

SUPPORTING INFORMATION

Tuning Inner-Ear Tip-Link Affinity Through Alternatively Spliced Variants of Protocadherin-15

Yoshie Narui and Marcos Sotomayor*

The Ohio State University, Department of Chemistry and Biochemistry, Columbus, Ohio 43210

Figure S1. Expression and purification of pcdh15 variants.

Figure S2. Purification and validation of cdh23-pcdh15(N2) complex.

Figure S3. MALS analysis verifies monomeric state of pcdh15(N5).

Figure S4. Analytical SEC analysis of pcdh15 variants and complexes.

Figure S5. Unbinding trajectory paths for canonical and noncanonical complexes.

Figure S6. Scheme depicting potential pcdh15-pcdh15 dimers that may interact with cdh23.

Table S1. Summary of SMD simulations.

VARIANT

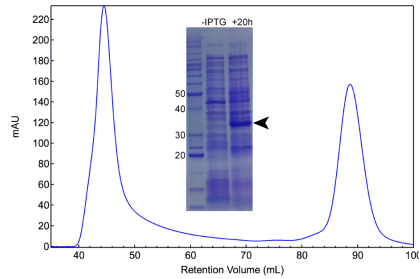
AMINO ACID SEQUENCE

SEC/SDS GEL

N1

QYDDDWQYEDCKLARGGPPATIVAID
EESRNGTILVDNMLIKGTAGGPDPTIE
 LSLKDNVDYVWLLDPVKQMLFLNSTG
 RVLDRDPPMNIHSIVVQVCVNKKVVG
 TVIYHEVRIVVRDRNDNSPTFKHESYY
 ATVNELTPVGTTFITGFGSDNGATDID
 DGPNGQIEYVIQYNPEDPTSNDTFEIP
 LMLTGNVVLKRLNYEDKTRYYVIIQA
 NDRAQNLNERRTTTTTLTVD

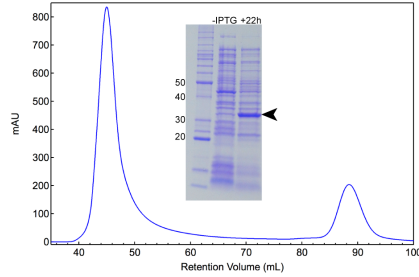
UniProt entry:
 Q99PJ1-1
 Accession no.:
 AAG53891
 MW=26.2 kDa;
 233 aa



N2

QYDDCKLARGGPPATIVAIDEESRN
 GTILVDNMLIKGTAGGPDPTIELSLKD
 NVDYVWLLDPVKQMLFLNSTGRVLD
 RDPPMNIHSIVVQVCVNKKVGTVIY
 HEVRIVVRDRNDNSPTFKHESYYATV
 NELTPVGTTFITGFGSDNGATDIDDGP
 NGQIEYVIQYNPEDPTSNDTFEIPLML
 TGNVVLKRLNYEDKTRYYVIIQAND
 RAQNLNERRTTTTTLTVD

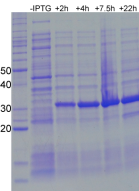
UniProt entry:
 Q99PJ1-2
 Accession no.:
 DQ354396
 MW=25.5 kDa;
 228 aa



N3

QYDDGTILVDNMLIKGTAGGPDPTIELS
 LKDNVDYVWLLDPVKQMLFLNSTGRV
 LDRDPPMNIHSIVVQVCVNKKVGTVI
 YHEVRIVVRDRNDNSPTFKHESYYATV
 NELTPVGTTFITGFGSDNGATDIDDGP
 NGQIEYVIQYNPEDPTSNDTFEIPLMLT
 GNVVLKRLNYEDKTRYYVIIQANDRA
 QNLNERRTTTTTLTVD

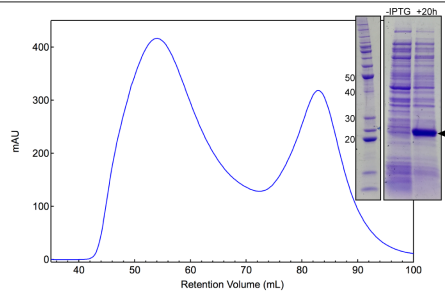
UniProt entry:
 Q99PJ1-8
 Accession no.:
 DQ354402
 MW=23.3 kDa;
 206 aa



N4

QYDDCKLARGGPPATIVAIDEESRN
 GTILVDNMLIKGTAGGPDPTIELSLKD
 NVDYVWLLDPVKQMLFLNSTGRVLD
 RDPPMNIHSIVVQVCVNKKVGTVIY
 HEVRIVVRDRNDNSPTFKHESYYATV
 NELTPVGTTFITGFGSDNGATDIDDGP
 NGQIEYVIQYNPEDPDRAQNLNERRT
 TTTTTLVD

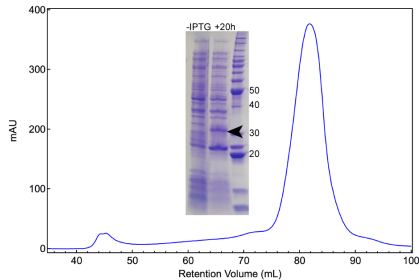
UniProt entry:
 Q99PJ1-7
 Accession no.:
 DQ354401
 MW=21.1 kDa;
 191 aa



N5

MTMAEQDNGHPLPAFASLHIEILDEN
 NQSPYFTMPYQGYILESAPVGTATISE
 SLNLTPLRIVALDKDIEDVPPGGVPT
 KDPELHLFLNDYTSVFTVTPTGITRYL
 TLLQPVDREEQQTYYFLITAFDGVQE
 SEPVVVNIQVMDANDNTPTTFPEISYD
 VYVYDMSPGDSVIQLTAVDADEGSN
 GEISYEILVGGKGFVINKTGLVSIAP
 GVELIVGQTYALTVAQSDNAPPAERR
 HSICTVYIEVLPNNQSPPRF

Accession no.:
 DQ354407
 MW=28.3 kDa;
 260 aa



N6

QYDDHSICTVYIEVLPNNQSPPRFP
 QLMYSLEVSEAMRIGAILNLQATDRE
 GDPITYAIENGDPQRFNLSETTGILS
 LGKALDRESTDRIYLIVTASDGRPDGT
 STATVNIIVTDVNDNAPVFDPLYLPRNL
 SVVEEENAFVGVQVRATDPDAGING
 QVHYSLGNFNNLFRITSNYSIYAVKL
 NREARDHYELVVVATDGAHVPRHSTL
 TLYIKVLDIDDNSPVF

UniProt entry:
 Q99PJ1-24
 Accession no.:
 AAY24693
 MW=25.1 kDa;
 228 aa

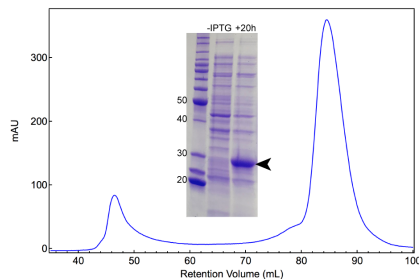


Figure S1. Expression and purification of pcdh15 variants. For each of the variants studied, the UniProt entry code, accession number, and predicted molecular weight are provided in the leftmost column. The amino acid sequence of the two N-terminal EC repeats included in the construct, excluding the start codon (Met) and the hexahistidine tag (including any additional residues due to cloning) are provided in the center column. The sequence is color coded to highlight the end of exon 2 in red, exon 3 in cyan, and exon 4 in purple. The right column shows SDS gels before and after inducing protein expression, and SEC chromatograms of refolded pcdh15 variants. Peaks of aggregated protein are expected near ~45-50 mL and homogenous, refolded two EC repeat constructs are expected at a retention volume between 80-90 mL (Superdex 200 16/600 column).

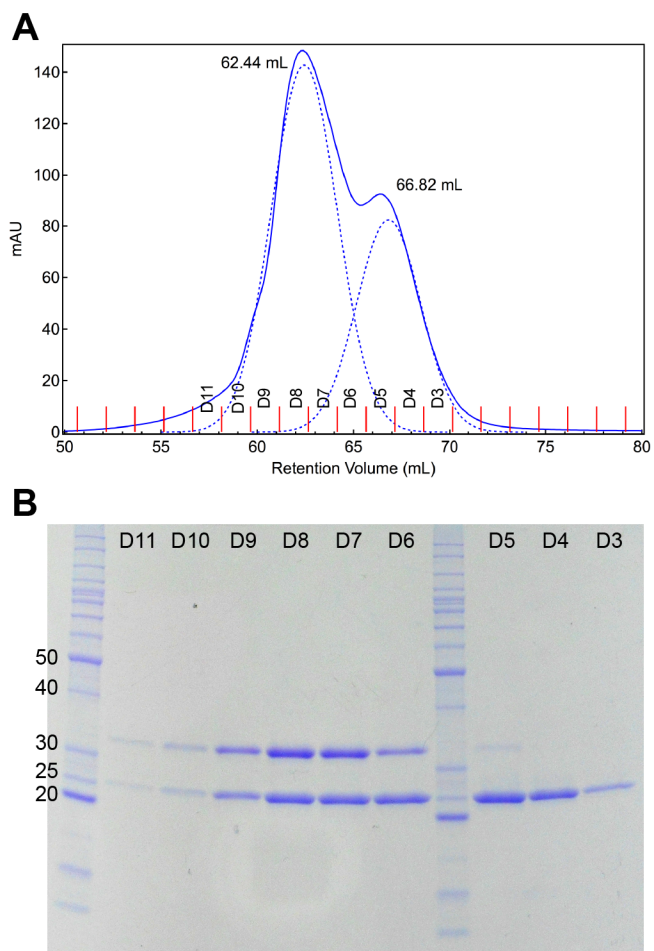


Figure S2. Purification and validation of cdh23-pcdh15(N2) complex. (A) SEC chromatogram displays a new peak at 62.44 mL, which represents the cdh23-pcdh15(N2) complex. The peak at 66.82 mL represents excess cdh23. Dotted lines are theoretical fits used to estimate retention volumes. The sample was analyzed on a Superdex 75 16/600 column. (B) Fractions from the SEC run were analyzed by SDS PAGE. Fractions D9-D6 were combined, concentrated, and utilized for crystal growth.

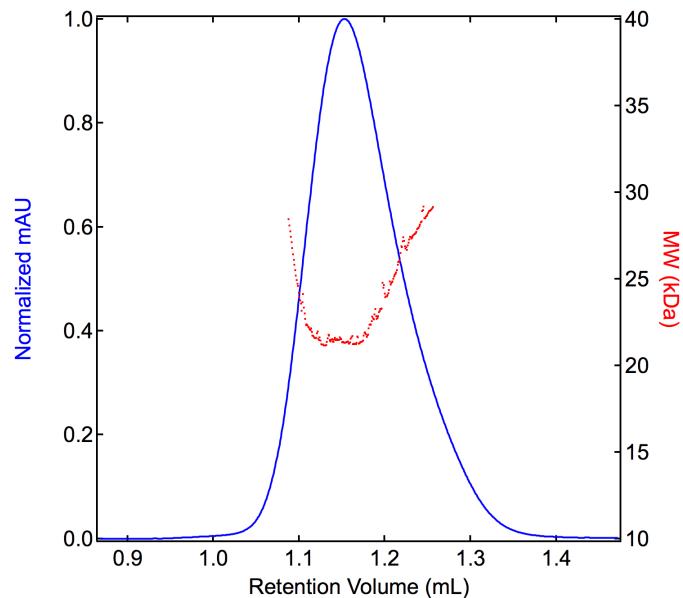


Figure S3. MALS analysis verifies monomeric state of pcdh15(N5). The refolded sample of pcdh15(N5) was analyzed on the AKTAMicro system connected in series with a Wyatt miniDAWN TREOS system. Protein absorbance at 280 nm was monitored (blue trace) as was light scattering from the sample, which was subsequently converted into molecular weight using a rod-like model (red dots). The predicted monomer mass of variant N5 is 28.3 kDa, while the measured value was ~ 26.0 kDa, which does not approach the molecular weight of a dimeric protein.

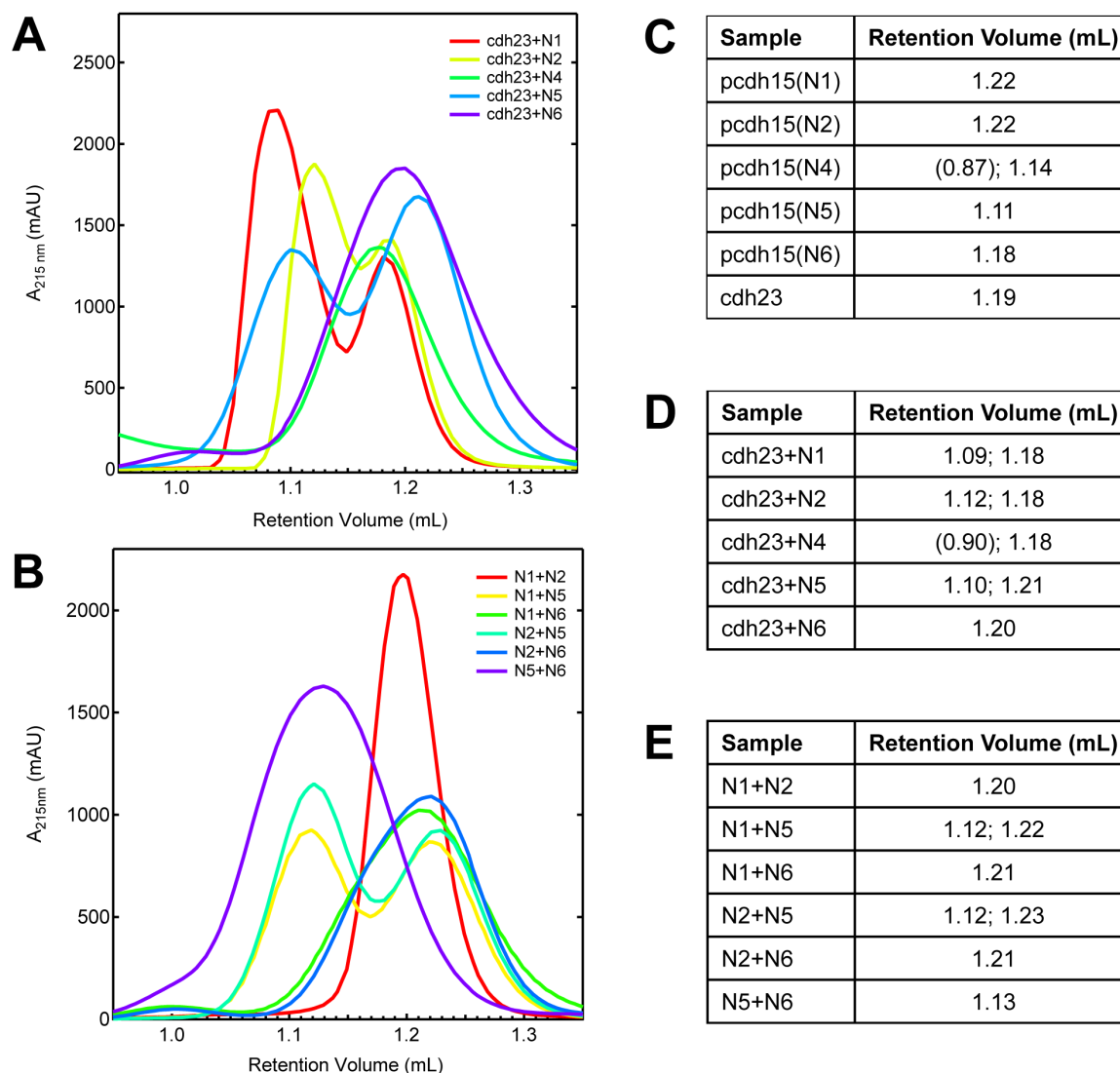


Figure S4. Analytical SEC analysis of pcdh15 variants and complexes. (A) Heterophilic mixtures of cdh23 and pcdh15. (B) Pseudo-heterophilic mixtures of pcdh15 variants. (C-E) Tables of peak retention volumes for individual proteins (C), heterophilic complexes (D), and pseudo-heterophilic complexes (E) of pcdh15 variants. Peaks in parentheses are present in the chromatogram but not visible in the plots due to the axis scale.

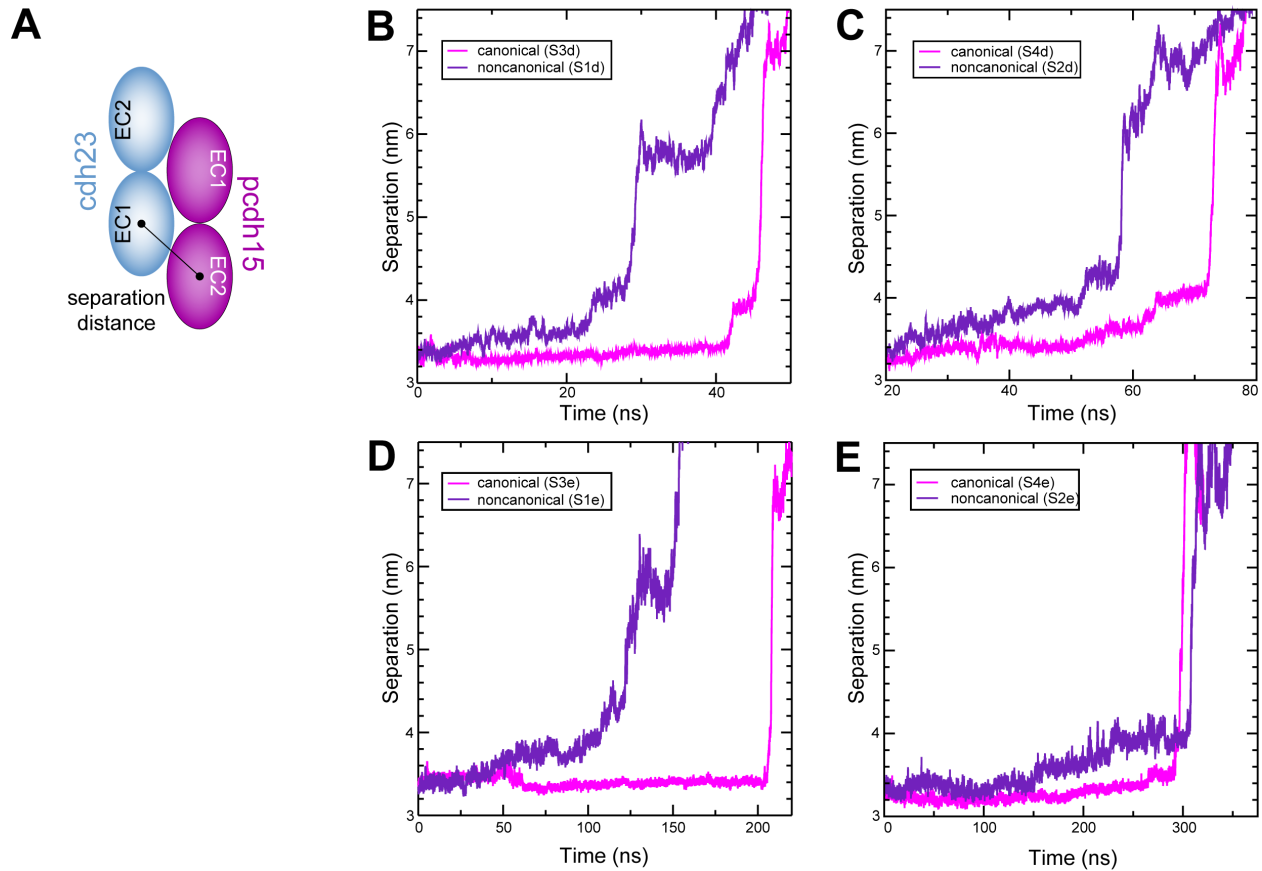


Figure S5. Unbinding trajectory paths for canonical and noncanonical complexes. (A) Separation distance between the centers of mass of cdh23 EC1 and pcdh15 EC2 plotted as a function of time for pulling at 0.1 nm/ns after the complexes were equilibrated for (B) 1 ns, and (C) 10 ns. Similar data are shown when the complexes were pulled at 0.02 nm/ns and equilibrated for (D) 1 ns, and (E) 10 ns. Simulations with low unbinding force (S1d and S1e) follow a distinct unbinding path in which EC1 of cdh23 separates earlier from pcdh15.

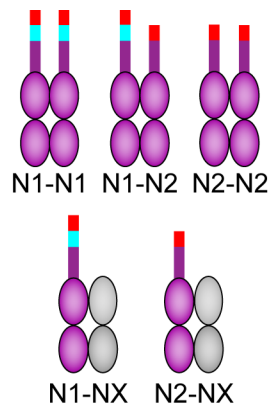


Figure S6. Scheme depicting potential pcdh15-*pcdh15* dimers that may interact with *cdh23*. Five different combinations of PCDH15 may be observed in hair cells. NX represents other *pcdh15* variants, such as N3, which could also bind with N1 or N2. These dimers could form heterotetrameric complexes with *cdh23*.

Table S1. Summary of SMD simulations.

Label	PDB	t_{sim} (ns)	Type	Start	Speed (nm/ns)	Average Peak Force (pN) ^b	Size (#atoms)	Size (nm ³)
S1a	4XXW	1	EQ ^a	–	–		153,825	27.9 x 8.5 x 7.0
S1b		2	SMD	S1a	10	1409.5		
S1c		6.5	SMD	S1a	1	669.8		
S1d		98.9	SMD	S1a	0.1	474.3		
S1e		222.3	SMD	S1a	0.02	397.1		
S2a	4XXW	20	EQ ^a	S1a	–		153,825	27.9 x 8.5 x 7.0
S2b		4	SMD	S2a	10	1259.5		
S2c		20.2	SMD	S2a	1	765.9		
S2d		127.0	SMD	S2a	0.1	694.7		
S2e		357.8	SMD	S2a	0.02	634.3		
S3a	4AQ8	1	EQ ^a	–	–		161,038	28 x 8.2 x 7.5
S3b		4	SMD	S3a	10	1246.8		
S3c		20	SMD	S3a	1	685.6		
S3d		111.2	SMD	S3a	0.1	794.4		
S3e		221.8	SMD	S3a	0.02	774.9		
S4a	4AQ8	20	EQ ^a	S3a	–		161,038	28 x 8.2 x 7.5
S4b		4	SMD	S4a	10	1340.8		
S4c		19.4	SMD	S4a	1	802.0		
S4d		91.7	SMD	S4a	0.1	579.1		
S4e		343.0	SMD	S4a	0.02	591.3		
Total:		1846.1						

^aEQ indicates equilibration on the system where the backbone was unconstrained.

^bAverage peak force is calculated from the peak force measured on both C-terminal atoms used (using 50-ps running averages).