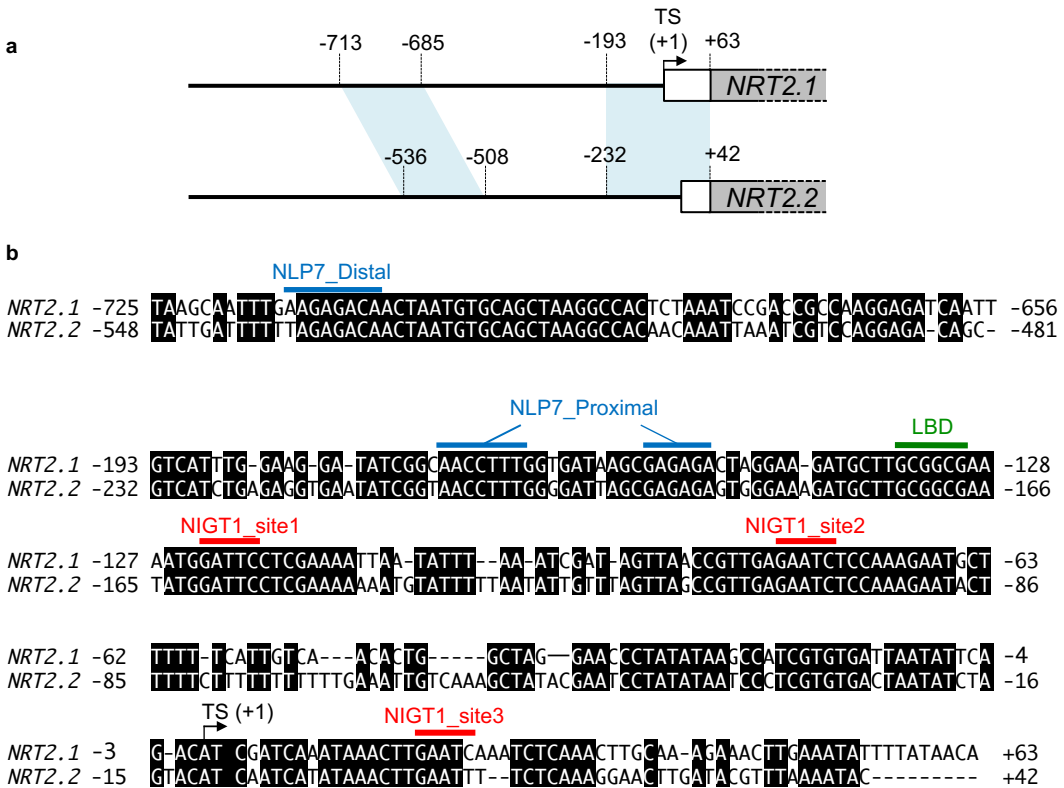


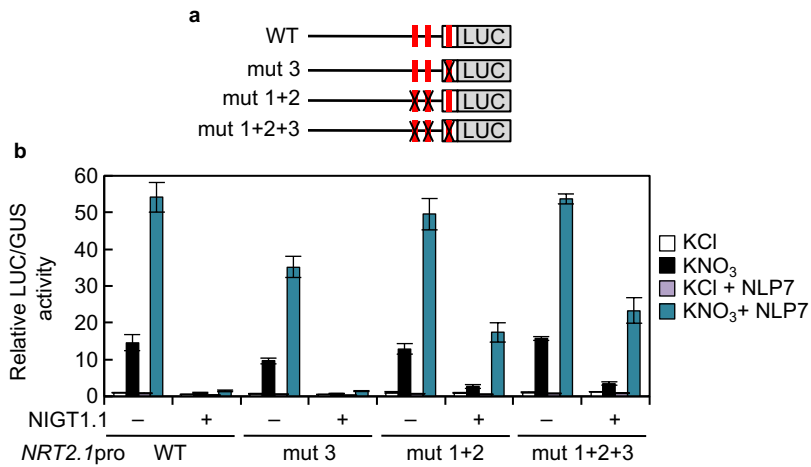
Supplementary Information

**A NIGT1-centered transcriptional cascade regulates nitrate signalling
and incorporates phosphorus starvation signals in Arabidopsis**

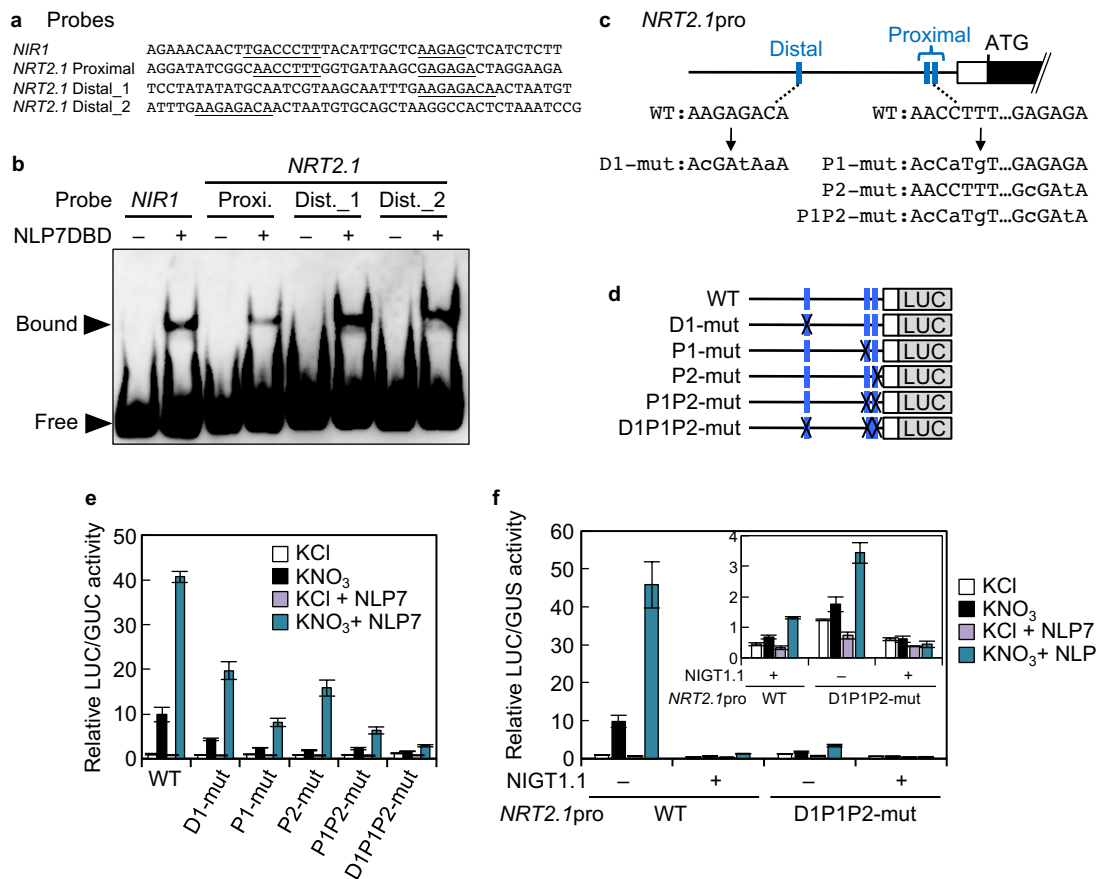
Maeda et al.



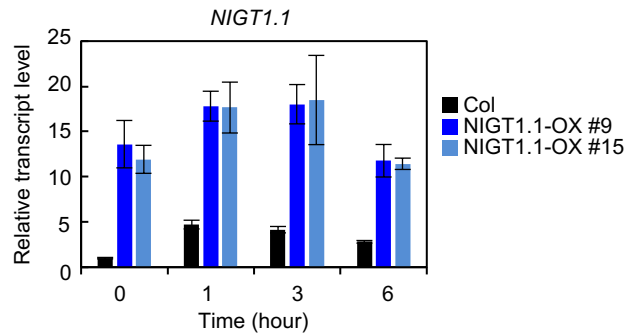
Supplementary Figure 1 | Alignment of the *NRT2.1* and *NRT2.2* promoters. Positions (a) and sequences (b) of two regions that are homologous between the *NRT2.1* and *NRT2.2* promoter sequences. In b, blue, red and green bars above the *NRT2.1* promoter sequence indicate sequences similar to the consensus sequences for NLP-, NIGT1-, and LBD-binding, respectively. The distal NLP-binding site (NLP7_Distal) is only similar to a part of the NLP-binding sequence in the *NIR1* promoter¹. These sequences are conserved in the *NRT2.1* and *NRT2.2* promoters. Numbers indicate nucleotide positions relative to the transcription start site (TS) of *NRT2.1*. The 5'-untranslated regions of *NRT2.1* cDNAs registered in the database (accession numbers: Z97058, AF093754 and AF019748) are 63, 49 bp and 44 bp in length. The position of the 63th nucleotide upstream of the translational start codon is adopted as a putative TS, and numbers are relative to the putative TS.



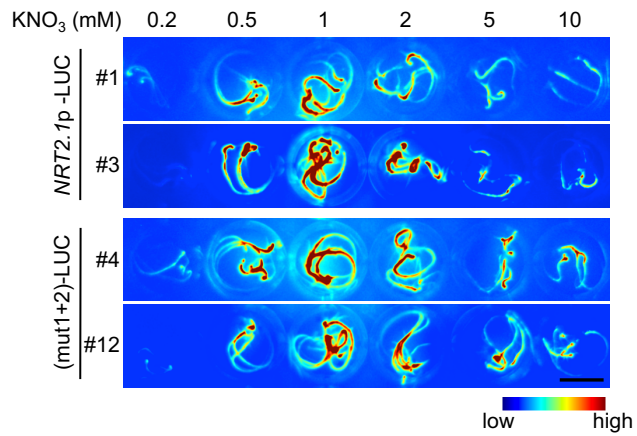
Supplementary Figure 2 | Effects of mutations within the putative NIGT1-binding site. (a) Reporter constructs. X indicates disrupted NIGT1 sites. (b) LUC activity from protoplasts that were co-transfected with the *LUC* gene fused to the wild-type or mutant *NRT2.1* promoter and the NLP7 expression vector or an empty vector and incubated in the presence of 1 mM KCl or KNO₃. LUC activity was normalised with GUS activity from the reference UBQ10-GUS plasmid and data are means ± s.d. of three biological replicates.



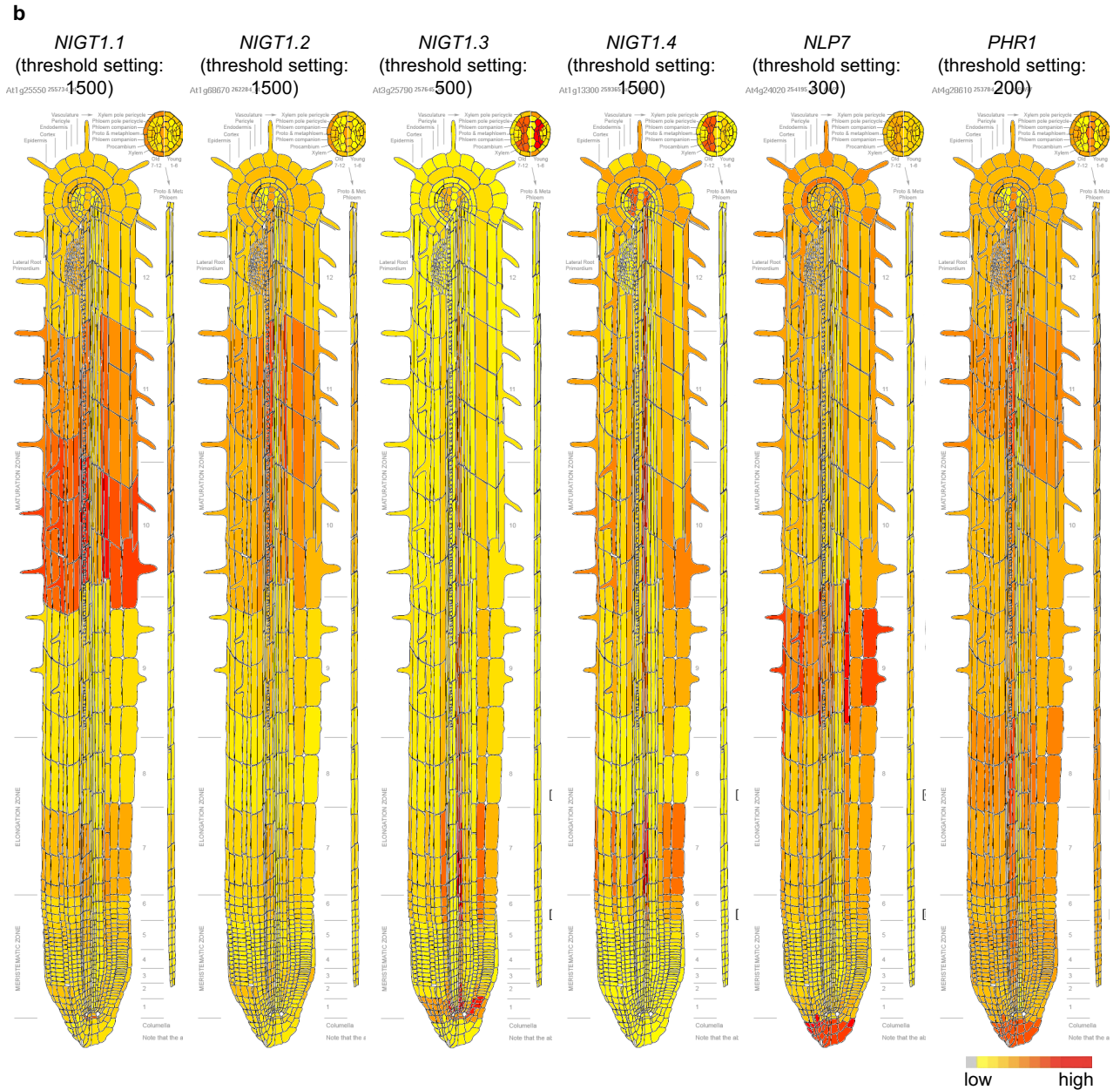
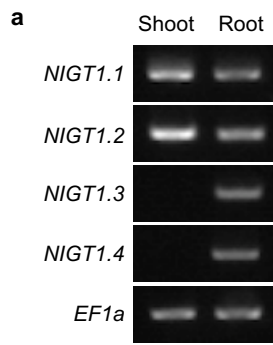
Supplementary Figure 3 | Identification of NLP-binding sites in the *NRT2.1* promoter. (a) DNA probes used in EMSA. (b) EMSA with the DNA fragments from the *NRT2.1* promoter and the DNA-binding domain of NLP7 (NLP7DBD). Distal_1 and Distal_2 probes feature the same distal NLP-binding sequence (see Supplementary Figure 1b). The NLP-binding sequence from the *NIR1* promoter¹ was used as a positive control. (c) A schematic representation of the 1.3 kb *NRT2.1* promoter. A horizontal line and a white box indicate the 5' upstream and the 5' untranslated regions, respectively. Blue boxes indicate two putative NLP-binding sites. Mutations in the mutated *NRT2.1* promoters are shown below the wild-type (WT) sequences. (d) Reporter constructs harbouring mutations on the NLP-binding sites. Mutated sites are indicated with X. (e,f) Protoplast transient assays for investigation of nitrate- and NLP7-dependent activation (e) and NIGT1.1-dependent repression (f) of mutated *NRT2.1* promoters. Protoplasts co-transfected with a reporter construct and the NLP7 or NIGT1.1 expression vector or an empty vector were incubated in the presence of 1 mM KCl or KNO₃. In e,f, LUC activity was normalised with GUS activity from the reference UBQ10-GUS plasmid and data are means ± s.d. of three biological replicates.



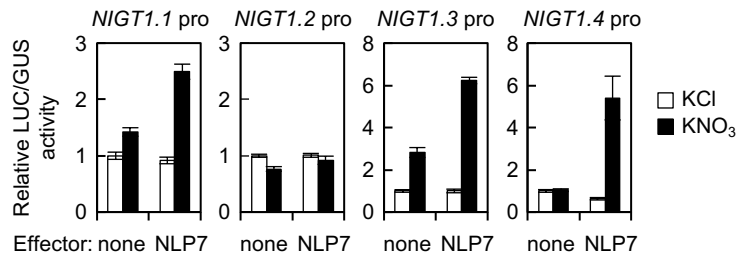
Supplementary Figure 4 | Levels of *NIGT1.1* transcripts in transgenic *Arabidopsis* lines harbouring the *NIGT1.1* gene under the control of the *UBQ10* promoter. Seedlings of *Arabidopsis* Col and two independent transgenic lines were grown with ammonium medium for 5 days, treated with KNO_3 (final concentration, 10 mM) to induce endogenous *NIGT1.1* expression, and then collected at the indicated time points. Levels of the *NIGT1.1* transcripts were quantified by RT-qPCR. The values for the NIGT1.1-OX lines indicate total levels of transcripts from endogenous and introduced *NIGT1.1* genes, whereas the values for Col seedlings indicate solely the levels of transcripts from endogenous *NIGT1.1*. Values were normalised with those of *UBQ10*, and means \pm s.d. of biological triplicates are shown.



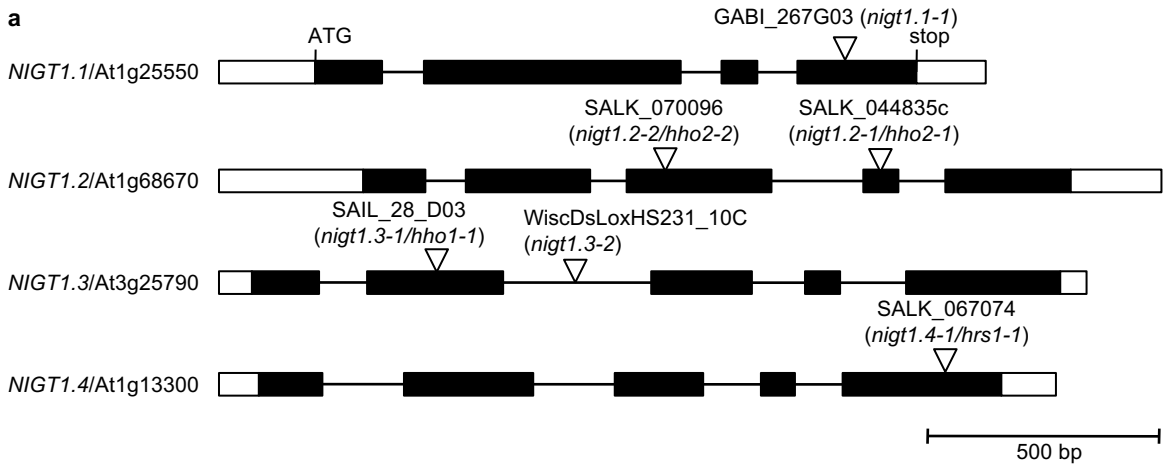
Supplementary Figure 5 | Effects of the disruption of the NIGT1-binding sites on *NRT2.1* promoter activity in the presence of different concentrations of KNO_3 . Three seedlings of transgenic lines harbouring the *LUC* gene fused to the wild-type (WT) or the mutated (mut1+2) *NRT2.1* promoter were grown with indicated concentrations of KNO_3 for 6 days. Images of LUC activity *in vivo* were captured in two independent transgenic lines. Scale bar, 1 cm.



Supplementary Figure 6 | Expression of NIGT1-clade genes. (a) Expression in shoots and roots. RT-PCR analysis was performed with cDNA prepared from RNA isolated from shoots or roots of 14-day-old seedlings grown on 1/2MS plates supplemented with 1% sucrose. *EF1a* is a housekeeping gene and was used as a control. (b) Expression in roots. Images for expression of NIGT1-clade genes, *NLP7* and *PHR1* in roots, which are from Arabidopsis eFP Browser² (<http://bbc.botany.utoronto.ca/efp/cgi-bin/efpWeb.cgi>).

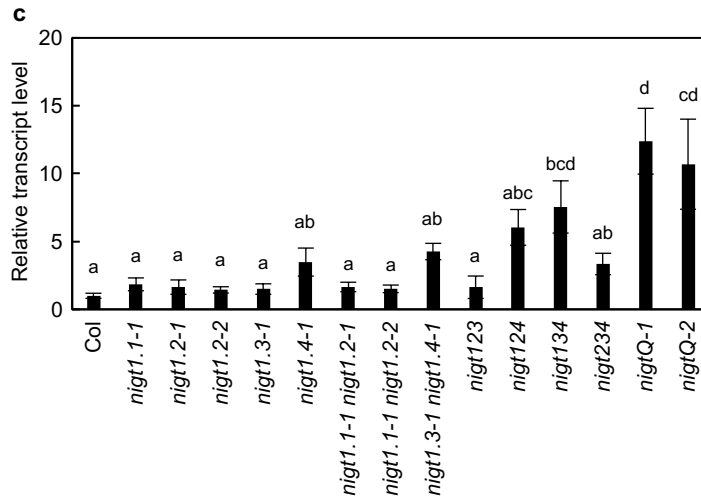


Supplementary Figure 7 | Activation of *NIGT1* gene promoters by nitrate and NLP7. Activation of promoters of *NIGT1* genes by nitrate and co-expression of NLP7 in protoplasts. The reporter plasmid containing the *LUC* gene downstream of the 1 kb promoter sequence of *NIGT1.1*, *1.2*, *1.3*, or *1.4* was co-transfected into protoplasts isolated from nitrogen-starved plants, together with the expression vector for NLP7 (NLP7) or the empty vector (none). UBQ10-GUS plasmid was included in all transfection reactions for the normalisation of transfection efficiency. Transfected protoplasts were incubated overnight with 10 mM KCl or KNO₃. LUC activity was normalised with GUS activity, and data are means ± s.d. (n=3).

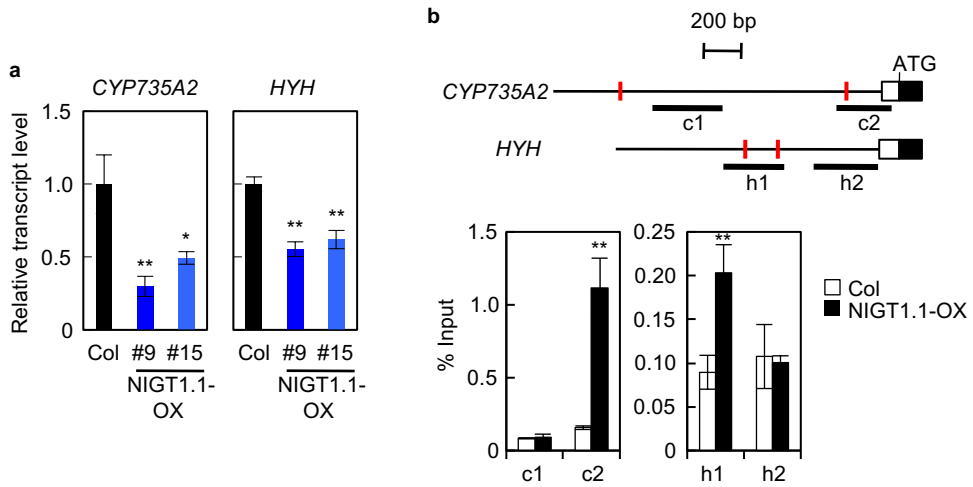


b

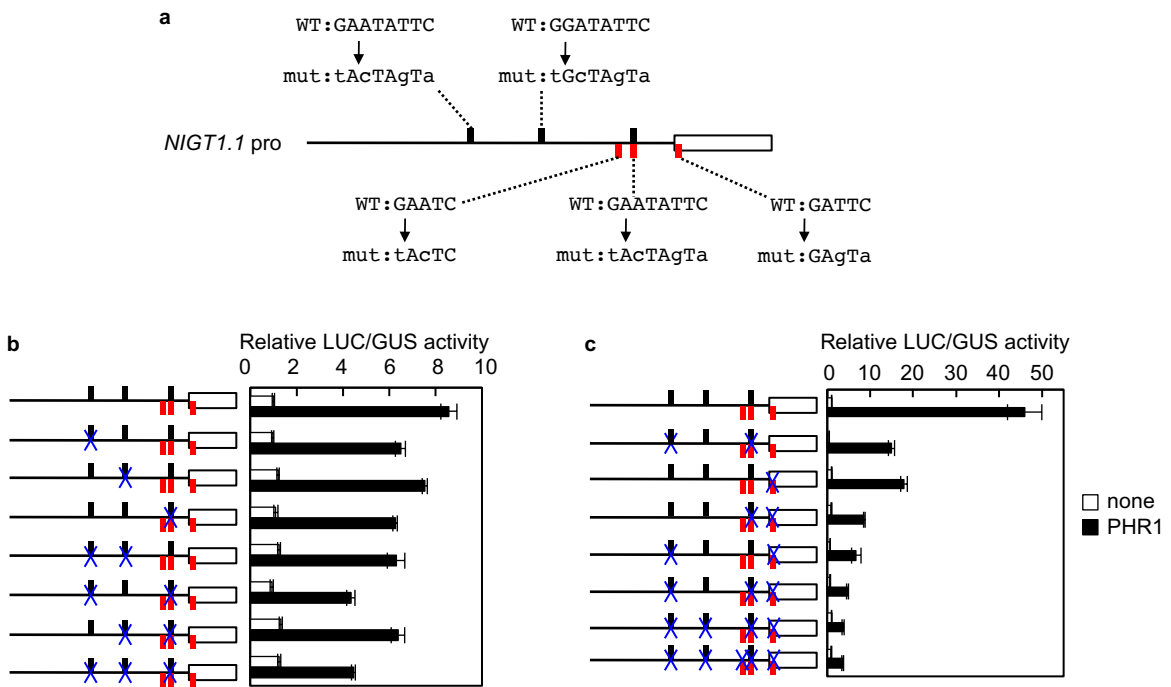
	<i>NIGT1.1</i>	<i>NIGT1.2</i>	<i>NIGT1.3</i>	<i>NIGT1.4</i>
<i>nigtQ-1</i>	GABI_267G03	SALK_044835c	SAIL_28_D03	SALK_067074
<i>nigtQ-2</i>	GABI_267G03	SALK_070096	SAIL_28_D03	SALK_067074
<i>nigtQ-3</i>	GABI_267G03	SALK_044835c	WiscDsLoxHS231_10C	SALK_067074
<i>nigt123</i>	GABI_267G03	SALK_044835c	WiscDsLoxHS231_10C	-
<i>nigt124</i>	GABI_267G03	SALK_070096	-	SALK_067074
<i>nigt134</i>	GABI_267G03	-	WiscDsLoxHS231_10C	SALK_067074
<i>nigt234</i>	-	SALK_070096	WiscDsLoxHS231_10C	SALK_067074



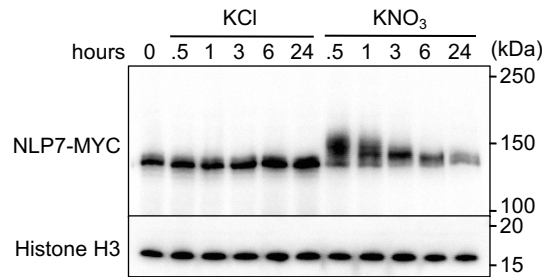
Supplementary Figure 8 | *nigt1* T-DNA lines. (a) Positions of T-DNA insertions in *NIGT1* genes. Boxes and horizontal lines indicate exons and introns, respectively. White and black boxes indicate untranslated and coding regions, respectively. (b) Combination of T-DNA insertion alleles in the *nigt1* quadruple mutants Q-1, Q-2 and Q-3, and triple mutants (*nigt123*, *124*, *134* and *234*). (c) *NRT2.1* transcript levels in single, double, triple and quadruple *nigt1* mutants. Plants were grown on 1/2MS plates supplemented with 1% sucrose for 9 days. Values are mean \pm s.d. ($n=3$). Bars not sharing the same letter designation are significantly different from each other (Tukey's HSD, $p < 0.05$).



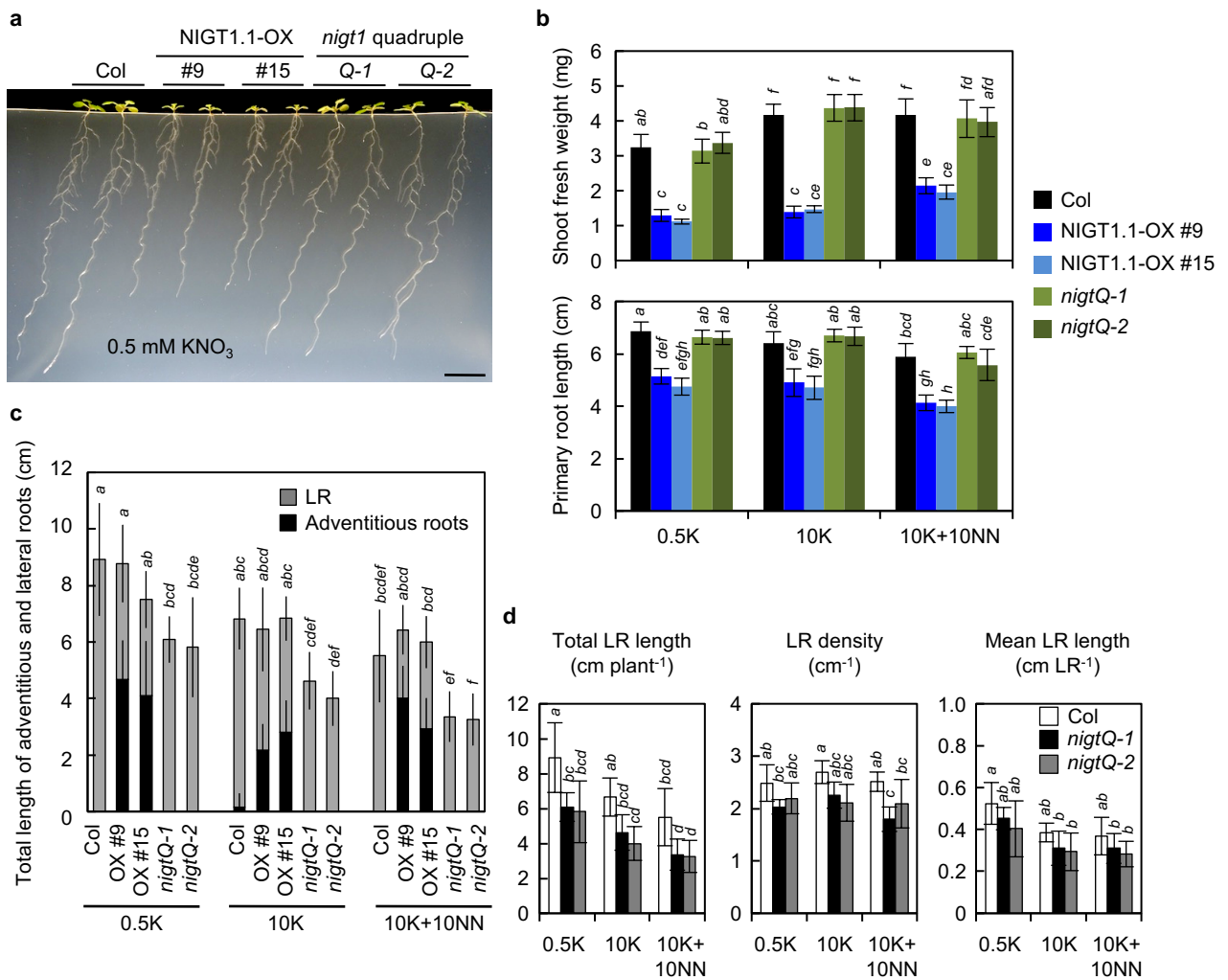
Supplementary Figure 9 | Targets of NIGT1 proteins include CYP735A2 and HYH. (a) Reduced expression of *CYP735A2* and *HYH* in the NIGT1.1 overexpressor. RNA from seedlings of Arabidopsis Col and NIGT1.1-OX lines (#9 and #15) was used for RT-qPCR. Data are means of three biological samples \pm s.d. * $p < 0.05$, ** $p < 0.01$ by two-tailed *t*-test compared with corresponding control values obtained with Col seedlings. (b) Binding of NIGT1.1 to the *CYP735A2* and *HYH* promoters *in vivo*. ChIP was performed with seedlings of Col and the NIGT1.1-OX line. Two different regions were amplified by qPCR with immunoprecipitated DNA. Data are means of four biological samples \pm s.d. ** $p < 0.01$ by one-tailed *t*-test compared with corresponding control values obtained with Col seedlings.



Supplementary Figure 10 | Transactivation of mutant *NIGT1.1* promoters by PHR1. (a) A schematic representation of the 1 kb *NIGT1.1* promoter. A horizontal line and a white box indicate the upstream sequence and the 5' untranslated region, respectively. Black and red boxes indicate the P1BS sequence (5'-GNATATNC-3') and the conserved NIGT1-binding sequences (5'-GAATC-3' and 5'-GAATATTC-3'), respectively. Mutated sequences (mut) are shown below the wild-type (WT) sequences. (b,c) Transactivation of mutated *NIGT1.1* promoters by PHR1 in protoplasts. Mutated sites are indicated with blue X. LUC activity was normalised with GUS activity, and data are means \pm s.d. of three biological replicates.



Supplementary Figure 11 | Decreases in the NLP7 protein level after nitrate treatment. Seedlings expressing MYC-tagged NLP7³ were grown in nitrogen-free 1/10MS solution supplemented with 0.5 mM ammonium succinate [1/10 MS salts, 0.1 g l⁻¹ MES-KOH (pH5.7), 0.5% sucrose, 0.5 mM ammonium succinate] for 4 days and treated with 10 mM KCl or KNO₃. Seedlings were collected at indicated time points after the onset of the treatment and used for immunoblot analysis with anti-MYC or anti-Histone H3 antibodies. NLP7 proteins in the active form (retarded bands) more rapidly disappeared, compared with ones in the inactive form.



Supplementary Figure 12 | Growth of NIGT1.1-overexpressing plants and *nigt1* quadruple mutants. (a) Image of seedlings grown on a plate containing 0.5 mM KNO₃ as N source. Scale bar, 1 cm. Shoot fresh weight and primary root length (b), total length of adventitious roots and lateral roots (LR) (c), and total LR length, LR density, and mean length of LR (d) of Arabidopsis Col, NIGT1.1-OX, and *nigt1* mutants grown under three different N-nutrient conditions (0.5K, 0.5 mM KNO₃; 10K, 10 mM KNO₃; 10K+10NN, 10 mM KNO₃+10 mM NH₄NO₃). Seedlings were initially grown on 1/2MS plates containing 9.4 mM KNO₃ and 10.3 mM NH₄NO₃ as N sources for 3 days, and then grown on test plates containing the indicated N source for 6 days. NIGT1.1-OX plants developed unusually long adventitious roots. Values are means ± s.d. (*n*=8). Bars not sharing the same letter designation are significantly different from each other (Tukey's HSD, *p* < 0.05). In c, the statistical test was performed for the total length of adventitious and lateral roots.

Supplementary Table 1. Cytokinin contents

pmol g ⁻¹ FW	Parental line (4xNRE-min-GUS)		NLP6-SUPRD #7		NLP6-SUPRD #14		Col		nlp6 nlp7-1		nigtQ-3		nigtQ-3+NIGT1.2		GFP-OX		NIGT1.2GFP-OX	
	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃	KCl	KNO ₃
iZ	N.D.	1.080 ± 0.174	N.D.	0.185 ± 0.048	0.028 ± 0.055	0.207 ± 0.054	0.138 ± 0.048	0.917 ± 0.088	0.024 ± 0.049	0.209 ± 0.162	0.027 ± 0.053	0.883 ± 0.104	0.035 ± 0.048	1.034 ± 0.073	0.085 ± 0.058	0.931 ± 0.087	N.D.	0.587 ± 0.099
iZR	0.384 ± 0.054	13.024 ± 1.270	0.563 ± 0.060	2.337 ± 0.330	0.539 ± 0.082	2.950 ± 0.271	0.866 ± 0.131	13.244 ± 0.768	0.280 ± 0.019	4.656 ± 0.208	0.607 ± 0.117	13.797 ± 0.547	0.387 ± 0.055	12.322 ± 0.483	0.274 ± 0.171	9.350 ± 0.234	0.507 ± 0.036	8.371 ± 0.301
iZRPs	2.039 ± 0.295	62.875 ± 5.681	2.447 ± 0.227	11.716 ± 0.732	2.366 ± 0.092	14.422 ± 1.041	2.972 ± 0.454	45.753 ± 3.624	1.473 ± 0.210	19.228 ± 1.267	3.167 ± 0.150	68.848 ± 3.023	1.620 ± 0.146	52.452 ± 2.730	2.055 ± 0.279	56.292 ± 3.314	3.057 ± 0.501	44.819 ± 4.076
cZ	0.394 ± 0.053	0.305 ± 0.045	0.499 ± 0.107	0.247 ± 0.057	0.292 ± 0.041	0.216 ± 0.017	0.884 ± 0.040	0.324 ± 0.047	0.416 ± 0.045	0.152 ± 0.053	0.531 ± 0.068	0.282 ± 0.029	0.599 ± 0.079	0.142 ± 0.135	0.540 ± 0.078	0.250 ± 0.063	0.169 ± 0.195	0.123 ± 0.142
cZR	0.783 ± 0.136	0.586 ± 0.069	0.852 ± 0.056	0.613 ± 0.070	0.884 ± 0.050	0.589 ± 0.047	1.026 ± 0.097	0.762 ± 0.043	0.655 ± 0.107	0.475 ± 0.035	0.961 ± 0.090	0.615 ± 0.069	0.787 ± 0.027	0.566 ± 0.048	0.617 ± 0.162	0.700 ± 0.107	1.036 ± 0.148	0.863 ± 0.214
cZRPs	4.662 ± 0.272	2.423 ± 0.144	5.782 ± 0.569	2.907 ± 0.143	5.248 ± 0.186	2.501 ± 0.115	5.539 ± 0.247	2.555 ± 0.136	3.987 ± 0.202	1.841 ± 0.088	5.797 ± 0.538	2.497 ± 0.066	4.549 ± 0.277	2.045 ± 0.176	4.274 ± 0.190	2.483 ± 0.237	7.415 ± 0.516	3.558 ± 0.083
DZ	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	0.072	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
DZR	N.D.	0.227 ± 0.031	N.D.	N.D.	N.D.	N.D.	N.D.	0.344 ± 0.023	N.D.	N.D.	N.D.	0.255 ± 0.039	N.D.	0.210 ± 0.015	N.D.	0.102 ± 0.027	N.D.	N.D.
DZRPs	N.D.	0.681 ± 0.112	N.D.	0.116 ± 0.015	N.D.	0.158 ± 0.033	0.089 ± 0.015	0.648 ± 0.051	N.D.	0.135 ± 0.007	N.D.	0.730 ± 0.053	N.D.	0.602 ± 0.048	N.D.	0.402 ± 0.020	N.D.	0.325 ± 0.061
iP	0.463 ± 0.054	0.641 ± 0.474	0.450 ± 0.528	0.683 ± 0.461	0.451 ± 0.307	0.728 ± 0.199	0.827 ± 0.097	0.622 ± 0.078	0.385 ± 0.449	0.175 ± 0.350	0.909 ± 1.054	1.642 ± 0.482	0.914 ± 0.072	1.226 ± 0.359	1.551 ± 0.144	1.404 ± 0.557	1.767 ± 0.337	0.990 ± 1.353
iPR	0.222 ± 0.071	0.736 ± 0.036	0.224 ± 0.027	0.403 ± 0.078	0.224 ± 0.065	0.278 ± 0.058	0.455 ± 0.068	1.148 ± 0.141	0.183 ± 0.025	0.574 ± 0.022	0.256 ± 0.028	0.780 ± 0.063	0.277 ± 0.024	0.631 ± 0.106	0.125 ± 0.106	0.697 ± 0.124	0.482 ± 0.135	1.057 ± 0.113
iPRPs	8.260 ± 0.773	29.220 ± 2.078	7.599 ± 0.428	13.323 ± 1.288	6.861 ± 0.197	10.440 ± 1.023	16.620 ± 1.121	36.881 ± 2.360	6.870 ± 0.809	32.559 ± 1.063	11.222 ± 0.749	45.613 ± 2.847	9.233 ± 0.640	32.888 ± 1.573	14.473 ± 0.816	35.240 ± 3.318	15.145 ± 1.691	36.537 ± 2.446
iZ7G	1.178 ± 0.169	4.477 ± 0.249	1.444 ± 0.122	1.737 ± 0.115	1.395 ± 0.042	1.822 ± 0.017	1.834 ± 0.110	4.204 ± 0.190	1.091 ± 0.154	1.943 ± 0.089	1.332 ± 0.097	3.604 ± 0.166	1.372 ± 0.031	5.074 ± 0.183	1.282 ± 0.213	4.322 ± 0.150	0.896 ± 0.080	2.991 ± 0.102
iZ9G	0.246 ± 0.103	1.199 ± 0.248	0.380 ± 0.024	0.407 ± 0.075	0.201 ± 0.067	0.340 ± 0.037	0.448 ± 0.040	0.889 ± 0.190	0.167 ± 0.088	0.555 ± 0.113	0.139 ± 0.161	0.911 ± 0.098	0.316 ± 0.088	1.138 ± 0.153	0.212 ± 0.032	1.082 ± 0.046	0.090 ± 0.064	0.425 ± 0.134
iZOG	1.484 ± 0.159	5.685 ± 0.311	1.598 ± 0.193	1.962 ± 0.366	1.437 ± 0.231	1.803 ± 0.111	2.584 ± 0.292	5.40 ± 0.487	1.439 ± 0.303	3.037 ± 0.382	2.168 ± 0.453	5.600 ± 0.549	1.715 ± 0.333	6.732 ± 0.422	2.301 ± 0.276	9.145 ± 1.366	2.029 ± 0.551	8.351 ± 0.125
cZOG	7.268 ± 0.494	6.382 ± 0.917	9.290 ± 1.201	7.887 ± 0.733	8.233 ± 0.553	6.519 ± 0.355	8.088 ± 0.624	5.768 ± 0.928	6.220 ± 0.856	5.166 ± 0.231	5.975 ± 0.897	3.797 ± 0.561	7.989 ± 0.891	5.725 ± 0.276	7.233 ± 1.215	5.749 ± 0.922	4.818 ± 0.465	5.059 ± 0.963
iZROG	0.019 ± 0.039	0.375 ± 0.077	0.049 ± 0.033	0.082 ± 0.055	0.038 ± 0.028	0.057 ± 0.045	0.122 ± 0.030	0.532 ± 0.037	0.047 ± 0.035	0.112 ± 0.033	0.049 ± 0.034	0.299 ± 0.088	0.044 ± 0.026	0.281 ± 0.088	0.056 ± 0.039	0.336 ± 0.064	0.062 ± 0.074	0.078 ± 0.098
cZROG	0.431 ± 0.048	0.467 ± 0.056	0.415 ± 0.025	0.398 ± 0.049	0.462 ± 0.016	0.413 ± 0.030	0.570 ± 0.046	0.538 ± 0.031	0.349 ± 0.058	0.328 ± 0.021	0.386 ± 0.043	0.381 ± 0.035	0.373 ± 0.041	0.339 ± 0.035	0.348 ± 0.030	0.364 ± 0.068	0.365 ± 0.092	0.388 ± 0.033
iZRPsoG	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	0.071 ± 0.008	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
cZRPsoG	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
DZ9G	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.
iP7G	106.665 ± 5.996	110.080 ± 2.521	85.748 ± 3.400	82.763 ± 2.025	73.019 ± 4.185	72.792 ± 3.633	94.843 ± 2.322	93.157 ± 2.279	91.659 ± 3.060	96.398 ± 2.955	96.447 ± 2.128	98.204 ± 2.697	86.118 ± 3.911	90.078 ± 3.277	101.625 ± 5.550	113.177 ± 5.441	81.982 ± 5.689	87.289 ± 4.285
iP9G	2.284 ± 0.082	2.398 ± 0.153	1.685 ± 0.089	1.731 ± 0.099	1.432 ± 0.101	1.421 ± 0.115	1.758 ± 0.060	1.925 ± 0.077	2.042 ± 0.113	2.126 ± 0.077	1.844 ± 0.067	1.963 ± 0.150	1.854 ± 0.103	1.828 ± 0.071	2.096 ± 0.049	2.249 ± 0.220	1.352 ± 0.115	1.499 ± 0.099
iZ-type	5.350 ± 0.732	88.715 ± 6.592	6.481 ± 0.327	18.426 ± 1.343	6.003 ± 0.176	21.602 ± 1.377	8.965 ± 0.854	70.943 ± 4.787	4.521 ± 0.466	29.740 ± 1.165	7.488 ± 0.473	93.942 ± 3.119	5.488 ± 0.265	79.034 ± 3.270	6.265 ± 0.571	81.458 ± 3.798	6.641 ± 1.144	65.622 ± 4.057
iP-type	117.893 ± 6.386	143.075 ± 2.455	95.706 ± 3.014	98.903 ± 2.188	81.986 ± 4.350	85.660 ± 3.185	114.503 ± 2.591	133.733 ± 4.550	101.140 ± 3.345	131.833 ± 4.203	110.678 ± 2.402	148.203 ± 5.742	98.395 ± 3.827	126.651 ± 4.872	110.870 ± 5.987	152.768 ± 9.255	100.728 ± 6.902	127.371 ± 6.850
iZ-type precursor	2.423 ± 0.347	75.899 ± 6.701	3.010 ± 0.285	14.053 ± 0.934	2.905 ± 0.100	17.372 ± 1.293	3.838 ± 0.536	58.998 ± 4.304	1.752 ± 0.198	23.884 ± 1.446	3.773 ± 0.063	82.645 ± 3.498	2.006 ± 0.160	64.774 ± 3.094	2.329 ± 0.435	65.642 ± 3.313	3.565 ± 0.503	53.190 ± 4.246
iP-type precursor	8.482 ± 0.763	29.956 ± 2.072	7.823 ± 0.415	13.726 ± 1.361	7.084 ± 0.189	10.718 ± 1.033	17.075 ± 1.137	38.029 ± 2.459	7.054 ± 0.795	33.133 ± 1.060	11.478 ± 0.730	46.393 ± 2.893	9.510 ± 0.651	33.519 ± 1.637	14.598 ± 0.847	35.938 ± 3.425	15.627 ± 1.705	37.594 ± 2.401

4xNRE-ming-GUS. The parental line of NLP6-SUPRD lines. N.D., not detected.

iZ, *trans*-zeatin; iZR, iZ riboside; iZRPs, iZ ribotides; cZ, *cis*-zeatin; cZR, cZ riboside; cZRPs, cZ ribotides; DZ, dihydrozeatin; DZR, DZ riboside; DZRPs, DZ ribotides; iP, N⁶-(Δ^2 -isopentenyl)adenine; iPR, iP riboside; iPRPs, iP ribotides; iZ7G, iZ-7-N-glucoside; iZ9G, iZ-9-N-glucoside; iZOG, iZ-O-glucoside; cZOG, cZ-O-glucoside; iZROG, iZR-O-glucoside; cZROG, cZR-O-glucoside; DZ9G, DZ-9-N-glucoside; iP7G, iP-7-N-glucoside; iP9G, iP-9-N-glucoside.

Supplementary Table 2. Primer list

Amplicon	AGI code	Sequence
Cloning for transient assay		
<i>NRT2.1</i> promoter	At1g08090	CGCAAGCTTCACGTCACGAGATTGATCGA CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NIGT1-site_mut1)	At1g08090	GCAAGATGAGGGGAGGGTAGAGCG TTTTTCGAGTACTCCATTTTCGCCGCAAGCATCT GAAAATGGAGTACTCGAAAATTAATTTAAA CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NIGT1-site_mut2)	At1g08090	GCAAGATGAGGGGAGGGTAGAGCG TCTTTGGAGAGTATCAACGGTTAACTATCGATT CCGTTGATACTCCAAAGAATGCTTTTTTCA CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NIGT1-site_mut3)	At1g08090	GCAAGATGAGGGGAGGGTAGAGCG TGAGATTGAGTAAAGTTTATTTGATCGATGA TAACTTTACTCAAATCTCAAAGTCAAAA CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NLP-site_D1mut)	At1g08090	CGCAAGCTTCACGTCACGAGATTGATCGA ATTAGTTTTATCGTCAAATGCTTACGATGTCAT AATTTGACGATAAACTAATGTGCAGTAAAGCC CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NLP-site_P1mut)	At1g08090	CGCAAGCTTCACGTCACGAGATTGATCGA ATCACACATGGTCCGATATCCTTCCAAATGAC TATCGGCACCATGTGGTATAAGCGAGAGACTAGG CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NLP-site_P2mut)	At1g08090	CGCAAGCTTCACGTCACGAGATTGATCGA TTCTAGTATCGCGCTTATCACCAGGTTTGC GATAAGCGGATCTAGGAAGATGCTTCCGG CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NRT2.1</i> promoter (NLP-site_P1P2mut)	At1g08090	CGCAAGCTTCACGTCACGAGATTGATCGA ATCGCGCTTATCACACATGGTCCGATATCCTTCCAAATGAC CCATGTGGTATAAGCGGATAGGAAAGATGCTTCCGG CGTCCATGGTTATAAAATATTTCAAGTTTCT
<i>NIGT1.1</i> promoter	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT GTGCCATGGTGAACAGATCTAGACTCGTTTTTG
<i>NIGT1.2/HHO2</i> promoter	At1g68670	CTCGGATCCGAGGAATATGAAAGGTCGGATATG GTGCCATGGTTATTTCAAAGGGCACTAAGAACA
<i>NIGT1.3/HHO1</i> promoter	At3g25790	CTCGGATCCGGATATAACAACAGCATAGCATAT GTGCCATGGTGGTTAAGGGACAAAACAAAACC
<i>NIGT1.4/HRS1</i> promoter	At1g13300	CTCGGATCCAATGAAGGTGTTATTAGTAGTGCC GTGCCATGGTGAATCTTATAGGACTTAATTTAA
<i>NIGT1.1</i> promoter mut_P1BSn -655 to -648 from ATG GAATATTC to TACTAGTA	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT GTGGAGTACTAGTACTCGAGAGTGGGAGAACTAAG CTCGAGTACTAGTACTCCACTTTTGCCAAAACCTCTT GTGCCATGGTGAACAGATCTAGACTCGTTTTTG
<i>NIGT1.1</i> promoter mut_P1BS3 -502 to -495 from ATG GGATATTC to TGCTAGTA	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT GAAGAATACTAGCAATAGAGATTGGAATCAAGATATG TCTCTATTGCTAGTATTCTTCACTCGTTTGTATTACAA GTGCCATGGTGAACAGATCTAGACTCGTTTTTG
<i>NIGT1.1</i> promoter mut_P1BSc -299 to -292 from ATG GAATATTC to TACTAGTA	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT CAGGGATACTAGTAGGGATATGGATTCCCTTCTTT ATCCCTACTAGTATCCCTGTATCAAACAAACCTCAGC GTGCCATGGTGAACAGATCTTAGACTCGTTTTTG
<i>NIGT1.1</i> promoter mut_NBS2 -315 to -311 from ATG GAATC to TACTC	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT GATATGTGGAGTACTCTTTCCATATCTATCGAGC AAGGAAGTACTCCACATATCCCGAATATCTCCCT GTGCCATGGTGAACAGATCTTAGACTCGTTTTTG
<i>NIGT1.1</i> promoter mut_NBS1 -202 to -198 from ATG GATTC to GAGTA	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT CTAAAACCTACTCTTTGATGTTTAAATTTATCCGG TCAAGAAGAGTAGGTTTTAGGTACGCGACGGCCGA GTGCCATGGTGAACAGATCTTAGACTCGTTTTTG
<i>NIGT1.1</i> promoter mut_NBS2 on mut P1BSc	At1g25550	CTCGGATCCGAATACATACTCAAATAGACACTT GATATGTGGAGTACTCTTTCCATATCTATCGAGC AAGGAAGTACTCCACATATCCCTACTAGTATCCCT GTGCCATGGTGAACAGATCTTAGACTCGTTTTTG
<i>NIGT1.1</i> CDS	At1g25550	TATGGATCCATGGGTATGATGATGTTCAAGAGCGG TCTAGGCCTTGAAGAGGAGAAAGGTGTGG
<i>NIGT1.2/HHO2</i> CDS	At1g68670	TATGGATCCATGGGTATGATGATGATGATGATTAC TCTAGGCCTTGACACAGGATAGAAGTATTTGT
<i>NIGT1.3/HHO1</i> CDS	At3g25790	TATGGATCCATGGGTATGATGATGATGATGATGATG TCTAGGCCTTATTTCTGACGTAATGATTACGG
<i>NIGT1.4/HRS1</i> CDS	At1g13300	TATGGATCCATGGGTATGATGATGATGATGATGATG TCTAGGCCTTGATATTTTGTCTTTCTG
LBD37 CDS	At5g67420	ATATCCATGGATGAGCTGCAATGGTTGC GGGCAGGCCAACAAGGTTAAGCAAC
PHR1 CDS	At4g28610	GTGCCATGGAGGCTCGTCCAGTTCATAGA GTCAGGCCTATTATCGATTTTGGGACGCTTTGGCTG
RT-qPCR		
<i>NIGT1.1</i>	At1g25550	TCGATGGTTAACTAATGACGAAGT TTACTACCGGAGTAGCTGGTCTTC
<i>NIGT1.1</i> (endogenous only)	At1g25550	ACACACACACCACACCTCT ACGAACATCTTGGTTACAAAACA
<i>NIGT1.2/HHO2</i>	At1g68670	TAGACTTCACACAAGAAGGCCAG AATTGGGTTGTTGCTGATTCC
<i>NIGT1.3/HHO1</i>	At3g25790	GCCAGAAGAGGGCAGAGAAA ATGAGGGCCACCAAGTTGTT

Supplementary Table 2. Primer list (continued)

Amplicon	AGI code	Sequence
RT-qPCR		
<i>NIGT1.4/HRS1</i>	At1g13330	CTAACACGGAAACTCTCAACG CGGTAGTCTTGCCCGTAGAGTA
RT-qPCR		
<i>NRT2.1</i>	At1g08090	TGAGCAGGAGAAGCAGAAGA TTGTTGGGTGTGTTCTCAGG
<i>NAR2.1/NRT3.1</i>	At5g50200	CTACGGGTTGATGCCATTG GTCGGTGCTGTGCATAGG
<i>CYP735A2</i>	At1g67110	TGGACTCTCATGCTCCTTGC ACAACCTTGCCGGACCTCAT
<i>HYH</i>	At3g17609	CAGAGTATCAGCGCAACAAGC TCTCTTCGAGCTGGTCATTGT
<i>UBQ10^a</i>	At4g05320	GGCCTTGATAATCCCTGATGAATAAG AAAGAGATAACAGGAACGGAAACATAGT
<i>IPS1</i>	At3g09922	AGACTGCAGAAAGCTGATTCAGA TTGCCCAATTCTAGAGGGAGA
<i>PEX4^b</i>	At5g25760	CTGCCACTCAGGGAATCTTAA TTTGCCATTGAATTGAACCC
RT-PCR		
<i>NIGT1.1</i>	At1g25550	TGGAACCAATCACCAGATCC AGTAGAAGATGTTGCCGGAGAT
<i>NIGT1.2/HHO2</i>	At1g68670	TGATGGTGAGATGGATTACG ACCACCCCTTGTGGCTACATCGGA
<i>NIGT1.3/HHO1</i>	At3g25790	CGAGCTTCTCTTTGCGTAG CTTTTCTTTGAGCGATCCTG
<i>NIGT1.4/HRS1</i>	At1g13330	GTCGCAATTGCATGACGCTTC GTAGAAGAGACATGCCGG
<i>EF1a</i>	At5g60390	ATGCCCCAGGACATCGATTTCAT TTGGCGCACCCCTAGCTGGATCA
EMSA probes		
<i>NRT2.1</i> promoter fragment p1_1st	At1g08090	CACGTCAGCGAGATTGATCG AGGAAACAGCTATGACCATGCAAAAGAAAAACGGCAAACTAAATA
<i>NRT2.1</i> promoter fragment p1_2nd	At1g08090	CACGTCAGCGAGATTGATCG Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p2_1st	At1g08090	CTGTTTTATTTAGTTTGGCGTTTTTC AGGAAACAGCTATGACCATGTAGTTGCTCTTCAAATTGCTTAC
<i>NRT2.1</i> promoter fragment p2_2nd	At1g08090	CTGTTTTATTTAGTTTGGCGTTTTTC Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p3_1st	At1g08090	GCAATCGTAAGCAATTTGAAGAGA AGGAAACAGCTATGACCATGACACTTAATTTGAACGTGTAAC
<i>NRT2.1</i> promoter fragment p3_2nd	At1g08090	GCAATCGTAAGCAATTTGAAGAGA Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p4_1st	At1g08090	CATGCTGTTACAGCAGTTCAAAT AGGAAACAGCTATGACCATGAATGACCGCTCTACCCCTCCCTCA
<i>NRT2.1</i> promoter fragment p4_2nd	At1g08090	CATGCTGTTACAGCAGTTCAAAT Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p5_1st	At1g08090	GCAAGATGAGGGGAGGGTAGAGCG AGGAAACAGCTATGACCATGTGTTATAAAAATTTCAAGTTTCT
<i>NRT2.1</i> promoter fragment p5_2nd	At1g08090	GCAAGATGAGGGGAGGGTAGAGCG Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p5-1_1st	At1g08090	CACGTCAGCGAGATTGATCG AGGAAACAGCTATGACCATGCAACGGTTAACTATCGATTAAATAT
<i>NRT2.1</i> promoter fragment p5-1_2nd	At1g08090	CACGTCAGCGAGATTGATCG Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p5-2_1st	At1g08090	AATATTTAAATCGATAGTTAACCGTTG AGGAAACAGCTATGACCATGCGATGCTGAATTAATCACACG
<i>NRT2.1</i> promoter fragment p5-2_2nd	At1g08090	AATATTTAAATCGATAGTTAACCGTTG Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> promoter fragment p5-3_1st	At1g08090	ATTGTCAACACTGGCTAGGAACCC AGGAAACAGCTATGACCATGTGTTATAAAAATTTCAAGTTTCT
<i>NRT2.1</i> promoter fragment p5-3_2nd	At1g08090	ATTGTCAACACTGGCTAGGAACCC Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>OsNIGT1_1st</i>		CGACGACACCTTCATTTTTATTTTC AGGAAACAGCTATGACCATGTTGCTGGGGTGAGGAAGGAAGAA
<i>OsNIGT1_2nd</i>		CGACGACACCTTCATTTTTATTTTC Biotin-ACACAGGAAACAGCTATGACCATG (BIO-M13)
<i>NRT2.1</i> Proximal_1st	At1g08090	GTCTAGACTGCAGCTGTGCGAGGATATCGCAACCTTTGG GCAAGCTTGTGAGCATGCGGATCTTCTAGTCTCTCGCTTA
<i>NRT2.1</i> Proximal_2nd		Biotin-GTCTAGACTGCAGCTGTGCGA (BIO-OligoZ) Biotin-GCAAGCTTGTGAGCATGCGGA (BIO-oligoY)
<i>NRT2.1</i> Distal1_1st	At1g08090	GTCTAGACTGCAGCTGTGCGATCCTATATGCAATCGTAAG GCAAGCTTGTGAGCATGCGGACATTAGTTGCTCTCAAATTG
<i>NRT2.1</i> Distal1_2nd		Biotin-GTCTAGACTGCAGCTGTGCGA (BIO-OligoZ) Biotin-GCAAGCTTGTGAGCATGCGGA (BIO-oligoY)
<i>NRT2.1</i> Distal2_1st	At1g08090	GTCTAGACTGCAGCTGTGCGATTTGAAGAGACAATAATGTG GCAAGCTTGTGAGCATGCGGATTTAGAGTGGCCCTAGCTGC
<i>NRT2.1</i> Distal2_2nd		Biotin-GTCTAGACTGCAGCTGTGCGA (BIO-OligoZ) Biotin-GCAAGCTTGTGAGCATGCGGA (BIO-oligoY)
Cloning for recombinant expression		
<i>NIGT1.1</i> DBD	At1g25550	CGTCCATGGCTGGGGTGGAAAAGATTTGAGGAG CGTCTCGAGACCACCGGTCTTACTACCGGAGT
Binary plasmid construction		
<i>NIGT1.1</i> CDS	At1g25550	CGTCCATGGCTATGTTCAAGAGCGGTGACATGGAT TCTAGGCCTTGAAGAGGAAGAGGTGTGG

Supplementary Table 2. Primer list (continued)

Amplicon	AGI code	Sequence
ChIP-qPCR		
<i>NRT2.1</i> pro_1	At1g08090	TGACACGTCAGCGAGATTGA CGGTGTGAAAAATTGAAATCGTTGA
<i>NRT2.1</i> pro_2	At1g08090	GATGAAGACATGTACACACACGA ACATACAGTGATACAGGATAGGCT
<i>NRT2.1</i> pro_3	At1g08090	TGGTGAAGAGGACCAAACTTT GCTCACCAGTAGAATCACCCA
<i>NRT2.1</i> pro_4	At1g08090	GGGTAGAGCGGTCATTGGGA GCTCACCAGTAGAATCACCCA
<i>NAR2.1</i> pro_1	At5g50200	CAGTGAACGGTTACCTGAGGA AGGGTTGATGTTTGTAGGCTGT
<i>NAR2.1</i> pro_2	At5g50200	TGAAAGCAAGACCTAAACCCA GCCAAAGGATGAGATATGAGTATTTGT
<i>NIGT1.1</i> pro_1	At1g25550	TGCTGATCCGATCGTTGT CCATGGATGTGAAGACCACCA
<i>NIGT1.1</i> pro_2	At1g25550	ACATCATCGCCAGCTCGATA GGCAGAGAAAAGGAGTTGTT
<i>NIGT1.2/HHO2</i> pro_3	At1g68670	GGAATCGAACCACCAAAGAA GCTAAATGTTACGCGCATA
<i>NIGT1.2/HHO2</i> pro_4	At1g68670	ACGTCAGTTCCAGCCATAC GAAGTGAAGAGAAATCGAGGTC
<i>NIGT1.3/HHO1</i> pro_5	At3g25790	CTCGAGACTCGAATAGGTGGA TATGTTCCGGTGATGTCCAA
<i>NIGT1.3/HHO1</i> pro_6	At3g25790	CCCAAATAAATAAAGTTCACACGA GGCTCTATAGGGTTTCTCAAGG
<i>NIGT1.4/HRS1</i> pro_7	At1g13300	GGCGACAGGAAAGAACT TCCTCATGCATTCCTCAAC
<i>NIGT1.4/HRS1</i> pro_8	At1g13300	GGTCTCTAATAAGTATCCA AGGATCAGTATTTTGAAGTGA
<i>IPS1</i> pro_9	At3g09922	CCTGACAGCTCAAGGCATTA TGATGGAACAAGGGGAGTC
<i>IPS1</i> pro_10	At3g09922	GTGGTTGCATGCTTGTTTA TTAGGGTAAAAAGGGGCATC
<i>CYP735A2_c1</i>	At1g67110	GAATGTGGTAATAGCGAATATGAGACA CTTATCTACTTGGGTGATTGAAATC
<i>CYP735A2_c2</i>	At1g67110	CTAGATAGCAACCACTTCCACAA CAAGATTTGAGAAAGAGAAAAGAGAT
<i>HYH_h1</i>	At3g17609	TGTTTATGAGTATTTCCCGC AGTCAATCATCCGAAGCATTT
<i>HYH_h2</i>	At3g17609	CATGCAAAATAGATACACACACATA CTCAAAGTCTGCAACTCGTTCA
Genotyping of T-DNA lines		
T-DNA allele of GABI_267G03	At1g25550	CCCATTGGAGCGAATGTAGACAC (GABI LBP) AGTAGAAGATGTTCCGGGAGAT
WT allele for GABI_267G03	At1g25550	TGGAACCAATCACCAGATCC AGTAGAAGATGTTCCGGGAGAT
T-DNA allele of SALK_044835C	At1g68670	AGCAATCGCAGTCGCATACGCAT ATTTTGCCGATTTCGGAAC-(Lb1.3)
WT allele for SALK_044835C	At1g68670	AGCAATCGCAGTCGCATACGCAT ACCACCCTTGTGGCTACATCGGA
T-DNA allele of SALK_070096	At1g68670	TGATGGTGGAGATGGATTACG ATTTTGCCGATTTCGGAAC-(Lb1.3)
WT allele for SALK_070096	At1g68670	TGATGGTGGAGATGGATTACG GTTTTCTATGCGTATGCGGACTG
T-DNA allele of SAIL_28_D03	At3g25790	GAAATGGATAAATAGCCTTGCTTC (pCSA110_LB3) CTTTTCTTGTGAGCGATCCTG
WT allele for SAIL_28_D03	At3g25790	CSAGCTTCTCTTTGCGTAG CTTTTCTTGTGAGCGATCCTG
T-DNA allele of WiscDsLxHS231_10C	At3g25790	TGATCCATGTAGATTTCCGGACATGAAG (L4) CTTTTCTTGTGAGCGATCCTG
WT allele for WiscDsLxHS231_10C	At3g25790	CGAGCTTCTCTTTGCGTAG CTTTTCTTGTGAGCGATCCTG
T-DNA allele of SALK_067074	At1g13300	ATTTTGCCGATTTCGGAAC (pROK2_LB1.3) TCTTTTTTTCTTCCACTTGCCTAC
WT allele for SALK_067074	At1g13300	GTCGCAATTGCATAGACGCTTC TCTTTTTTTCTTCCACTTGCCTAC
T-DNA allele (<i>nlp6</i>) of SALK_036557	At1g64530	GTGCTGCAGTTATCAATTATCGATTTGGGACGCTT CTCGGTACCAGGGCAGTTCTTTGTAATTGGAT
WT allele (<i>NLP6</i>) for SALK_036557	At1g64530	GTGAGCCCTAGTTACTGTGCCATCAAAGCCAAG TCCTTAGAGATTTGTGCTGGATGA
T-DNA allele (<i>nlp7-1</i>) of SALK_026134	At4g24020	GTGCTGCAGTTATCAATTATCGATTTGGGACGCTT GTCAGGCCCTCAATCTCCAGTGTCTCGCAGGA
WT allele (<i>NLP7</i>) for SALK_026134	At4g24020	GTTGAATTCGCCACTGTTAACGGTGTGGTTAAG GTCAGGCCCTCAATCTCCAGTGTCTCGCAGGA
T-DNA allele (<i>phr1</i>) of SALK_067629C	At4g28610	CGCTGGACCGCTTGTGCAACT (pROK2_LB1) GTGCTGCAGTTATCAATTATCGATTTGGGACGCTT
WT allele (<i>PHR1</i>) for SALK_067629C	At4g28610	GTGCCATGGAGCTCGTCCAGTTCATAGA GTGCTGCAGTTATCAATTATCGATTTGGGACGCTT
T-DNA allele (<i>phl1</i>) of SAIL_731_B09	At5g29000	GTCGGATCCATGACTCTGGCTAATGATTTCCGGA TAGCATCTGAATTTCAATAACCACTCGATACAC (pCSA110-LB3)
WT allele (<i>PHL1</i>) for SAIL_731_B09	At5g29000	GTCGGATCCATGACTCTGGCTAATGATTTCCGGA GTCAGGCCATCTCTCTGACACGTTTCTTGA

^aCzechowski T, Stitt M, Altmann T, Udvardi MK, Scheible WR. Genome-wide identification and testing of superior reference genes for transcript normalization in Arabidopsis. *Plant Physiol.* 139, 5-17 (2005)

Supplementary References

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