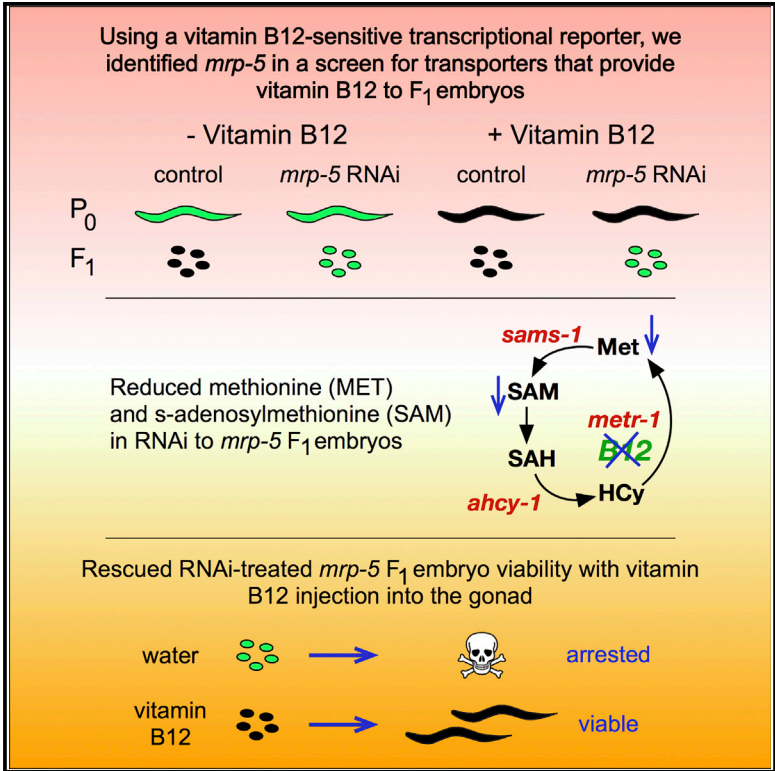


Cell Reports

C. elegans MRP-5 Exports Vitamin B12 from Mother to Offspring to Support Embryonic Development

Graphical Abstract



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In Brief

How dietary vitamin B12 is transported from the mother to developing offspring is unknown. Na et al. demonstrate that the ABC transporter *MRP-5* (multidrug resistance protein 5) transports vitamin B12 from the intestine of *C. elegans* mothers to the offspring to promote embryonic viability and development

Highlights

- ABC transporter MRP-5 transports vitamin B12 from mother to offspring in *C. elegans*
- *mrp-5* mutant embryonic lethality is rescued by vitamin B12 injection into gonad
- RNAi of *mrp-5* reduces S-adenosylmethionine and methionine content in F1 embryos



C. elegans MRP-5 Exports Vitamin B12 from Mother to Offspring to Support Embryonic Development

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SUMMARY

Vitamin B12 functions as a cofactor for methionine synthase to produce the anabolic methyl donor S-adenosylmethionine (SAM) and for methylmalonyl-CoA mutase to catabolize the short-chain fatty acid propionate. In the nematode *Caenorhabditis elegans*, maternally supplied vitamin B12 is required for the development of offspring. However, the mechanism for exporting vitamin B12 from the mother to the offspring is not yet known. Here, we use RNAi of more than 200 transporters with a vitamin B12-sensor transgene to identify the ABC transporter MRP-5 as a candidate vitamin B12 exporter. We show that the injection of vitamin B12 into the gonad of *mrp-5* deficient mothers rescues embryonic lethality in the offspring. Altogether, our findings identify a maternal mechanism for the transit of an essential vitamin to support the development of the next generation.

INTRODUCTION

Maternal micronutrient status during pregnancy greatly affects embryonic development and fetal health (Dror and Allen, 2008; Fall et al., 2003; Owens and Fall, 2008; Skjærven et al., 2016). For instance, the intake of folate supplements by pregnant women reduces the occurrence of neural tube defects in newborns (Czeizel and Dudás, 1992; Viswanathan et al., 2017). Folate functions together with vitamin B12, or cobalamin, in the one carbon cycle to recycle methionine from homocysteine (Moreno-García et al., 2013) (Figure 1A). One of the main functions of the one-carbon cycle is to produce S-adenosylmethionine (SAM), a major methyl donor required to synthesize phosphatidylcholine, an important component of cellular membranes, and to modify histones and DNA to ensure proper genomic functions (Rosenblatt and Whitehead, 1999). The other function of vitamin B12 is as a cofactor in the breakdown of the short-chain fatty acid propionate (Shchelochkov et al., 2016) (Figure 1A). Humans obtain vitamin B12 through the ingestion of animal products in which this stable micronutrient is efficiently stored (Martens et al., 2002). Deficiency of vitamin B12 in newborns is mostly caused either by maternal depletion or malab-

sorption, although rare recessive inborn errors of vitamin B12 processing or utilization have also been identified (Rosenblatt and Cooper, 1990; Rosenblatt and Whitehead, 1999).

Recently, the nematode *Caenorhabditis elegans* has been established as a powerful model for studying vitamin B12-dependent processes (Yilmaz and Walhout, 2014). *C. elegans* is a bacterivore that can be fed individual bacterial species and strains that can elicit different effects on the animal's life history traits and gene expression. For instance, when fed *Comamonas aquatica* DA1877 bacteria, the animals develop faster, have reduced fecundity, and have a shorter lifespan relative to animals fed the standard laboratory diet of *Escherichia coli* OP50 (MacNeil et al., 2013; Watson et al., 2013). These dietary differences are due, at least in part, to the fact that *Comamonas* synthesizes vitamin B12, whereas *E. coli* does not (Watson et al., 2014). Dietary vitamin B12 is required to support *C. elegans* development, as the animal itself cannot synthesize this cofactor (Bito et al., 2013; Watson et al., 2014). Vitamin B12 is ingested by the mother and needs to be exported from the intestine into the gonad to support the development of her offspring. However, the mechanism by which this occurs is not yet known. Here, we report that the ABC transporter *mrp-5* controls vitamin B12 export from the intestine to support *C. elegans* embryonic development.

RESULTS

An RNAi Screen Identifies the ABC Transporter MRP-5 as a Candidate to Export Vitamin B12 from the *C. elegans* Intestine

We have previously established the *Pacdh-1::GFP* transgenic *C. elegans* strain as a reporter of dietary vitamin B12 status: in this strain, GFP is highly expressed when vitamin B12 is low and repressed when levels of this micronutrient are high (Arda et al., 2010; MacNeil et al., 2013; Watson et al., 2014; Watson et al., 2016). GFP expression in this strain is under the control of the promoter of the acyl-coenzyme A (CoA) dehydrogenase *acdH-1*. This gene encodes the first enzyme of an alternate propionate breakdown pathway, or propionate shunt, which does not require vitamin B12 and is transcriptionally activated when this cofactor is in low supply (Watson et al., 2016)(Figure 1A). We reasoned that a defect in vitamin B12 transport from the mother to her offspring would result in parental generation (P)0 animals with low levels of GFP in the presence of vitamin



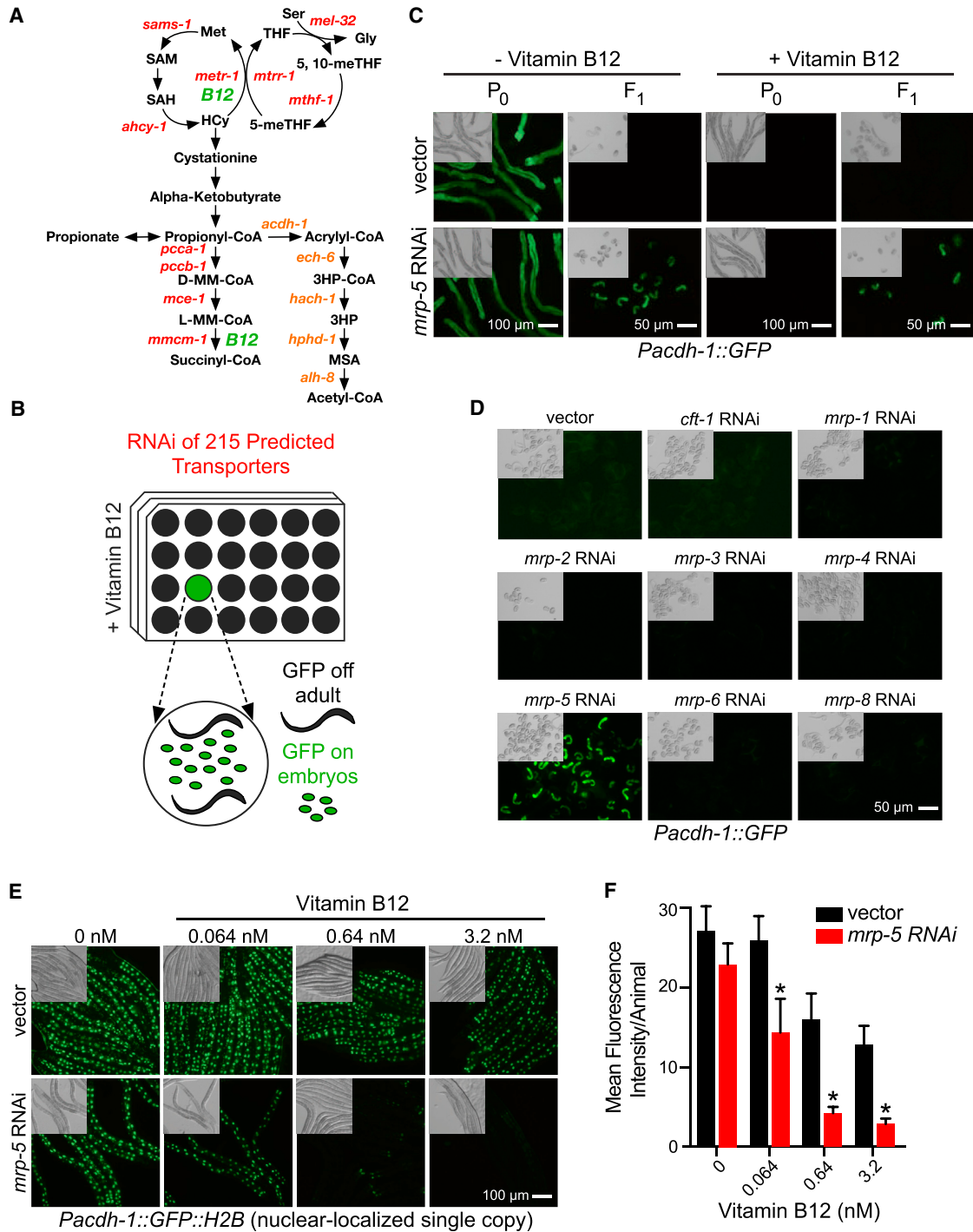


Figure 1. A *C. elegans* Transporter RNAi Screen Identifies *mrp-5* as a Potential Vitamin B12 Exporter

(A) Cartoon of the two vitamin-B12-dependent metabolic pathways. Gene names are from *C. elegans*. 3HP, 3-hydroxypropionate; 5-meTHF, 5-methyltetrahydrofolate; 5,10-meTHF, 5,10-methylenetetrahydrofolate; D-MM-CoA, D-methylmalonyl-CoA; Gly, glycine; HcY, homocysteine; L-MM-CoA, L-methylmalonyl-CoA; Met, methionine; MSA, malonic semialdehyde; SAH, S-adenosylhomocysteine; SAM, S-adenosylmethionine; THF, tetrahydrofolate. The *acdH-1* branch in orange text indicates the vitamin-B12-independent propionate shunt.

(B) Diagram of *C. elegans* transporter RNAi library screen. RNAi-treated animals that displayed GFP fluorescence in the F1, but not the P0, generation in the presence of 64 nM vitamin B12 were considered hits.

(C) Fluorescence and differential interference contrast (DIC) microscopy images of *Pacdh-1::GFP* animals subjected to RNAi of *mrp-5* in *E. coli* HT115 compared to vector control at P0 and F1 generations in the presence or absence of 64 nM vitamin B12. Scale bars, 50 μ m for embryos and 100 μ m for adults.

(legend continued on next page)

B12 and F1 embryos with high levels of GFP expression. Further, these embryos would be expected to have developmental defects, because vitamin B12 is required to produce SAM, which is needed to generate biomass and, thus, support development (Watson et al., 2014). We performed RNAi knockdown of 215 predicted *C. elegans* transporters of the ABC transporter and solute carrier families by feeding animals *E. coli* HT115 bacteria expressing double-stranded RNA in the presence of 64 nM vitamin B12 (Figure 1B). We identified a single gene, *mrp-5*, that, when knocked down, resulted in mothers with low levels of GFP expression and dead embryos with high GFP expression (Figure 1C). *mrp-5* encodes an ABC transporter belonging to the multi-drug resistance protein (MRP) family (Korolnek et al., 2014). MRP proteins are highly similar, and the *C. elegans* genome encodes nine family members (<http://www.wormbase.org>). To test for specificity, we retested eight of the nine members of this family for which RNAi clones were available and found that only *mrp-5* RNAi caused activation of GFP expression in the F1 generation (Figure 1D). We then titrated vitamin B12 from 6.4 nM to 6.4 μ M and found that GFP expression remains activated in *mrp-5* RNAi embryos even under very high vitamin B12 conditions (Figure S1A).

MRP-5 is expressed in the intestine (Korolnek et al., 2014) (Figure S2). If *mrp-5*, indeed, encodes a transporter that exports vitamin B12 from the *C. elegans* intestine to support embryonic development, this would lead to the prediction that higher levels of vitamin B12 should be retained in the intestine when the transporter is perturbed. To test this prediction, we devised a highly sensitive experimental setup by using (1) a single-copy *Pacdh-1::GFP::H2B* strain that expresses GFP in the nucleus, enabling easier monitoring of differences in GFP levels, and (2) an RNAi-compatible strain of *E. coli* OP50 (Xiao et al., 2015), because animals fed this diet express higher levels of GFP than those fed *E. coli* HT115 (Figure S1B). Importantly, RNAi of *mrp-5* is equally efficient in *E. coli* HT115 and *E. coli* OP50, as measured by qRT-PCR (Figure S1C). We found that much lower concentrations of vitamin B12 were sufficient to repress intestinal GFP in *mrp-5* RNAi animals compared to vector control animals (Figures 1E and 1F). This finding suggests that vitamin B12 is retained in the intestine when *mrp-5* is knocked down. Together, these findings indicate that *mrp-5* encodes a transporter that exports vitamin B12 from the intestine of the mother to support embryonic development of her offspring.

Injection of Vitamin B12 into the Gonad Rescues Embryonic Lethality Caused by Loss of *mrp-5*

We predicted that direct supplementation of vitamin B12 into the gonad should bypass the requirement for vitamin B12 export from the intestine and rescue embryonic lethality caused by perturbation of *mrp-5*. To test this prediction, we injected

either vitamin B12 or water (vehicle) as a control into the gonad of *Pacdh-1::GFP* animals that were exposed to *mrp-5* or vector control RNAi. If the vitamin B12 injection into the gonad bypassed MRP-5-dependent export, we expected to observe live animals with low GFP expression, while injecting water would lead to dead embryos with high GFP expression (Figure 2A). For this experiment, we used the *E. coli* OP50 RNAi strains we generated (see also Figure 1E), because transgenic *Pacdh-1::GFP* embryos express high levels of GFP, while they express low GFP levels when fed *E. coli* HT115 bacteria (Figure S1). Using *E. coli* OP50 thus enabled us to observe a decrease in GFP expression, while using *E. coli* HT115 would not.

Injection of vitamin B12 into the gonad of *mrp-5* RNAi animals fully and specifically rescued the embryonic viability of their offspring (Figure 2B). In addition, in *Pacdh-1::GFP* transgenic animals exposed to *mrp-5* RNAi, injecting vitamin B12 into the gonad resulted in developed larvae with low levels of GFP, while injection of water resulted in dead embryos with high GFP expression (Figure 2B). We recapitulated these findings with *mrp-5(ok2067)* deletion mutant animals. Since *mrp-5* is an essential gene, the *mrp-5* deletion allele is maintained genetically balanced with *+/svT1[lon-2(e678)]* (Figure 2C). Heterozygous *mrp-5 (+/-)* mothers injected with vitamin B12 into their gonad produced *mrp-5 (-/-)* offspring that survived to the adult stage, while animals injected with vehicle control produced *mrp-5 (-/-)* animals that arrested at the L1–L2 stage (Figure 2D). Importantly, the offspring of the rescued F1 *mrp-5 (-/-)* animals again exhibited larval arrest, suggesting that, in the absence of *mrp-5*, vitamin B12 cannot be passed on to the next (F2) generation. Altogether, these findings show that vitamin B12 injected into the gonad of a *mrp-5 (+/-)* mother can rescue embryonic lethality in her offspring.

Heme Rescues Embryonic Lethality in *mrp-5* RNAi Animals

A previous study proposed that *mrp-5* encodes a heme transporter (Korolnek et al., 2014). This finding was supported by the observation that embryonic lethality in *mrp-5* RNAi animals could be rescued by feeding the animals high doses of heme (Korolnek et al., 2014). How could either feeding of heme or injection of vitamin B12 rescue embryonic lethality of *mrp-5* RNAi or mutant animals? ABC transporters are known to be capable of transporting numerous similar types of molecules (Lee and Rosenbaum, 2017; Locher, 2016; Slot et al., 2011). Heme and vitamin B12 share some structural similarities; heme is an iron-containing porphyrin, while vitamin B12 is related to porphyrins and contains cobalt as a metal ion (Figure 3A). Thus, it is possible that MRP-5 transports both molecules. However, if lack of heme and vitamin B12 explains the embryonic lethality of *mrp-5* RNAi

(D) Fluorescence and DIC microscopy images of *Pacdh-1::GFP* animals subjected to RNAi of *mrp* family members in *E. coli* HT115 compared to control RNAi in the F1 generation. An RNAi clone for *mrp-7* was not available. Scale bar, 50 μ m.

(E) Fluorescence microscopy images of single-copy insertion *Pacdh-1::GFP::H2B* animals subjected to *mrp-5* RNAi in *E. coli* OP50 compared to control RNAi with low-dose titration of vitamin B12. Scale bar, 100 μ m.

(F) Images from (E) were quantitated using ImageJ to determine the mean fluorescence intensity per animal. At least five animals were analyzed per condition. Error bars represent SD. The p value was determined by unpaired multiple t tests, with a false discovery rate of 1% (*p < 0.0001).

See also Figures S1 and S2.

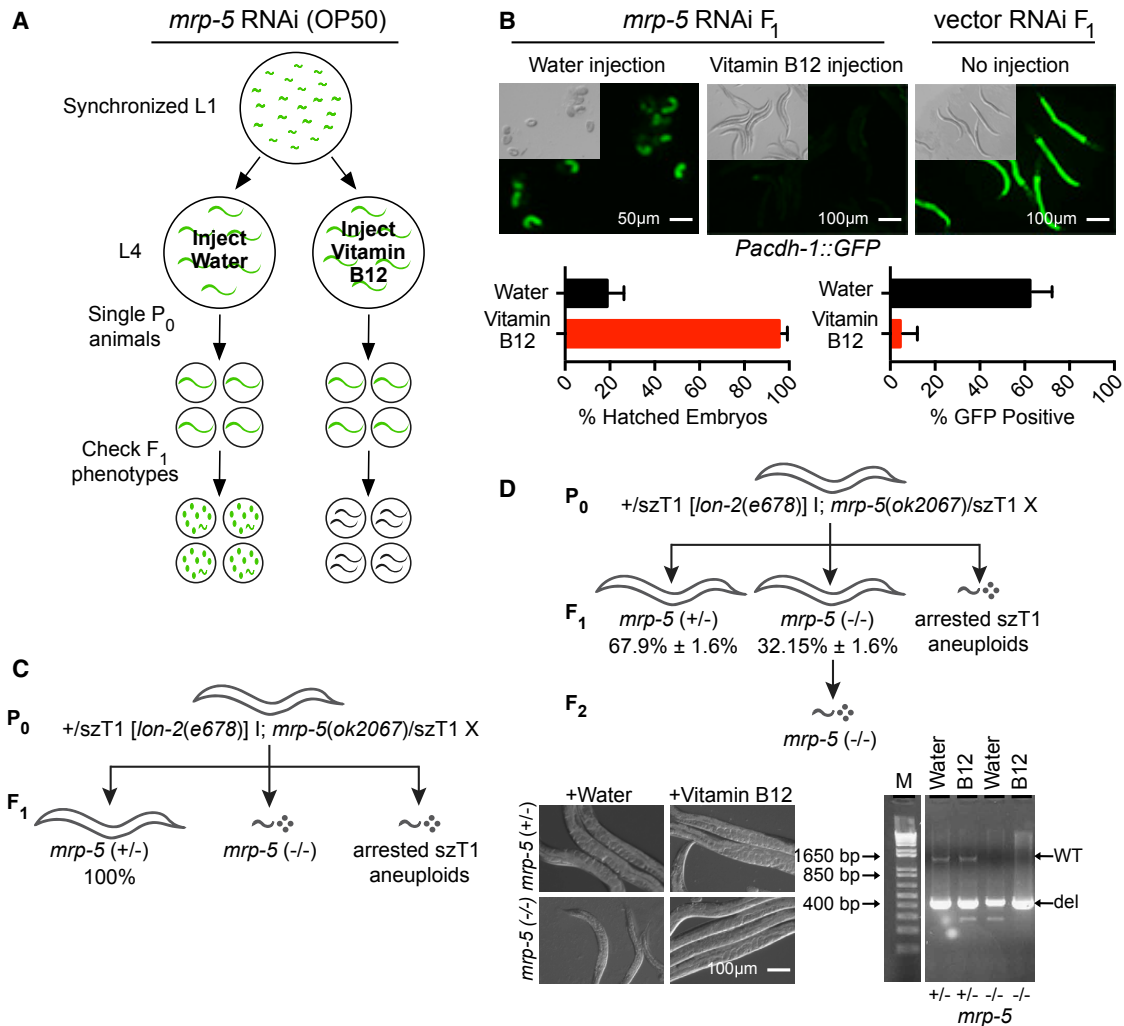


Figure 2. Injection of Vitamin B12 into the Gonad of *mrp-5*-Deficient Mothers Rescues Lethality in the Offspring

(A) Experimental design. Briefly, either 3.2 mM vitamin B12 or water (vehicle) as a control was injected into the gonad of *Pacdh-1::GFP* mothers that were exposed to *mrp-5* or vector control RNAi. If the vitamin B12 injection rescues embryonic lethality, live offspring will show low GFP expression, while water-injected animals will show dead embryos with high GFP expression.

(B) Top: DIC and fluorescence microscopy images of embryos from *Pacdh-1::GFP* mothers subjected to *mrp-5* RNAi in *E. coli* OP50 injected into the gonad with water (left panel) or with 3.2 mM vitamin B12 (middle panel). Animals treated with vector control RNAi without injection are shown as a control (right panel). Scale bars, 50 µm for embryos and 100 µm for adults. Bottom: quantification of lethality and GFP expression in embryos from *Pacdh-1::GFP* animals described above. Error bars represent SD of the average of three independent experiments.

(C) Diagram of genotypes and phenotypes of the offspring of genetically balanced *mrp-5 (+/-)* mutant animals. All P0 adult animals are heterozygous for *mrp-5*. F1 larval arrest or embryonic lethality occurs in *mrp-5 (-/-)* or szT1 aneuploid animals.

(D) Top: diagram of genotypes and phenotypes of viable offspring from genetically balanced *mrp-5 (+/-)* mutant animals after vitamin B12 injection. All P0 adult animals are heterozygous for *mrp-5*. Live F1 animals can be either heterozygous or homozygous for the *mrp-5(ok2067)* deletion allele. The average percentage and SD of the total number of L1-L2 animals is listed below the genotype. F2 larval arrest or embryonic lethality occurs in all offspring of the F2 generation of *mrp-5 (-/-)* animals. Bottom left: DIC microscopy images of *mrp-5 (+/-)* and *mrp-5 (-/-)* animals with or without vitamin B12 injection. Scale bar, 100 µm. Bottom right: agarose gel image of the genotyping results of heterozygous and homozygous *mrp-5* L1-L2 animals with and without injection of vitamin B12.

animals, one would expect that neither cofactor alone would be sufficient for rescue. Our observation that embryonic lethality can be rescued by injection of vitamin B12 into the gonad indicates that lack of vitamin B12 rather than heme may be the cause of embryonic lethality when MRP-5 function is perturbed. To further examine the effects of heme on animal development, we repeated the experiment published in the previous study

(Korolnek et al., 2014) and confirmed that feeding 500 µM heme can rescue embryonic lethality in the offspring of *mrp-5* RNAi animals (Figure 3B). Interestingly, however, such a high dose of heme was toxic to developing control animals, arresting most animals at the L1-L2 stage, whereas high doses of vitamin B12 were not toxic (Figure S3). Importantly, one can argue that feeding heme does not actually test for transport, because the

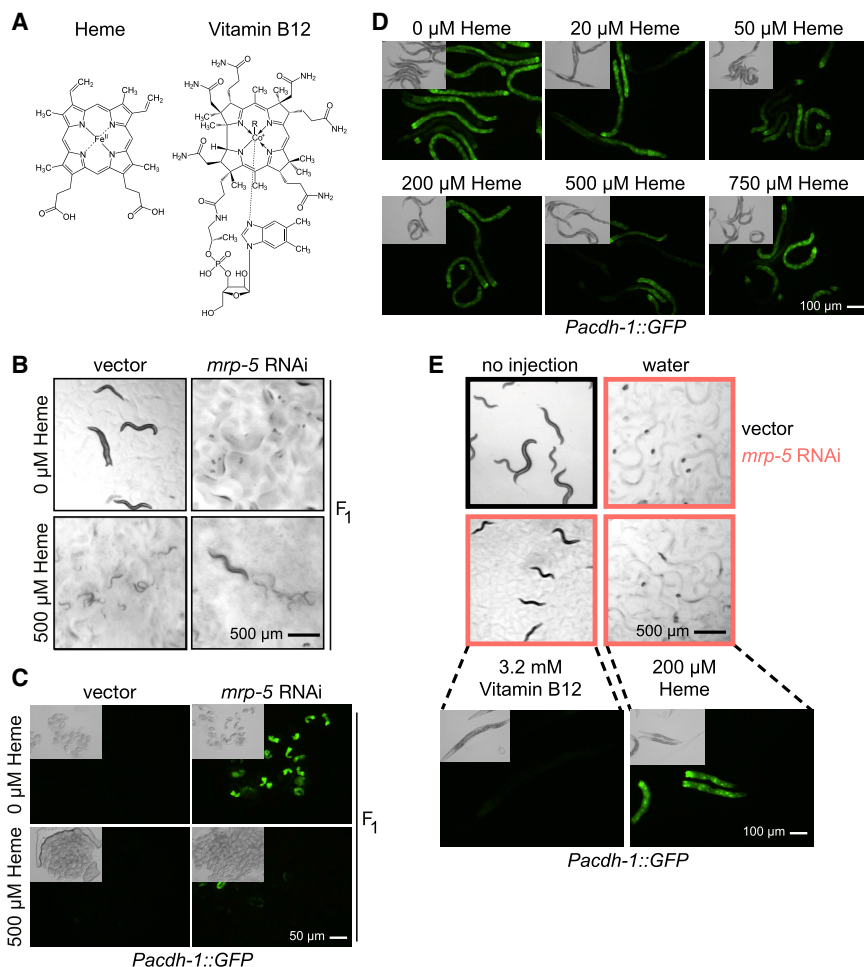


Figure 3. High Concentrations of Heme Rescue Embryonic Lethality in *mrp-5* RNAi Animals but Do Not Affect *acdh-1* Promoter Activity

(A) Heme and vitamin B12 are porphyrin or porphyrin-like molecules with structural similarities. (B) Bright-field images of feeding 500 μ M heme to *mrp-5* RNAi-treated mothers on *E. coli* HT115 rescues viability in her offspring and allows them to develop to the L2 stage and later but causes L1 larval arrest and toxicity in vector-treated mothers. 24 singled mothers plus their offspring were analyzed for each condition. Scale bar, 500 μ m.

(C) Fluorescence and DIC microscopy images show that feeding 500 μ M heme to *mrp-5* RNAi-treated mothers on *E. coli* HT115 reduces *Pacdh-1::GFP* reporter activity in embryos but not in the embryos of vector control mothers. 50 L1 mothers were seeded onto 6-cm plates, and F1 embryos were collected for imaging. Scale bar, 50 μ m.

(D) Fluorescence and DIC microscopy images demonstrate that feeding heme to P0 mothers with *E. coli* OP50 does not affect *Pacdh-1::GFP* reporter activity. Scale bar, 100 μ m.

(E) Bright-field (top) and fluorescence and DIC microscopy images (bottom) shows that injection of 200 μ M heme into *mrp-5* RNAi-treated mothers with *E. coli* HT115 does not rescue embryonic lethality or larval arrest in the offspring, while injection of vitamin B12 leads to viable offspring and repression of the *acdh-1* promoter. Scale bars, 500 μ m for bright-field microscopy and 100 μ m for fluorescence microscopy. See also Figure S3.

cofactor would be stuck in the intestine when *mrp-5* is perturbed, which would, therefore, not result in the rescue of embryonic development in the offspring.

One explanation for the rescue of embryonic development by feeding heme is that this increases vitamin B12 passage through intestinal membranes. Therefore, we next asked whether heme affects the vitamin-B12-responsive *Pacdh-1* reporter. We supplemented increasing concentrations of heme and found that the reporter was unaffected by heme in adult animals (Figure 3D). As noted earlier, embryos from *Pacdh-1::GFP* mothers treated with *mrp-5* RNAi without supplemented heme expressed high levels of GFP, suggesting that vitamin B12 cannot enter the embryos and repress the reporter. Importantly, however, GFP levels were greatly reduced in embryos from *Pacdh-1::GFP/mrp-5* RNAi animals supplemented with 500 μ M heme (Figure 3C). These findings suggest that supplying a high dose of heme facilitates the export of vitamin B12 from the intestine and, therefore, that the cause for embryonic lethality is lack of vitamin B12 and not heme. Indeed, we found that, while injection of vitamin B12 into the gonad rescued embryonic development in *Pacdh-1::GFP/mrp-5* RNAi animals, direct injection of 200 μ M heme did not rescue. Further, the arrested L1 larvae from *Pacdh-1::GFP/mrp-5* RNAi animals injected with 200 μ M heme

expressed high levels of GFP, indicating low vitamin B12 levels (Figure 3E).

Methionine and SAM Content Is Reduced in Offspring from *mrp-5* RNAi Mothers

Although vitamin B12 is a cofactor for two different metabolic enzymes (Figure 1A), our previous studies demonstrated that the developmental delay caused by vitamin B12 deficiency is mainly due to its function as a cofactor for the methionine synthase enzyme METR-1, which generates the methyl donor SAM from methionine (Watson et al., 2014). To determine whether the lack of vitamin B12 or the lack of heme in *mrp-5*-deficient animals results in defects in the SAM cycle, we measured the levels of methionine and SAM in embryos from mothers treated with *mrp-5* or vector control RNAi. We reasoned that differences in methionine and SAM content would be most clear in the presence of supplemented vitamin B12, because the *E. coli* diet is naturally low in this cofactor. Indeed, we detected a reduced methionine and SAM content in embryos from mothers treated with *mrp-5* RNAi relative to control animals in the presence of vitamin B12 (Figures 4A and 4B). Further, SAM levels could be restored to wild-type levels by feeding the animals with 500 μ M heme. These results suggest that a defect in the SAM

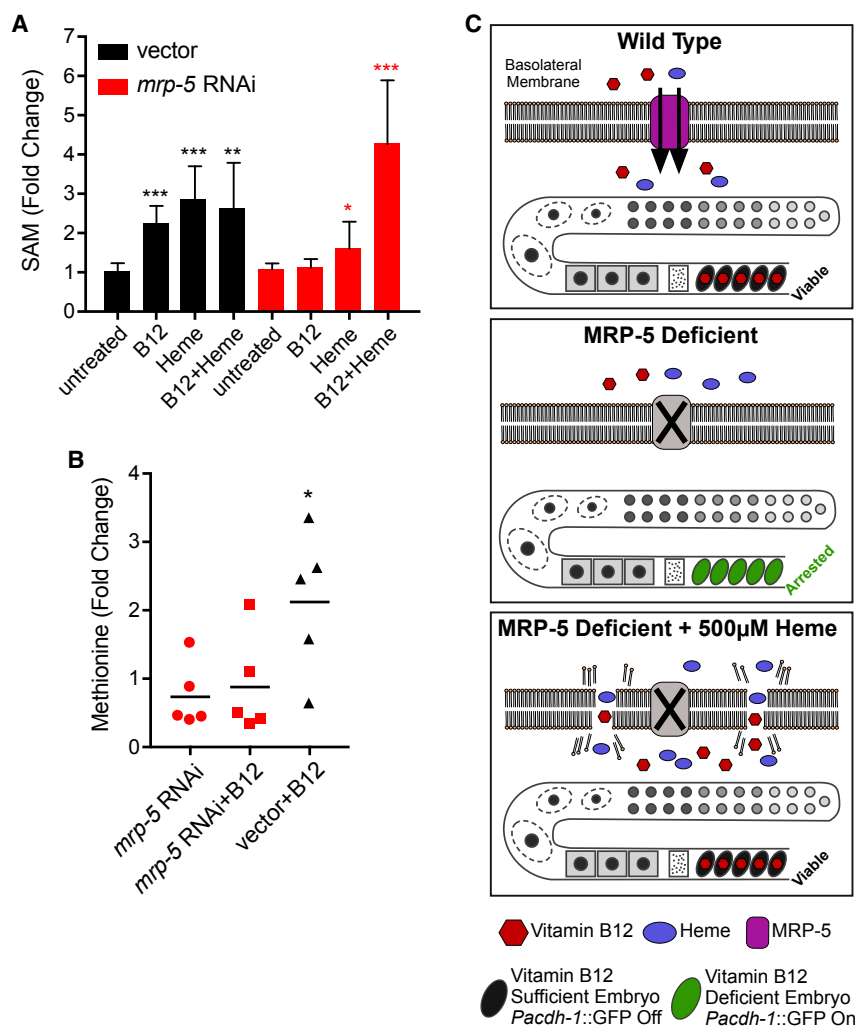


Figure 4. Methionine and S-Adenosylmethionine Content Is Reduced in Offspring from Mothers Treated with *mrp-5* RNAi

(A) SAM content was measured in the embryos of vector or *mrp-5* RNAi-treated mothers on *E. coli* HT115. SAM content was greatly reduced in *mrp-5*-deficient embryos compared to vector control. Error bars represent SD of the average of three independent experiments. The p value was determined by Student's t test relative to untreated embryos (*p < 0.05; **p < 0.001; ***p < 0.0001).

(B) Methionine content was measured by GC-MS of vector control or *mrp-5* RNAi-treated mothers fed *E. coli* HT115 with and without 64 nM vitamin B12 supplementation. Each dot represents a biological replicate, and values are relative to vector RNAi without vitamin B12 supplementation. The p value was determined by Student's t test (*p < 0.05).

(C) Model of MRP-5 function. In wild-type animals, both vitamin B12 and heme are exported from the intestine by MRP-5. Vitamin B12 can enter the embryo and repress the *acdH-1* reporter (top). In the absence of MRP-5, neither vitamin B12 nor heme can cross the intestinal membrane, the embryos arrest, and the *acdH-1* reporter remains active (middle). The presence of 500 µM heme causes membrane permeability in *mrp-5*-deficient mothers, leading to vitamin B12 crossing the intestine membrane, entering the embryo to rescue lethality, and repressing the *acdH-1* promoter (bottom).

(20 µM) (Rao et al., 2005), and high doses of heme are toxic to many systems (Chiabrando et al., 2014). Indeed, supplementing wild-type animals with a dose of 500 µM heme was toxic to *C. elegans* as well, whereas high doses of vitamin B12 were not (Figures 3B and S3). One mechanism of heme toxicity is by enhanced membrane

permeability (Chiabrando et al., 2014). Therefore, we propose that supplementing *mrp-5* RNAi animals with high concentrations of heme leads to increased intestinal membrane permeability, enabling both heme and vitamin B12 to pass through without the need for a dedicated transporter (Figure 4C).

DISCUSSION

We identified MRP-5 as a candidate transporter that exports vitamin B12 from the *C. elegans* intestine of the mother to support the development of her offspring. MRP-5 is a member of the *C. elegans* ABC transporter family, and these transporters are known to function as exporters of metabolites and drugs (Locher, 2016; Sheps et al., 2004; Slot et al., 2011). MRP-5 has previously been reported to export heme in *C. elegans* (Korolnek et al., 2014). *C. elegans* can synthesize neither vitamin B12 nor heme and depends on dietary supplementation of both cofactors. Loss or reduction of *mrp-5* by RNAi knockdown or by genetic mutation results in embryonic or larval lethality. As reported previously, we confirm that this lethality can be rescued by feeding the mother high concentrations of heme. However, it can also be rescued by injecting vitamin B12 directly into the gonad, bypassing the need for import from the intestine and, perhaps, other tissues. Wild-type *C. elegans* requires only low doses of heme

permeability (Chiabrando et al., 2014). Therefore, we propose that supplementing *mrp-5* RNAi animals with high concentrations of heme leads to increased intestinal membrane permeability, enabling both heme and vitamin B12 to pass through without the need for a dedicated transporter (Figure 4C).

More than 15 human genes involved in vitamin B12 processing have been identified (Nielsen et al., 2012), and four of these encode transporters. One of these is the ABC transporter MRP-1 (or ABCC1), which is localized to intestinal epithelial cells. ABC transporters in human and *C. elegans* are highly similar in sequence and structure. Therefore, we propose that *C. elegans* MRP-5 may be a functional ortholog of human MRP1 to export vitamin B12 from the intestine to surrounding tissues.

EXPERIMENTAL PROCEDURES

C. elegans Strains

All *C. elegans* strains were maintained at standard laboratory conditions as described previously (Brenner, 1974), and strain N2 was used as wild-type. Construction of VL749 (wwIs24 [*Pacdh-1::GFP* + *unc-119(+)*]) strain has been previously described (Arda et al., 2010). Strain VL1168 (*wwSi1/Pacdh-1::GFP::H2B unc-119(+)* II;*avr-14(ad1302)* I; *unc-119(ed3)* III; *avr-15(ad1051)*;

glc-1(pk54) *V* was generated by *mos1*-mediated single copy insertion (Frokjaer-Jensen et al., 2008). Strain *wwl47* [*mnp5p::MRP-5::GFP + rol-6(+)*] was generated by fusion PCR (<http://www.wormbook.org>). The *mnp-5* mutant strain VC1599 (+/szT1 [*lon-2(e678)*]; *mnp-5(ok2067)/szT1 X*) was obtained from the *C. elegans* Genetics Center (CGC). Bacterial strains *E. coli* OP50, HT115, and *Comamonas aquatica* DA1877 were obtained from CGC. The *E. coli* OP50 RNAi-compatible bacterial strain was generously provided by the Xu lab (Xiao et al., 2015).

RNAi Screen

The Kyoto Encyclopedia of Genes and Genomes (KEGG) database was used to predict *C. elegans* ABC transporters and solute carrier transporters (Kanehisa et al., 2016). The transporter RNAi mini-library was generated by selecting 215 clones from both the ORFeome and Ahringer RNAi libraries (Kamath et al., 2003; Rual et al., 2004). RNAi screening was carried out on 24-well nematode growth medium (NGM) agar plates containing 2 mM isopropyl β -D-1-thiogalactopyranoside (IPTG), 50 μ g/mL ampicillin with or without 64 nM Ado-Cbl (vitamin B12). *Pacdh-1::GFP* animals were grown on regular NGM plates with *E. coli* OP50 without Ado-Cbl supplementation for several generations before harvesting embryos. Embryos were incubated in M9 buffer overnight to obtain synchronized L1 animals. Approximately 15 L1 animals were added to each well containing individual RNAi clones. Animal phenotypes were observed after 80 hr of incubation, when the F1 generation was at the young adult stage.

Quantitation of Fluorescence

The animal body was outlined, and fluorescence intensity of the animal was determined using ImageJ (NIH). At least five animals per condition were analyzed, and the mean fluorescence per animals was determined.

C. elegans qRT-PCR Experiments

Animals were synchronized by L1 arrest and grown on ampicillin and IPTG plates seeded with either *E. coli* HT115 (vector RNAi and *mnp-5* RNAi) or *E. coli* OP50 (vector RNAi and *mnp-5* RNAi). Approximately 1,000 adult animals were harvested for each condition, in biological duplicate. Animals were washed in M9 buffer, and total RNA was isolated using TRIzol (Invitrogen) followed by DNase I treatment and cleanup using QIAGEN RNeasy columns. cDNA was generated from 1 μ g RNA using random primer and M-MuLV reverse transcriptase (New England Biolabs). qPCR was performed in technical duplicates per gene per condition using the Applied Biosystems StepOnePlus Real-Time PCR System and the Fast SYBR Green Master Mix (Thermo Fisher Scientific). Relative transcript abundance was determined using the $\Delta\Delta$ Ct method and normalized to averaged *ama-1* mRNA expression levels.

Vitamin B12 Injections

Animals were treated with *mnp-5* RNAi generated in an *E. coli* OP50 RNAi compatible strain. Vitamin B12 (3.2 mM) was injected into the gonad of L4 animals using a Narishige microinjection arm attached to the body of a Nikon Eclipse Ti inverted microscope. Sterile water was injected into the control group. After injection, animals were singled and treated with the same RNAi (*mnp-5* or vector). Phenotypes were scored after 2 days. *mnp-5(ok2067)* heterozygotes were singled at the L4 stage. Vitamin B12 was injected as described earlier. After injection, animals were singled, and the phenotype was scored after 5 days.

SAM Measurement

SAM was measured using the Bridge-It S-Adenosyl Methionine Fluorescence Assay Kit purchased from Mediomics following the manufacturer's recommendations.

Heme Solution Preparation

Hemin is the oxidized form of heme and must be dissolved in acid to make the active form of heme. 10 mM hemin solution was prepared as described previously (Hickman and Winston, 2007). 6.5 mg/mL hemin (Sigma-Aldrich) was dissolved in 0.1 M NaOH and incubated at 37°C for 1 hr. 1 M Tris (pH 7.5) was then added to a final concentration of 0.1 M. The pH was adjusted using

HCl, and the hemin solution was stored at 4°C, protected from light, and used within 2 days.

Relative Quantification of Methionine Using Gas Chromatography-Mass Spectrometry

Approximately 150,000 embryos were homogenized with 0.5 mL 200- to 300- μ m acid-washed glass beads (Sigma-Aldrich) in 1 mL 80% methanol using a FastPrep-24 bead beater (MP Biomedicals), with intermittent cooling in dry ice/ethanol bath. Samples were then extracted on dry ice for 15 min and centrifuged for 10 min at 20,000 \times g, and 250 μ L supernatant was dried under vacuum using a SpeedVac concentrator SPD111V (Thermo Fisher Scientific). Derivatization of dried samples was performed first with 20 μ L 20 mg/mL methoxyamine hydrochloride (Sigma-Aldrich) in pyridine at 37°C for 90 min, followed by the addition of 50 μ L N-methyl-N-(trimethylsilyl)trifluoroacetamide (Sigma-Aldrich) and incubation for 3 hr at 37°C. The derivatization reaction was completed by incubation for 5 hr at room temperature. Measurements were performed on an Agilent 7890B/5977B single quadrupole GC-MS (gas chromatography-mass spectrometry) equipped with an HP-5ms Ultra Inert capillary column (30 m \times 0.25 mm \times 0.25 μ m). The inlet temperature was set to 230°C, the transfer line was at 280°C, and the MS source and quadrupole were at 230°C and 150°C, respectively. The trimethylsilyl derivative of methionine was quantified as a 176-m/z ion with two qualifier ions of 128 and 293 m/z. Relative quantification of peak areas was done using samples within a linear response range, after mean normalization to total metabolites and blank subtraction.

SUPPLEMENTAL INFORMATION

Supplemental information includes Supplemental Experimental Procedures and three figures and can be found with this article online at <https://doi.org/10.1016/j.celrep.2018.02.100>.

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AUTHOR CONTRIBUTIONS

Conceptualization, H.N. and A.J.M.W.; Investigation, H.N. and O.P.; Methodology, H.N.; Resources, G.E.G.; Writing – Original Draft, H.N. and A.J.M.W.; Writing – Review & Editing, H.N. and A.J.M.W.; Visualization, H.N. and A.J.M.W.; Funding Acquisition, A.J.M.W.; Supervision, A.J.M.W.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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Supplemental Information

***C. elegans* MRP-5 Exports Vitamin B12
from Mother to Offspring
to Support Embryonic Development**

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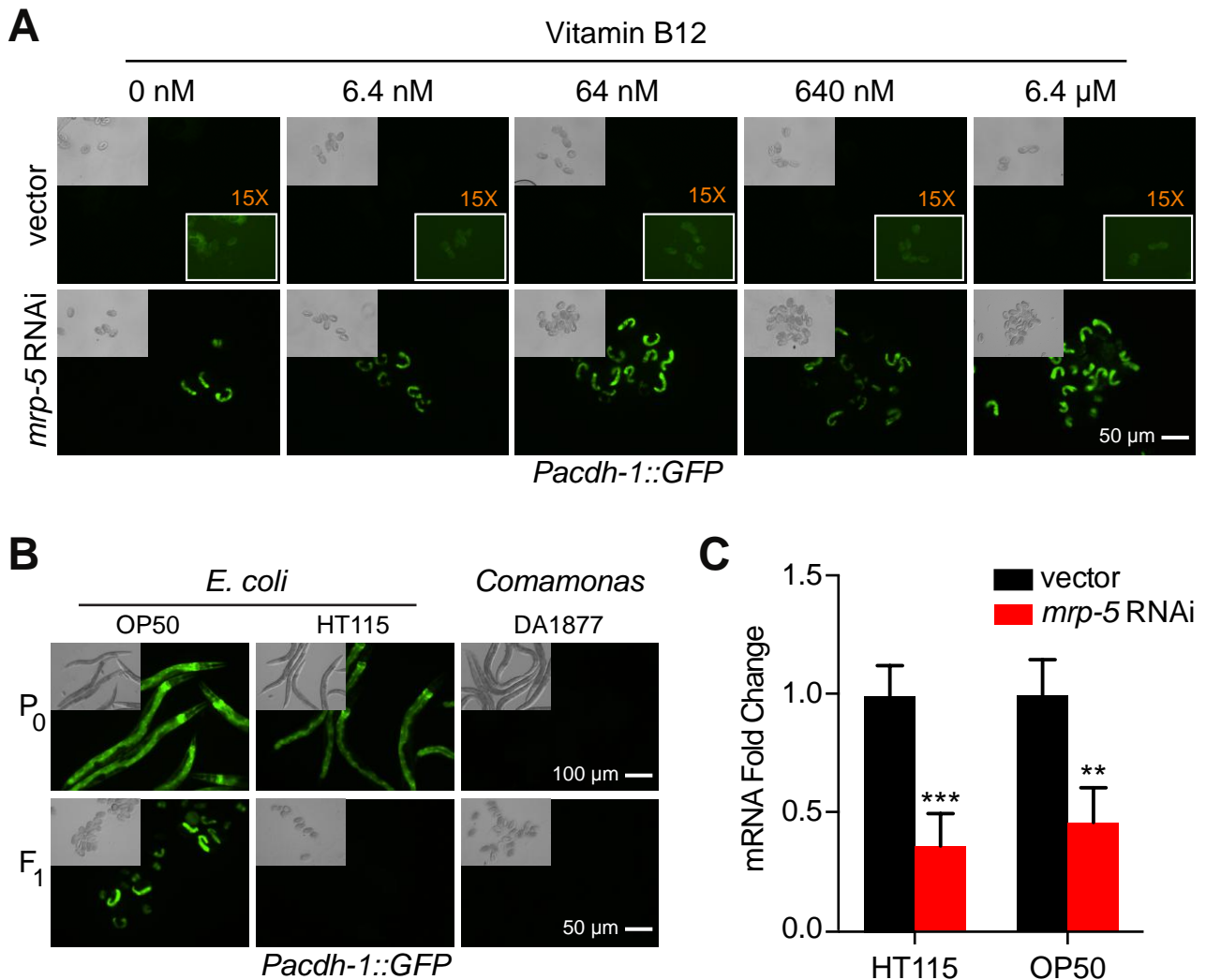


Figure S1. RNAi of *mrp-5* retains vitamin B12 in the intestine of P0 mothers, Related to Figure 1.

(A) Fluorescence and DIC microscopy images of embryos from *Pacdh-1::GFP* mothers subjected to vector control RNAi or *mrp-5* RNAi in *E. coli* HT115 and supplemented with increasing concentrations of vitamin B12. In the upper row, the inset box in the lower right indicates 15 times the exposure time. Scale bar, 50 μ m.

(B) Fluorescence and DIC microscopy images of *Pacdh-1::GFP* animals fed *E. coli* OP50, *E. coli* HT115 or *C. aquatica* DA1877 at P0 and F1 generations. Scale bars, 50 μ m for embryos and 100 μ m for adults.

(C) RT-qPCR of *mrp-5* showing similar RNAi efficiency in *E. coli* HT115 and *E. coli* OP50-fed animals relative to vector control. Error bars represent standard deviation. The p value was determined by unpaired student's t-test (***) p < 0.0001, ** p < 0.0005).

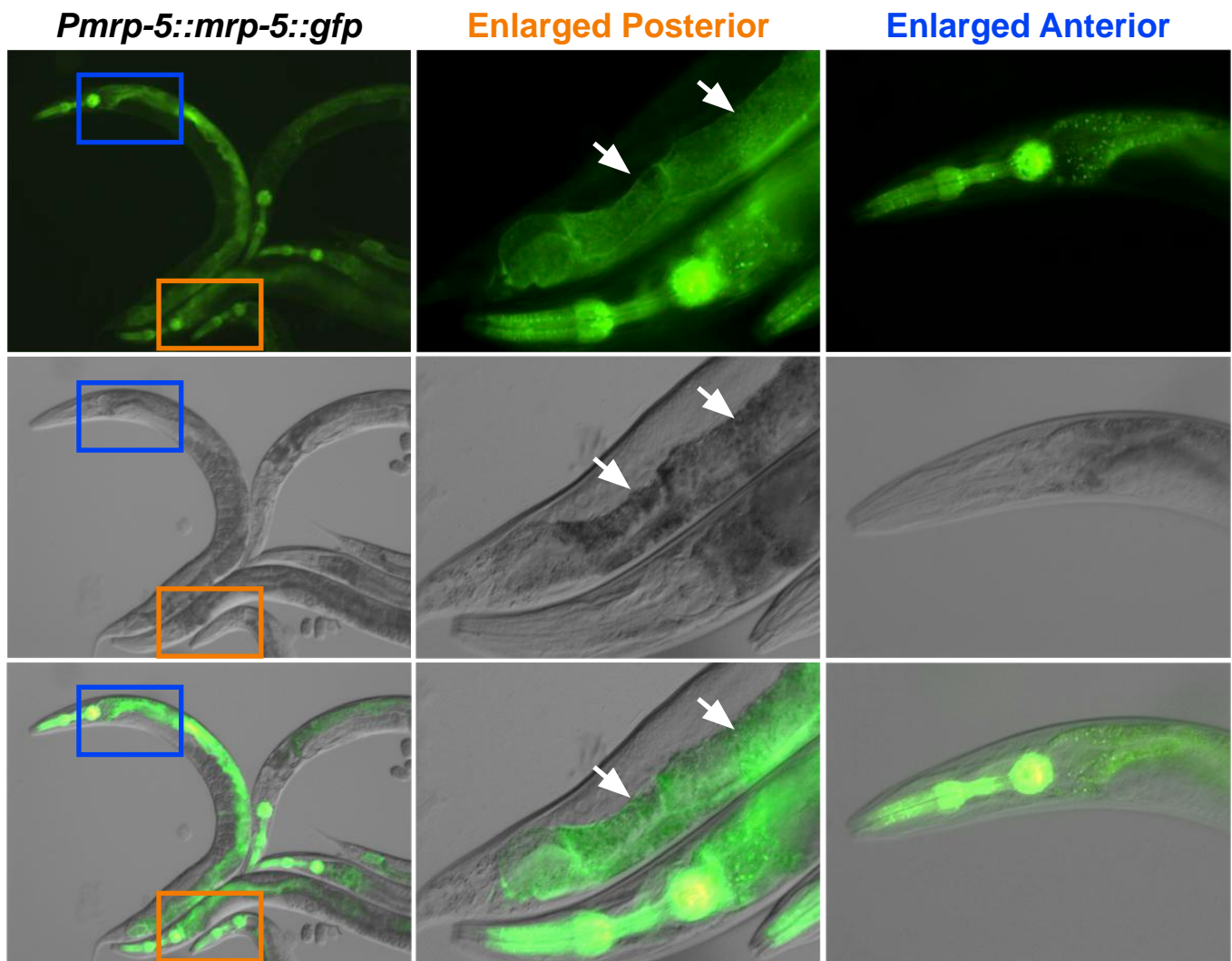


Figure S2. MRP-5 is expressed in the intestine, Related to Figure 1.

Fluorescence (top), DIC (middle), and overlay (bottom) microscopy images of GFP expression in *Pmrp-5::mrp-5::GFP* transgenic animals. The white arrowheads in the middle panels indicate intestinal cells.

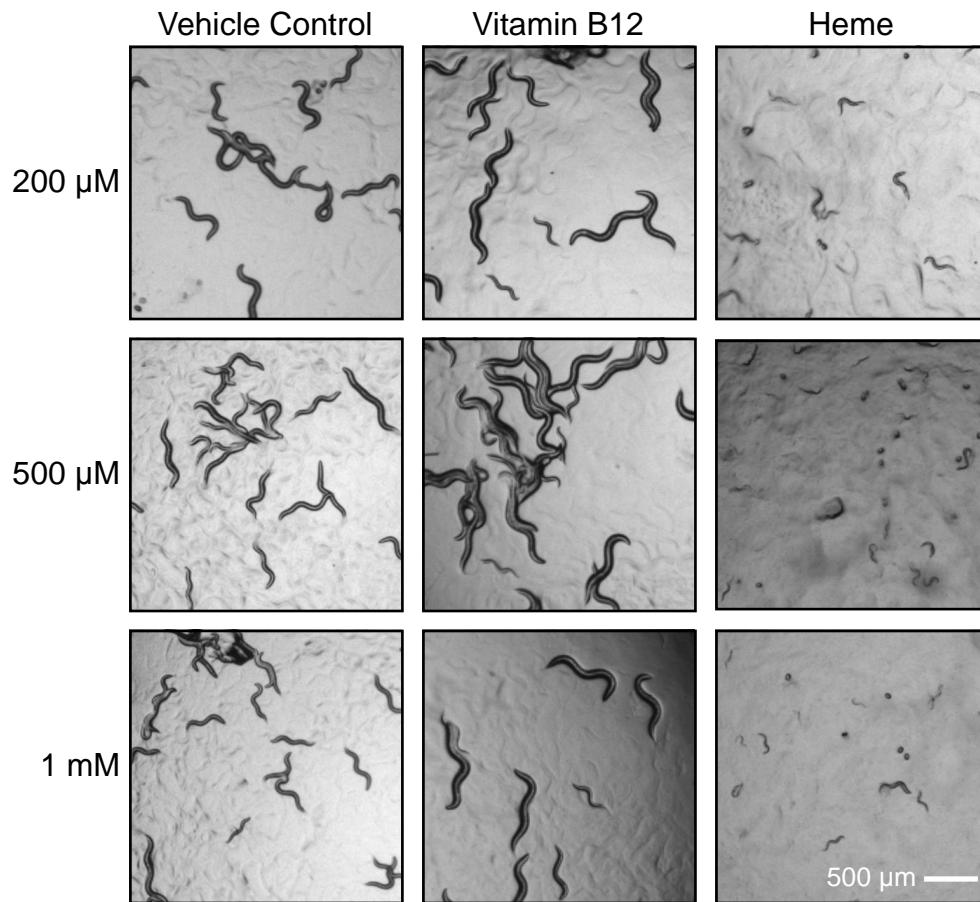


Figure S3. High doses of heme cause arrest at the L1-L2 stage, while high doses of vitamin B12 does not affect development, Related to Figure 3.

Bright field microscopy images of feeding various concentrations of heme, vitamin B12, or vehicle control (water) to wild type mothers on *E. coli* OP50 demonstrates viable offspring while developing on vitamin B12 or vehicle control, but causes larval arrest and toxicity in heme fed mothers. Scale bar, 500 μm.

RESOURCE TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Chemicals and Commercial Assays		
Adenosyl Cobalamin	Sigma Aldrich	Cat#: C0884
Hemin	Sigma Aldrich	Cat#: H9039
Glass beads	Sigma Aldrich	Cat#: G1277
Methoxyamine hydrochloride	Sigma Aldrich	Cat#: 226904
N-Methyl-N-(trimethylsilyl)trifluoroacetamide	Sigma Aldrich	Cat#: 69479
Isopropyl β -D-1 thiogalactopyranoside (IPTG)	US Biological	Cat#: I8500
TRIzol Reagent	Invitrogen	Cat#: 15596026
M-MuLV Reverse Transcriptase	NEB	Cat#: M0253S
Fast SYBR Green Master Mix	Thermo Fisher Scientific	Cat#: 4385612
RNeasy Mini Kit	Qiagen	Cat#: 74104
Bridge-It® S-Adenosyl Methionine (SAM) Fluorescence Assay Kit	Mediomics, LLC	Cat#: SKU: 1-1-1003
Experimental Models: Organisms/Strains		
<i>Caenorhabditis elegans</i> N2	Caenorhabditis Genetics Center (CGC); http://cgc.umn.edu/	N/A
<i>C. elegans</i> , wwIs24 [<i>acdh-1p::GFP + unc-119(+)</i>]	Arda <i>et al.</i> , 2010	Strain VL749
<i>C. elegans</i> , wwSi1 [<i>acdh-1p::GFP::H2B + unc-119(+)</i> II; <i>avr-14(ad1302)</i> I; <i>unc-119(ed3)</i> III; <i>avr-15(ad1051)</i> ; <i>glc-1(pk54)</i> V]	This study	Strain VL1168
<i>C. elegans</i> , +/szT1 [<i>lon-2(e678)</i> I; <i>mrp-5(ok2067)/szT1</i> X]	CGC	Strain VL1599
<i>C. elegans</i> , wwIs47 [<i>mrp5p::MRP-5::GFP + rol-6(+)</i>]	This study	
<i>Escherichia coli</i> OP50	CGC	N/A
<i>Escherichia coli</i> HT115	CGC	N/A
<i>Escherichia coli</i> OP50 RNAi compatible	Xiao <i>et al.</i> , 2015	N/A
<i>Comamonas aquatica</i> DA1877	CGC	N/A
<i>Escherichia coli</i> HT115 Ahringer RNAi Library	Kamath <i>et al.</i> , 2003	N/A
<i>Escherichia coli</i> HT115 ORFeome RNAi Library	Reboul <i>et al.</i> , 2001	N/A
Oligonucleotides		
<i>mrp-5</i> qPCR forward	This study	GACTGTCAGG GGGCTACCTA

<i>mrp-5</i> qPCR reverse	This study	AACAGCTCCAA TAACCGCGA
<i>ama-1</i> qPCR forward	This study	CGGATGGAGG AGCATCGCCG
<i>ama-1</i> qPCR reverse	This study	CAGCGGCTGG GGAAGTTGGC
Software, Algorithms and Instruments		
Eclipse Ti Inverted Microscope	Nikon	N/A
Eclipse 90i Inverted Microscope	Nikon	N/A
SpeedVac concentrator SPD111V	Thermo Fisher Scientific	N/A
FastPrep-24 bead beater	MP Biomedicals	Cat#: 116004500
ImageJ	NIH	N/A
StepOnePlus Real-Time PCR System	Applied Biosystems	Cat#: 4376600
DS Ri1 Camera	Nikon	N/A
EVOS FL Auto Imaging System	Invitrogen	N/A
FemtoJet [®] 4i electronic microinjector	Eppendorf	N/A
7890B/5977B quadrupole GC- MS	Agilent	N/A
Microinjection Arm	Narishige	N/A