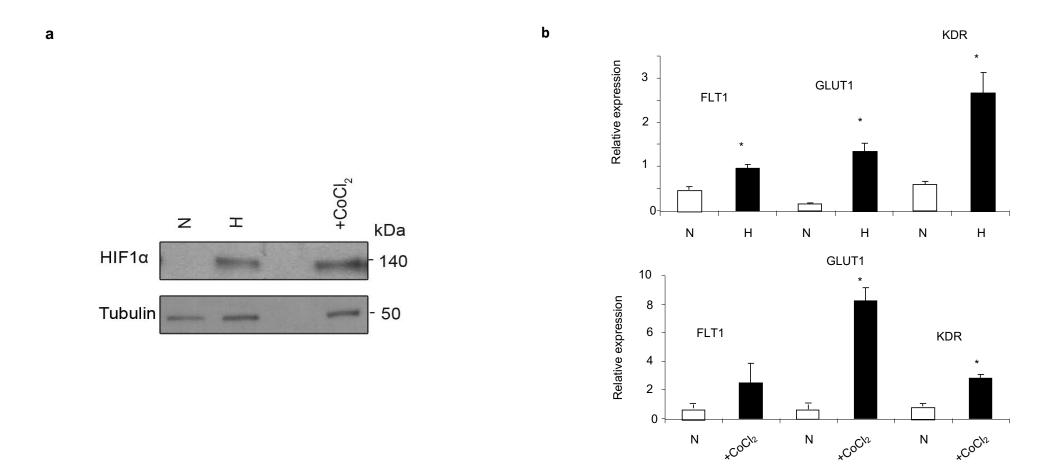
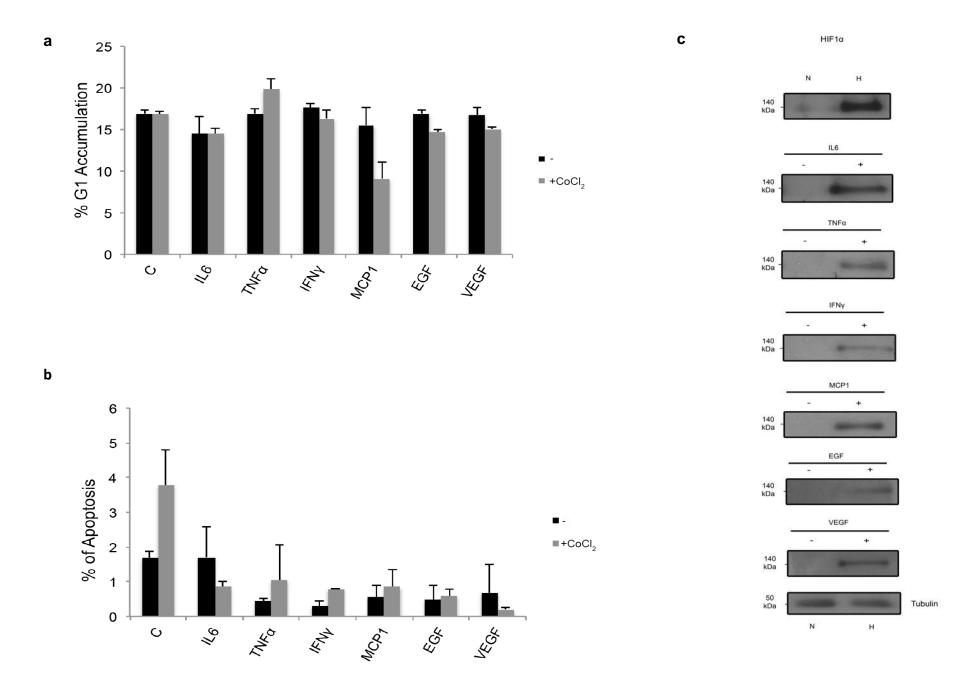
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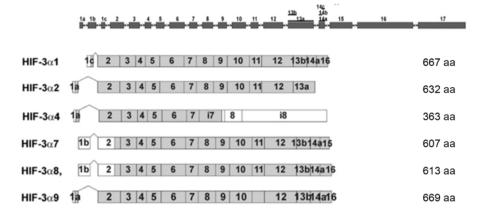
Pro-inflammatory cytokines activate hypoxia-inducible factor 3a via epigenetic changes in mesenchymal stromal/stem cells

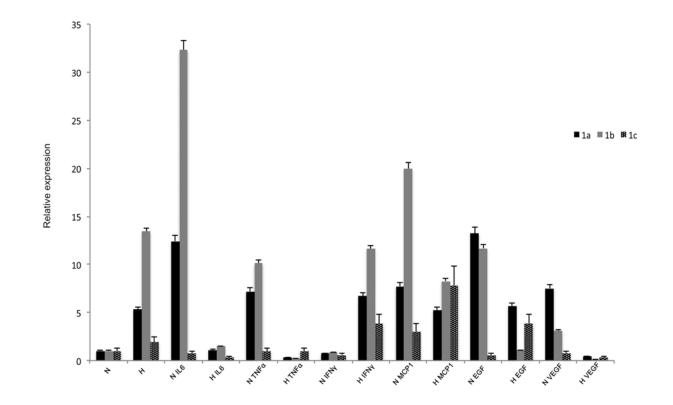
Francesca Cuomo^a, Antonietta Coppola^a, Chiara Botti^{a,§}, Ciro Maione^a, Amalia Forte^b, Lucia Scisciola^a, Giuseppina Liguori^c, Ilaria Caiafa^a, Matilde Valeria Ursini^d, Umberto Galderisi^b, Marilena Cipollaro^b, Lucia Altucci^a and Gilda Cobellis^a

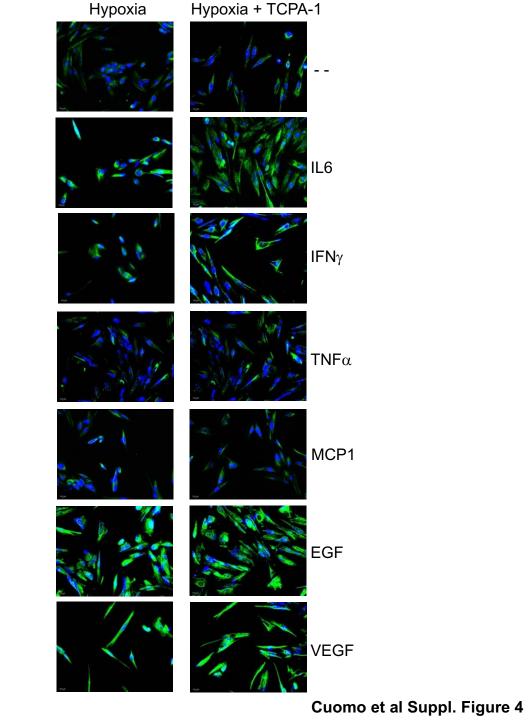




Cuomo et al Suppl. Figure 2







b

+CoCl₂

FΝγ

MCP1

EGF

VEGF

kDa

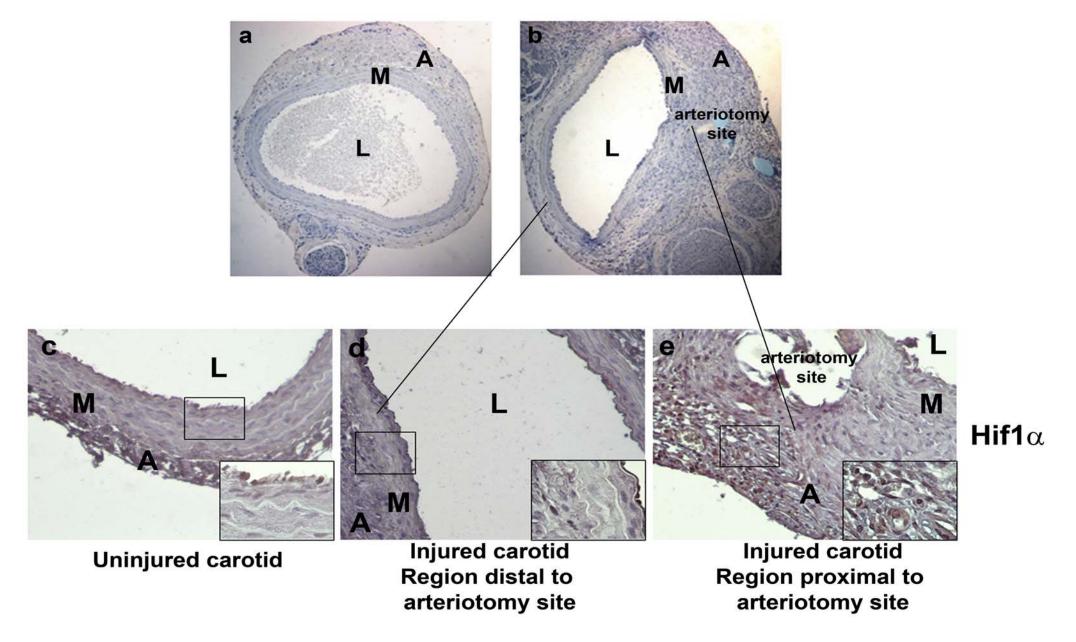
43

а

TCPA-1

IkBa

Erk 1/2



Supplementary Figure Legends

Supplementary Figure 1: a) Immunoblotting analysis of HIF1a protein in hMSC cells cultured either in 1%O2 conditions generated by GasPack method (H) or in presence of CoCl2 for 24 h. Total proteins were extracted and WB probed with antibodies against HIF1α. Tubulin was used as loading control. Protein molecular weights are shown. b) Expression analysis of FLT1, GLUT1 and KDR mRNAs in hMSC cells cultured either in 1%O2 conditions generated by GasPack method (H) or in presence of CoCl2 for 24 h. Gene expression data are reported as 2-DDCt method, normalized to housekeeping gene (beta-actin mRNA) and ALU sequences. Data are expressed as means ± SEM. Statistical significance is reported: *: p <0.05

Supplementary Figure 2: a) G1-phase accumulation: hMSCs were cultured in normoxic (N) and hypoxic (H) conditions and treated with IL6, TNF α , IFN γ MCP1, EGF, and VEGF for 24 h and analysed by flow cytometry using CellQuest software. b) Quantification of apoptotic cells in pre-G1 phase was performed by ModFit software. c) Immunoblotting analysis of HIF1a protein in hMSC cells cultured either in normoxic conditions or in presence of CoCl2 for 24 h in absence or in presence of indicated cytokines. Total proteins were extracted and WB probed with antibodies against HIF1 α . Tubulin was used as loading control. Protein molecular weights are shown. Images derived from different part of the same gel and cropped for layout reasons and included in Supplementary Information.

Supplementary Figure 3: a) Schematic representation of the HIF3A locus and relative isoforms, modified from 15.

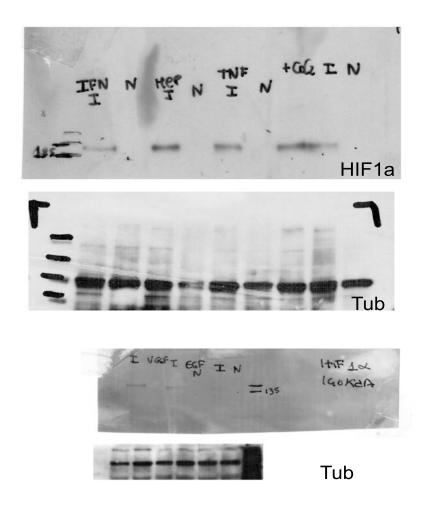
Supplementary Figure 4: a) Immunoblotting analysis of IkBa in hMSCs cultured in CoCl2- induced hypoxia conditions and treated with IL6, IFNg, TNFα, MCP1, EGF and VEGF for 24h, after an incubation with TCPA-1 for 1hr. b) Immunofluorescence analysis of HIF3a protein in hMSCs cultured in hypoxia in absence and in presence of indicated cytokines and probed with antibodies against HIF3a. Cells were pre-treated with TCPA-1 for 1h before cytokines supplementation. Scale bars: 10μm)

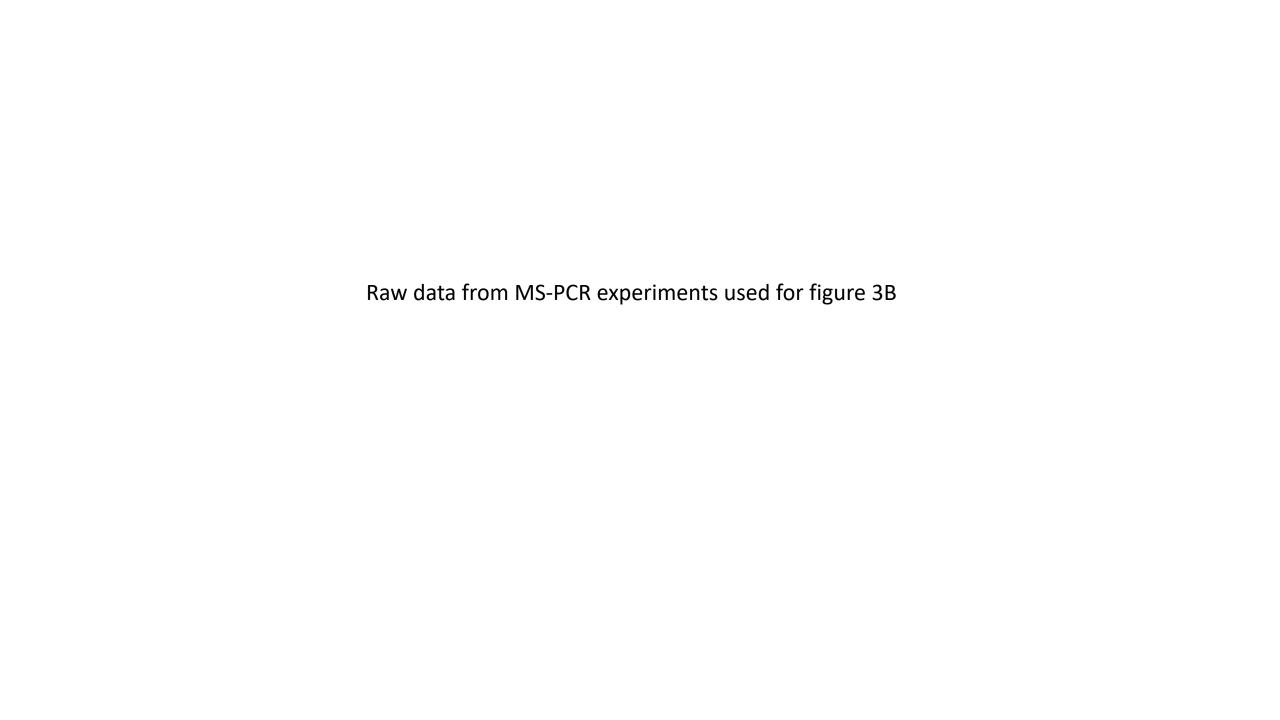
Supplementary Figure 5: Immunohistochemical analysis of Hif1α expression in uninjured rat carotids and in carotids from Wistar male rats harvested 7 days after arteriotomy. a: uninjured rat carotid; b: arteriotomy-injured rat carotid harvested 7 days after injury, haematoxylin staining. The injury site is indicated in b, where arteriotomy is followed by the application of an 8.0 polypropylene stitch (light blue); c-e: representative immunohistochemical staining of Hif1α in uninjured rat carotid (c) and in injured carotids harvested 7 days after arteriotomy, showing a carotid region distal (d) and proximal (e) to the arteriotomy site. a,b: 10x magnification; c-e: 20x magnification; small insets: 40x magnification of selected areas enclosed in black rectangles, representative of nuclei positive to Hif1α in intima, adventitia, *vasa vasorum* and perivascular tissue. Brown staining corresponds to target protein expression. Nuclei were counterstained with haematoxylin. L: lumen; M: media; A: adventitia.

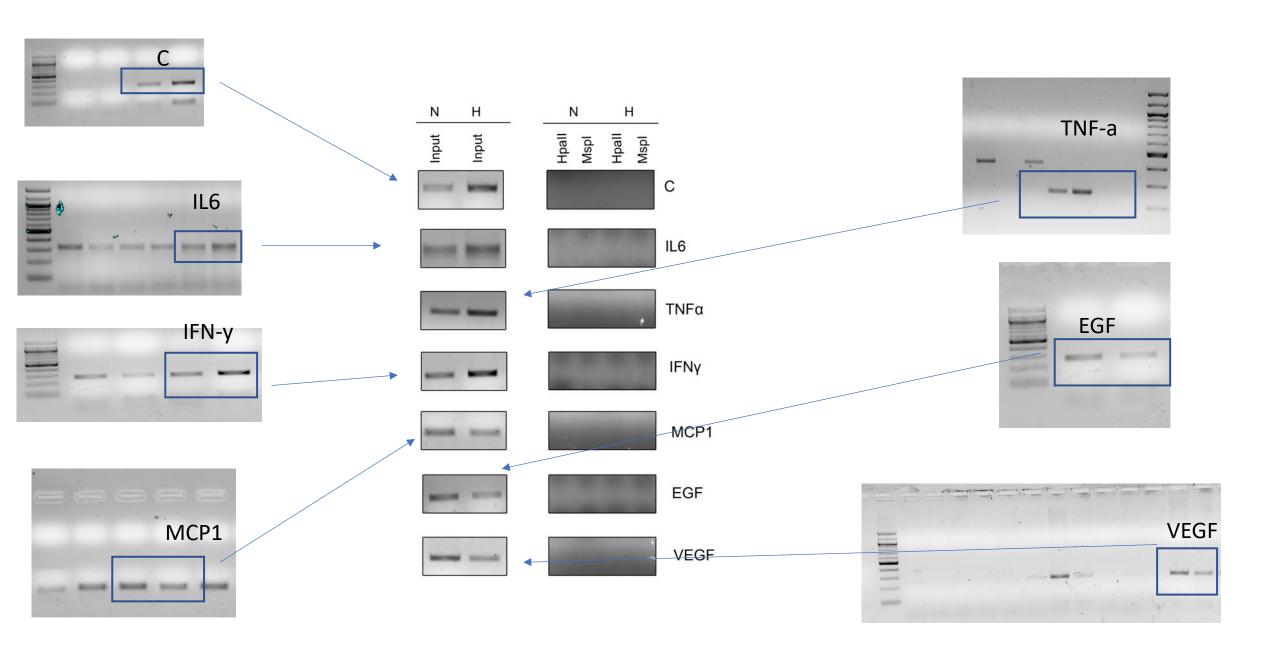
Supplementary Figure 7: Raw data (8 panels) from MS-PCR experiments used for figure 3B

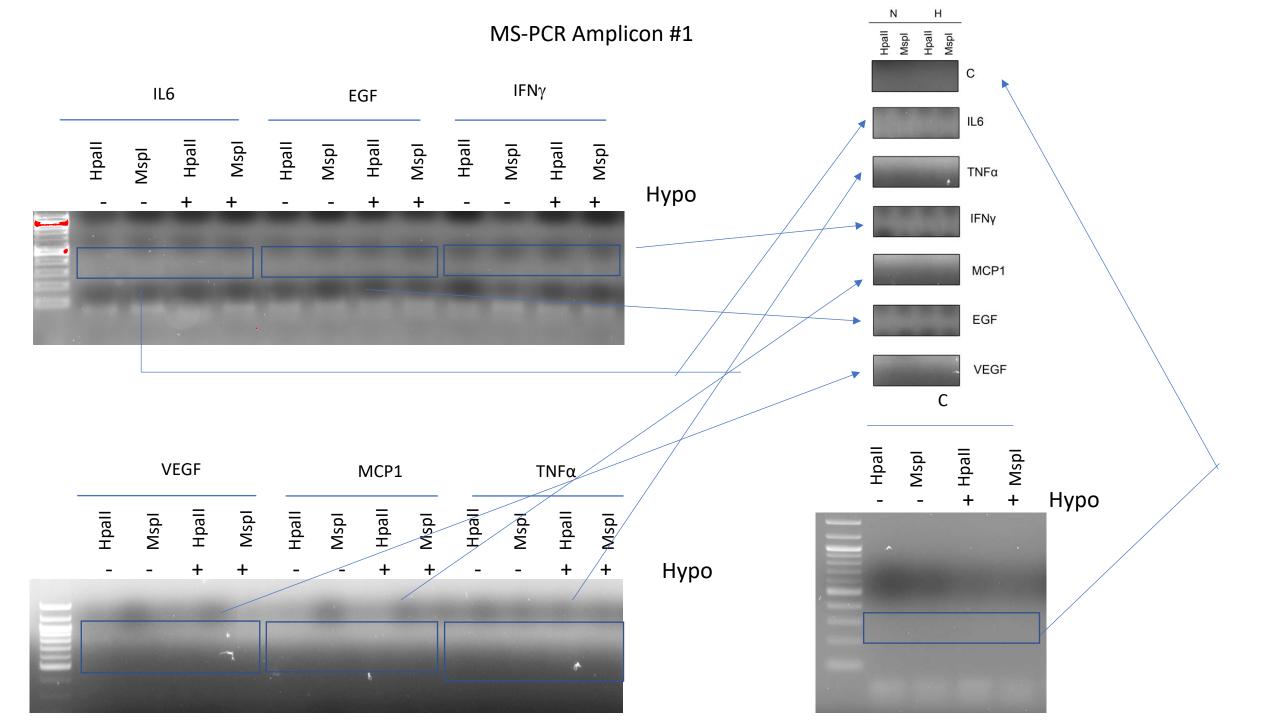
Supplementary Figure 8: Raw data (8 panels) from replicate MS-PCR experiment.

Raw data of HIF1a expression in normoxic, hypoxic and CoCl₂-induced hypoxia in Suppl. Figure 2

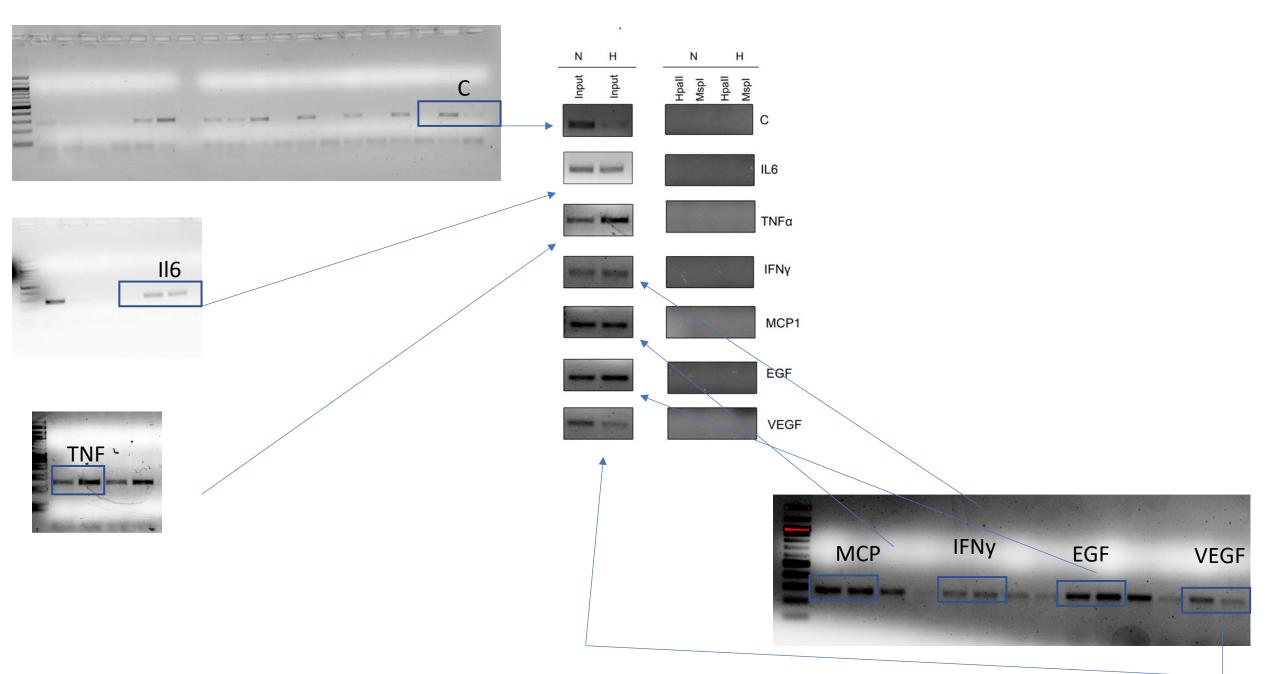


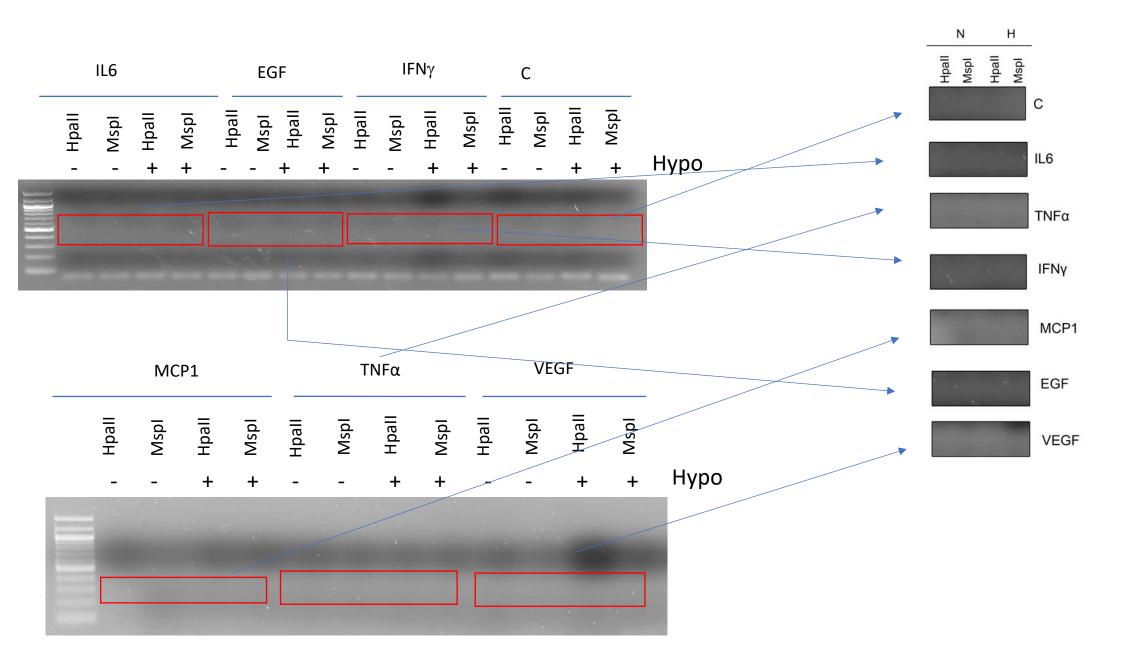




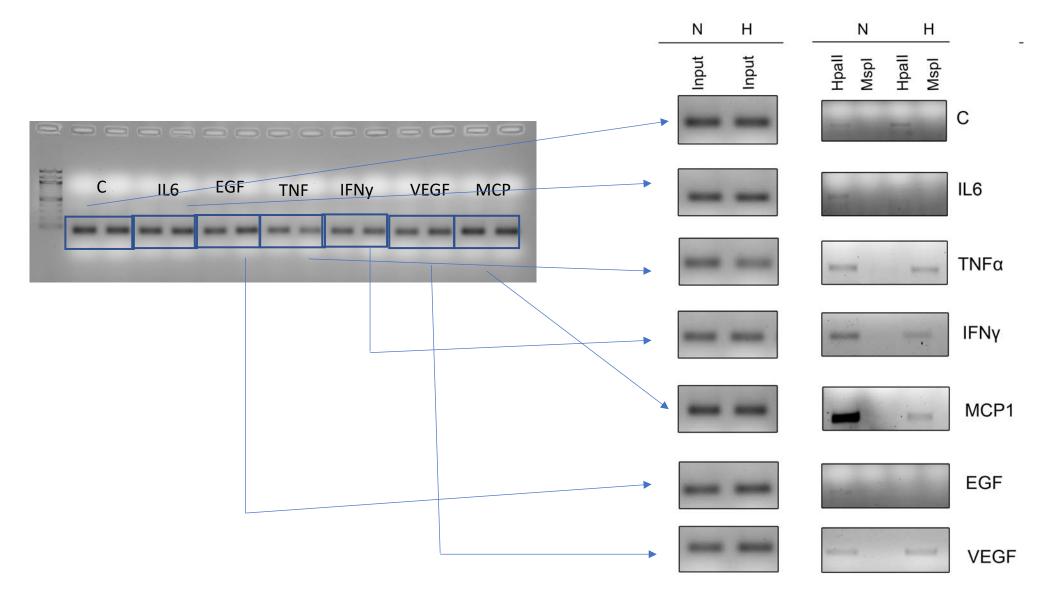


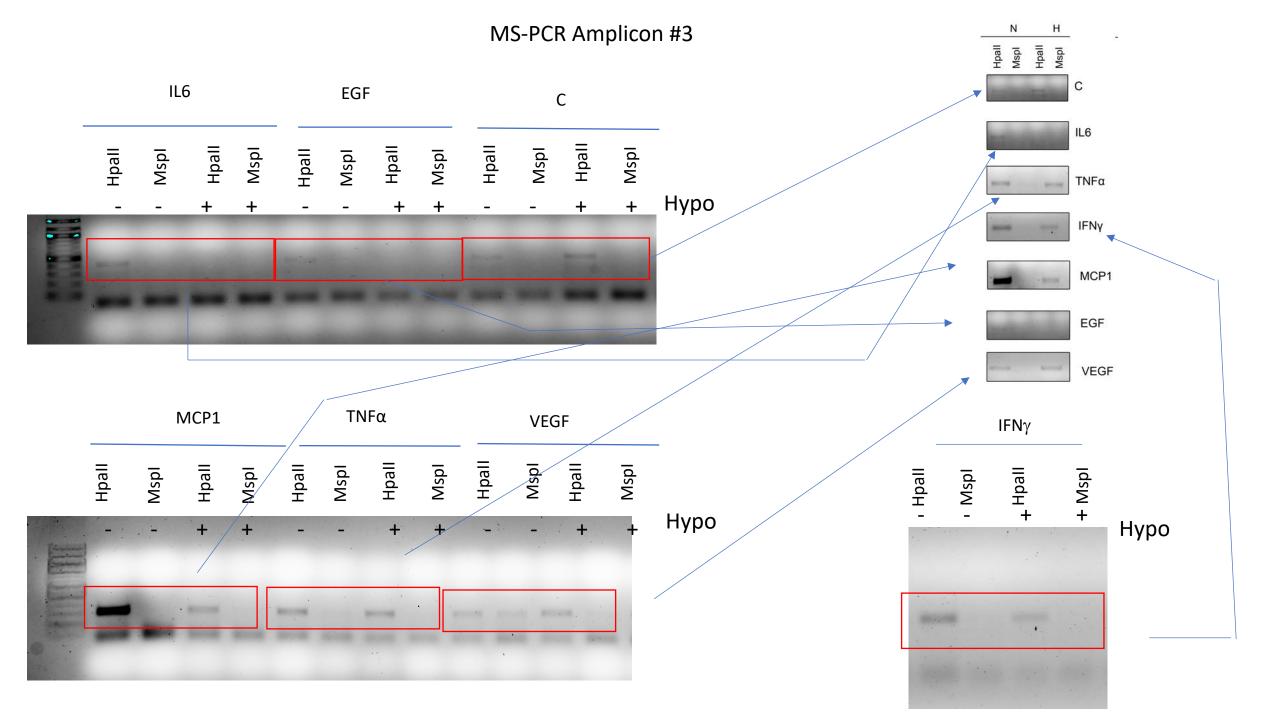
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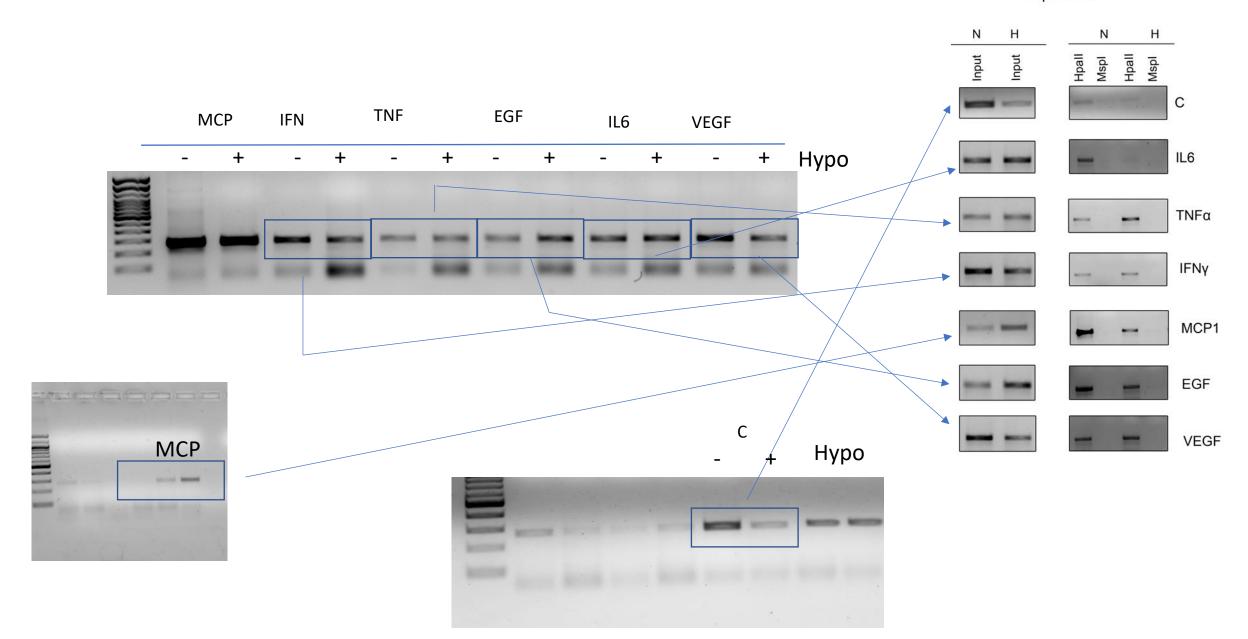


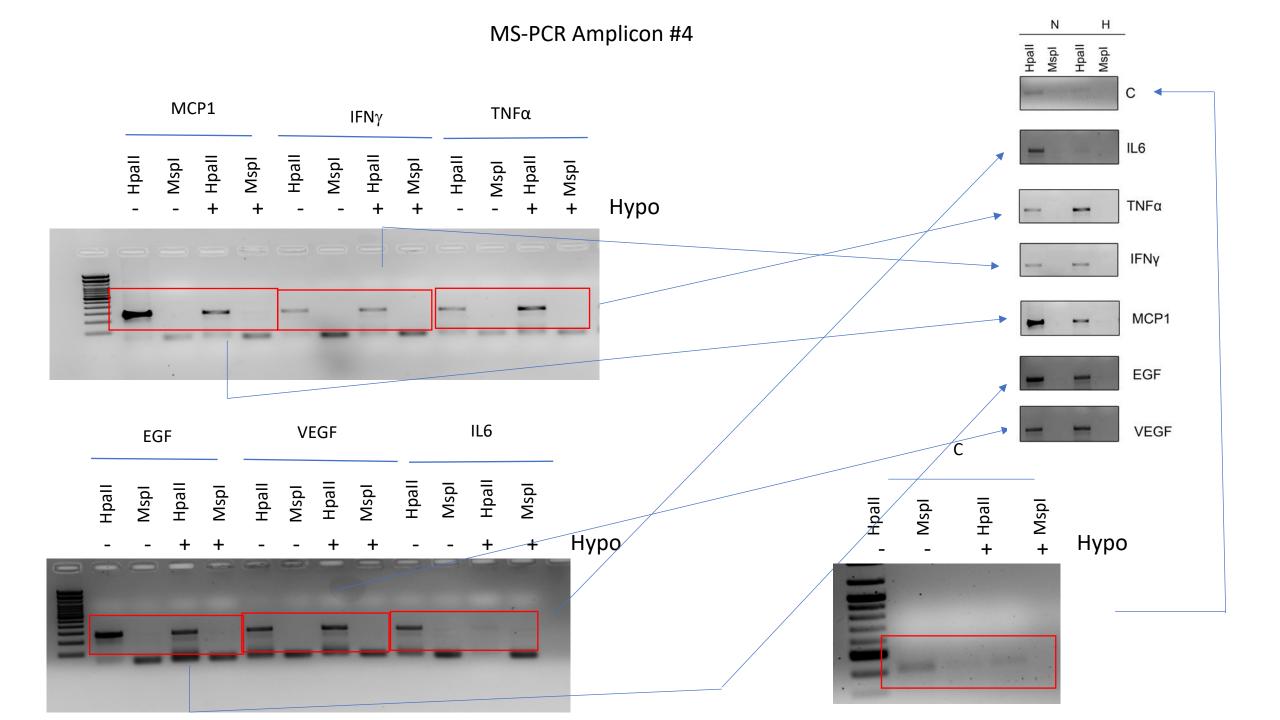
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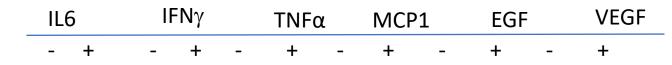


amplicon #4



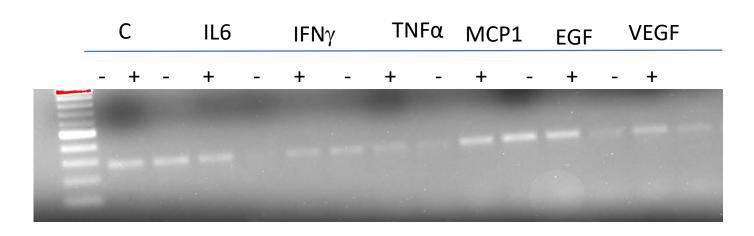


AMPLICON INPUT #1

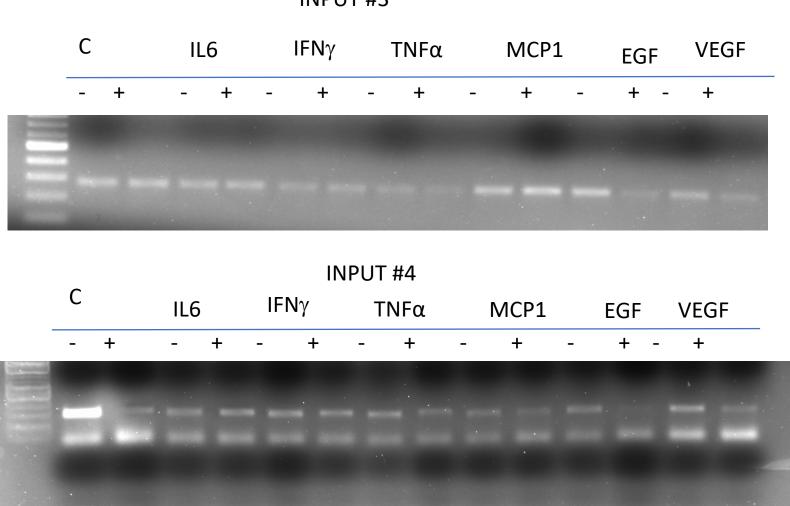




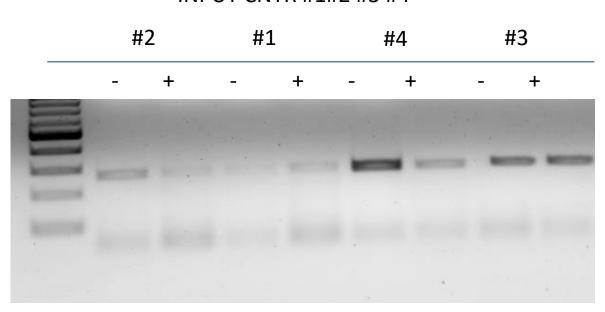
AMPLICON INPUT #2

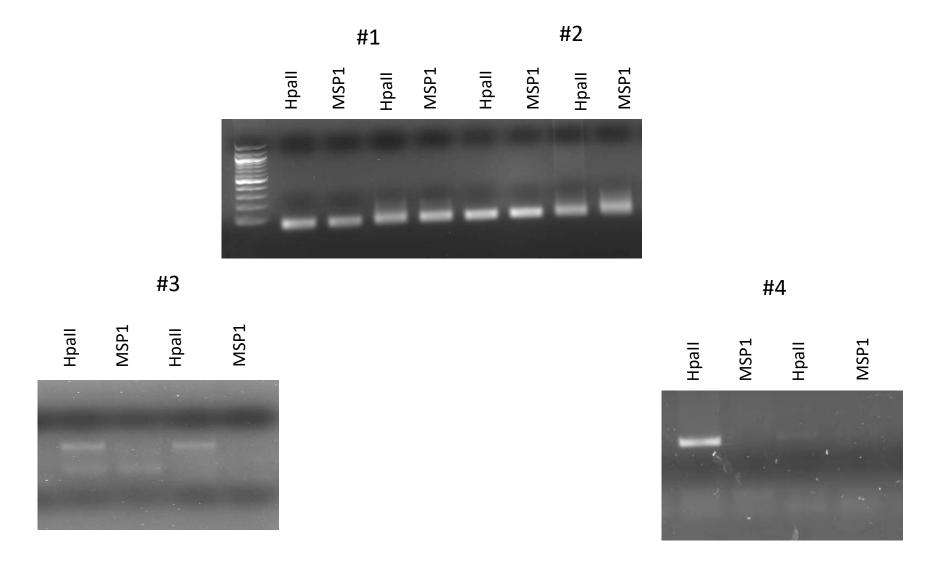












Raw Replicate data from MS-PCR experiment: amplicon #1

_	l		EGF	:		I	Ι ΓΝ γ					
	Нраш	MSP1	Нраш	MSP1	Hpall	MSP1	Hpall	MSP1	Hpall	MSP1	Hpall	MSP1
							Ħ		Ħ			
				•		d			m	M		

	VEG	F			M	CP1		TNF			
Hpall	MSP1										

Raw Replicate data from MS-PCR experiment: amplicon #2

	IL6					EGF			ΙΕΝγ				С				
-	Hpall	0.00	MSFI	Hpall	MSP1												
										•							

	MC	:P1			٦	ΓΝFα		VEGF			
Hpall	MSP1										
											Ξ

Raw Replicate data from MS-PCR experiment: amplicon #3

= 7							VEGF				
Hpall MSP1	Hpall	MSP1	Нраш	MSP1	Нраш	MSP1	Hpall	MSP1	Нраш	MSP1	
								ij		u	

_		MCI	P1			TNF	α		VEGF			
	Hpall	MSP1	Hpall	MSP1	Нраш	MSP1	Hpall	MSP1	Hpall	MSP1	Hpall	MSP1

Raw Replicate data from MS-PCR experiment: amplicon #4

	ſ	MCP1			V	EGF			EGF			
Нраш	MSP1	Hpall	MSP1									
				*		-	7 4			,		

	IFNγ	′			TNF				IL6				
Hpall	MSP1	Hpall	MSP1	Hpall	MSP1	Нраш	MSP1	Нраш	MSP1	Нраш	MSP1		