SUPPLEMENTARY INFORMATION

Species-specific host factors rather than virus-intrinsic virulence determine primate lentiviral pathogenicity

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Supplementary Figure 1. Functional characterization and SIVagm proviral expression of the SIVgsn Vpu and HIV-1 NA7 Nef proteins. (a) Modulation of CD4 and AGM tetherin by HIV-1 WITO and SIVgsn71 Vpu. FACS analysis of HEK293T cells cotransfected with CD4 or AGM tetherin expression vectors and pCG plasmids expressing eGFP alone or together with the indicated *vpu* allele. (b) Human PBMCs or CD4+ T cells were stimulated with CD3/CD28 beads for three days, transduced with VSV-G pseudotyped vpu- and env-defective HIV-1 NL4-3 IRES eGFP constructs containing the indicated *nef* alleles and examined by flow cytometric analysis 2 days later. (c) Upmodulation of CD74 cell surface expression by HIV-1 NA7 and SIVagm Sab Nef proteins. THP-1 cells or human PBMCs were transduced and analyzed by flow cytometry for CD74 expression as described in panel b. Each symbol represents the result obtained in one independent experiment. (d) Western blot analysis of cellular extracts of HEK293T cells transfected with the indicated mutant SIVagm vpu Kozak variants. (e) Infectious virus production by HEK293T cells transfected with the indicated SIVagm GU1N constructs. Shown are average values (±SEM) of triplicate infections. (f) Modulation of CD3, MHC-I, CD28 and AGM tetherin in AGM PBMCs. The cells were stimulated with PHA for three days and then spinfected with the different SIVagm IMC. Surface expression by flow cytometric analysis was performed 2 days later.



Supplementary Figure 2. Replication of chimeric SIVagm constructs *in vivo*. (a) Viral RNA copy numbers in AGMs infected with the four SIVagm constructs. (b) Cell-associated viral loads in lymph nodes (Ln) at the indicated time points. 12 wpi ewas not determined for the GU group (c) Proviral copy numbers in blood-derived PBMCs at later time points (>170 wpi). Panel b and c show the mean \pm SEM.



Supplementary Figure 3. Humoral immune response and cellular activation in WT and chimeric SIVagm infection. (a) Humoral immune response to SIVagm as determined by Western immunoblot. Viral proteins were separated by SDS-PAGE, transferred to a nitrocellulose membrane, and subjected to immunoblot with sera obtained from the infected AGMs at the indicated weeks post-infection (wpi). P, preserum; gp, glycoprotein. (b) Changes in the numbers of activated (CD69+ and HLA-DR+) and proliferating (Ki67+) CD4+ and CD8+ T cells compared to the average numbers detected prior to infection (100%). (c-e) Frequencies of (c) naïve, (c) memory and (e) activated naïve T cell subsets at later time points (\geq 170 wpi). Shown are individual data points with the mean \pm SEM.



Figure S4b



Figure S4c



Supplementary Figure 4. Phylogenetic relationship of SIVagm sequences. (a-c) Phylogenetic analyses of sequences derived from the (a) WT, (b) 1N and (c) GU1N groups of infected AGMs. Panel b also includes sequences derived from the GU exposed animal 14627 that showed an unusual increase in plasma vRNA loads after >200 wpi. Maximum-likelihood phylograms were inferred using PhyML (Guindon et al. 2010) based on evolutionary substitution models favored by jModeltest (Daribba et al. 2012) for 3' half SGA sequences derived from the indicated animals at 121 or 223 wpi. Numbers at nodes are percent bootstrap support (only values >65% are shown) and asterisks indicate alleles selected for functional anaylsis of Vpu and/or Nef.

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Supplementary Figure 5. Alignment of Nef sequences derived from WT infected AGMs and AGM14634. (a) The WT SIVagmSab Nef sequence is shown on top and several known functional domains are indicated. The numbers represent the frequency of the respective sequence and the total number of *nef* alleles analyzed. (b) Alignment of Nef-fusions detected in AGM14634.



Supplementary Figure 6. Distribution of SIVagm GU in gut during the chronic phase of infection. Confocal images of SIV-infected cells (red), nucleus (blue) in duodenum (a), ileum (b) and jejunum (c) are shown. Representative SIV-specific in situ hybridization during chronic infection, demonstrating productively infected cells in duodenum, ileum and jejunum. The enlargements display representative examples of the shape of the SIV RNA positive cells. Duodenum sections from an uninfected monkey were used as control to demonstrate the specificity of the SIV probes. Pictures were obtained using a Leica SP8 confocal microscope and processed with ImageJ software.



Supplementary Figure 7. Uncropped scans of the Western blots shown in the indicated figures.