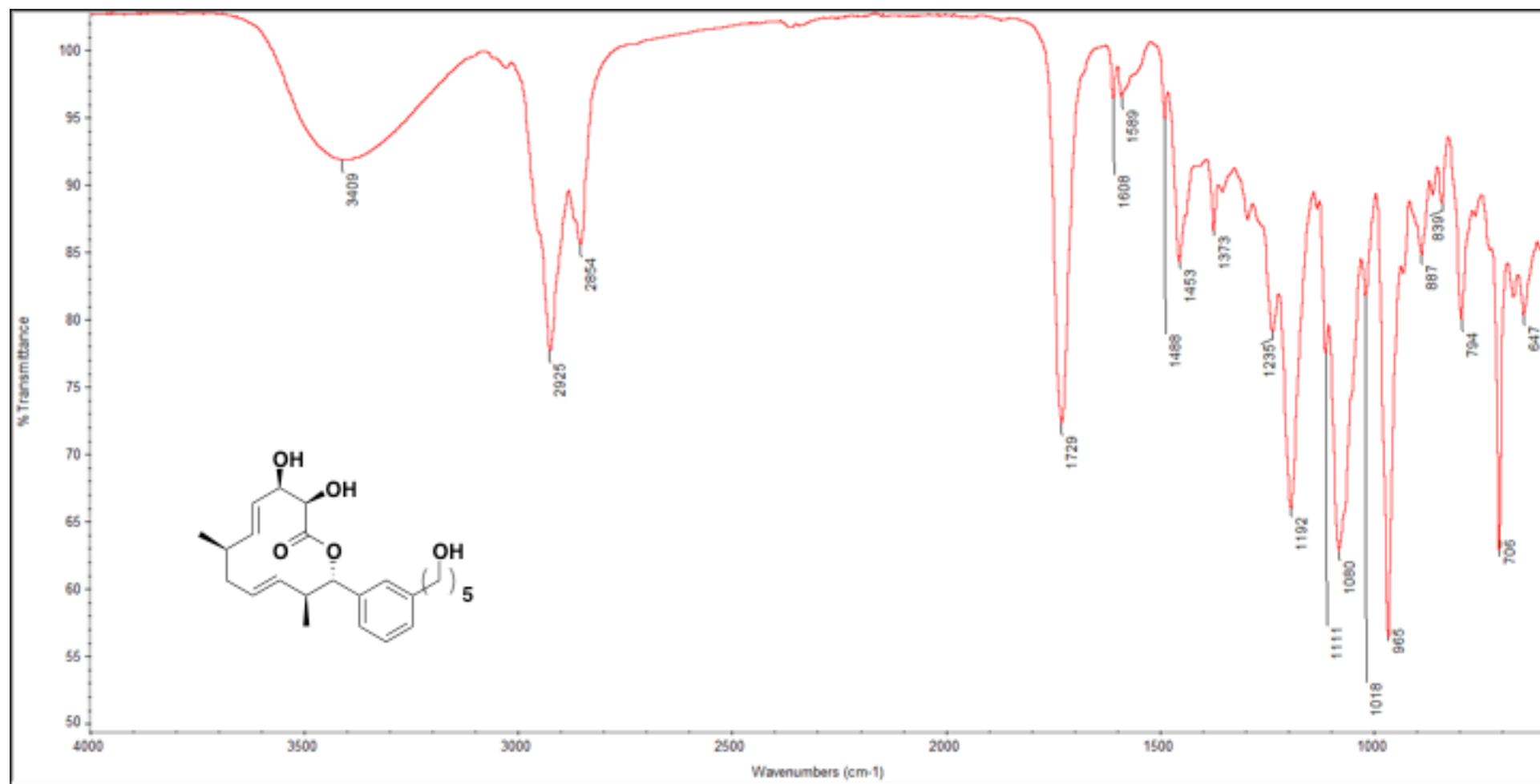




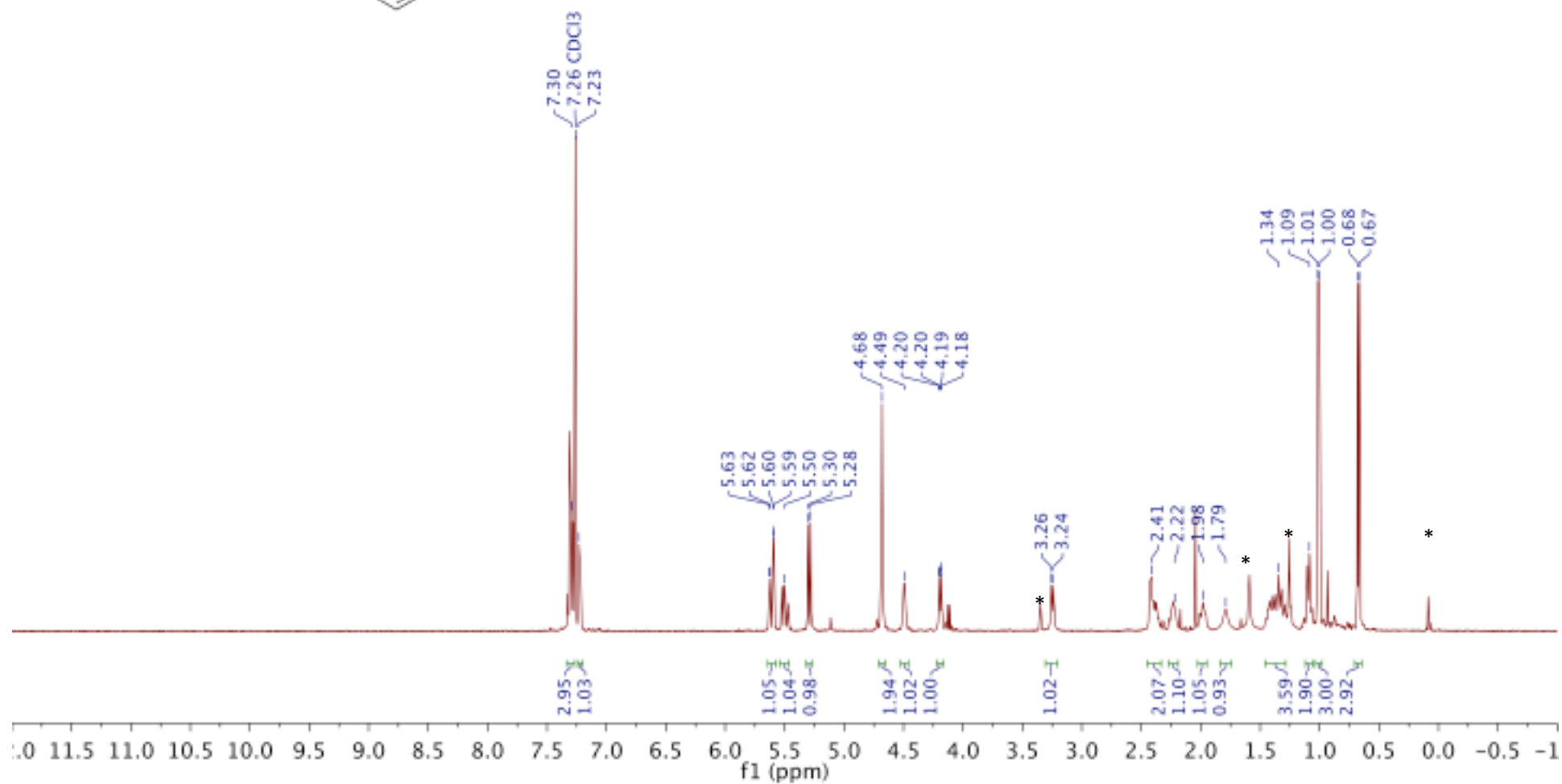
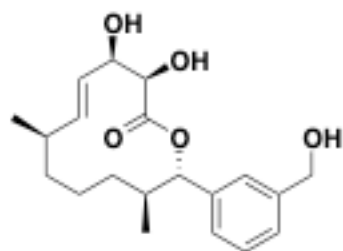


*Impurities present in spectrum

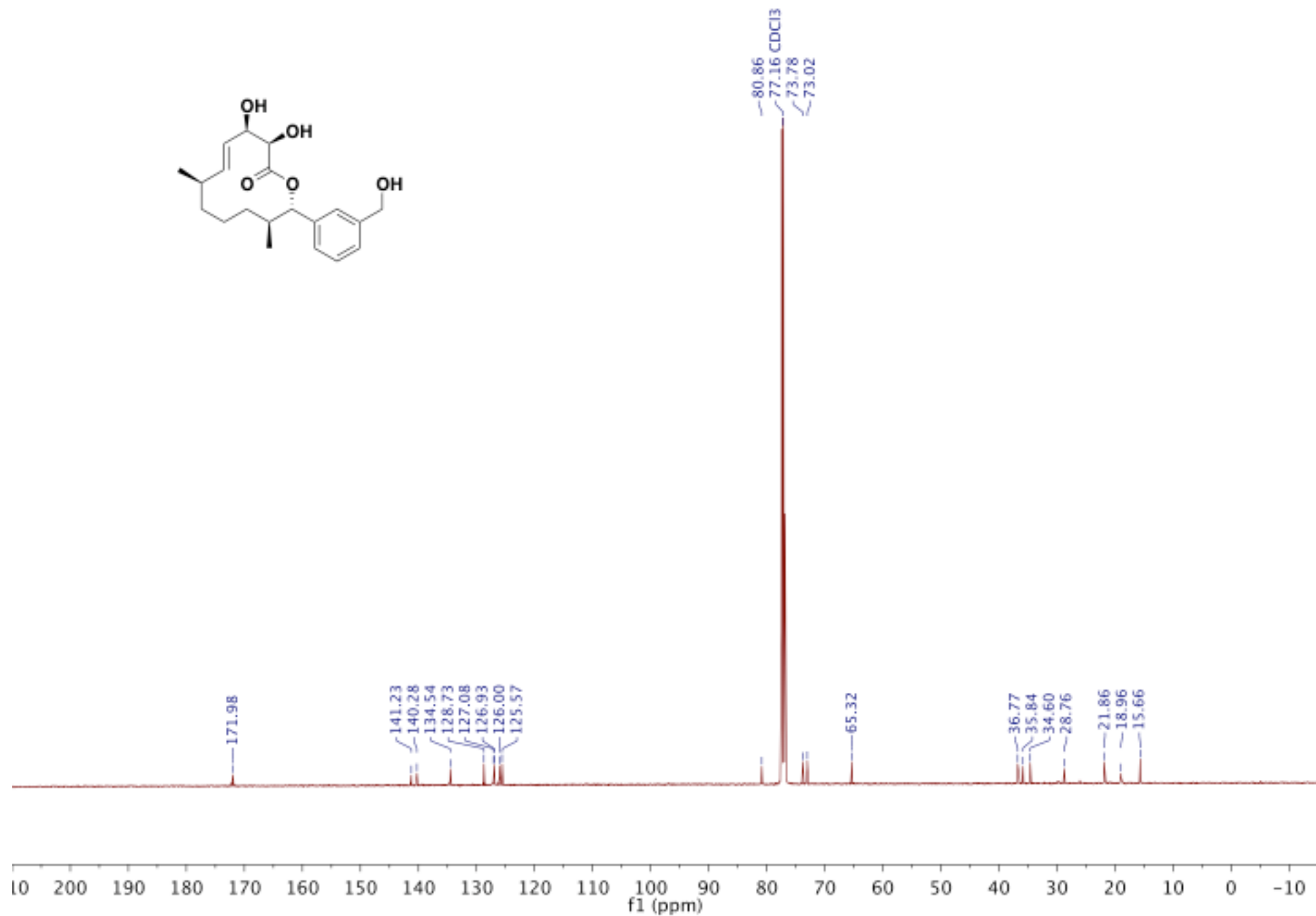
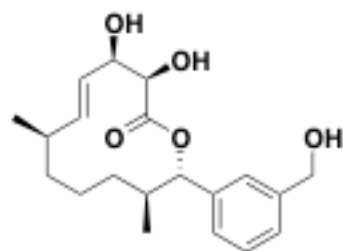


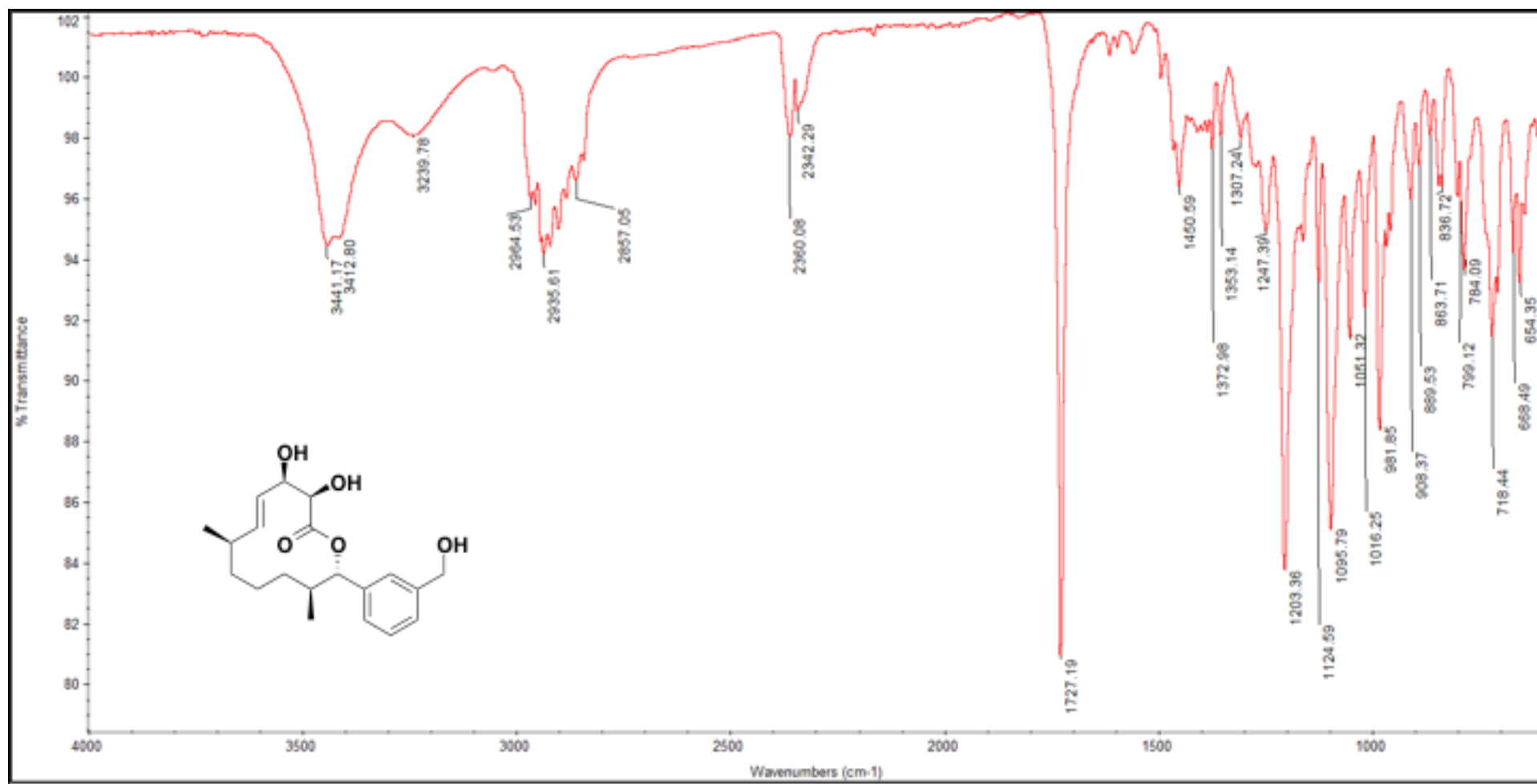


(-)-B2: ¹H NMR (500 MHz, CDCl₃)

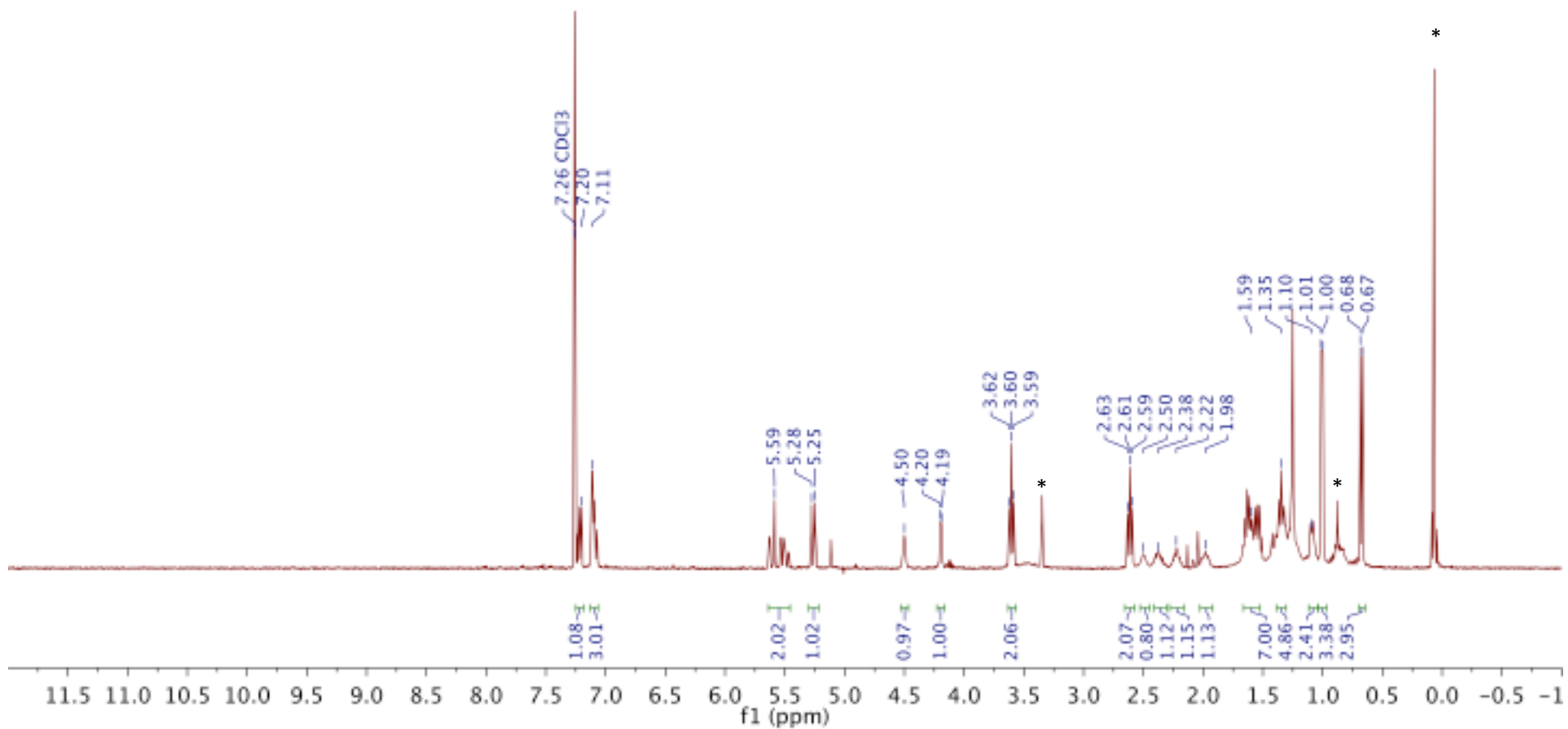
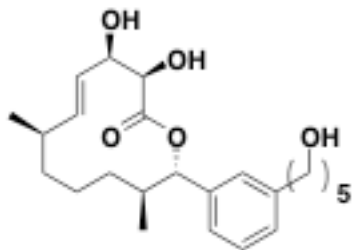


*Impurities present in spectrum

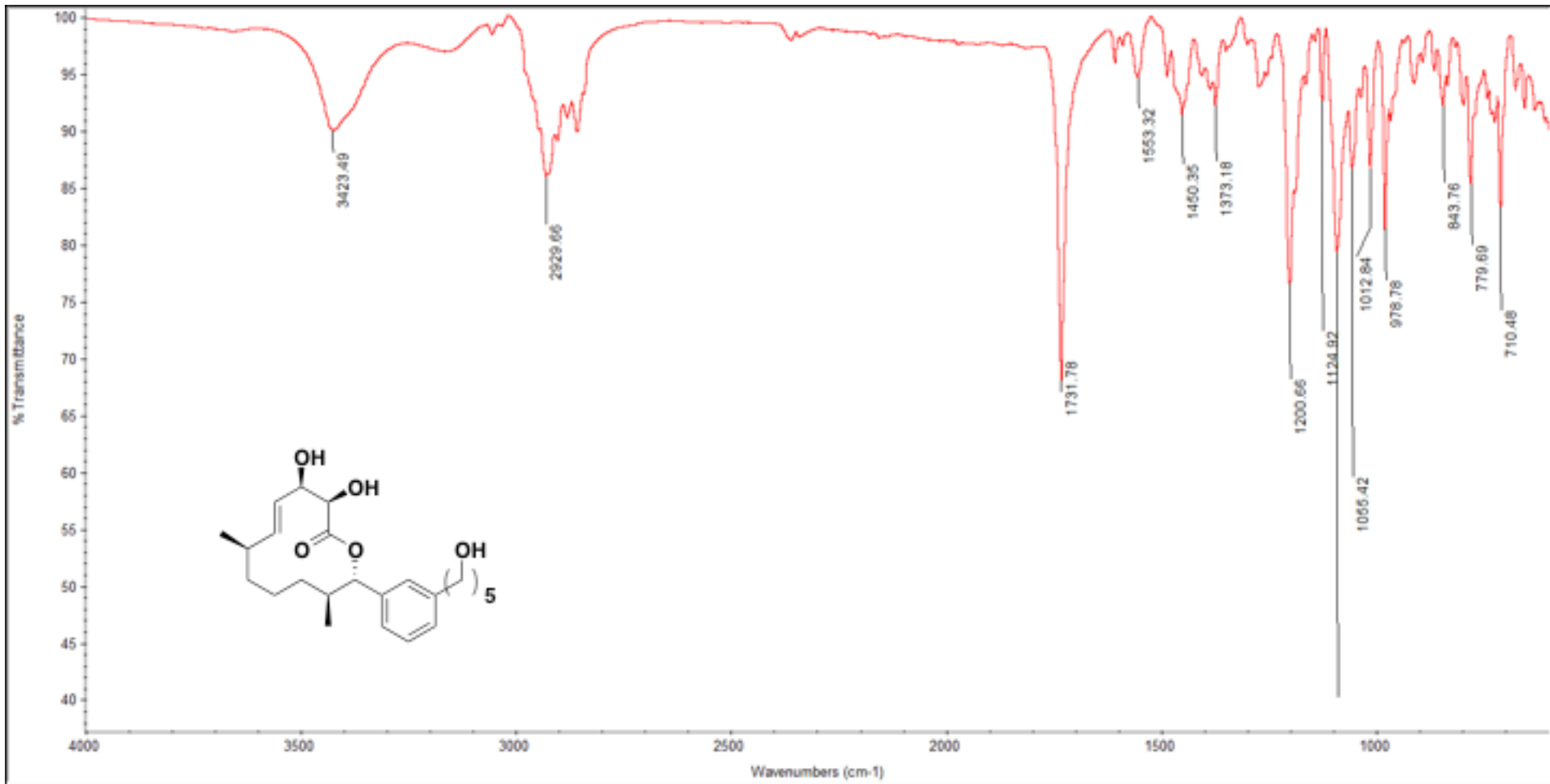




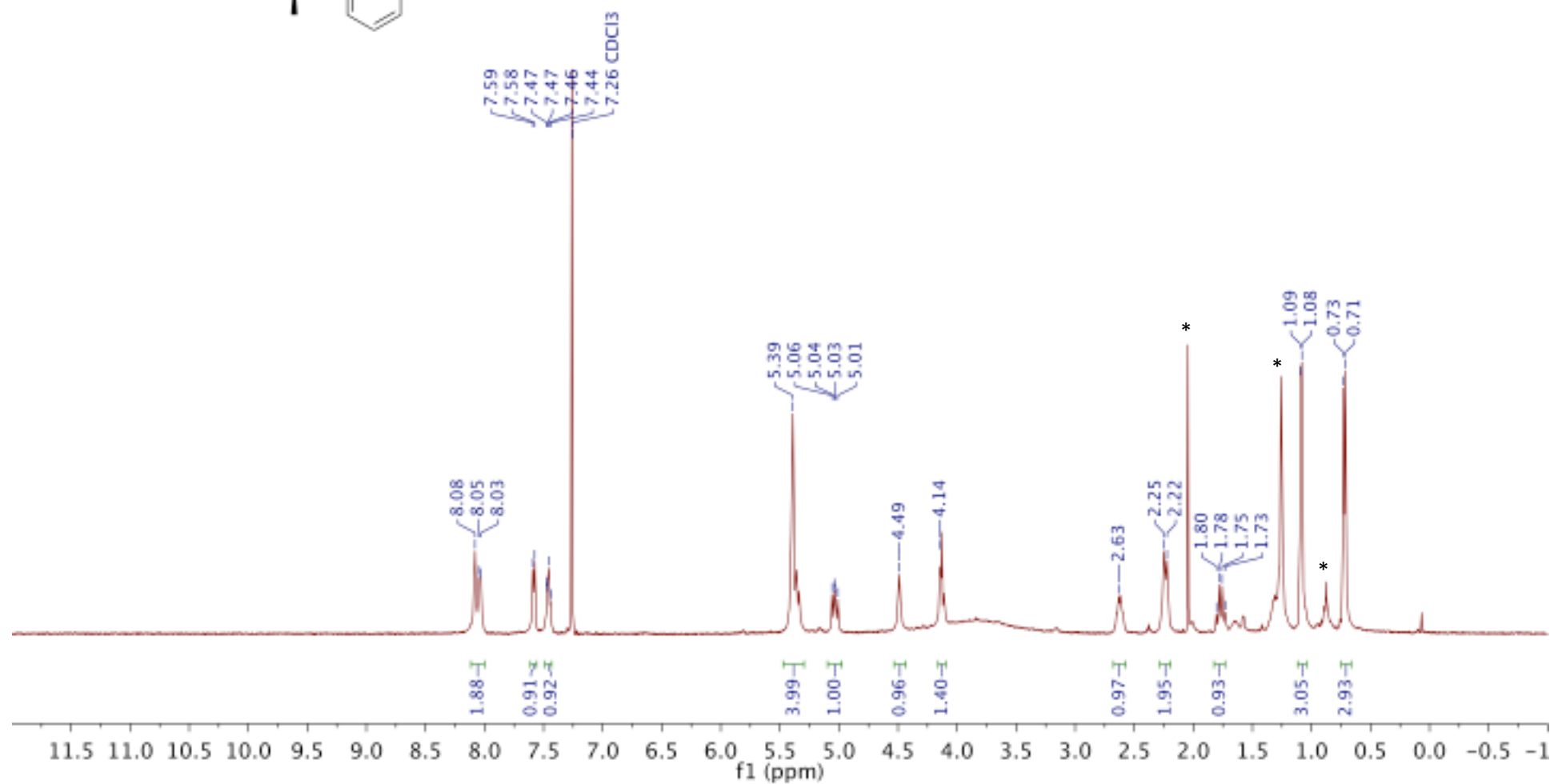
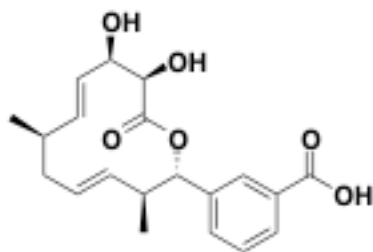
(-)-B4: ¹H NMR (500 MHz, CDCl₃)



*Impurities present in spectrum

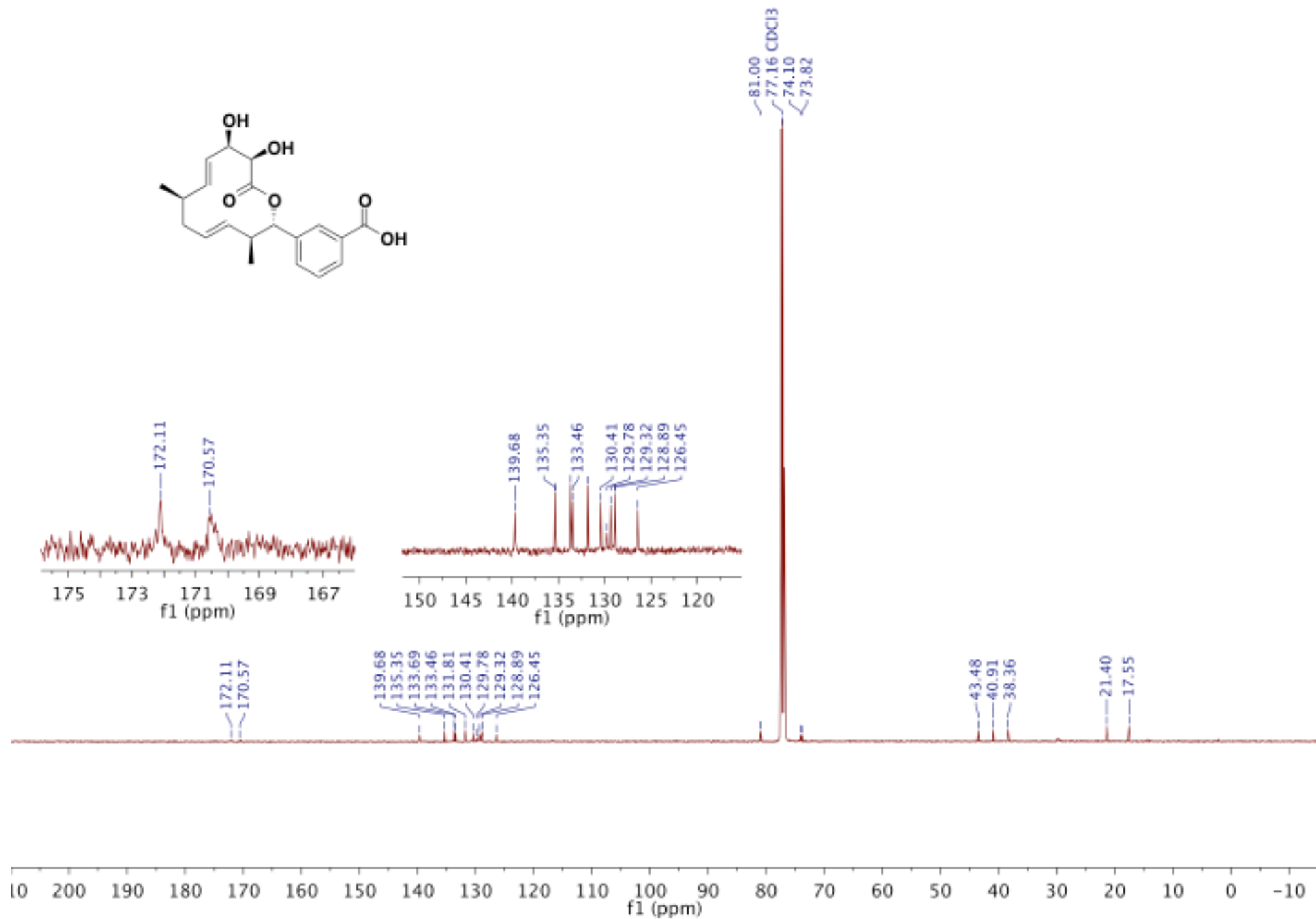
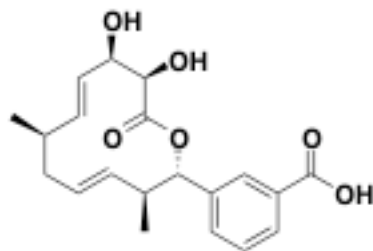


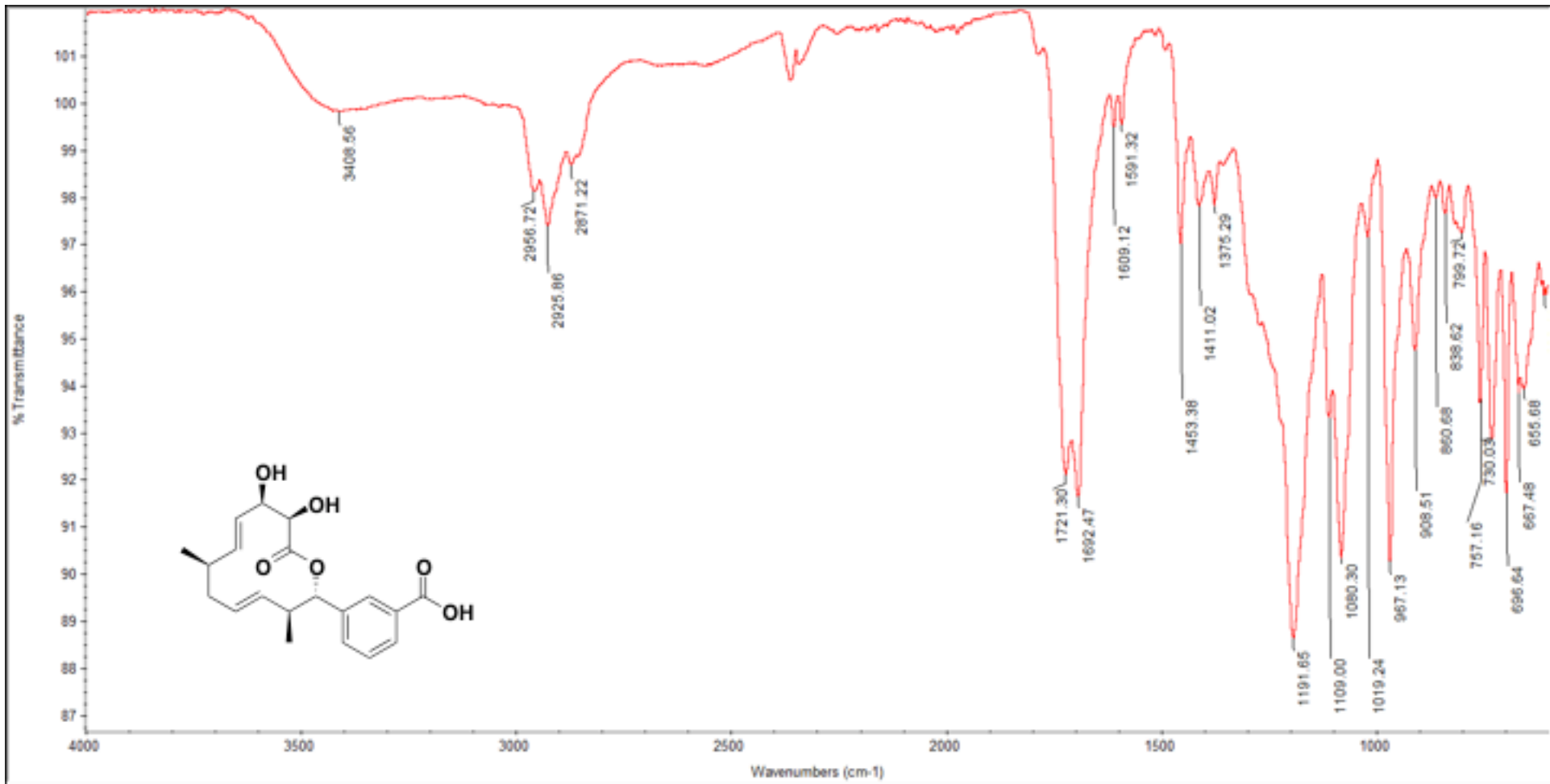
(-)-D1: ^1H NMR (500 MHz, CDCl_3)



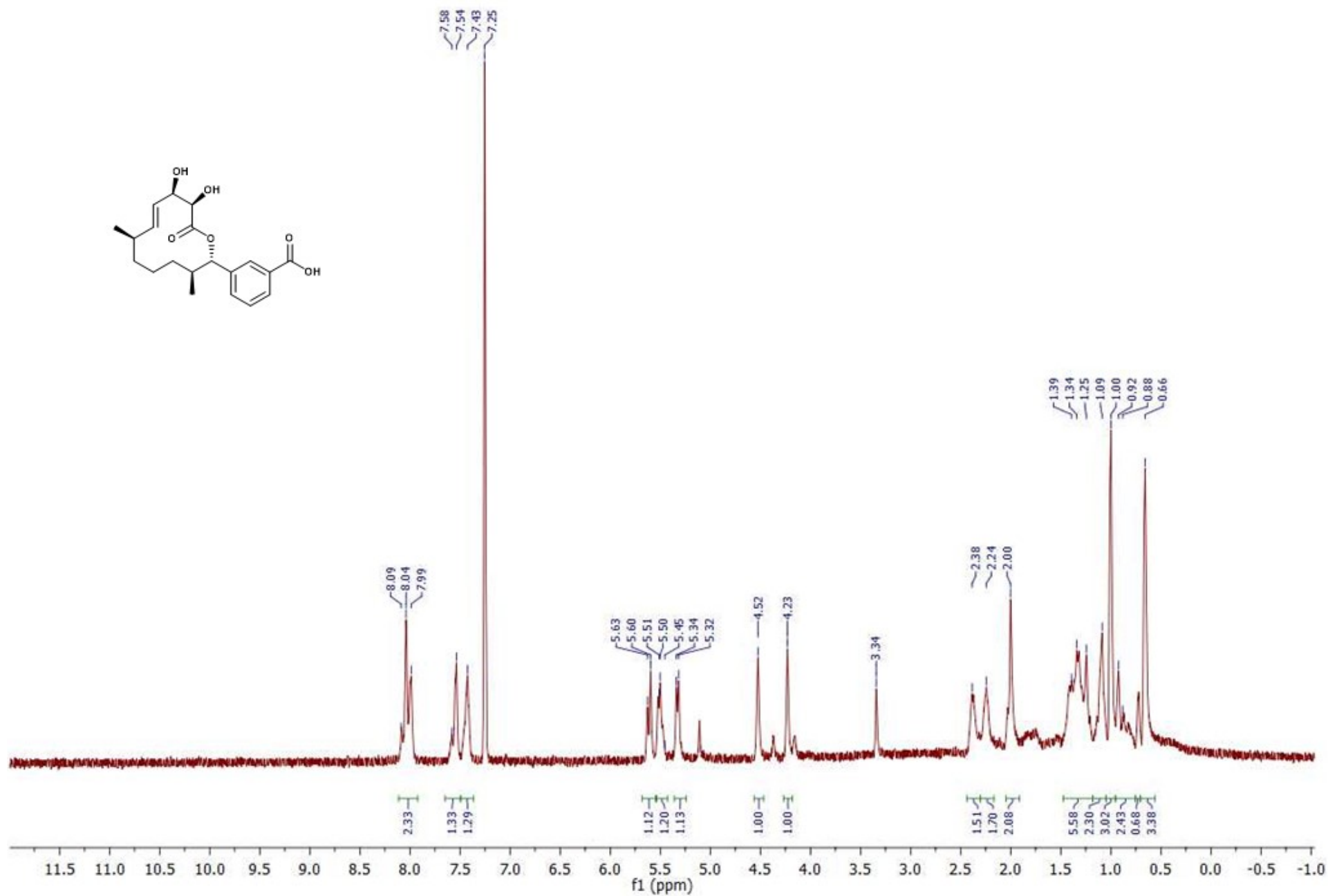
*Impurities present in spectrum

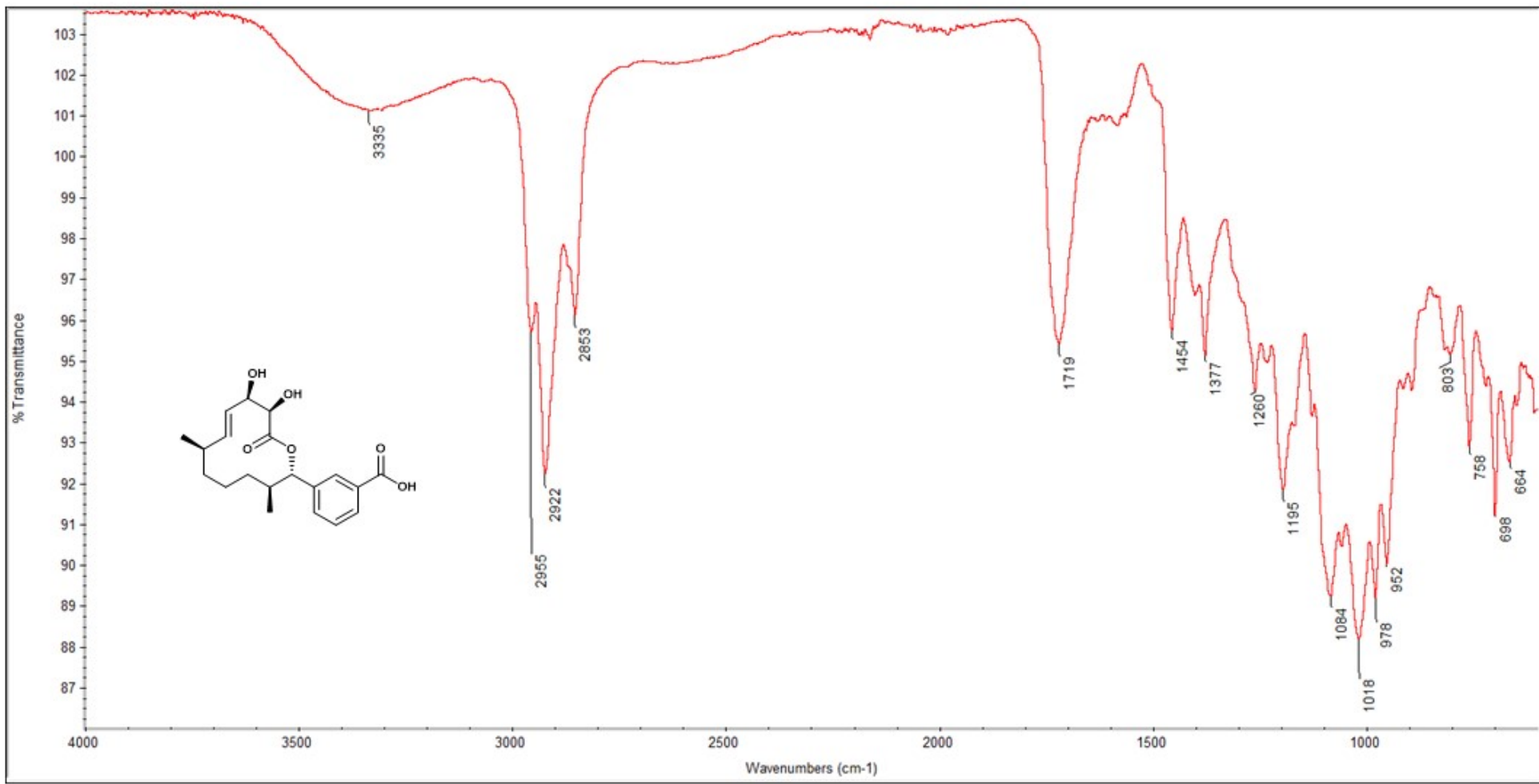
(-)-D1: ^{13}C NMR (126 MHz, CDCl_3)



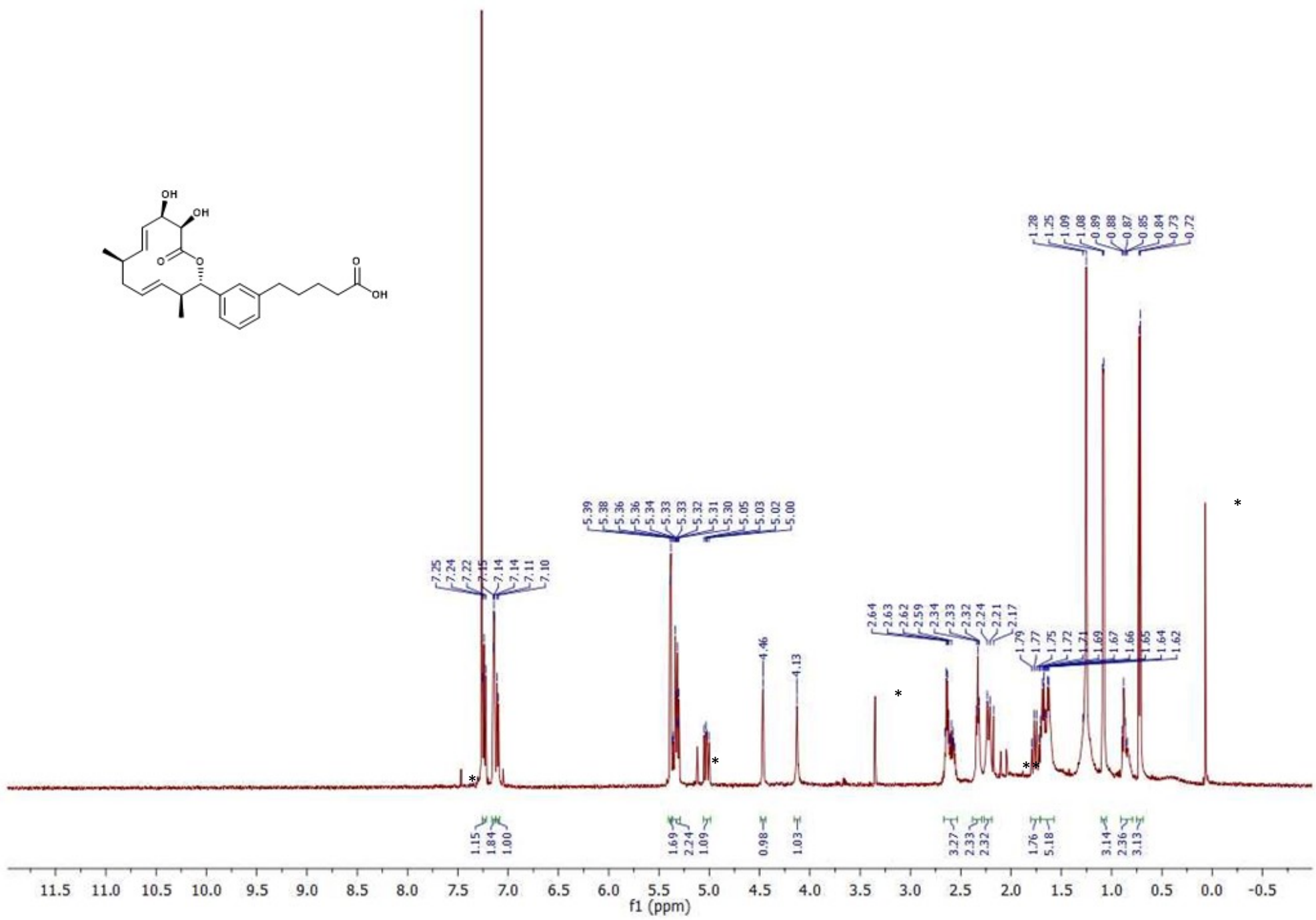
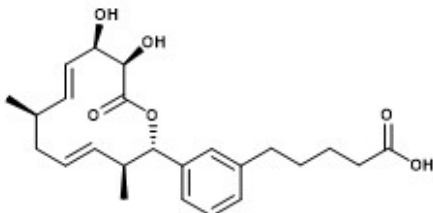


(-)-D2: ¹H NMR (500 MHz, CDCl₃)



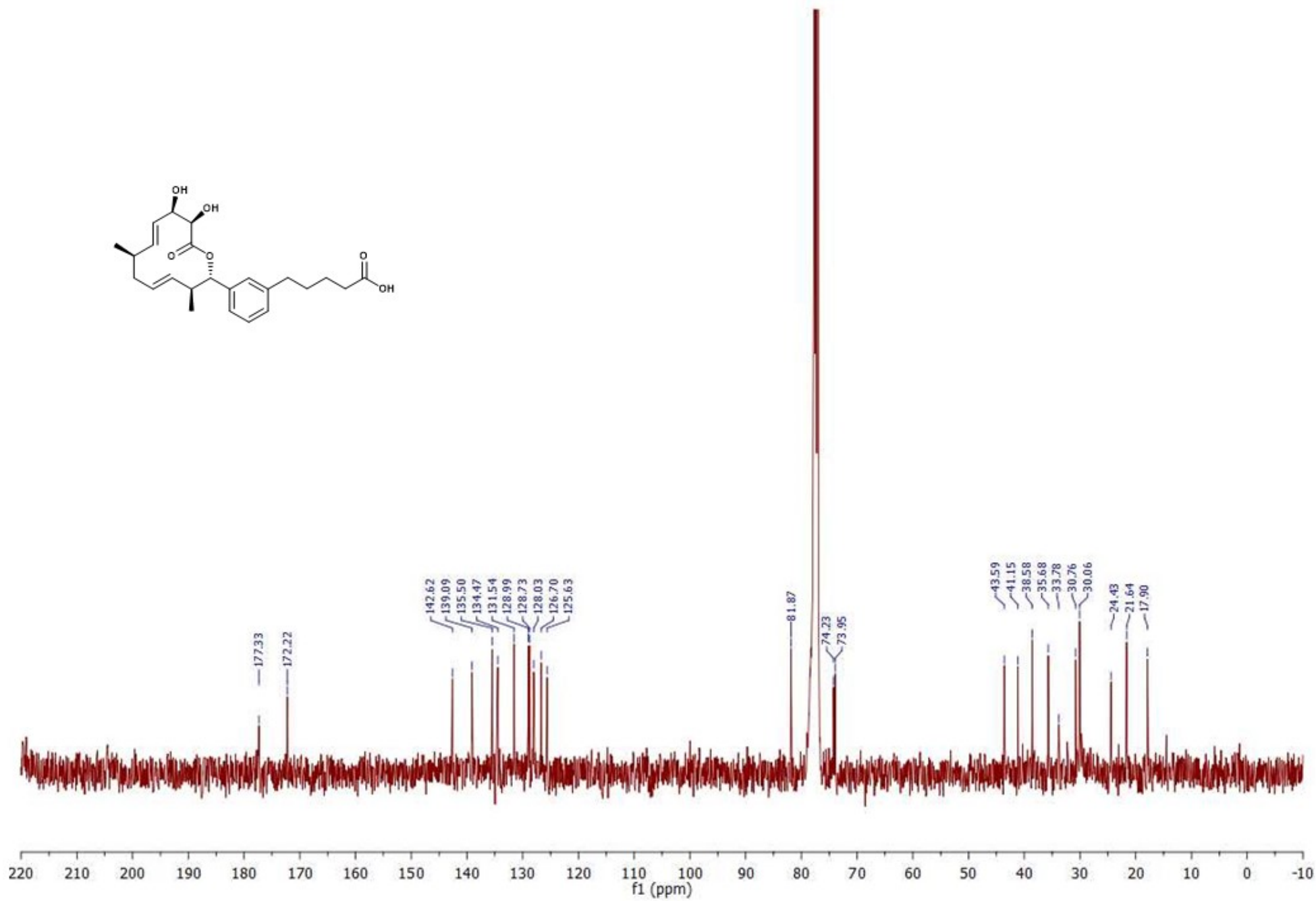
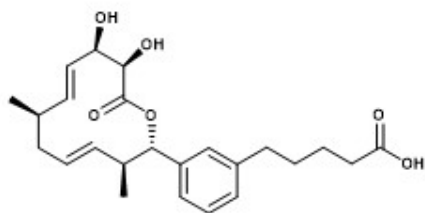


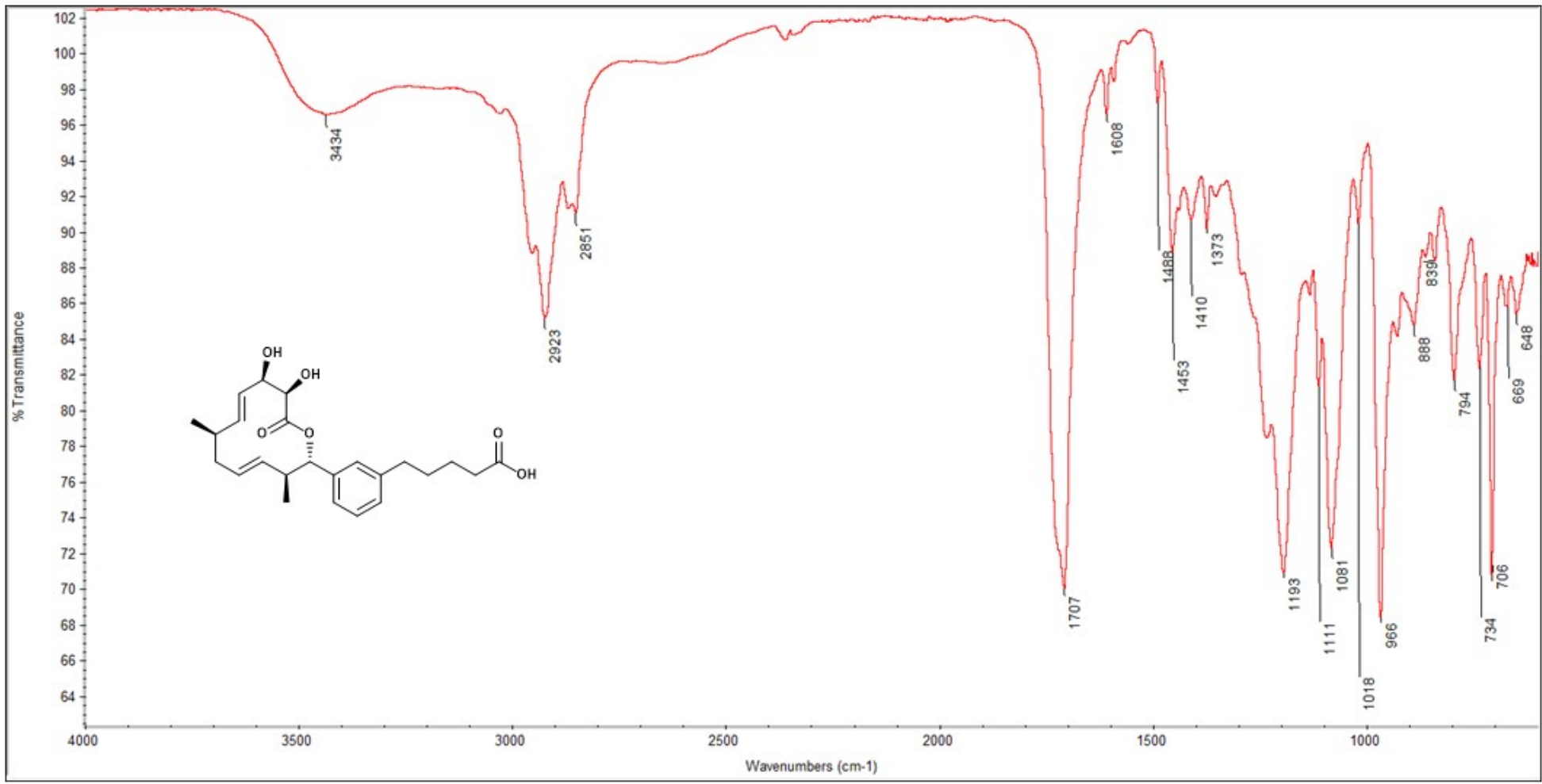
(-)-D3: ¹H NMR (500 MHz, CDCl₃)
 (-)-D3: ¹H NMR (500 MHz, CDCl₃)



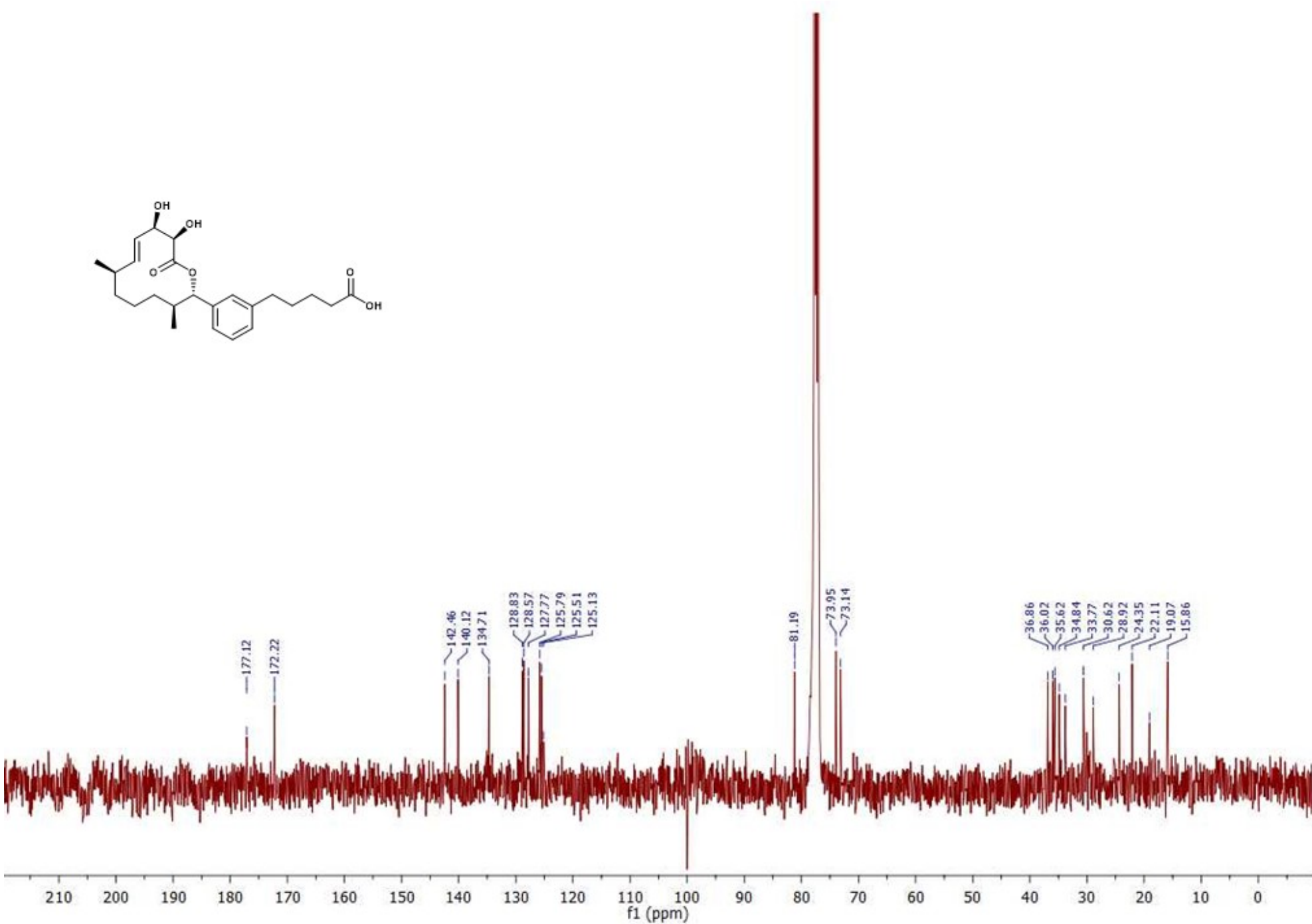
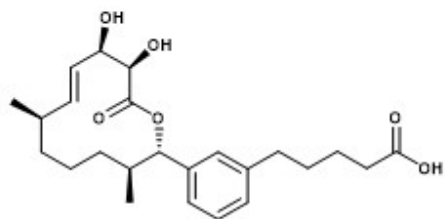
*Impurities present in spectrum

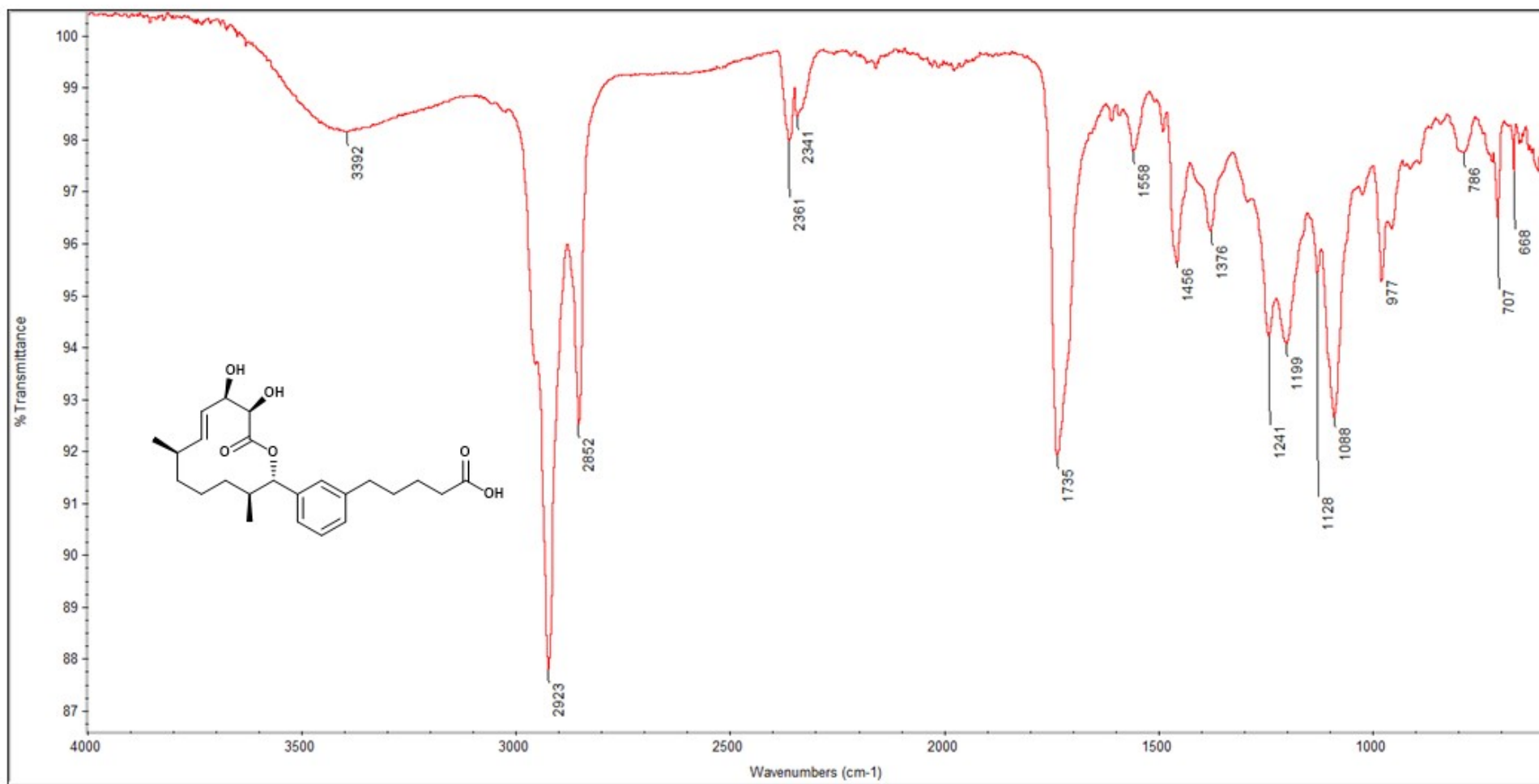
(-)-D3: ^{13}C NMR (126 MHz, CDCl_3)



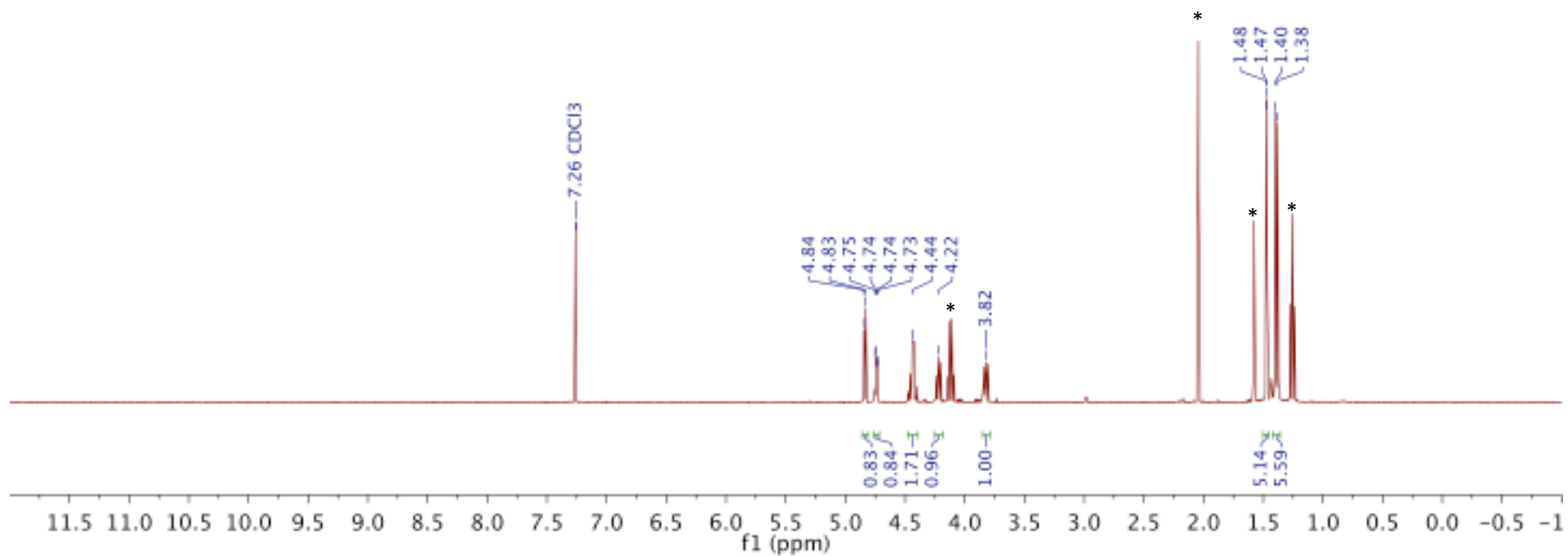
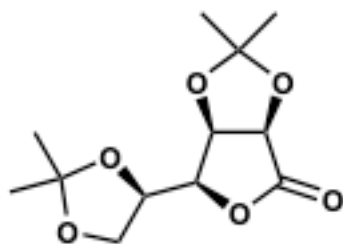


(-)-D4: ^{13}C NMR (126 MHz, CDCl_3)



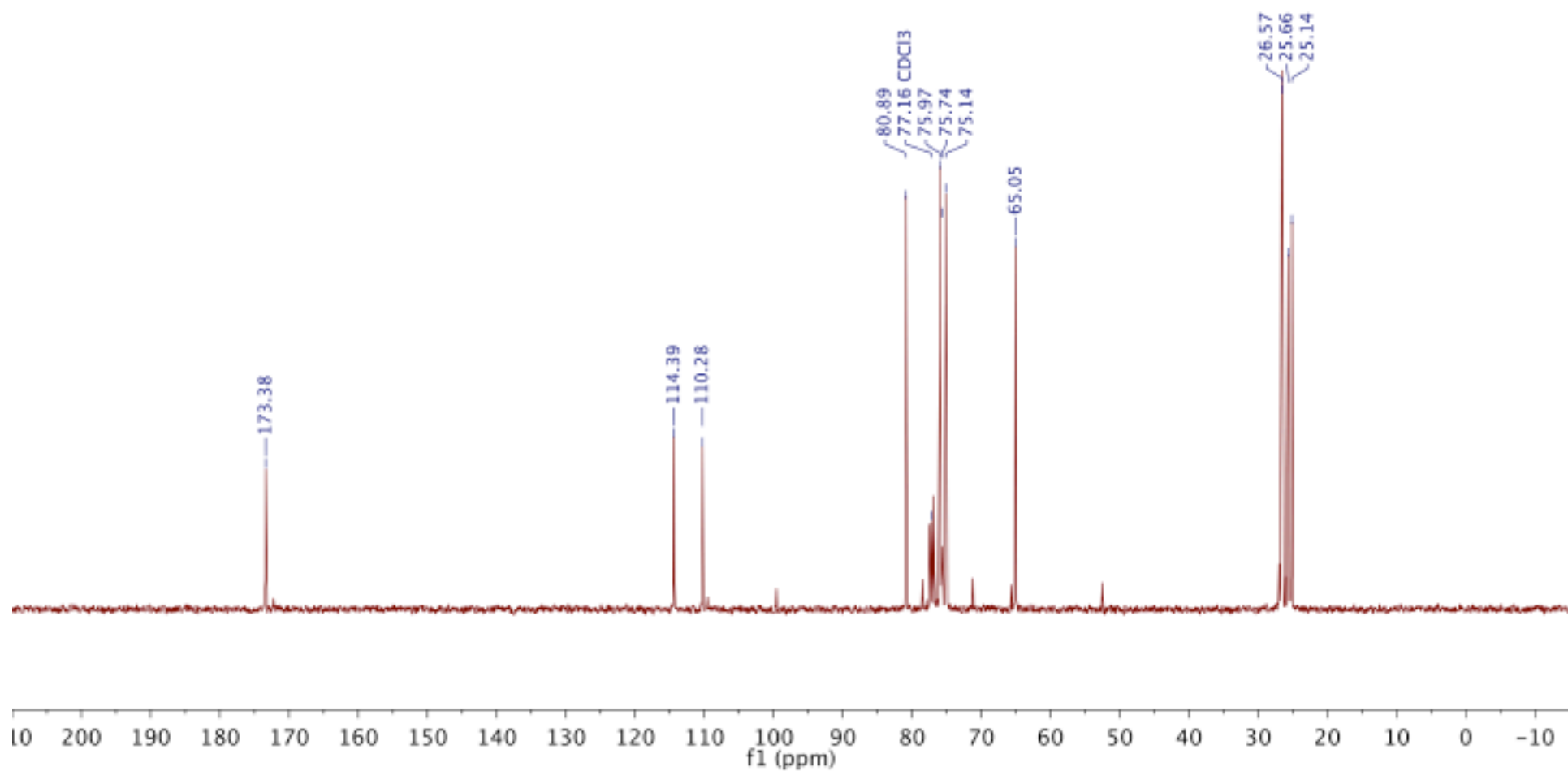
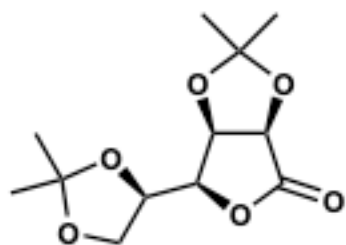


12: ¹H NMR (500 MHz, CDCl₃)

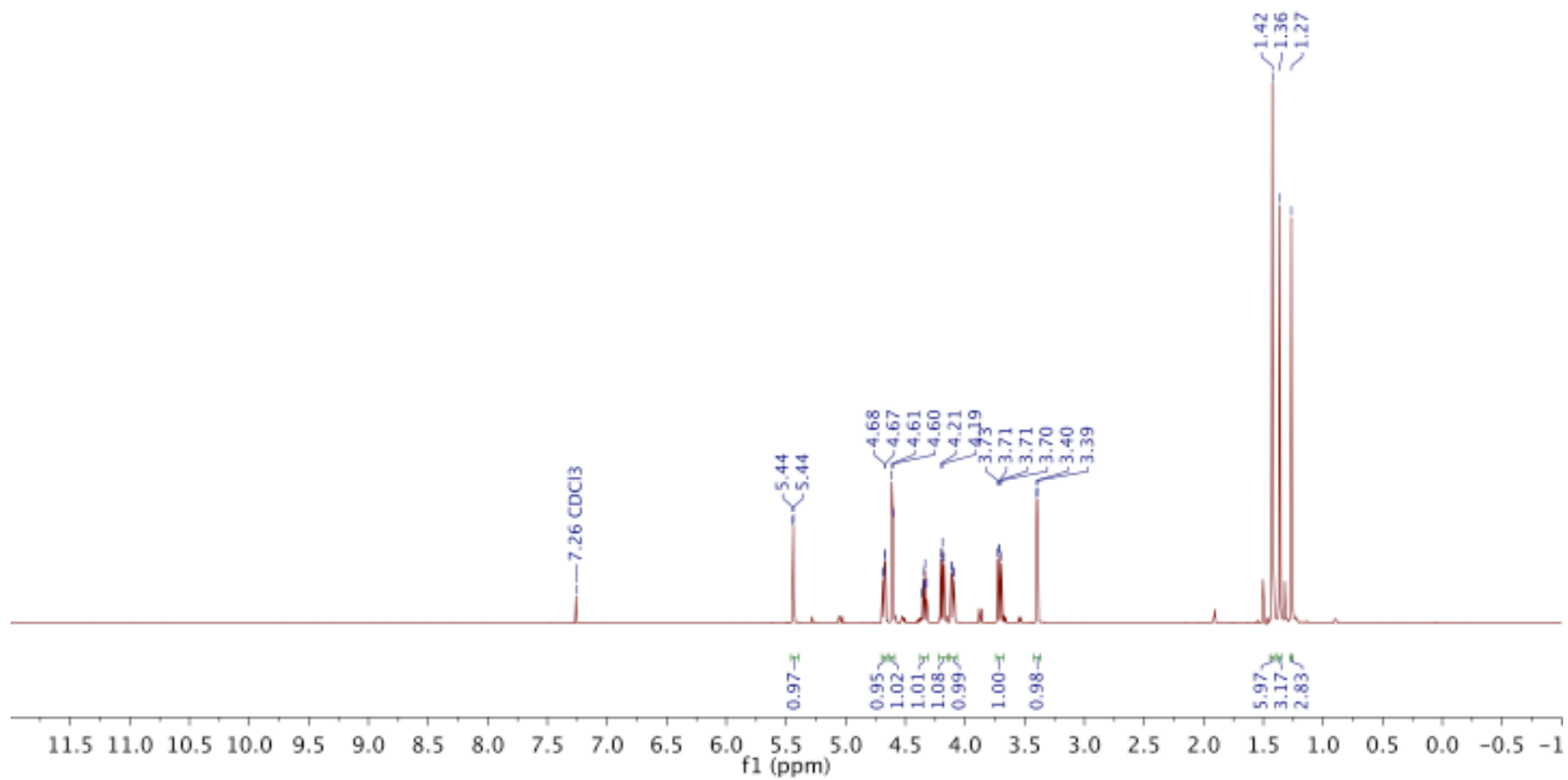
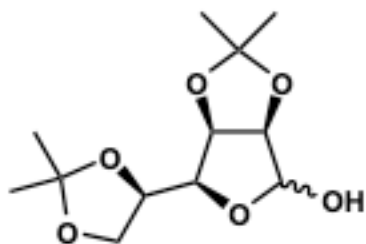


*Impurities present in spectrum

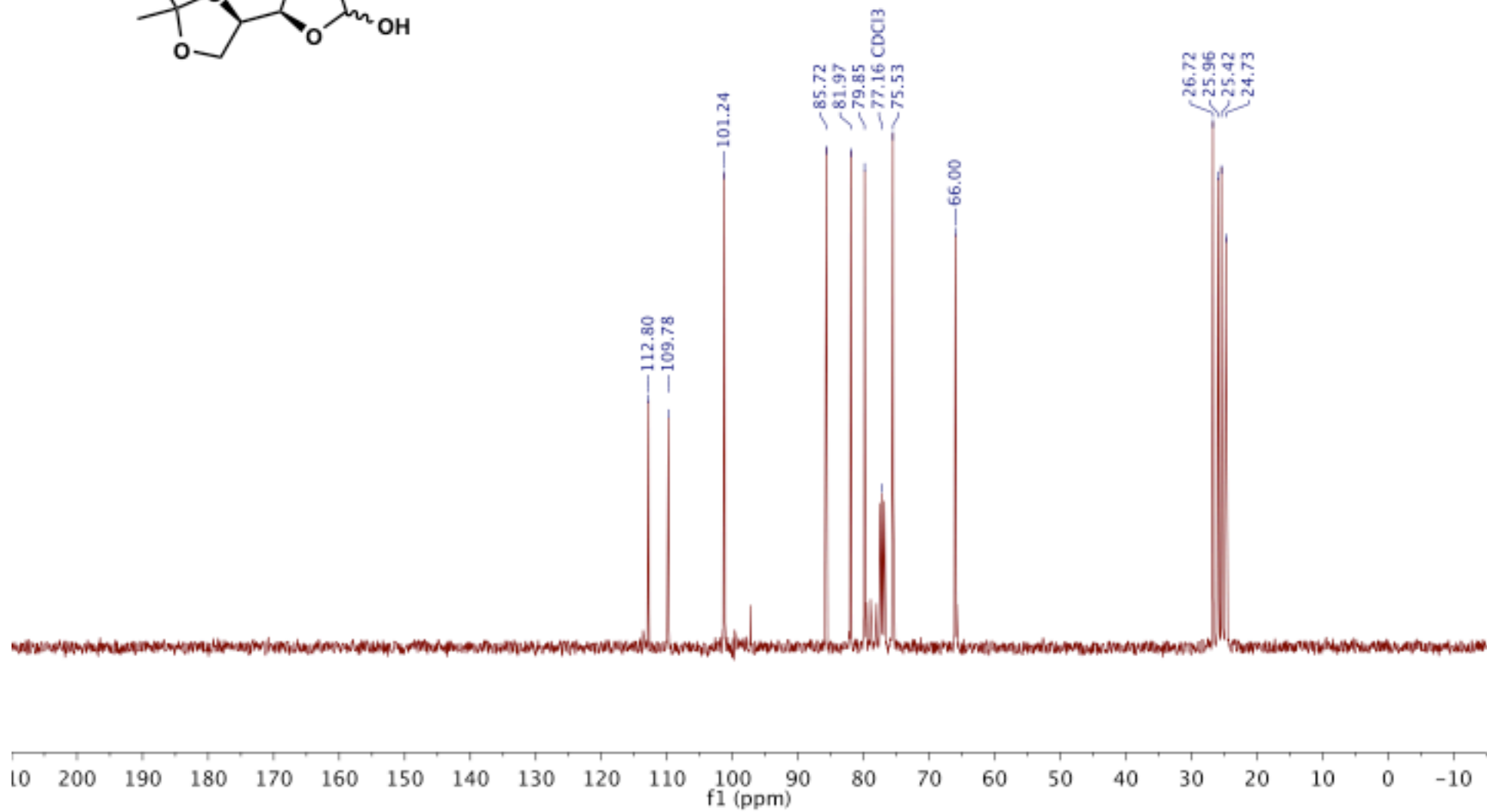
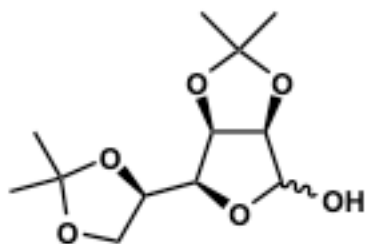
12: ¹³C NMR (126 MHz, CDCl₃)



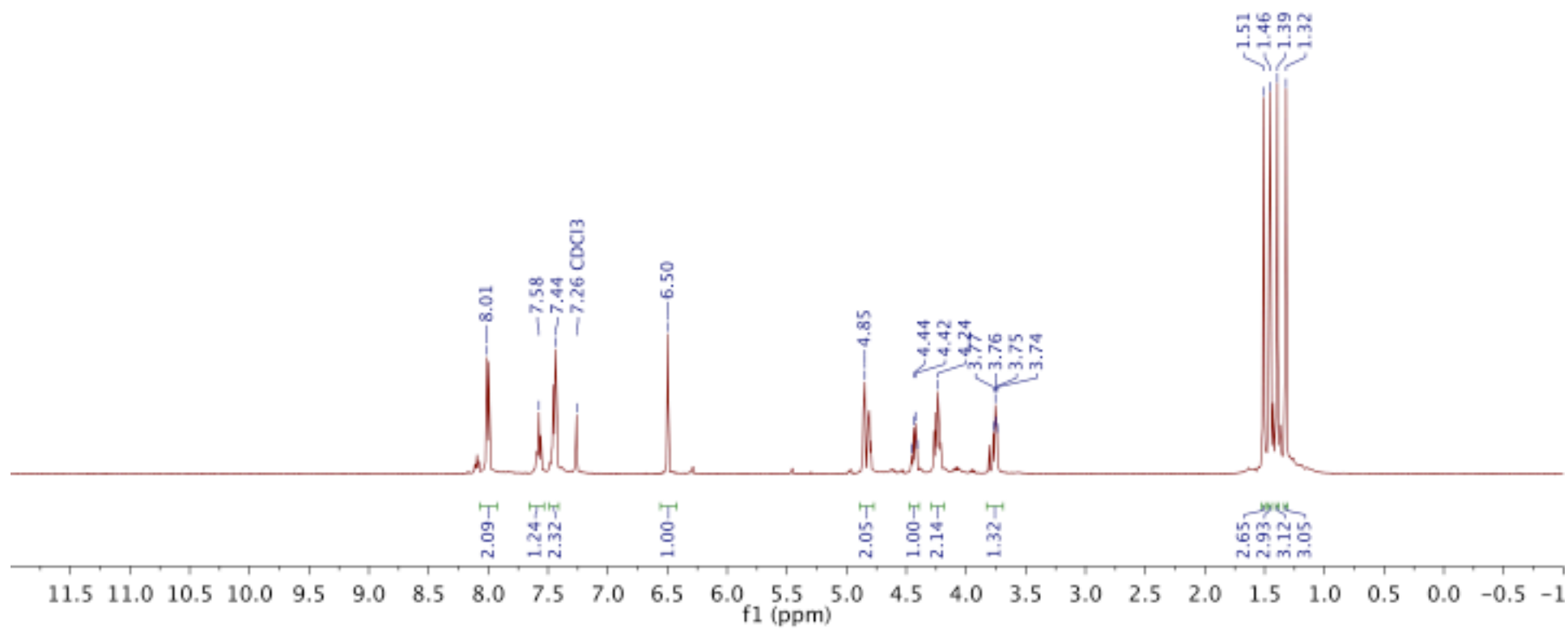
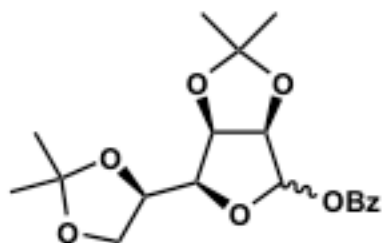
13: ^1H NMR (500 MHz, CDCl_3)



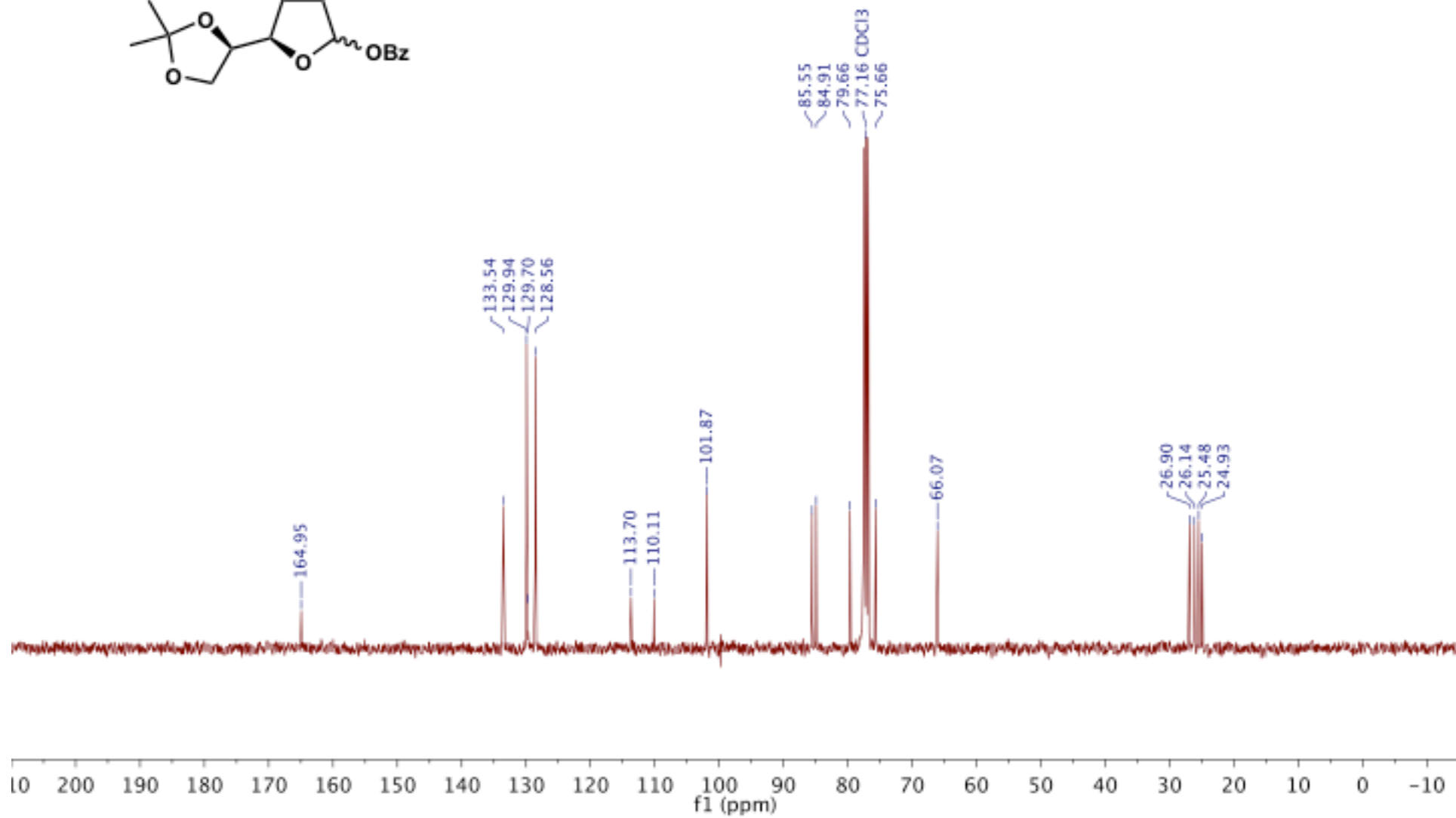
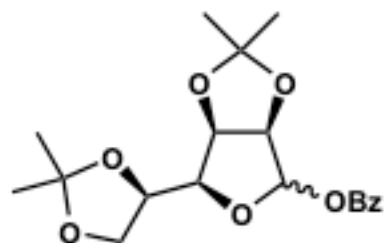
13: ^{13}C NMR (126 MHz, CDCl_3)



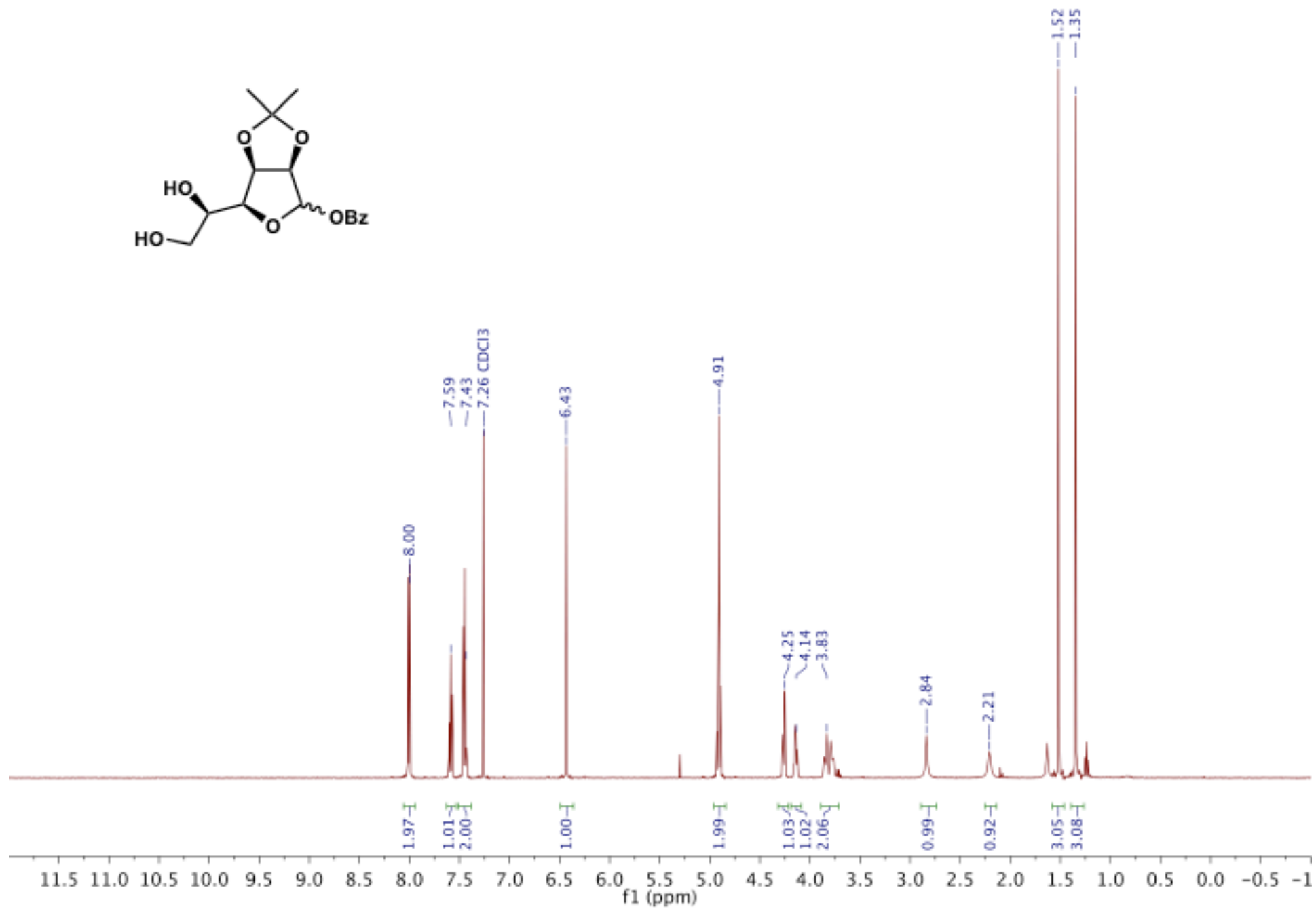
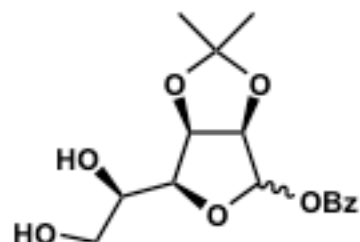
14: ¹H NMR (500 MHz, CDCl₃)



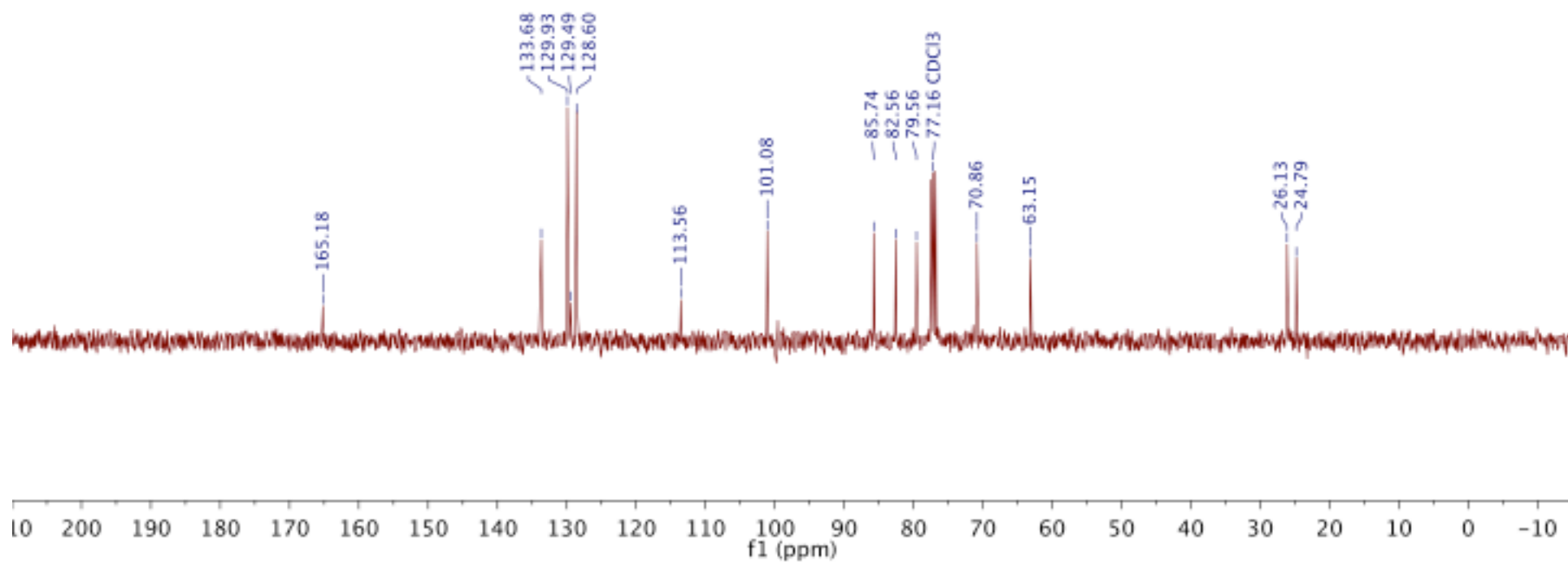
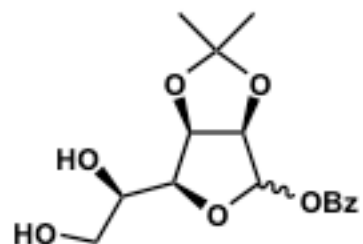
14: ^{13}C NMR (126 MHz, CDCl_3)



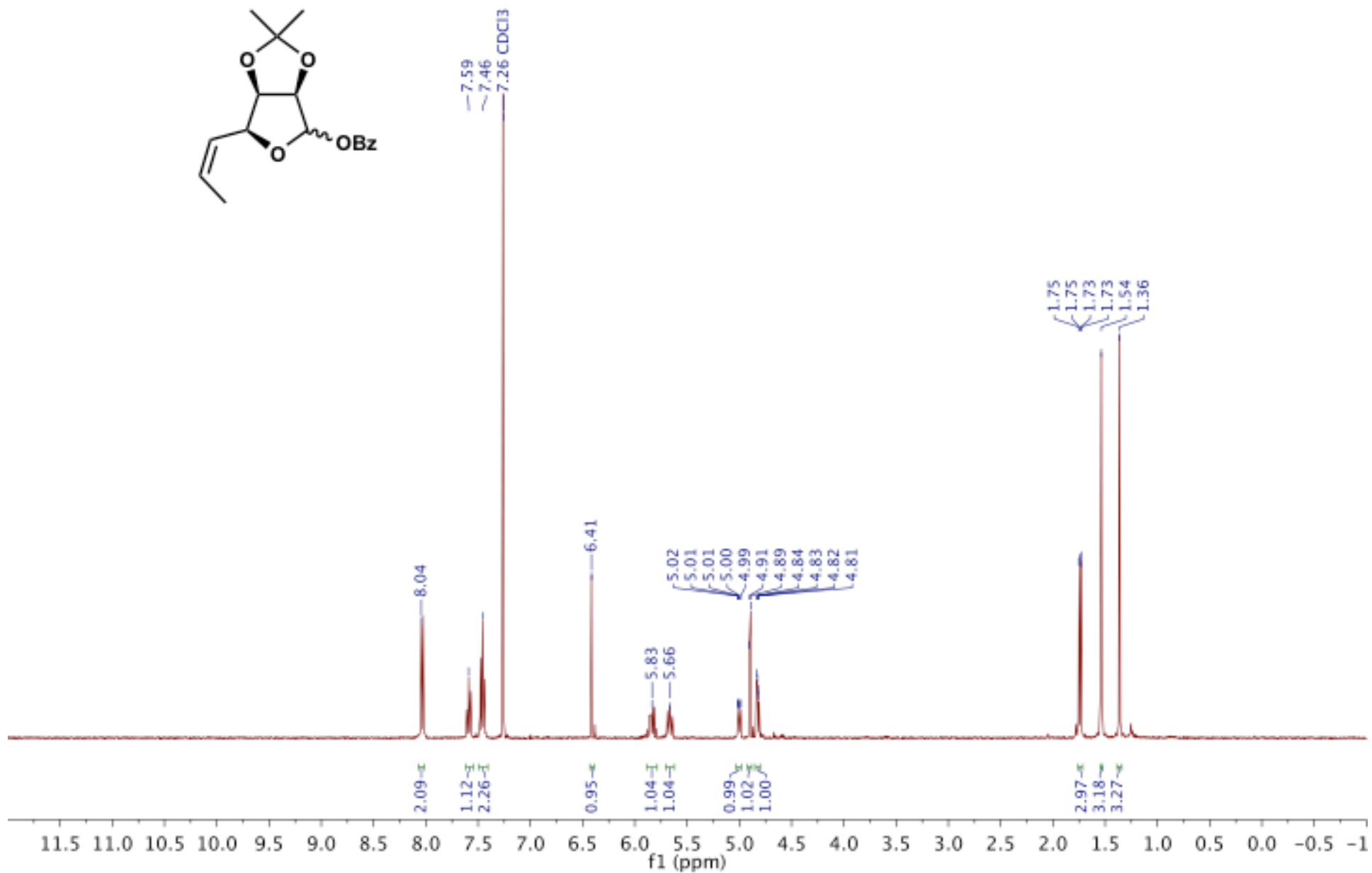
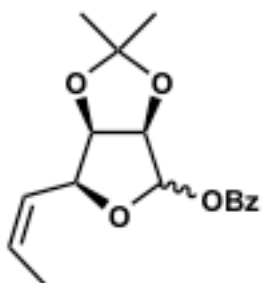
15: ^1H NMR (500 MHz, CDCl_3)



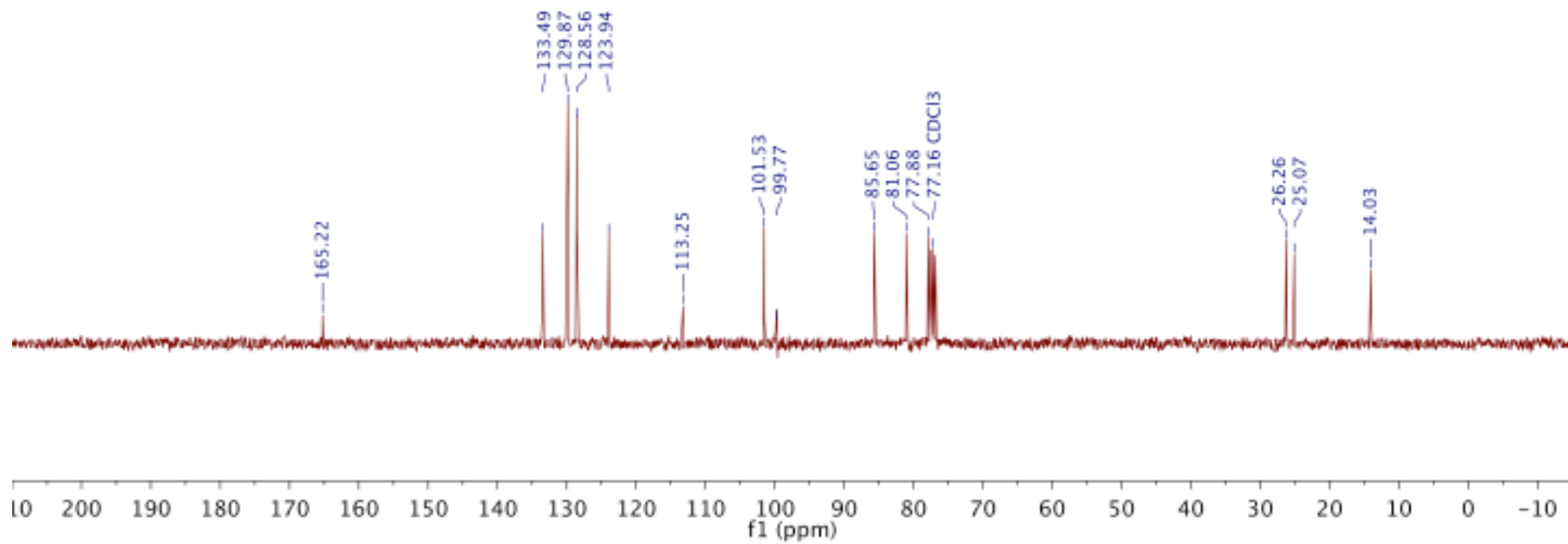
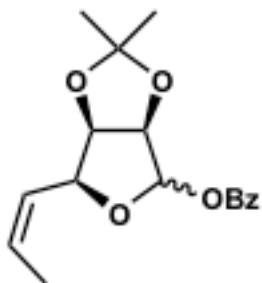
15: ^{13}C NMR (126 MHz, CDCl_3)

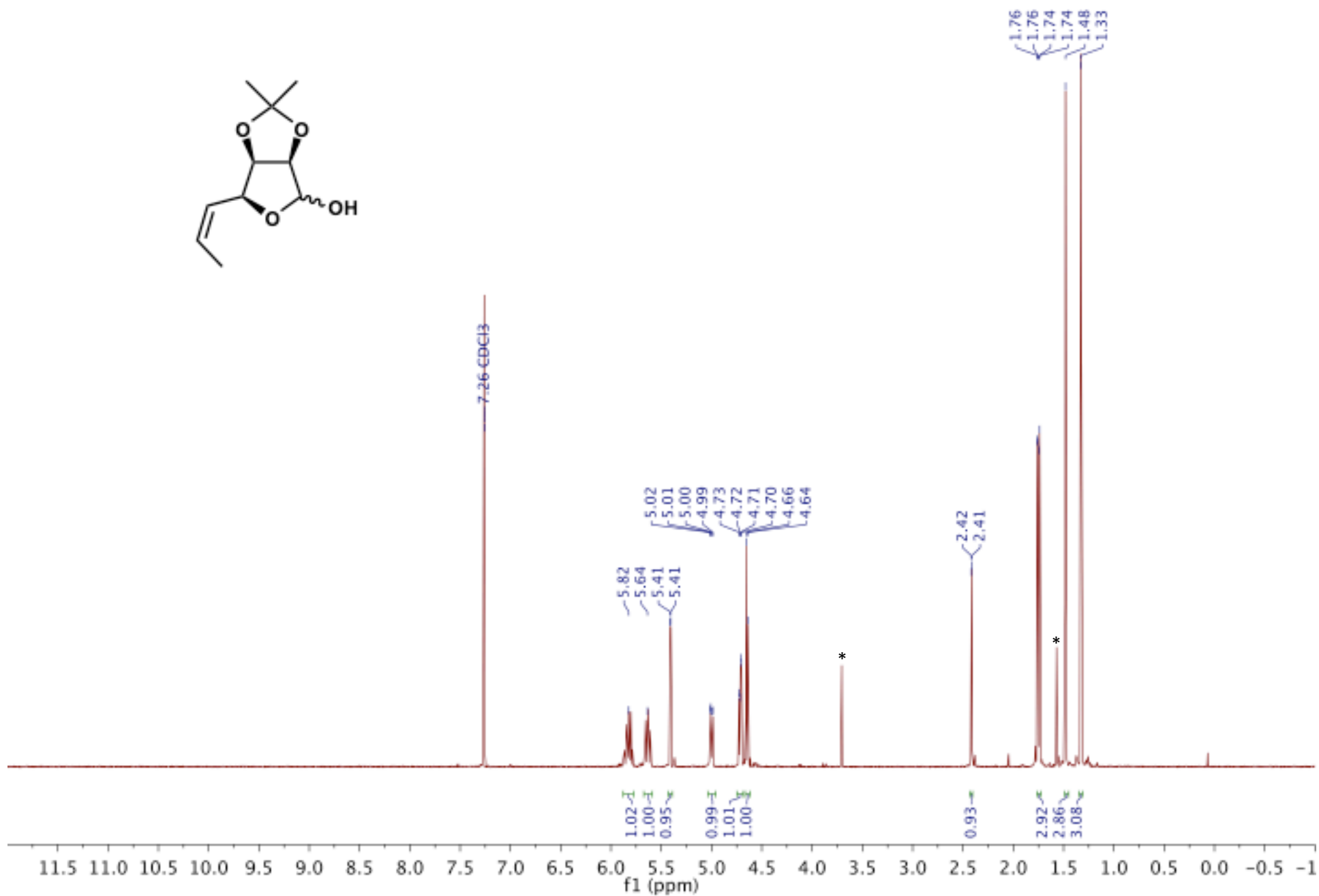
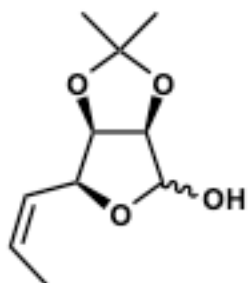


1: ¹H NMR (500 MHz, CDCl₃)

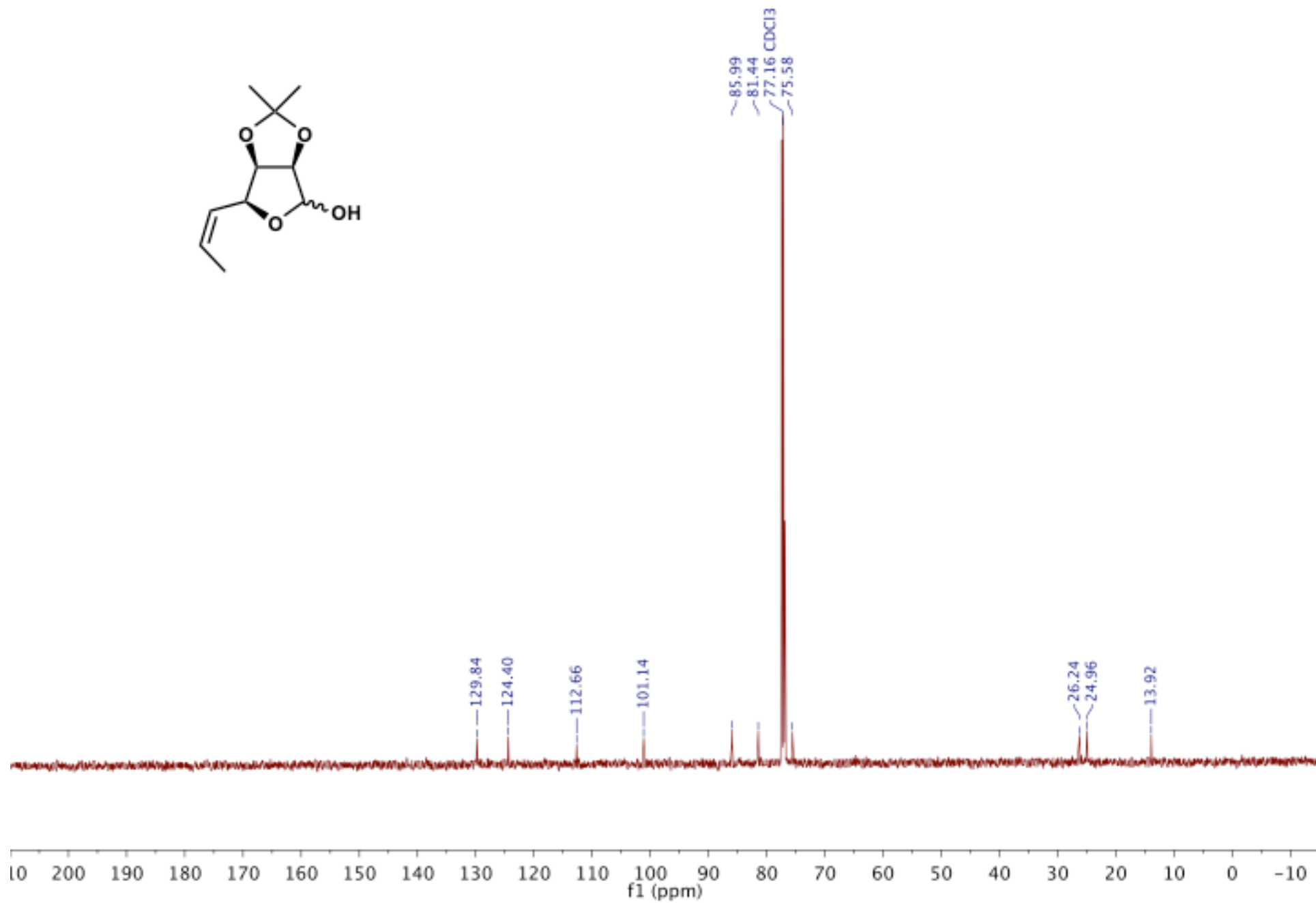
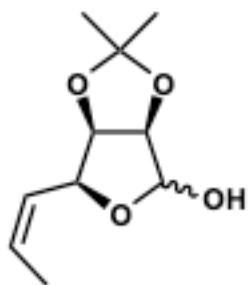


1: ^{13}C NMR (126 MHz, CDCl_3)

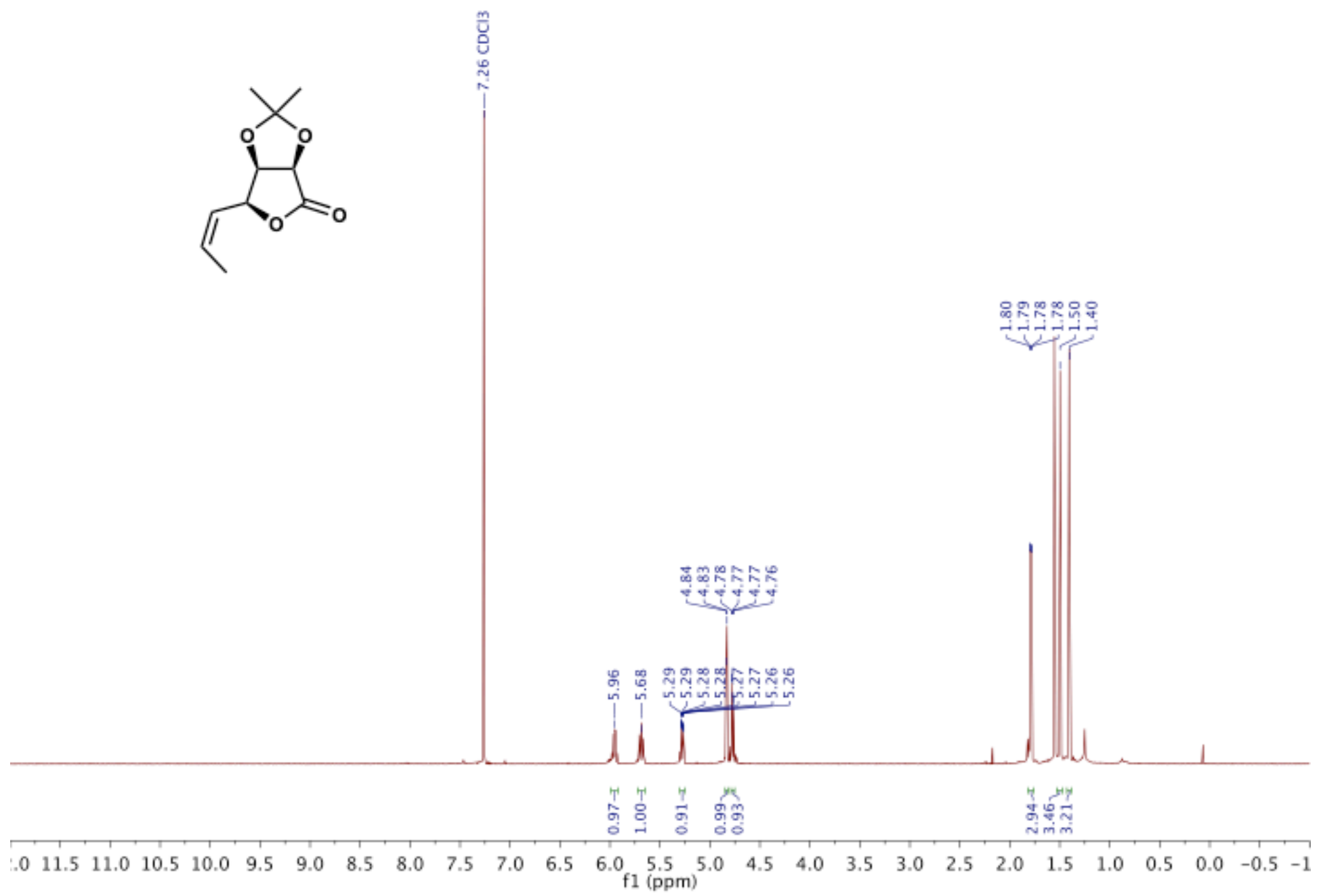
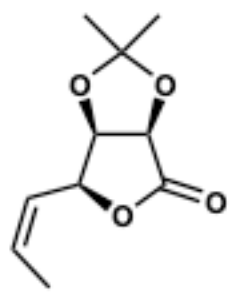




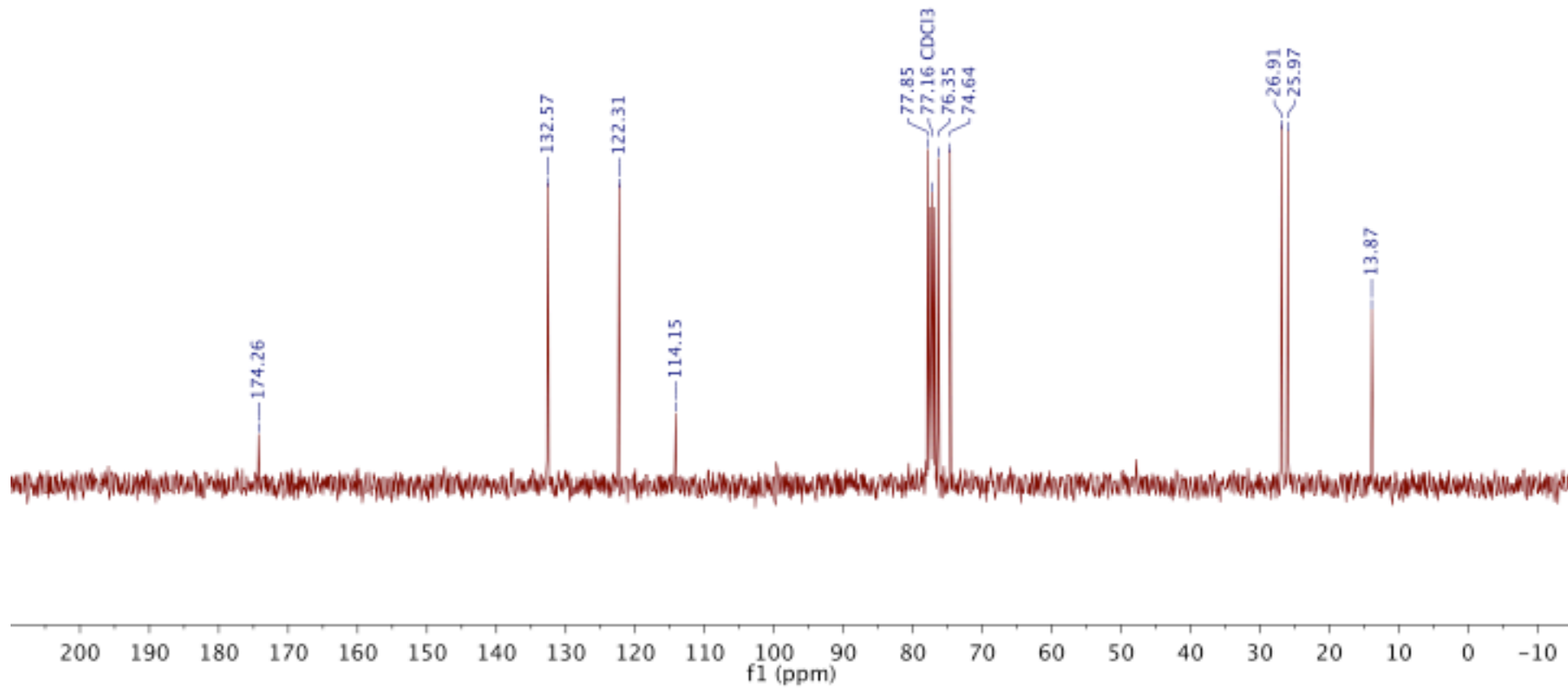
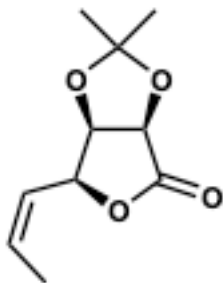
8: ^{13}C NMR (126 MHz, CDCl_3)



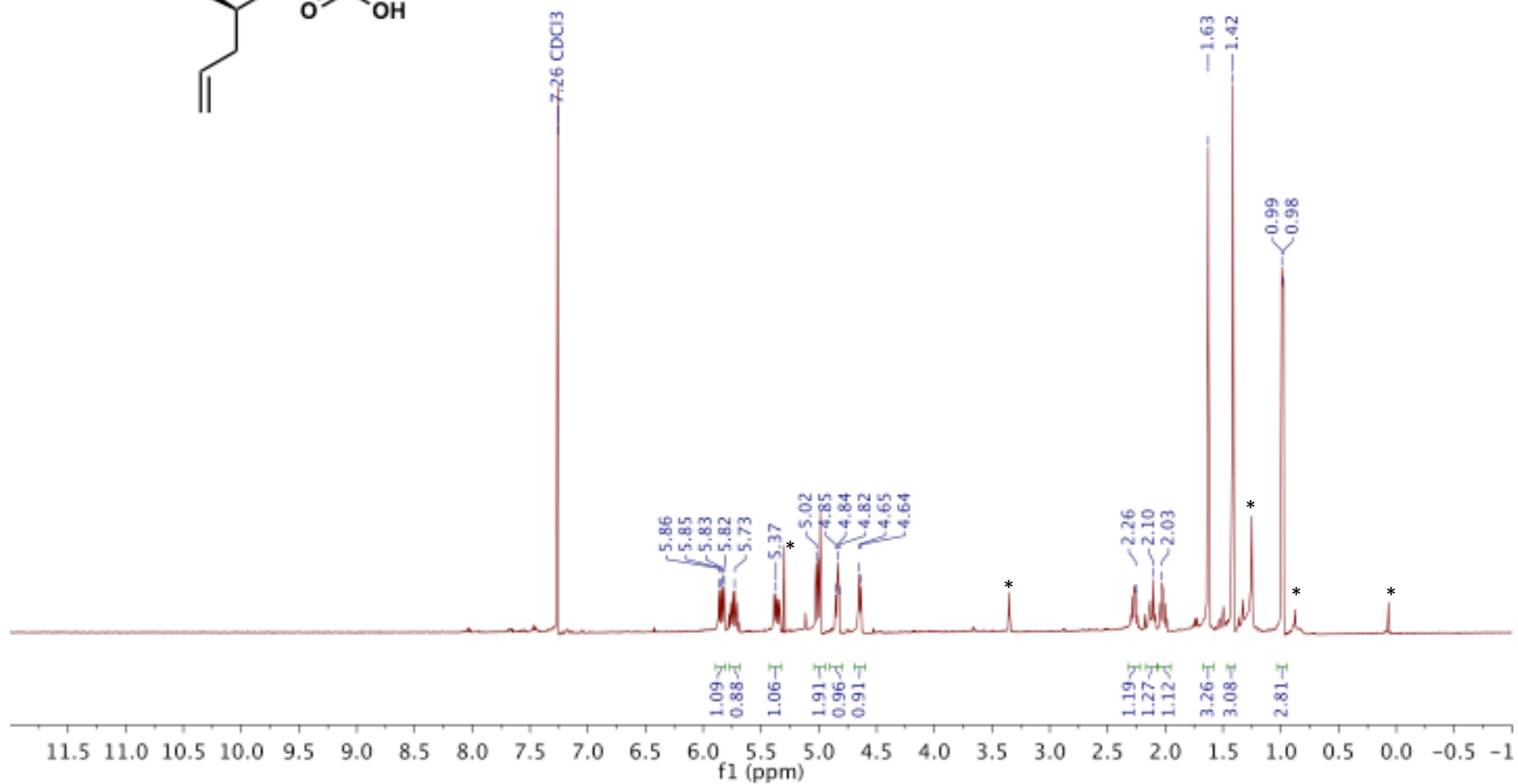
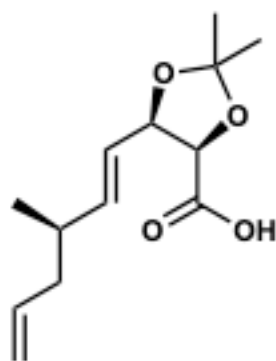
2: ¹H NMR (500 MHz, CDCl₃)



2: ^{13}C NMR (126 MHz, CDCl_3)

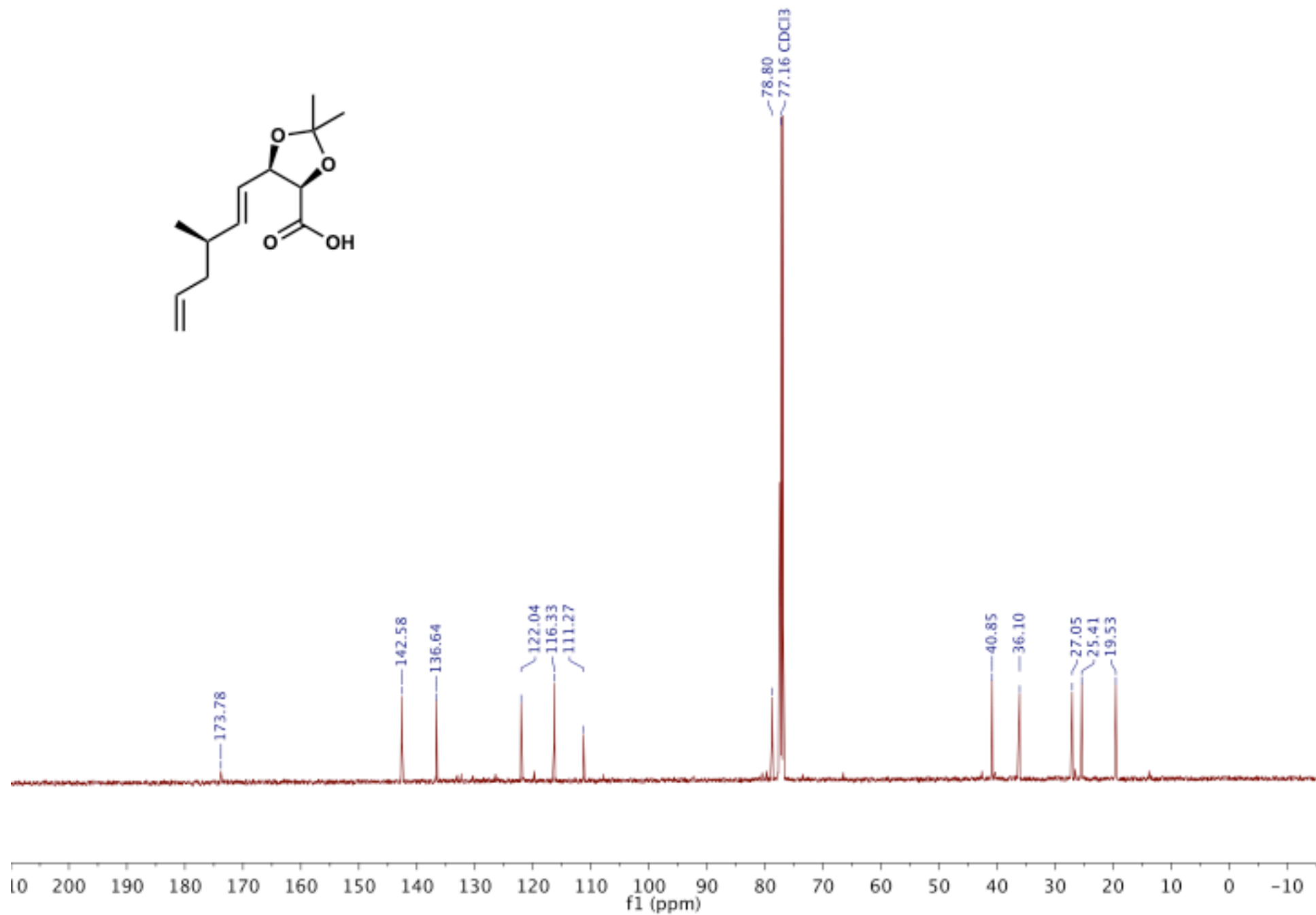
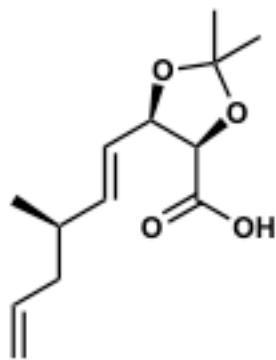


3: ¹H NMR (500 MHz, CDCl₃)

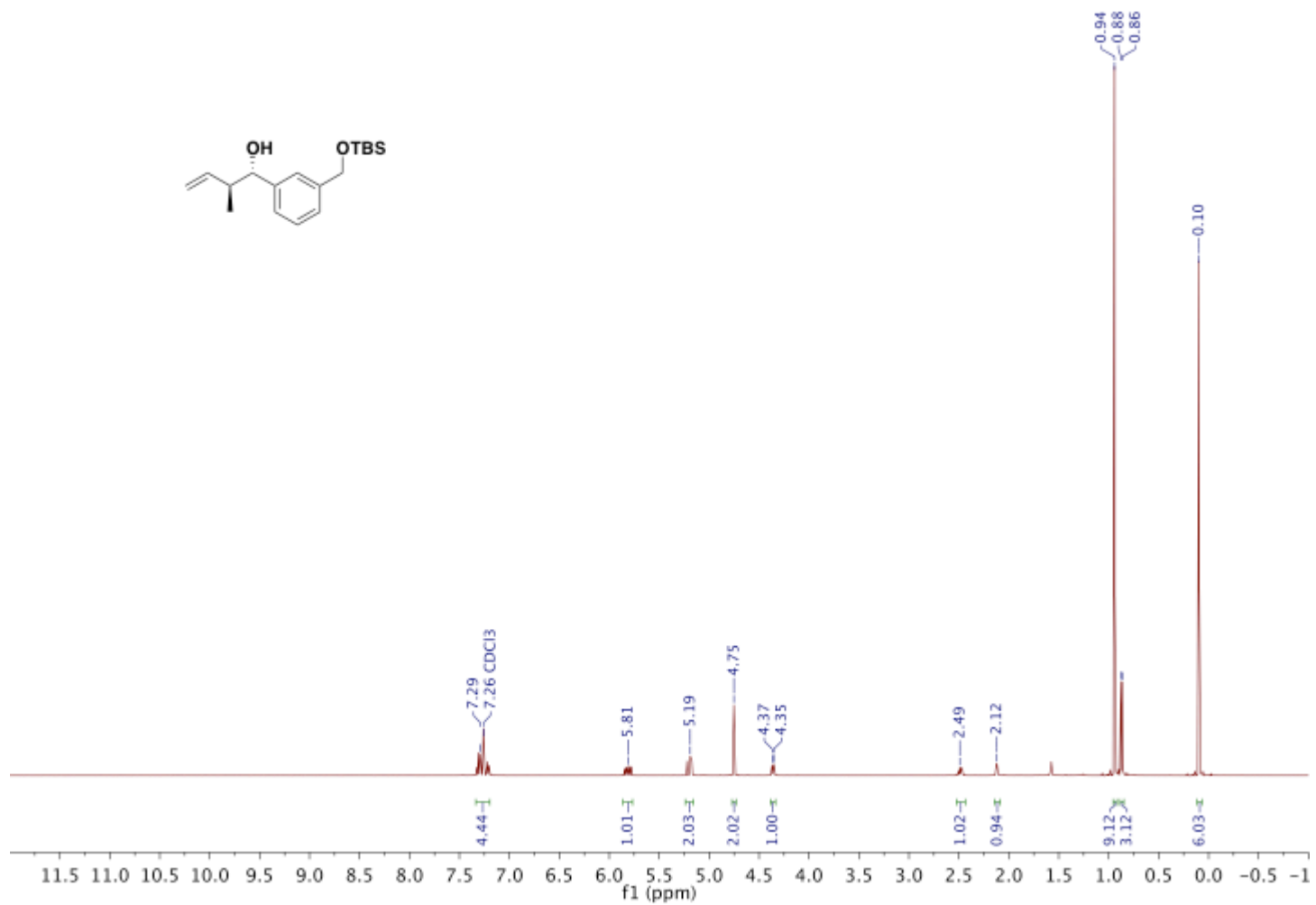


*Impurities present in spectrum

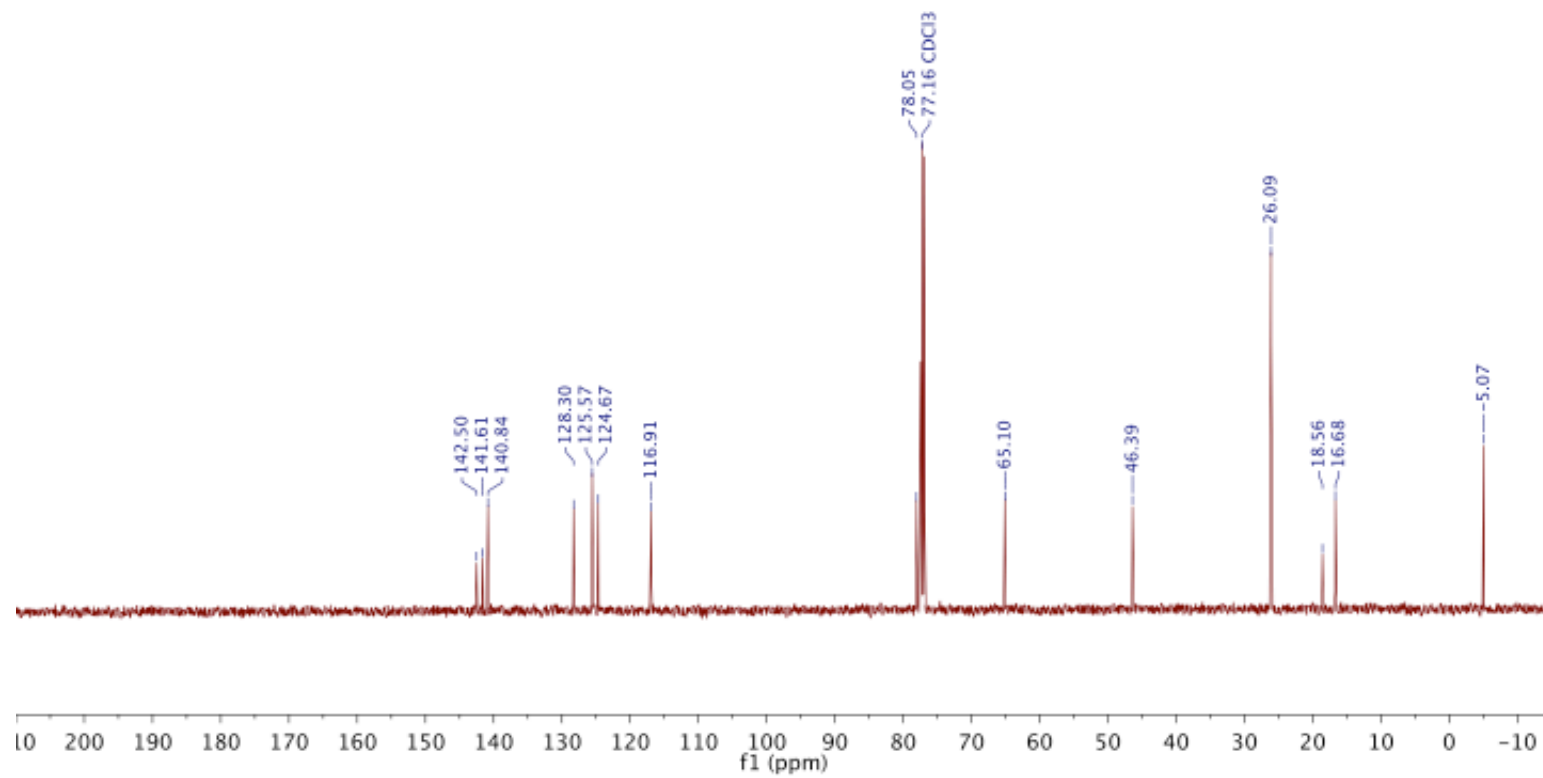
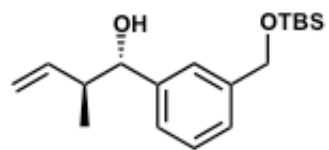
3: ^{13}C NMR (126 MHz, CDCl_3)

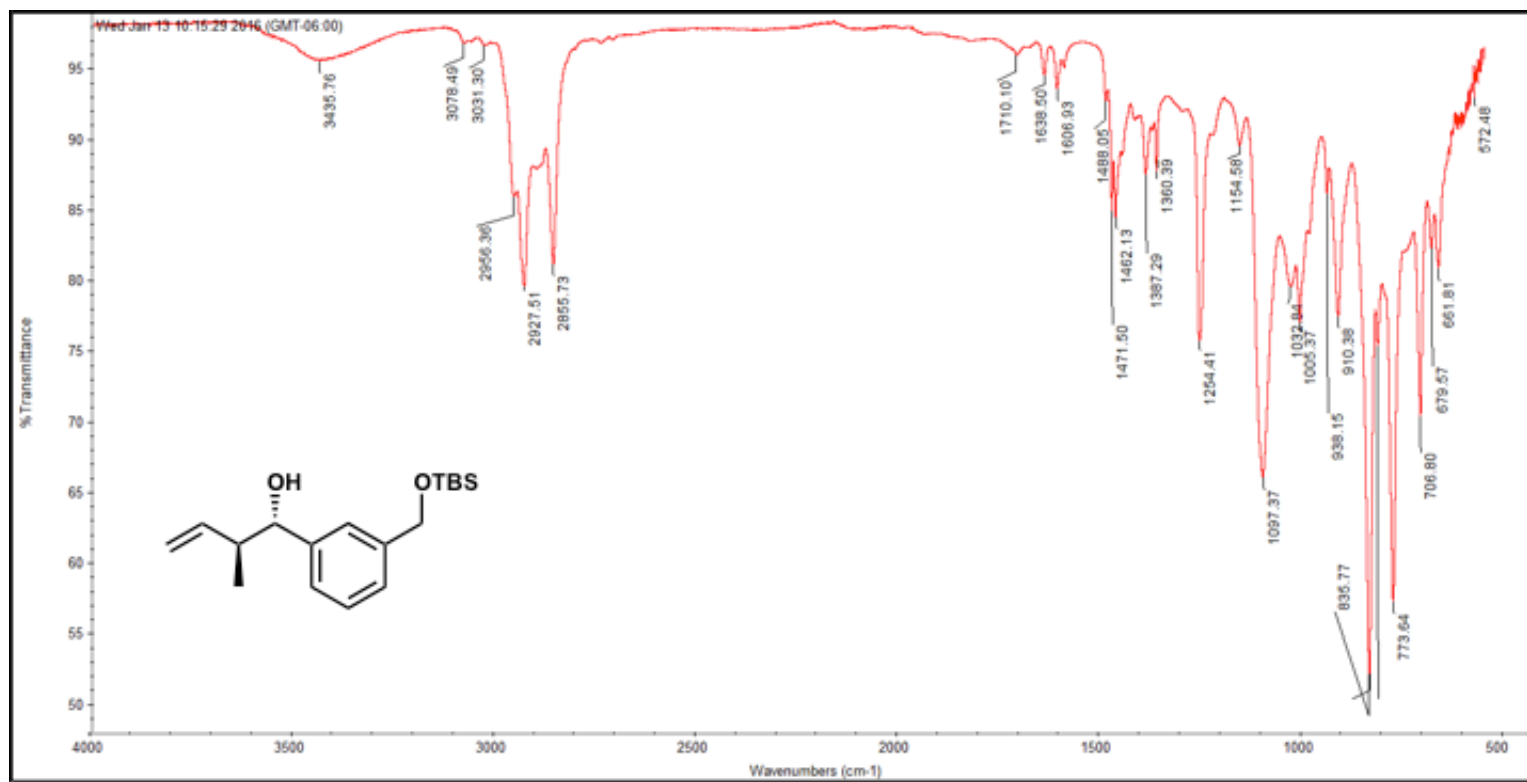


(-)-5a: ^1H NMR (500 MHz, CDCl_3)

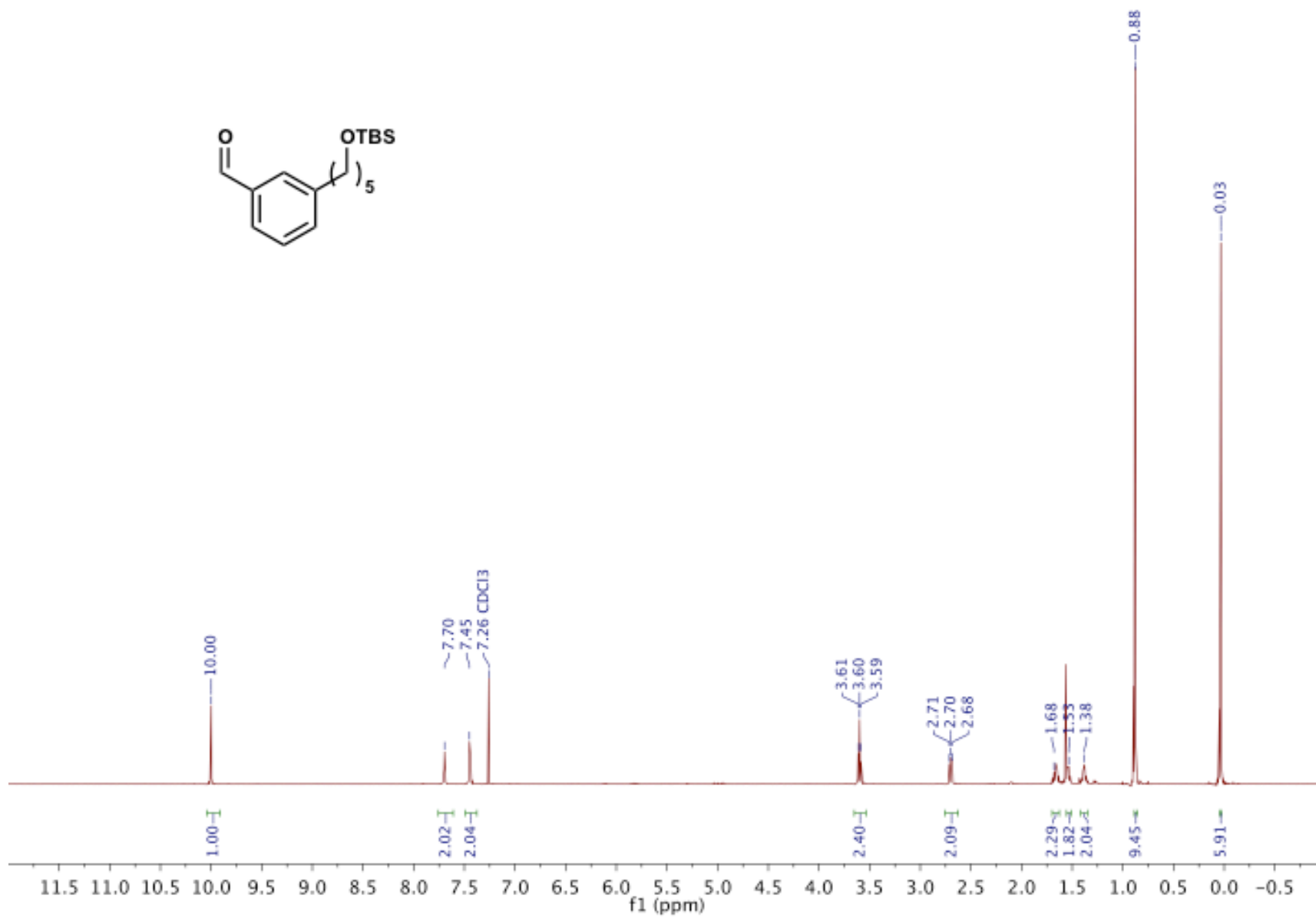
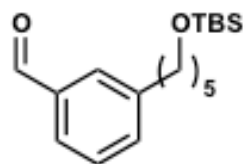


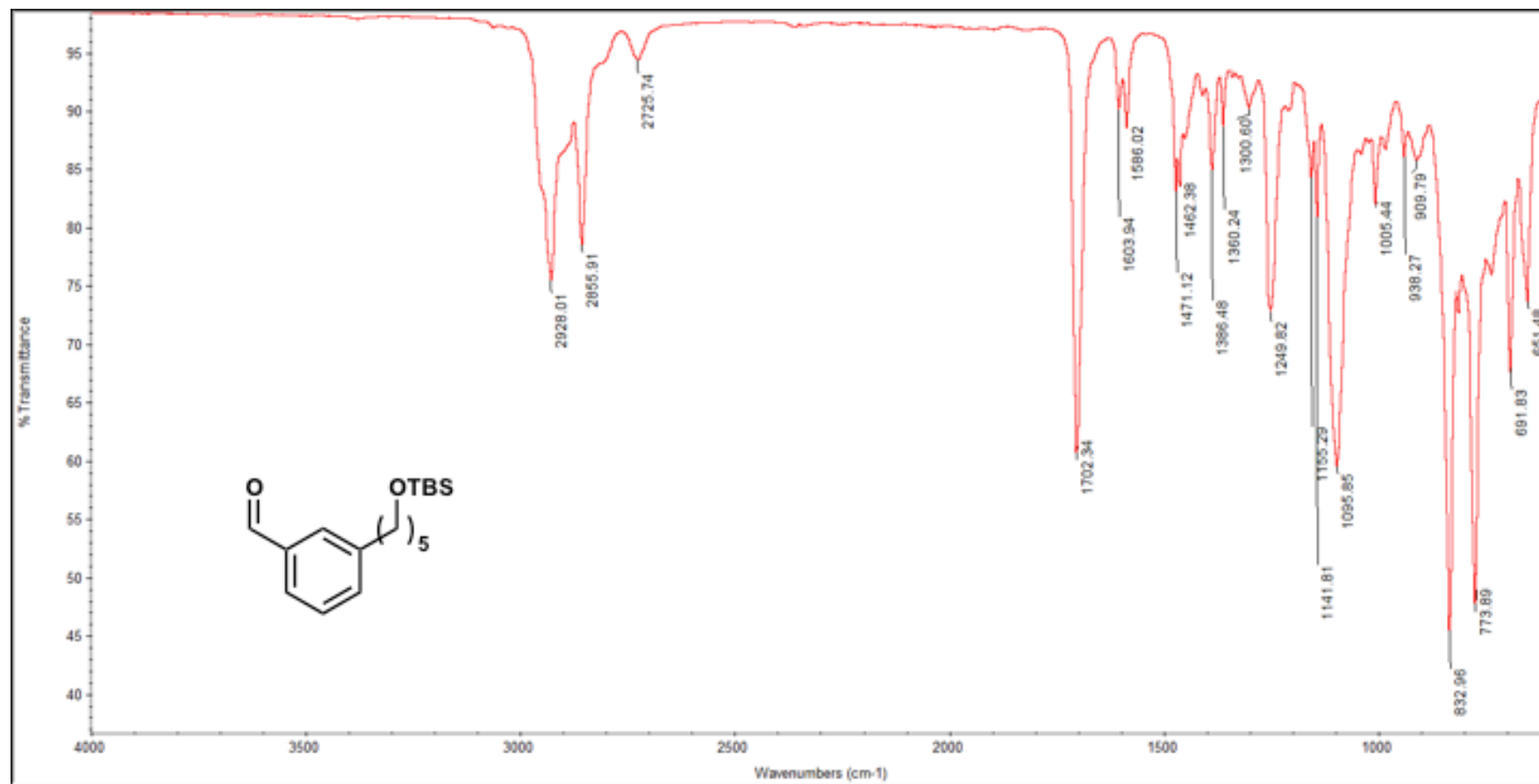
(-)-5a: ^{13}C NMR (126 MHz, CDCl_3)

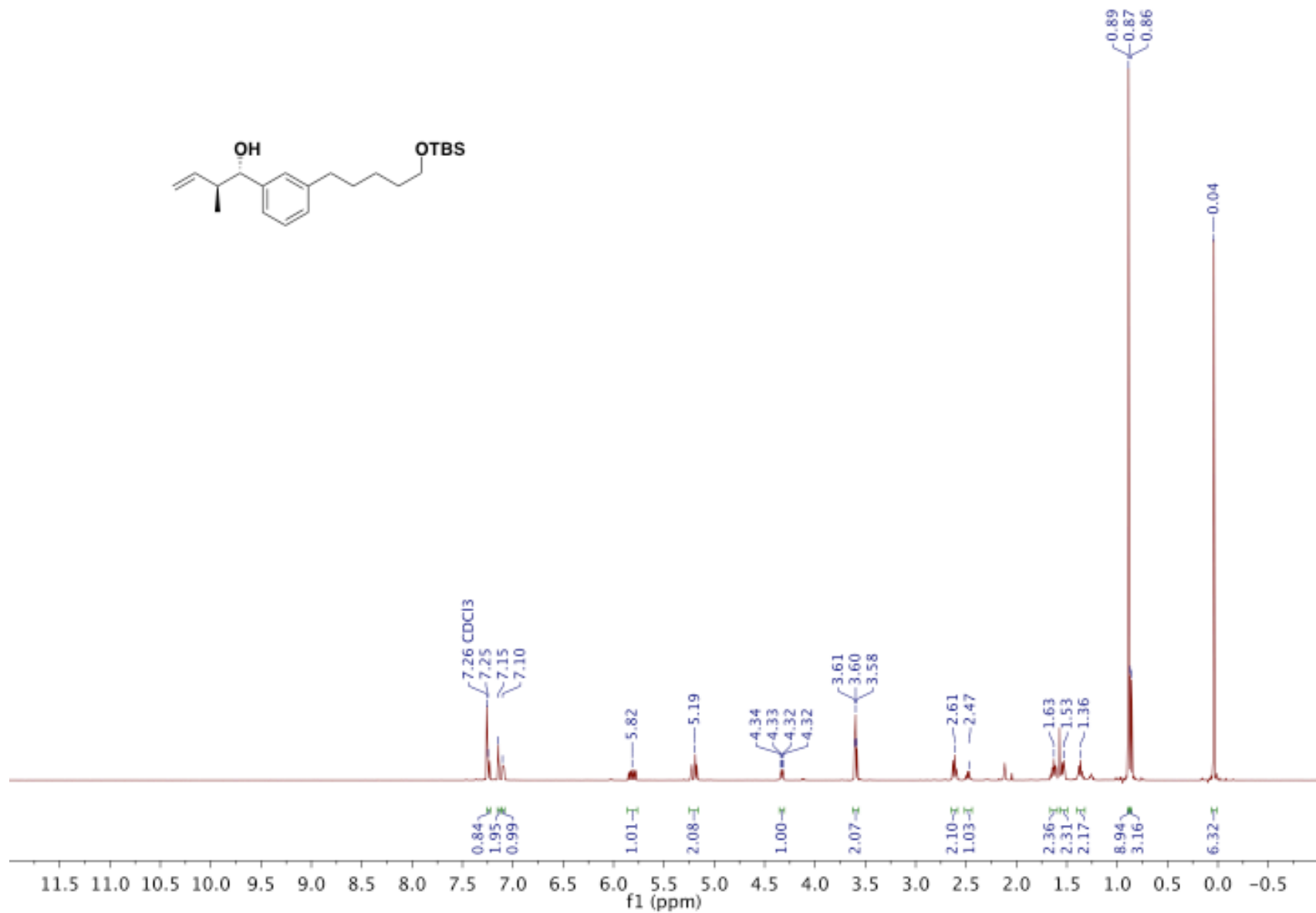
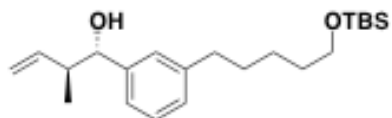




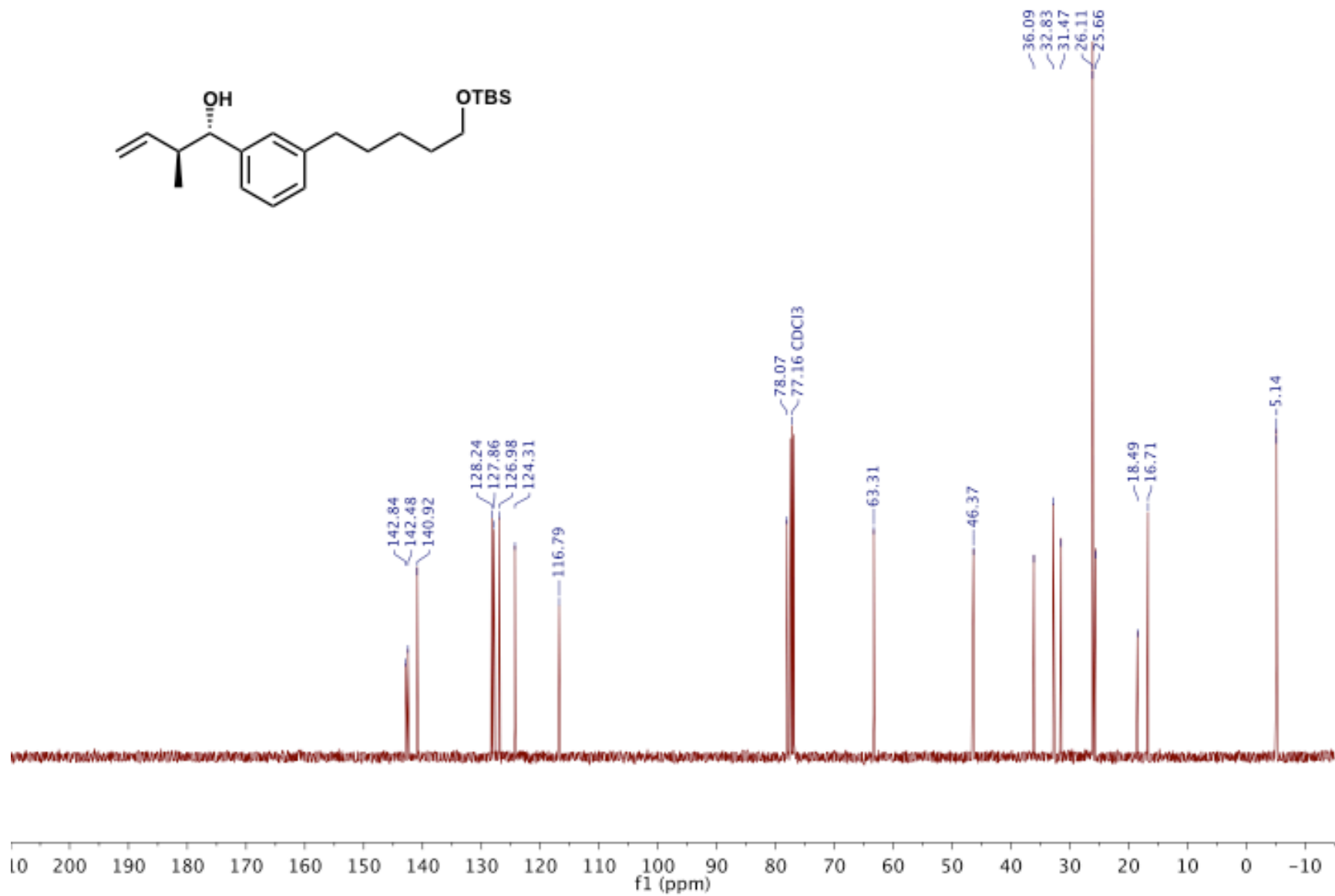
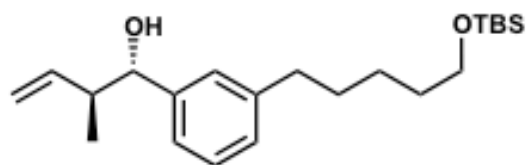
4b: ¹H NMR (500 MHz, CDCl₃)

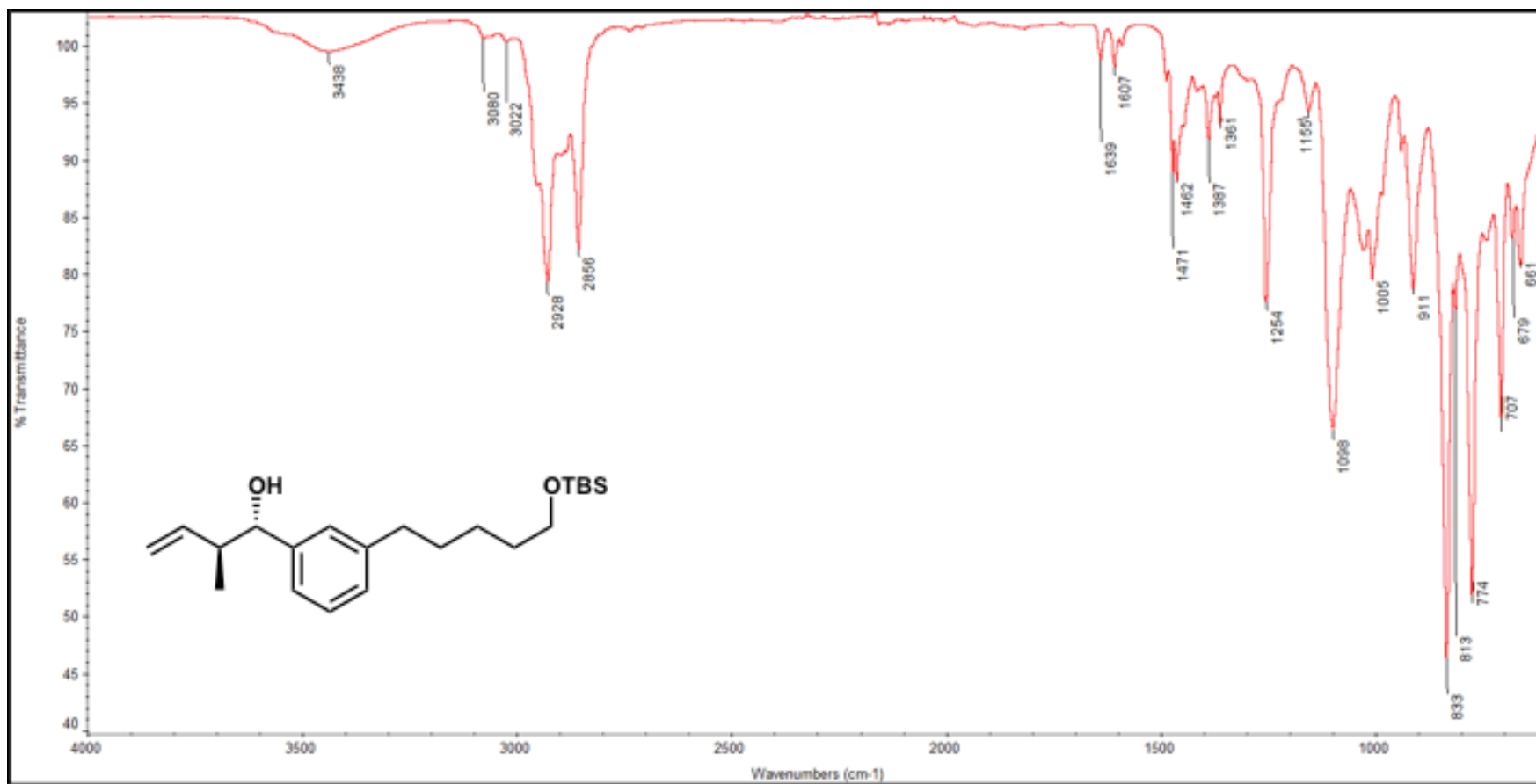






(-)-5b: ¹³C NMR (126 MHz, CDCl₃)





6. References

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