Supporting Information for

Sterically shielded, stabilized nitrile imine for rapid bioorthogonal protein labeling in live cells

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Table S1. Product distribution after photoirradiating the substituted tetrazoles in the presence of equal molar ratio of acrylamide and glutathione in phosphate buffer/acetonitrile $(1:1)^{a}$



	Tetrazole structureProduct distribution (%) ^b		(%) ^b		
		а	b	с	d
	1: R = H	47	51	2	0
	2: $R = CH_3$	50	48	2	0
N.N.OMe	3: $R = CH_2OH$	0	0	0	100
	4: R = COOH	0	0	0	100
	5: $\mathbf{R} = \mathbf{CH}_2\mathbf{CN}$	38	42	20	0
R N=N	6: $\mathbf{R} = \mathbf{CH}_2\mathbf{OCH}_3$	55	45	0	0
1-10	7: $\mathbf{R} = \mathbf{CH}_2\mathbf{SCH}_3$	0	70	0	30
	$8: R = CH_2N(CH_3)_2$	0	0	0	100
	9: $R = CH_2CH_2OCH_3$	51	49	0	0
	10: $R = CH(CH_3)OCH_3$	38	42	20	0
R ³	11: $R^1 = CH_3$; $R^2 = H$; $R^3 = H$	0	0	100	0
R ² N N	12: $R^1 = CH_3$; $R^2 = H$; $R^3 = Cl$	0	0	100	0
	13: $R^1 = CH_3$; $R^2 = OCH_3$; $R^3 = H$	$R^2 = OCH_3; R^3 = H$ 0 0 100	0		
R ¹ \ 11-14	14: $R^1 = CH_3$; $R^2 = OCH_3$; $R^3 = OCH_3$	0	0	100	0
R^2 OMe	15: $R^1 = R^2 = CH_3$	10	10	80	0
N.N.OUMe	16: $R^1 = R^2 = OCH_3$	58	12	30	0
[∖] N=Ń R ¹ 15-17	17: $R^1 = R^2 = CH_2OCH_3$	32	46	22	0
	18: Ar = 2'-furan	37	58	5	0
	19: $Ar = 2$ '-thiophene	36	56	8	0
	20: Ar = 2'- <i>N</i> -Boc-pyrrole	58	39	3	0
Ar N=Ń	21: Ar = 2'-methoxybenzene	56	41	3	0
18-23	22: Ar = 2'-pyridine	1	1	0	98
	23: Ar = 2'-pyrrole	3	5	92	0
	24: $Ar^1 = Ar^2 = o-2$ '-furan	30	59	11	0
25	25: $Ar^1 = Ar^2 = o-2$ '-thiophene	27	63	10	0
Ar ² OMe	26: $Ar^1 = Ar^2 = o-2$ '- <i>N</i> -Boc-pyrrole	92 2 6 0			
	27: $Ar^1 = Ar^2 = o - 2$ '-pyrrole 1 1 9	98	0		
Ar ¹ 24-29	28: $Ar^1 = Ar_2 = o-2'-N-Moc-pyrrole$	78	20	2	0
	29: $Ar^1 = o-2$ '- <i>N</i> -Boc-pyrrole; $Ar^2 = p-2$ '- <i>N</i> -Boc-pyrrole	74	24	2	0

^{*a*} A solution of 25 μ M tetrazole and 500 μ M each of acrylamide and glutathione (reduced form; GSH) in 500 μ L phosphate buffer/acetonitrile (1:1), pH 7.4, was photoirradiated with a handheld 302-nm UV lamp at room temperature for 1 min. ^{*b*} The product distribution was analyzed by reverse-phase HPLC with absorbance set at 254 nm. The product peaks were assigned by comparing the trace of the product mixture to those of control reactions using either acrylamide or glutathione.



Figure S1. Fluorescence-based interference study of the cycloaddition reaction by glutathione in acetonitrile/phosphate buffer (1:1). (a) Reaction scheme. (b) Fluorescence spectra of the reaction involving tetrazole **26** and Sph in the presence of 0.01–10 mM glutathione; $\lambda_{ex} = 405$ nm. (c) Fluorescence spectra of the reaction involving tetrazole **1** and Sph in the presence of 0.01–10 mM glutathione; $\lambda_{ex} = 405$ nm. (d) Quantification of the inhibitory effect of glutathione on the formation of fluorescent pyrazoline adduct. The fluorescence measurements were repeated three times for each condition. Grey bar = **26**, orange bar = **1**.

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Figure S2. Fluorescence-based interference study of the cycloaddition reaction by glutamic acid in acetonitrile/phosphate buffer (1:1). (a) Reaction scheme. (b) Fluorescence spectra of the reaction involving tetrazole **26** and Sph in the presence of 0.01–10 mM glutamic acid; $\lambda_{ex} = 405$ nm. (c) Fluorescence spectra of the reaction involving tetrazole **1** and Sph in the presence of 0.01–10 mM glutamic acid; $\lambda_{ex} = 405$ nm. (d) Quantification of the inhibitory effect of glutamic acid on the formation of the fluorescent pyrazoline adduct. The fluorescence measurements were repeated three times for each condition. Grey bar = **26**, orange bar = **1**.



Figure S3. Fluorescence-based interference study of the cycloaddition reaction by a mixture of nucleophilic amino acids (methionine, cysteine, lysine, tryptophan, arginine, and tyrosine; 0.3 mM each) in acetonitrile/phosphate buffer (1:1). (a) Reaction scheme. (b) Fluorescence spectra of the reaction involving tetrazole 26 and Sph in the presence (red dashed line) and absence (black dashed line) of the amino acid mixture; $\lambda_{ex} = 405$ nm. (c) Fluorescence spectra of the reaction involving tetrazole 1 and Sph in the presence (red dashed line) of the amino acid mixture; $\lambda_{ex} = 405$ nm. (c) Fluorescence spectra of the reaction involving tetrazole 1 and Sph in the presence (red dashed line) of the amino acid mixture; $\lambda_{ex} = 405$ nm.

Table S2. Kinetic characterization of the cycloaddition reactions of tetrazole **1** or **26** with various dipolarophiles based on fluorescence of the pyrazoline adducts, and photophysical properties of the pyrazolines a^{a}



Tetrazole	Dipolarophile (µM)	Pyrazoline		$k_2 (M^{-1} s^{-1})$
(µM)		$\lambda_{abs}(nm)$	$\lambda_{\mathrm{em}} (\mathrm{nm})^b$	
1 (10)	acrylamide (100)	360	508	2000 ± 300
1 (10)	dimethyl fumarate (50)	366	516	4700 ± 500
1 (10)	styrene (100)	ND^{c}	ND^{c}	/
1 (10)	norbornene (100)	381	504	2200 ± 700
1 (10)	1,3-Cp (100)	ND^{c}	ND^{c}	/
1 (10)	3,3-Cp (100)	375	495	2800 ± 300
1 (10)	Sph (50)	376	494	3100 ± 600
1 (10)	TCO (50)	343	508	3800 ± 700
26 (10)	acrylamide (100)	340	503	860 ± 110
26 (10)	dimethyl fumarate (50)	351	502	2400 ± 300
26 (10)	styrene (100)	346	508	1000 ± 100
26 (10)	norbornene (100)	353	518	1000 ± 250
26 (10)	1,3-Cp (100)	360	488	520 ± 140
26 (10)	3,3-Cp (100)	360	496	1400 ± 130
26 (10)	Sph (50)	375	495	2800 ± 200
26 (10)	TCO (50)	323	490	2300 ± 300

^aPerformed in a quartz cuvette for direct fluorescence measurement.

^bEmission maximum was recorded in phosphate buffer/acetonitrile (1:1). ^cpyrazoline was not detected.

synazonne was not detected.



Figure S4. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and acrylamide (100 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and acrylamide monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **26** and acrylamide fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.

a)



a)



Figure S5. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and dimethyl fumarate (50 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and dimethyl fumarate monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **1** and dimethyl fumarate monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **1** and dimethyl fumarate monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**d**, **e**) Plots of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.

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Figure S6. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and styrene (100 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and styrene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Plot of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constant, k_2 , was calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation. The fluorescence was not detected for tetrazole **1** under the same reaction condition.



Figure S7. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and norbornylene (100 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and norbornene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **26** and norbornene tetrazole **1** and norbornene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **1** and norbornene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**d**, **e**) Plots of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.



Figure S8. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and 1,3-cyclopropene (100 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and 1,3-cyclopropene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Plot of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $\mathbf{y} = (\mathbf{y}_0-\mathbf{a}) e^{\mathbf{kt}} + \mathbf{a}$, to give k. The second-order rate constant, k_2 , was calculated using the following equation: $k_2 = \mathbf{k}/[\text{dipolarophile}]$. Measurements were repeated three times at each time point to derive the mean and standard deviation. The fluorescence was not detected for tetrazole **1** under the same reaction condition.



Figure S9. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and 3,3-cyclopropene (100 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (**a**) Reaction scheme. (**b**) Time course of the cycloaddition reaction between tetrazole **26** and 3,3-cyclopropene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **1** and 3,3-cyclopropene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**c**) Time course of the cycloaddition reaction between tetrazole **1** and 3,3-cyclopropene monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (**d**, **e**) Plots of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.

a)





Figure S10. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole 26 (10 μ M) or 1 (10 µM) and spirohexene (Sph, 50 µM) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (a) Reaction scheme. (b) Time course of the cycloaddition reaction between tetrazole 26 and Sph monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (c) Time course of the cycloaddition reaction between tetrazole 1 and Sph monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (d, e) Plots of relative fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.

0

5

10

tim e/s

15

20

25

a)

0

10

20

tim e/s

30



Figure S11. Fluorescence-based kinetics measurement of the cycloaddition between tetrazole **26** (10 μ M) or **1** (10 μ M) and TCO-amine (50 μ M) in phosphate buffer/acetonitrile (1:1) under 302 nm photoirradiation. (a) Reaction scheme. (b) Time course of the cycloaddition reaction between tetrazole **26** and TCO-amine monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (c) Time course of the cycloaddition reaction between tetrazole tetrazole **1** and TCO-amine monitored by a spectrofluorometer ($\lambda_{ex} = 405$ nm). (c) Time course of the cycloaddition reaction between tetrazole fluorescence intensity vs. reaction time. The amounts of pyrazoline adduct were fitted to an exponential rise to maximum equation, $y = (y_0-a) e^{kt} + a$, to give k. The second-order rate constants, k_2 , were calculated

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using the following equation: $k_2 = k/[dipolarophile]$. Measurements were repeated three times at each time point to derive the mean and standard deviation.



Figure S12. Transition state isomers and activation energies for the methyl thiolate addition reaction to nitrile imine-**1**. The distances shown are in Å, and dihedral angles are in degree. Two possible TS for the addition of methyl thiolate to nitrile imine-**1** are presented. The *anti*- and *syn*- transition states of nucleophilic addition of thiols are defined by the dihedral angle around the forming C—S bond (\angle (C-S-C-N) = -137.8° and 46.1° for **TS3** and **TS3'**, respectively). *syn*-**TS** (**TS3'**) has a higher $\triangle G^{\ddagger}$ compared to the *anti*-**TS** (**TS3**) ($\triangle \Delta G^{\ddagger}_{\omega-B97X-D} = 1.2$ kcal mol⁻¹).

	Single crystal of 26
Empirical formula	C ₃₂ H ₃₄ N ₆ O ₅
Formula weight	582.65
Temperature/K	90
Crystal system	monoclinic
Space group	$P2_1/n$
a/Å	11.8321(5)
b/Å	13.4953(6)
c/Å	19.2839(9)
α/°	90
β/°	105.5800(10)
γ/°	90
Volume/Å ³	2966.1(2)
Z	4
$\rho_{calc}g/cm^3$	1.305
µ/mm⁻¹	0.090
F(000)	1232.0
Crystal size/mm ³	0.1 imes 0.08 imes 0.07
Radiation	MoKa ($\lambda = 0.71073$)
20 range for data collection/°	3.656 to 56.654
Index ranges	$-15 \le h \le 15, -18 \le k \le 17, -25 \le l \le 25$
Reflections collected	59012
Independent reflections	7373 [$R_{int} = 0.0256$, $R_{sigma} = 0.0132$]
Data/restraints/parameters	7373/0/395
Goodness-of-fit on F ²	1.025
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0374, wR_2 = 0.0963$
Final R indexes [all data]	$R_1 = 0.0412, wR_2 = 0.0997$
Largest diff. peak/hole / e Å ⁻³	0.39/-0.24

 Table S3. Crystal data and structure refinement for 26.

	Single crystal of 27
Empirical formula	$C_{22}H_{18}N_6O$
Formula weight	382.42
Temperature/K	90
Crystal system	monoclinic
Space group	$P2_1/n$
a/Å	18.7948(8)
b/Å	5.1334(2)
c/Å	21.0251(9)
$\alpha/^{\circ}$	90
β/°	116.3170(10)
γ/°	90
Volume/Å ³	1818.28(13)
Z	4
$\rho_{calc}g/cm^3$	1.397
µ/mm ⁻¹	0.091
F(000)	800.0
Crystal size/mm ³	$0.1\times0.08\times0.01$
Radiation	MoKa ($\lambda = 0.71073$)
2Θ range for data collection/°	3.892 to 56.708
Index ranges	$-25 \le h \le 25, \ -6 \le k \le 6, \ -28 \le l \le 28$
Reflections collected	36043
Independent reflections	4528 [$R_{int} = 0.0219, R_{sigma} = 0.0126$]
Data/restraints/parameters	4528/0/263
Goodness-of-fit on F ²	1.045
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0376, wR_2 = 0.0976$
Final R indexes [all data]	$R_1 = 0.0432$, $wR_2 = 0.1023$
Largest diff. peak/hole / e Å ⁻³	0.35/-0.23

 Table S4. Crystal data and structure refinement for 27.



c)



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f1 (ppm)



Figure S13. 2D NMR spectra of tetrazole 26. (a) Structure of tetrazole 26 with number assigned for all protons. (b) 2D COSY spectrum (500 MHz, 10 mM, CDCl₃) of tetrazole 26. (c) Partial zoomed-in 2D COSY spectrum of tetrazole 26. (d) 2D ROESY spectrum (500 MHz, 10 mM, CDCl₃, 25 °C) of tetrazole 26. (e) Partial zoomed-in 2D ROESY spectrum of tetrazole 26.











e)



g)



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Figure S14. Fluorescence-based determination of the half-lives of nitrile imine-1, -26, and -28 in phosphate buffer/acetonitrile (1:1). (a) Scheme showing measurement procedure: 10 µM tetrazole 1, 26 or 28 was photoirradiated with a handheld 302 nm UV lamp for 15 sec. The resulting solution was left unperturbed for the in situ generated nitrile imine to decay for a period of time before dimethyl fumarate was added with a final concentration of 500 µM to capture the residual nitrile imine to produce the fluorescent pyrazoline product. (b) Fluorescence spectra of the pyrazoline products derived from the reaction of dimethyl fumarate with the residual nitrile imine-1 after left unperturbed for various times; $\lambda_{ex} = 405$ nm. The zero-point sample was obtained by incubating dimethyl fumarate with tetrazole 1 prior to photoirradiation. (c) Plot of fluorescence intensity vs. delay time for nitrile imine-1. No fluorescence was detected after adding dimethyl fumarate, indicating that the half-life of nitrile imine-1 must be shorter than 7.5 sec. (d) Fluorescence spectra of the pyrazoline products derived from the reaction of dimethyl fumarate with the residual nitrile imine-26 after left unperturbed for various times; $\lambda_{ex} = 405$ nm. (e) Plot of fluorescent intensity vs. delay time for nitrile imine-26. Based on the curve-fitting, the half-life of nitrile imine-26 was determined to be 102 sec. (f) Fluorescence spectra of the pyrazoline products derived from the reaction of dimethyl fumarate with the residual nitrile imine-28 after left unperturbed for various times; $\lambda_{ex} = 405$ nm. (g) Plot of the fluorescence intensity vs. the delay time for nitrile imine-28. Based on the curve-fitting, the half-life of nitrile imine-28 was determined to be 21 sec. Each fluorescence measurement was repeated three times to obtain the mean values and standard deviations.

Ö HO HO 302 nm Ö ö OMe OMe 0 o N Sph 30 pyrazoline-30 b) c) ACN (%) 2.8x10⁶ PB -5 2.6x10⁶ MeOH 10 2.4x10⁶ DCM 4x10⁵ 20 ACN 2.2x10⁶ 35 0.25ACN 2.0x10⁶ 50 -EA 1.8x10⁶ 3x10⁴ Intensity (CPS) Intensity (CPS) 1.6x10⁶ 1.4x10⁶ 1.2x10⁶ 2x10⁶ 1.0x10⁶ 8.0x10⁵ 1x10⁶ 6.0x10⁵ 4.0x10⁵ 2.0x10^t 0.0 0 450 500 550 600 Wavelength (nm) 650 700 750 450 500 550 600 650 700 Wavelength (nm) d) 4x10⁵ -10 mM glu -5 mM glu 1 mM glu 0.1 mM glu 3x10⁵ 0 glu Intensity (CPS) 2x10⁶ 1x10⁵ 0 672 728 504 560 448 616

a)

Figure S15. Fluorescent properties of pyrazoline-30 under different conditions. (a) Scheme for pyrazoline-30 synthesis. (b) Fluorescence spectra of pyrazoline-30 (5 μ M) in different solvents: PB = phosphate buffer, DCM = dichloromethane, 0.25ACN = 25% acetonitrile in PB, EA = ethyl acetate. (c) Fluorescence spectra of pyrazoline-30 in PB containing varying amounts of acetonitrile (v/v). (d) Response of pyrazoline-30 fluorescence after adding varying amounts of glutamine. Pyrazoline-30 was dissolved in 20% ACN in phosphate buffer.

Wavelength (nm)



Figure S16. *In situ* synthesis of a pyrazoline-based QBP fluorescence sensor. (**a**) Scheme for reaction of a QBP mutant encoding SphK at position-160 (purple color) with a water-soluble tetrazole **30**. (**b**) ESI-MS analyses of QBP mutants. For QBP-N160SphK, calcd mass 26006 Da, found 26007 Da; for QBP-N160BocK, calcd mass 25970 Da, found 25971 Da. (**c**) SDS-PAGE analyses of the reaction products showing alkene-dependent fluorescent labeling of QBP protein. Top panel, Coomassie staining; bottom panel, in-gel fluorescence analysis.



sfGFP-Q204SphK

sfGFP-Q204Cy5

b)





a)



Figure S17. SDS-PAGE analysis of fluorescent labeling the SphK-encoded sfGFP in *E. coli* cell lysates. (a) Reaction scheme. (b) De-convoluted masses of sfGFP-Q204SphK and sfGFP-Q204BocK. The sfGFP construct we used here contains the T65G and Y66G mutations, which effectively knocks out the intrinsic GFP fluorescence. (c) SDS-PAGE analysis of the lysates of *E. coli* cells expressing either sfGFP-Q204SphK (lane 1) or sfGFP-Q204BocK (lane 2) after treating the lysates with the Cy5-conjugated tetrazole **31**: left panel, Coomassie blue staining; right panel, in-gel fluorescence imaging. Asterisks indicate weak background labeling, likely due to non-specific association of tetrazole **31** with these proteins in the cell lysates.



Figure S18. Bioorthogonal labeling of GCGR in live HEK 293T cells via tetrazine ligation. (**a**) Scheme for bioorthogonal labeling of a GCGR-GFP mutant encoding SphK at position-372 with a Cy5-functionalized tetrazine. (**b**) Confocal micrographs of HEK 293T cells expressing GCGR-GFP-H372SphK after treated with 1 μ M tetrazine-Cy5^[S1] for 5, 10, or 30 min in DMEM medium. Scale bar = 20 μ m.



Figure S19. Flow cytometry analysis of the GCGR-expressing HEK 293T cells after treated with the Cy5conjugated tetrazole **31**. (**a**) GCGR-H372SphK-expressing HEK 293T cells. (**b**) GCGR-GFP-H372SphKexpressing HEK 293T cells prior to treatment. (**c**) GCGR-GFP-H372SphK-expressing HEK 293T cells after treated with 500 nM tetrazole-Cy5 for 1 min. The cell number and mean Cy5 fluorescence (Y mean) in each quadrant are shown in the tables. The cell labeling efficiency is calculated as follows: % labeling = #cells in right top / (# cells in right bottom + # cells in right top). For GCGR-GFP-H372SphK, % labeling = 3037 / (3037 + 378) = 3037/3415= 89%.

Supplemental Methods

Expression and purification of unnatural amino acid-containing sfGFP mutants. BL21(DE3) cells were transformed with the pEVOL-PyIT-mmPyIKRS and pET-sfGFP-GGG-Q204TAG plasmids. The cells were recovered in 1 mL SOC medium following transformation and incubated at 37 °C for 1 hour before plating on LB agar plate containing chloramphenicol (Cam) (34 µg/mL) and ampicillin (Amp) (100 µg/mL). A 5-mL overnight culture from a single colony was used to inoculate 25 mL LB medium supplemented with Cam and Amp. Cells were grown at 37 °C in a shaker-incubator (250 rpm), and the protein expression was induced by adding 1 mM IPTG, 0.2% arabinose, and 1 mM SphK for sfGFP-Q204SphK or 1 mM BocK for sfGFP-Q204BocK when OD₆₀₀ reached 0.6. After 6-hour induction, cells were harvested, resuspended in a lysis buffer (50 mM NaH₂PO₄, 300 mM NaCl, 10 mM imidazole, pH 8.0), and sonicated in an ice/water bath ten times (10 sec each with 20 sec interval). The lysate was centrifuged (30 min, 4 °C). The supernatant was incubated with 0.1 mL Ni-NTA resin (Thermo HisPurTM) (2 hours, 25 °C). The slurry was then loaded to a column and the protein-bound resin was washed twice with 3 mL washing buffer (50 mM Na₂HPO₄, 300 mM NaCl, 50 mM imidazole, pH 8.0). The protein was eluted off the resin using the elution buffer (50 mM Na₂HPO₄, 300 mM NaCl, 250 mM imidazole, pH 8.0). The eluted fractions were collected, concentrated, and subjected to buffer-exchange to DPBS containing 100 µM TCEP using 10 kD MWCO spin column (Thermo Scientific). The protein identity was verified by LC/ESI-MS. For SDS-PAGE analysis, protein in cell lysates was added 25 µM tetrazole-Cy5 and the mixture was irradiated with a handheld 302-nm UV lamp for 1 min. The sample was then added with the loading buffer and heated to 100 °C for 10 min before loaded to the gel. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis of proteins was performed on an XCell SureLock Mini-Cell apparatus using the precast NuPAGE 4-12% Bis-Tris gels (Invitrogen). BenchMark Prestained Protein Ladder was applied to one lane of each gel for estimation of apparent molecular weight.

Cloning and mutagenesis. The cloning of GCGR-GFP and site-specific amber mutagenesis of glucagon receptor (GCGR) were carried out as reported previously.^[S1] The bacterial expression construct QBP-N160TAG was derived from pETtrio-1-AcKRS-pylT(TAA)-QBP141TAA in which position-141 was mutated back to Asn using the following primer pair: 5'–CTGCGTCAGTTCCCGAACATCGATAATGC CTATATGG–3' and 5'–GTTGGTGCCCAGTTCCATATAGGCATTATCGATGTTC–3'. Afterwards, Asn160 was mutated to TAG using the following primer pair: 5'–CGATACGCCATAGATTCTGTACTT CATCAAAACCG–3' and 5'–TGCAGAACGGCGTCTGCG–3'.

HEK 293T cell culture and transfection. HEK 293T cells were maintained in a growth medium containing Dulbecco's modified eagle medium (DMEM, Life Technologies) supplemented with 10% (v/v) fetal bovine serum (FBS, Life Technologies) and 10μ g/mL gentamicin (Life Technologies). Transfection was performed at 70-80% confluency using 3:1 reagent/DNA ratio of Lipofectamine 2000 (Life Technologies) with 2.5 μ g total DNA or polyethylenimine (PEI, Polysciences Inc.) with 3 μ g total DNA per 35-mm dish. For imaging experiments, cells were maintained in 35-mm glass bottom culture dishes using FluoroBrite DMEM medium (Life Technologies) supplemented with 10% FBS, 4 mM L-glutamine, and 25 mM HEPES.

General Information

Solvents and chemicals were purchased from commercial sources and used directly without further purification. N^e-Boc-lysine (BocK) was purchased from Chem-Impex (Cat. No. 00363). TCO-amine was purchased from Click Chemistry Tools. Sph and 3,3-cyclopropene were synthesized by following our previous report^[S2]. 1,3-Cylopropene was synthesized by following the reported procedure^[S3]. Flash chromatography was performed either manually with SiliCycle P60 silica gel (40-63 µm, 60 Å) or an automatic Yamazen AKROS flash system equipped with SiliaSep HP pre-packed columns. ¹H NMR spectra were recorded with Inova-300, -400 or -500 MHz spectrometers. Chemical shifts were reported in ppm using TMS or deuterated solvents as internal standards (TMS, 0.00; CDCl₃, 7.26; CD₃OD, 3.31; DMSO- d_6 , 2.50; acetone- d_6 , 2.05). Multiplicity was reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, brs = broad. ¹³C NMR spectra were recorded at 75.4 MHz, and chemical shifts were reported in ppm using deuterated solvents as internal standards (CDCl₃, 77.0; DMSO-d₆, 39.5; CD₃OD, 49.05; acetone-d₆, 30.0). UV-Vis absorption spectra were recorded using 1-cm quartz cuvette on a HP-8452 Diode Array Spectrophotometer. Fluorescence spectra were recorded using 1-cm quartz cuvette on a Horiba FluoroMax-4 spectrofluorometer at 25 °C. High resolution mass spectrometry was performed on a Thermo LTO-Orbitrap XL mass spectrometer equipped with collision cells for collision induced dissociation. Low resolution mass spectrometry was performed on a Thermo LCQ mass spectrometer.

Scheme S1



2-(4-Methoxyphenyl)-5-(o-tolyl)-2H-tetrazole (2): To a solution of benzenesulfonohydrazide (1.43 g, 8.3 mmol) in ethanol was added 2-methylbenzaldehyde (962 μ L, 8.3 mmol), and the mixture was stirred at room temperature for 3 hours. The solvent was removed and small amount of water was added to precipitate the solid, which was then collected by filtration and dried to give the sulfohydrazone intermediate as a white solid (2.1 g, 45%). <u>Preparation of diazonium salt</u>: to a mixed solution of 4-methoxyaniline (1.1 g, 9.0 mmol) in ethanol/water (2:1;10 mL,) cooled with an ice-water bath was added sequentially 2.0 mL concentrated HCl and 1.0 mL sodium nitrite (0.69 g, 10.0 mmol) solution in water, and the mixture was stirred for about 10 min before the color turned purple. To a solution of sulfohydrazone (1.0 g, 3.7 mmol) in pyridine (50 mL) cooled with an ice-salt bath was added the freshly prepared diazonium salt, and the mixture was stirred while temperature was allowed to gradually increase to room temperature. After about 6 hours, the solvent

was removed and DCM (50 mL) was added. The mixture was washed with 1 N HCl before the solvent was evaporated under reduced pressure. The residue was purified by silica gel flash chromatography using ethyl acetate/hexanes as eluent to give the title product as a colorless solid (435 mg, 45%): ¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, *J* = 9.1 Hz, 3H), 7.49–7.30 (m, 3H), 7.07 (d, *J* = 9.1 Hz, 2H), 3.90 (s, 3H), 2.71 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.28, 160.34, 137.48, 131.34, 129.90, 129.44, 126.28, 125.96, 124.24, 121.21, 114.57, 114.06, 55.55, 21.73; HRMS calcd for C₁₅H₁₄N₄O 267.1240 [M+H⁺], found 267.1235.

Methyl 2-formylbenzoate (S1): Compound S1 was synthesized as a colorless solid with 90% yield: ¹H NMR (400 MHz, CDCl₃) δ 10.61 (s, 1H), 7.88 (m, 2H), 7.62 (m, 2H), 3.99 (s, 3H).

Methyl 2-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)benzoate (S2): Compound S2 was synthesized using the same procedure as compound 2 with 45% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 9.2 Hz, 2H), 7.97–7.89 (m, 1H), 7.83–7.76 (m, 1H), 7.58 (m, 2H), 7.03 (d, *J* = 9.2 Hz, 2H), 3.86 (s, 3H), 3.80 (s, 3H).

(2-(2-(4-Methoxyphenyl)-2*H*-tetrazol-5-yl)phenyl)methanol (3): To a solution of tetrazole S2 (158 mg, 0.51 mmol) in 10 mL anhydrous THF at 0 °C was add LiAlH₄ (58 mg, 1.53 mmol). After TLC showed complete disappearance of the starting materials, the reaction was quenched by adding 1 mL methanol. The solvent was evaporated in vacuum, and the residue was purified by flash chromatography using ethyl acetate/hexanes as eluent to give the title product as a white solid (115 mg, 80%): ¹H NMR (400 MHz, CDCl₃) δ 8.17 (dd, *J* = 7.3, 1.6 Hz, 1H), 8.04 (d, *J* = 9.1 Hz, 2H), 7.55–7.47 (m, 1H), 7.47–7.39 (m, 2H), 7.01 (d, *J* = 9.1 Hz, 2H), 4.81 (d, *J* = 7.5 Hz, 2H), 4.60 (t, *J* = 7.5 Hz, 1H), 3.84 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.43, 160.62, 139.87, 130.63, 130.43, 129.90, 129.43, 128.12, 125.50, 121.26, 114.63, 64.25, 55.53; HRMS calcd for C₁₅H₁₄N₄O₂ 283.1190 [M+H⁺], found 283.1188.

5-(2-(Methoxymethyl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (6): To a solution of alcohol **3** (53 mg, 0.19 mmol) in anhydrous THF at 0 °C was added NaH (60%, 23 mg, 0.56 mmol), and the mixture was stirred at room temperature for 10 min. Then tetrabutylammonium iodide (14 mg, 0.038 mmol) and methyl iodide (35 μ L, 0.56 mmol) were added. The mixture was stirred at room temperature until the starting material was completely consumed. Afterwards, several drops of saturated NH₄Cl solution were added followed by addition of water. The solution was extracted with ethyl acetate, and the organic layer was collected and evaporated. The residue was purified by flash chromatography using ethyl acetate/hexanes as eluent to give the title product as a white solid (50 mg, 90%): ¹H NMR (500 MHz, CDCl₃) δ 8.18 (d, *J* = 7.7 Hz, 1H), 8.11 (d, *J* = 9.0 Hz, 2H), 7.70 (d, *J* = 7.7 Hz, 1H), 7.52 (t, *J* = 7.5 Hz, 1H), 7.44 (t, *J* = 7.5 Hz, 1H), 7.07 (d, *J* = 9.0 Hz, 2H), 4.97 (s, 2H), 3.90 (s, 3H), 3.48 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 164.56, 160.43, 137.67, 130.17, 129.39, 128.08, 127.46, 124.91, 121.24, 114.62, 72.59, 58.54, 55.57; HRMS calcd for C₁₆H₁₆N₄O₂ 297.1346 [M+H⁺], found 297.1344.

5-(2-(Bromomethyl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (S3): To a solution of alcohol **3** (55 mg, 0.20 mmol) in DCM cooled to 0 °C was added PPh₃ (102 mg, 0.39 mmol) and NBS (69 mg, 0.39 mmol), and the reaction mixture was stirred overnight. The mixture was filtered through a thin pad of neutral alumina, and the filtrate was concentrated to give a yellow solid. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (57 mg, 85%): ¹H NMR (400 MHz, CDCl₃) δ 8.23 (dd, J = 5.5, 3.6 Hz, 1H), 8.10 (d, J = 9.1 Hz, 2H), 7.54 (dd, J = 5.4, 3.6 Hz, 1H), 7.44 (dd, J = 5.8, 3.4 Hz, 2H), 7.04 (d, J = 9.1 Hz, 2H), 5.14 (s, 2H), 3.85 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.84, 160.43, 136.53, 131.60, 130.43, 130.16, 129.88, 128.83, 125.81, 121.21, 114.59, 55.54, 32.48; HRMS calcd for C₁₅H₁₃BrN₄O 345.0346 [M+H⁺], found 345.0344.

2-(4-Methoxyphenyl)-5-(2-((methylthio)methyl)phenyl)-2H-tetrazole (7): To a solution of Me₂S₂ (4 g, 43.6 mmol) in 50 mL anhydrous THF was added sodium (0.5 g, 21.7 mmol), and the mixture was stirred

at room temperature under argon overnight. The precipitated white solid was filtered, washed with diethyl ether, and dried to give sodium thiomethylate. To a solution of tetrazole **S3** (20 mg, 0.058 mmol) in ethanol (5 mL) was added sodium thiomethylate (8.3 mg, 0.116 mmol) solution in 2 mL ethanol. The suspension was stirred while refluxing for 1 hour. The reaction mixture was cooled to room temperature, and the mixture was diluted with water and extracted with diethyl ether. The organic layer was collected, concentrated, and purified by silica gel flash chromatography to give the desired product as a white solid (15 mg, 81%): ¹H NMR (400 MHz, CDCl₃) δ 8.20 (d, *J* = 6.6 Hz, 1H), 8.11 (d, *J* = 9.1 Hz, 2H), 7.47–7.34 (m, 3H), 7.06 (d, *J* = 9.1 Hz, 2H), 4.26 (s, 2H), 3.89 (s, 3H), 2.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.66, 160.48, 137.69, 130.91, 130.41, 130.21, 129.92, 127.38, 126.02, 121.33, 114.68, 55.64, 36.73, 15.10; HRMS calcd for C₁₆H₁₆N₄OS 313.1118 [M+H⁺], found 313.1112.

1-(2-(2-(4-Methoxyphenyl)-2*H*-tetrazol-5-yl)phenyl)-*N*,*N*-dimethylmethanamine (8): To a solution of tetrazole S3 (24 mg, 0.07 mmol) in acetonitrile was added dimethyl amine (40%; 88 µL, 0.7 mmol), and the reaction mixture was stirred at room temperature for 1 hour. The reaction mixture was diluted with water and extracted with DCM, and the organic layer was washed with brine, dried over Na₂SO₄, and concentrated to give the title compound as a white solid (20 mg, 95%): ¹H NMR (400 MHz, CDCl₃) δ 8.11–8.07 (m, 3H), 7.59 (d, *J* = 7.4 Hz, 1H), 7.50–7.36 (m, 2H), 7.06 (d, *J* = 9.1 Hz, 2H), 3.90 (d, *J* = 9.6 Hz, 5H), 2.25 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 165.06, 160.40, 138.27, 130.75, 130.02, 129.91, 127.25, 126.65, 121.30, 114.65, 77.30, 76.99, 76.67, 61.53, 55.62, 45.51; HRMS calcd for C₁₇H₁₉N₅O 310.1662 [M+H⁺], found 310.1664.

2-(2-(4-Methoxyphenyl)-2*H***-tetrazol-5-yl)benzoic acid (4):** Compound 4 was synthesized as a colorless solid with 92% yield from **S2**: ¹H NMR (500 MHz, acetone-d₆) δ 8.04 (d, *J* = 8.5 Hz, 2H), 7.91 (d, *J* = 7.3 Hz, 1H), 7.84 (d, *J* = 7.3 Hz, 1H), 7.70–7.56 (m, 2H), 7.11 (d, *J* = 7.3 Hz, 2H), 3.87 (s, 3H).

Scheme S2



2-(2-(4-Methoxyphenyl)-2*H***-tetrazol-5-yl)phenyl)acetonitrile (5):** To a solution of tetrazole **S3** (100 mg, 0.29 mmol) in DMF (10 mL) was added NaCN (43 mg, 0.87 mmol), and the suspension was stirred at 80 °C overnight. After cooling to room temperature, the reaction mixture was added water and extracted with diethyl ether. The ether layer was concentrated, and the resulting residue was purified by silica gel flash chromatography to give the desired product as a white solid (63 mg, 75%): ¹H NMR (500 MHz, CDCl₃) δ 8.32 (dd, *J* = 7.2, 1.7 Hz, 1H), 8.10 (d, *J* = 9.0 Hz, 2H), 7.66 (d, *J* = 6.9 Hz, 1H), 7.61–7.44 (m, 2H), 7.07 (d, *J* = 9.0 Hz, 2H), 4.36 (s, 2H), 3.90 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 163.91, 160.69, 130.83, 130.04, 129.92, 128.95, 128.62, 125.73, 121.40, 117.85, 114.75, 55.65, 23.52; HRMS calcd for C₁₆H₁₃N₅O 292.1193 [M+H⁺], found 292.1189.

5-(2-(2-Methoxyethyl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (9): To a round bottom flask was added tetrazole **5** (60 mg, 0.21 mmol), water (5.3 mL), acetic acid (6 mL), concentrated sulfuric acid (5 mL), and the mixture was stirred at 110 °C overnight. After cooling to room temperature, water was added and the mixture was extracted with ethyl acetate. The organic layer was collected, dried over Na₂SO₄, and concentrated to give the carboxylic acid intermediate. The acid was then dissolved in methanol and heated to reflux in the presence of catalytic amount of sulfuric acid until TLC showed complete conversion of the

carboxylic acid to the ester. Water was then added and the mixture was extracted by ethyl acetate. The organic layer was washed sequentially with water and saturated NaHCO₃ solution, dried over Na₂SO₄, and concentrated to give the ester intermediate. To the solution of ester in hydrous THF was added LiAlH₄ (20 mg, 0.5 mmol), the mixture was stirred at room temperature until TLC showed complete consumption of the starting material. The reaction was quenched by adding 1 mL methanol, and the mixture was filtered through Celite, washed with ethyl acetate, and concentrated to give the alcohol intermediate. Finally, to a solution of alcohol in anhydrous THF at 0 °C was added NaH (60%; 21 mg, 0.51 mmol), and the mixture was stirred at room temperature for 10 min. Then tetrabutylammonium iodide (13 mg, 0.036 mmol) and methyl iodide (105 μ L, 1.70 mmol) were added. The mixture was stirred at room temperature for 8 hours. Several drops of saturated NH₄Cl solution were added followed by water. The mixture was extracted with ethyl acetate. After evaporating the solvent under reduced pressure, the residue was purified by silica gel flash chromatography using ethyl acetate/hexanes as eluent to give the title compound as a white solid (33 mg, 52%): ¹H NMR (500 MHz, CDCl₃) δ 8.22–8.01 (m, 3H), 7.49–7.31 (m, 3H), 7.06 (d, J = 9.0 Hz, 2H), 3.89 (s, 3H), 3.69 (t, J = 7.1 Hz, 2H), 3.44–3.25 (m, 5H); ¹³C NMR (126 MHz, CDCl₃) δ 165.00, 160.44, 138.10, 131.43, 130.07, 129.92, 126.68, 126.32, 121.24, 114.67, 73.14, 58.56, 55.62, 34.49; HRMS calcd for C₁₇H₁₈N₄O₂ 311.1503 [M+H⁺], found 311.1538.

Scheme S3



2-(2-(4-Methoxyphenyl)-2*H***-tetrazol-5-yl)benzaldehyde (S4):** To a solution of tetrazole **3** (110 mg, 0.39 mmol) in DCM was added PCC (126 mg, 0.59 mmol), and the mixture was stirred at room temperature overnight. The solution was concentrated under reduced pressure, and the residue was purified by silica gel flash chromatography to give the desired product as a white solid (82 mg, 75%): ¹H NMR (500 MHz, CDCl₃) δ 10.81 (s, 1H), 8.24 (d, *J* = 7.8 Hz, 1H), 8.15–8.06 (m, 3H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.65 (t, *J* = 7.6 Hz, 1H), 7.09 (d, *J* = 8.9 Hz, 2H), 3.91 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.07, 163.04, 160.77, 134.67, 133.71, 130.47, 130.08, 128.97, 128.11, 121.40, 114.76, 55.66; HRMS calcd for C₁₅H₁₂N₄O₂ 281.1033 [M+H⁺], found 281.1030.

(±)1-(2-(2-(4-Methoxyphenyl)-2*H*-tetrazol-5-yl)phenyl)ethanol (S5): To a solution of tetrazole S4 (50 mg, 0.18 mmol) was in anhydrous THF was added methylmagnesium bromide (3 M in ether; 0.30 mL, 0.89 mmol), and the mixture was stirred at 50 °C under argon. After TLC showed complete disappearance of the starting materials, drops of saturated NH₄Cl solution was added followed by water. The mixture was then extracted with ethyl acetate, and the organic layer was separated and concentrated. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (47 mg, 88%): ¹H NMR (400 MHz, CDCl₃) δ 8.17–7.97 (m, 3H), 7.74 (dd, *J* = 7.7, 0.6 Hz, 1H), 7.53 (td, *J* = 7.6, 1.3 Hz, 1H), 7.43 (td, *J* = 7.6, 1.3 Hz, 1H), 7.07 (d, *J* = 9.2 Hz, 2H), 5.37 (dd, *J* = 6.4, 4.7 Hz, 1H), 3.92 (d, *J* = 6.5 Hz, 1H), 3.89 (s, 3H), 1.62 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.72, 160.68, 144.34, 130.74, 130.15, 129.97, 127.63, 126.35, 124.90, 121.37, 114.75, 66.64, 55.66, 22.45; HRMS calcd for C₁₆H₁₆N₄O₂ 297.1346 [M+H⁺], found 297.1343.

(±)5-(2-(1-Methoxyethyl)phenyl)-2-(4-methoxyphenyl)-2*H*-tetrazole (10): Tetrazole 10 was synthesized using the same etherification procedure as 6 with 90% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.19–7.99 (m, 3H), 7.74 (d, *J* = 7.8 Hz, 1H), 7.55 (t, *J* = 7.5 Hz, 1H), 7.42 (t, *J* = 7.5 Hz, 1H), 7.07 (d, *J* =

9.1 Hz, 2H), 5.20 (d, J = 6.2 Hz, 1H), 3.89 (s, 3H), 3.21 (s, 3H), 1.55 (d, J = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.60, 160.52, 143.55, 130.68, 130.34, 129.62, 127.26, 126.08, 125.13, 121.25, 114.71, 75.82, 56.60, 55.64, 23.86; HRMS calcd for C₁₇H₁₈N₄O₂ 311.1503 [M+H⁺], found 311.1538.

Scheme S4



2-(2-(4-Methoxyphenyl)-2H-tetrazol-5-yl)phenyl)propan-2-ol (S6): To a solution of tetrazole **S2** (60 mg, 0.19 mmol) in anhydrous THF was added methylmagnesium bromide (3 M in ether; 0.33 mL, 1.0 mmol), and the mixture was stirred at 70 °C under argon until TLC showed complete disappearance of the starting materials. Drops of saturated NH₄Cl solution were then added followed by water. The mixture was extracted with ethyl acetate and the organic layer was collected and concentrated to dryness. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (49 mg, 82%): ¹H NMR (400 MHz, CDCl₃) δ 8.18–8.02 (m, 2H), 7.92 (dd, *J* = 7.7, 1.4 Hz, 1H), 7.67 (dd, *J* = 8.0, 1.0 Hz, 1H), 7.50 (td, *J* = 7.7, 1.6 Hz, 1H), 7.41 (td, *J* = 7.5, 1.2 Hz, 1H), 7.15–6.97 (m, 2H), 5.52 (s, 1H), 3.89 (s, 3H), 1.61 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 166.75, 160.71, 147.51, 132.42, 130.41, 129.99, 127.08, 126.49, 124.73, 121.36, 114.72, 72.26, 55.62, 30.77; HRMS calcd for C₁₇H₁₆N₄O₂ 311.1502 [M+H⁺], found 311.1496.

2-(4-Methoxyphenyl)-5-(2-(2-methoxypropan-2-yl)phenyl)-2*H***-tetrazole (11): Tetrazole 11 was synthesized using the same etherification procedure as 6** with 90% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.09 (d, J = 9.2 Hz, 2H), 7.62–7.41 (m, 2H), 7.41–7.29 (m, 2H), 7.05 (d, J = 9.2 Hz, 2H), 3.88 (s, 3H), 2.89 (s, 3H), 1.63 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 166.90, 160.31, 146.06, 132.55, 130.54, 130.01, 127.17, 126.75, 126.27, 121.24, 114.61, 77.96, 55.61, 50.55, 28.97; HRMS calcd for C₁₈H₂₀N₄O₂ 325.1659 [M+H⁺], found 325.1654.

2-(2-Chloro-4-methoxyphenyl)-5-(2-(2-methoxypropan-2-yl)phenyl)-2H-tetrazole (12): Tetrazole 12 was synthesized using the same procedure as **11** except 2-chloro-4-methoxyaniline was used a starting material: ¹H NMR (500 MHz, CDCl₃) δ 7.58 (d, J = 8.9 Hz, 1H), 7.54–7.46 (m, 2H), 7.43–7.31 (m, 2H), 7.13 (d, J = 2.3 Hz, 1H), 6.97 (dd, J = 8.8, 2.3 Hz, 1H), 3.89 (s, 3H), 2.91 (s, 3H), 1.63 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 167.01, 161.33, 146.21, 132.64, 130.87, 130.04, 128.54, 127.16, 126.68, 125.96,

115.88, 113.31, 77.96, 55.93, 50.53, 28.91; HRMS calcd for $C_{18}H_{19}ClN_4O_2$ 359.1269 [M+H⁺], found 359.1265.

Methyl 2-formyl-5-methoxybenzoate (S7) Aldehyde **S7** was synthesized using a reported procedure^[S4]: ¹H NMR (300 MHz, CDCl₃) δ 10.48 (s, 1H), 7.95 (d, *J* = 6.0 Hz, 1H), 7.40 (s, 1H), 7.11 (d, *J* = 6.0 Hz, 1H), 3.98 (s, 3H), 3.90 (s, 3H).

5-(4-Methoxy-2-(2-methoxypropan-2-yl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (13): Tetrazole 13 was synthesized using the same procedure as 11 from aldehyde S5: ¹H NMR (500 MHz, CDCl₃) δ 8.08 (d, *J* = 9.1 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 1H), 7.05 (dd, *J* = 15.8, 5.7 Hz, 3H), 6.88 (dd, *J* = 8.4, 2.5 Hz, 1H), 3.88 (s, 6H), 2.90 (s, 3H), 1.61 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 166.75, 160.75, 160.26, 147.86, 133.89, 121.21, 118.49, 114.59, 113.98, 110.95, 77.92, 55.61, 55.33, 50.61, 28.79; HRMS calcd for C₁₉H₂₂N₄O₃ 355.1765 [M+H⁺], found 355.1759.

2-(2,4-Dimethoxyphenyl)-5-(4-methoxy-2-(2-methoxypropan-2-yl)phenyl)-2H-tetrazole (14): Tetrazole 14 was synthesized using the same procedure as 11 with S7 and 2,4-dimethoxyaniline as the starting materials: ¹H NMR (400 MHz, CDCl₃) δ 7.52 (d, *J* = 8.6 Hz, 1H), 7.36 (d, *J* = 8.4 Hz, 1H), 7.10 (d, *J* = 2.2 Hz, 1H), 6.87 (dd, *J* = 8.4, 2.2 Hz, 1H), 6.61 (dd, *J* = 11.8, 2.9 Hz, 2H), 3.89 (s, 3H), 3.85 (s, 3H), 2.95 (s, 3H), 1.59 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 166.36, 162.30, 160.67, 154.65, 147.93, 133.94, 127.77, 118.61, 113.75, 110.98, 104.45, 99.77, 77.99, 56.09, 55.71, 55.32, 50.60, 28.71; HRMS calcd for C₁₀H₂₄N₄O₄ 385.1870 [M+H⁺], found 385.1863.

Scheme S5



5-(2,6-Dimethoxyphenyl)-2-(4-methoxyphenyl)-2H-tetrazole (16): Tetrazole **16** was synthesized using the same procedure as tetrazole **2** with 60% yield: ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, *J* = 9.0 Hz, 2H), 7.41 (t, *J* = 8.4 Hz, 1H), 7.03 (d, *J* = 9.0 Hz, 2H), 6.66 (d, *J* = 8.5 Hz, 2H), 3.87 (s, 3H), 3.76 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 160.31, 159.69, 159.51, 131.93, 130.68, 121.49, 114.53, 105.84, 103.96, 56.03, 55.62; HRMS calcd for C₁₆H₁₆N₄O₃ 313.1295 [M+H⁺], found 313.1298.

5-(2,6-Dimethylphenyl)-2-(4-methoxyphenyl)-2*H***-tetrazole (15): Tetrazole 15 was synthesized using the same procedure as tetrazole 2 with 12% yield: ¹H NMR (400 MHz, CDCl₃) \delta 8.12 (d,** *J* **= 9.1 Hz, 2H), 7.28 (dd,** *J* **= 14.0, 6.4 Hz, 1H), 7.16 (d,** *J* **= 7.6 Hz, 2H), 7.06 (d,** *J* **= 9.1 Hz, 2H), 3.89 (s, 3H), 2.22 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) \delta 164.04, 160.46, 138.26, 129.67, 127.74, 127.10, 121.27, 114.68, 55.65, 20.49; HRMS calcd for C₁₆H₁₆N₄O 281.1397 [M+H⁺], found 281.1394.**

5-(2,6-Bis(methoxymethyl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (17): To a mixture of tetrazole 15 (127 mg, 0.45 mmol) in pyridine (2 mL), a solution of NaOH (182 mg, 4.54 mmol) in 6 mL water and solid KMnO4 (358 mg, 2.27 mmol) were added. The reaction mixture was refluxed with stirring overnight. Ethanol (20 mL) was then slowly added to the mixture over 10 min before the reaction was cooled down to room temperature. The mixture was filtered, washed with hot water, acidified with 1 N HCl, and extracted with ethyl acetate. The organic layer was separated and concentrated to give the crude dicarboxylic acid. To a solution of dicarboxylic acid in acetone was added methyl iodide (183 μ L, 2.94 mmol) and K₂CO₃ (406 mg, 2.94 mmol), and the mixture was refluxed for 8 hours. After cooled down, the solution was filtered through Celite and washed with ethyl acetate. The filtrate was concentrated to give the crude diester. To a solution of diester in hydrous THF was added LiAlH₄ (35 mg, 0.93 mmol), and the mixture was stirred at room temperature until all starting materials were consumed. The reaction was then quenched by adding 1 mL methanol. The mixture was filtered through Celite and washed with ethyl acetate. The filtrate was concentrated to give the crude diol. To a solution of diol in anhydrous THF at 0 °C was added NaH (60%; 44 mg, 1.08 mmol), and the mixture was stirred a room temperature for 10 min. Then tetrabutylammonium iodide (13 mg, 0.036 mmol) and methyl iodide (112 μ L, 1.80 mmol) were added. The mixture was stirred at room for 8 hours. Several drops of saturated NH₄Cl solution were then added followed by water. The mixture was extracted with ethyl acetate, and the organic layers were combined. After evaporating the solvent under reduced pressure, the residue was purified by silica gel flash chromatography using ethyl acetate/hexanes as eluent to give the title product as white solid (50 mg, 33%): ¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, J = 9.1 Hz, 2H), 7.54 (m, 3H), 7.07 (d, J = 9.1 Hz, 2H), 4.46 (s, 4H), 3.89 (s, 3H), 3.26 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 162.59, 160.56, 138.67, 130.13, 127.40, 124.77, 121.27, 114.72, 72.15, 58.20, 55.66; HRMS calcd for C₁₈H₂₀N₄O₃ 341.1608 [M+H⁺], found 341.1605.

Scheme S6



5-(2-Bromophenyl)-2-(4-methoxyphenyl)-2H-tetrazole (S8): Tetrazole **S8** was synthesized using the same procedure as tetrazole **2** with 55% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.10 (d, J = 9.0 Hz, 2H), 7.95 (d, J = 6.7 Hz, 1H), 7.75 (d, J = 7.9 Hz, 1H), 7.45 (t, J = 7.4 Hz, 1H), 7.33 (t, J = 7.1 Hz, 1H), 7.04 (d, J = 9.0 Hz, 2H), 3.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.92, 160.53, 134.13, 131.64, 131.25, 128.37, 127.42, 122.06, 121.40, 114.64, 55.60; HRMS calcd for C₁₄H₁₁BrN₄O 331.0189 [M+H⁺], found 331.0189.

5-(2-(Furan-2-yl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (18): To a mixture of tetrazole **S8** (100 mg, 0.30 mmol), furan-2-ylboronic acid (51 mg, 0.45 mmol), K₂CO₃ (125 mg, 0.91 mmol) and bis(triphenylphosphine)palladium(II) dichloride (11 mg, 0.015 mmol) was added DMF (3 mL) and H₂O (500 μ L) under argon. The reaction was stirred at 100 °C overnight under argon. The reaction mixture was then concentrated under reduced pressure, and the residue was purified by silica gel flash chromatography to give the desired product as a white solid (80 mg, 83%): ¹H NMR (400 MHz, CDCl₃) δ 8.04 (d, *J* = 9.0
Hz, 2H), 7.88–7.73 (m, 2H), 7.54 (dd, J = 7.7, 1.1 Hz, 1H), 7.46 (td, J = 7.5, 1.0 Hz, 1H), 7.36 (s, 1H), 7.03 (d, J = 9.0 Hz, 2H), 6.40 (d, J = 1.5 Hz, 1H), 6.32 (s, 1H), 3.88 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.90, 160.34, 152.31, 142.16, 130.89, 130.76, 130.30, 130.11, 128.63, 127.83, 124.91, 121.17, 114.55, 111.28, 108.68, 55.51; HRMS calcd for C₁₈H₁₄N₄O₂ 319.1190 [M+H⁺], found 319.1186.

2-(4-Methoxyphenyl)-5-(2-(thiophen-2-yl)phenyl)-2*H***-tetrazole (19): Tetrazole 19 was synthesized using the same procedure as tetrazole 18 with 85% yield: ¹H NMR (500 MHz, CDCl₃) \delta 7.95 (d,** *J* **= 9.0 Hz, 2H), 7.89 (d,** *J* **= 6.7 Hz, 1H), 7.61 (d,** *J* **= 7.0 Hz, 1H), 7.56–7.41 (m, 2H), 7.32–7.20 (m, 1H), 7.05–6.86 (m, 4H), 3.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) \delta 164.69, 160.37, 141.80, 134.67, 131.36, 130.61, 130.33, 130.04, 128.00, 127.07, 126.96, 126.66, 125.83, 121.21, 114.57, 55.58; HRMS calcd for C₁₈H₁₄N₄OS 335.0961 [M+H⁺], found 335.0958.**

tert-Butyl 2-(2-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)phenyl)-1*H*-pyrrole-1-carboxylate (20): Tetrazole 20 was synthesized using the same procedure as tetrazole 18 with 75% yield: ¹H NMR (500 MHz, CDCl₃) δ 8.36–8.25 (m, 1H), 7.95 (d, *J* = 9.0 Hz, 2H), 7.48 (dd, *J* = 9.9, 6.5 Hz, 4H), 7.01 (d, *J* = 9.0 Hz, 2H), 6.27 (s, 1H), 6.11 (s, 1H), 3.86 (s, 3H), 1.08 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 164.47, 160.29, 149.32, 134.21, 132.68, 131.52, 129.62, 128.49, 128.07, 127.47, 121.11, 121.00, 114.58, 113.93, 110.69, 82.98, 55.58, 27.23; HRMS calcd for C₂₃H₂₃N₅O₃ 418.1874 [M+H⁺], found 418.1871.

5-(2'-Methoxy-[1,1'-biphenyl]-2-yl)-2-(4-methoxyphenyl)-2*H***-tetrazole (21): Tetrazole 21 was synthesized using the same procedure as tetrazole 18 with 80% yield: ¹H NMR (400 MHz, CDCl₃) \delta 8.14 (dd, J = 7.5, 1.3 Hz, 1H), 7.77 (d, J = 9.2 Hz, 2H), 7.60–7.40 (m, 3H), 7.40–7.22 (m, 2H), 6.97–7.06 (m, 1H), 6.95 (d, J = 9.2 Hz, 2H), 6.83 (d, J = 8.2 Hz, 1H), 3.83 (s, 3H), 3.44 (s, 3H); ¹³C NMR (100 MHz, cdcl₃) \delta 165.20, 160.17, 156.47, 138.29, 131.30, 130.77, 130.44, 129.93, 129.14, 128.82, 127.51, 126.92, 120.95, 120.62, 114.50, 110.41, 55.54, 55.10; HRMS calcd for C₂₁H₁₈N₄O₂ 359.1503 [M+H⁺], found 359.1501.**

2-(2-(4-Methoxyphenyl)-2*H***-tetrazol-5-yl)phenyl)pyridine (22):** Tetrazole **22** was synthesized using the same procedure as tetrazole **18** with 78% yield: ¹H NMR (500 MHz, CDCl₃) δ 8.58 (d, *J* = 4.1 Hz, 1H), 8.08 (d, *J* = 7.3 Hz, 1H), 7.86 (d, *J* = 8.9 Hz, 2H), 7.71–7.61 (m, 2H), 7.61–7.53 (m, 2H), 7.34 (d, *J* = 7.8 Hz, 1H), 7.23 (d, *J* = 6.7 Hz, 1H), 6.98 (d, *J* = 8.9 Hz, 2H), 3.86 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 164.86, 160.30, 159.06, 149.15, 140.83, 135.97, 130.48, 130.36, 130.20, 130.14, 128.59, 125.97, 124.14, 121.98, 121.10, 114.53, 55.60; HRMS calcd for C₁₉H₁₅N₅O 330.1349 [M+H⁺], found 330.1344.

5-(2-(1*H***-Pyrrol-2-yl)phenyl)-2-(4-methoxyphenyl)-2***H***-tetrazole (23): To a solution of tetrazole 20 (35 mg, 0.084 mmol) in anhydrous THF was added NaOMe (25% in methanol; 100 µL, 0.42 mmol) at 0 °C slowly via syringe under argon. The solution was stirred at 0 °C for 10 min and then room temperature for 30 min. Water was then added, and the mixture was extracted with diethyl ether. The organic layer was collected and concentrated under reduced pressure. The resulting residue was purified by silica gel flash chromatography using ethyl acetate/hexanes as eluent to give the title compound as a white solid (20 mg, 75%): ¹H NMR (400 MHz, CDCl₃) \delta 10.39 (s, 1H), 8.05 (d,** *J* **= 9.1 Hz, 2H), 7.82 (dd,** *J* **= 7.8, 1.1 Hz, 1H), 7.74 (dd,** *J* **= 7.9, 0.9 Hz, 1H), 7.59–7.47 (m, 1H), 7.44–7.33 (m, 1H), 7.05 (d,** *J* **= 9.1 Hz, 2H), 6.87 (dd,** *J* **= 4.1, 2.6 Hz, 1H), 6.43 (dd,** *J* **= 4.3, 3.1 Hz, 1H), 6.32–6.18 (m, 1H), 3.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) \delta 165.70, 160.65, 133.12, 131.65, 130.47, 130.20, 130.11, 126.46, 123.55, 121.44, 118.88, 114.70, 109.50, 109.22, 55.61; HRMS calcd for C₁₈H₁₅N₅O 318.1349 [M+H⁺], found 318.1348.**

Scheme S7



5-(2,6-Dibromophenyl)-2-(4-methoxyphenyl)-2*H***-tetrazole (S9**): Tetrazole **S9** was synthesized using the same procedure as tetrazole **2** with 35% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.13 (d, *J* = 8.4 Hz, 2H), 7.69 (dd, *J* = 8.1, 0.7 Hz, 2H), 7.32–7.18 (m, 1H), 7.07 (d, *J* = 8.4 Hz, 2H), 3.90 (d, *J* = 0.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.28, 160.68, 132.39, 131.87, 130.74, 130.30, 125.36, 121.50, 114.71, 55.67; HRMS calcd for C₁₄H₁₀Br₂N₄O 408.9294 [M+H⁺], found 408.9287.

5-(2,6-Di(furan-2-yl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (24): To a mixture of tetrazole **S9** (25 mg, 0.06 mmol), furan-2-ylboronic acid (34 mg, 0.31 mmol), K₂CO₃ (42 mg, 0.31 mmol) and bis(triphenylphosphine)palladium(II) dichloride (4.3 mg, 0.006 mmol) were added DMF (2 mL) and H₂O (300 µL) under argon. The reaction was stirred at 90 °C overnight under argon. The reaction mixture was then cooled down and concentrated under reduced pressure. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (19 mg, 81%): ¹H NMR (400 MHz, CDCl₃) δ 8.08 (d, *J* = 9.1 Hz, 2H), 7.80 (d, *J* = 7.9 Hz, 2H), 7.61 (dd, *J* = 8.3, 7.6 Hz, 1H), 7.30 (dd, *J* = 1.7, 0.6 Hz, 2H), 7.05 (d, *J* = 9.2 Hz, 2H), 6.24 (dd, *J* = 3.4, 1.8 Hz, 2H), 5.75 (dd, *J* = 3.4, 0.6 Hz, 2H), 3.88 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.31, 160.57, 151.68, 142.35, 132.77, 130.42, 126.35, 121.37, 120.73, 114.70, 111.45, 109.02, 55.68; HRMS calcd for C₂₂H₁₆N₄O₃ 385.1295 [M+H⁺], found 385.1294.

5-(2,6-Di(thiophen-2-yl)phenyl)-2-(4-methoxyphenyl)-2H-tetrazole (25): Tetrazole **25** was synthesized using a similar procedure as tetrazole **24** with 80% yield: ¹H NMR (500 MHz, CDCl₃) δ 7.93 (d, *J* = 9.0 Hz, 2H), 7.65–7.55 (m, 3H), 7.20–7.13 (m, 2H), 7.00 (d, *J* = 9.0 Hz, 2H), 6.92–6.82 (m, 4H), 3.86 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 163.14, 160.49, 141.36, 136.80, 130.22, 129.89, 127.40, 127.04, 126.31, 121.43, 119.52, 114.62, 55.66; HRMS calcd for C₂₂H₁₆N₄OS₂ 417.0838 [M+H⁺], found 417.0835.

Di*tert*-butyl 2,2'-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)-1,3-phenylene)-bis(1*H*-pyrrole-1carboxylate) (26): Tetrazole 26 was synthesized using a similar procedure as tetrazole 24 with 70% yield: ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 9.0 Hz, 2H), 7.51–7.44 (m, 3H), 7.30 (s, 2H), 6.96 (d, *J* = 9.1 Hz, 2H), 6.08 (t, *J* = 3.1 Hz, 2H), 5.93 (s, 2H), 3.84 (s, 3H), 1.26 (s, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 163.31, 160.08, 149.18, 135.78, 131.89, 130.66, 130.35, 128.19, 121.11, 120.91, 119.51, 114.68, 114.47, 110.14, 83.12, 55.54, 27.42; Orbitrap LC-MS calcd for C₃₂H₃₄N₆O₅ 583.2663 [M+H⁺], found 583.2651.

5-(2,6-Di(1*H***-pyrrol-2-yl)phenyl)-2-(4-methoxyphenyl)-2***H***-tetrazole (27): Tetrazole 27 was synthesized using a similar procedure as tetrazole 23 with 65% yield: ¹H NMR (400 MHz, CDCl₃) \delta 8.60 (s, 2H), 7.91 (d,** *J* **= 8.8 Hz, 2H), 7.47 (s, 3H), 6.99 (d,** *J* **= 8.8 Hz, 2H), 6.64 (s, 2H), 6.08 (d,** *J* **= 11.6 Hz, 4H), 3.85 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) \delta 164.64, 160.69, 135.57, 130.66, 130.38, 130.10, 127.27, 121.90, 121.59, 119.04, 114.67, 109.25, 109.23, 55.65; HRMS calcd for C₂₂H₁₈N₆O 383.1615 [M+H⁺], found 383.1614.**

Scheme S8



Dimethyl 2,2'-(2-(4-methoxyphenyl)-2H-tetrazol-5-yl)-1,3-phenylene)bis(1H-pyrrole-1-carboxyl-ate) (28): To a solution of tetrazole **27** (25 mg, 0.065 mmol) in acetonitrile (5 mL) was added dimethyl dicarbonate (21 μ L, 0.196 mmol) and DMAP (1.6 mg, 0.013 mmol), and the mixture was stirred at room temperature under argon for 24 hours. The solution was then concentrated, and the residue was purified by silica gel flash chromatography to give the desired product as a white solid (6.5 mg, 20%): ¹H NMR (400 MHz, CDCl₃) δ 7.79 (d, *J* = 9.0 Hz, 2H), 7.56–7.44 (m, 3H), 7.25 (d, *J* = 6.4 Hz, 2H), 6.96 (d, *J* = 9.1 Hz, 2H), 6.11–5.99 (br, 4H), 3.84 (s, 3H), 3.73 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 163.49, 160.22, 151.06, 134.97, 132.33, 130.46, 130.28, 128.58, 128.25, 121.24, 121.17, 115.12, 114.48, 111.00, 55.57, 53.73; HRMS calcd for C₂₆H₂₂N₆O₅ 499.1724 [M+H⁺], found 499.1708.

Di*tert*-butyl **2,2'-(4-(2-(4-methoxyphenyl)-2H-tetrazol-5-yl)-1,3-phenylene)bis(1H-pyrrole-1carboxylate) (29): Tetrazole 29 was synthesized using a similar procedure as tetrazole 26 except 2,4dibromobenzaldehyde was used as the starting material: ¹H NMR (400 MHz, CDCl₃) \delta 8.31 (d,** *J* **= 7.9 Hz, 1H), 7.95 (d,** *J* **= 9.1 Hz, 2H), 7.55–7.44 (m, 3H), 7.37 (dd,** *J* **= 3.3, 1.8 Hz, 1H), 7.01 (d,** *J* **= 9.1 Hz, 2H), 6.33–6.22 (m, 3H), 6.14 (dd,** *J* **= 3.2, 1.8 Hz, 1H), 3.87 (s, 3H), 1.44 (s, 9H), 1.11 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) \delta 164.37, 160.27, 149.26, 149.11, 135.61, 134.01, 133.42, 132.62, 131.99, 130.38, 128.55, 127.69, 126.00, 123.17, 121.11, 120.99, 115.14, 114.57, 113.88, 110.70, 110.67, 83.87, 83.01, 55.57, 27.71, 27.25; HRMS calcd for C₃₂H₃₄N₆O₅ 583.2663 [M+H⁺], found 583.2651.**

Scheme S9



Methyl 3,5-dibromo-4-(bromomethyl)benzoate (S10): Compound **S10** was synthesized by following a literature procedure^[S5]. To a solution of methyl 3,5-dibromo-4-methylbenzoate (154 mg, 0.5 mmol) in carbon tetrachloride was added NBS (107 mg, 0.6 mmol) followed by benzoyl peroxide (70%; 17 mg, 0.05 mmol). The mixture was refluxed until TLC showed complete disappearance of the starting materials. The reaction was then cooled down to room temperature, and the solvent was removed under reduced pressure. The residue was redissolved in ethyl acetate, washed with water, and concentrated. The residue was purified

by silica gel flash chromatography to give the desired product as a white solid (182 mg, 95%): ¹H NMR (400 MHz, CDCl₃) δ 8.20 (s, 2H), 4.83 (s, 2H), 3.93 (s, 3H).

Methyl 3,5-dibromo-4-formylbenzoate (S11): To a solution of **S10** (180 mg, 0.47 mmol) in acetonitrile was added 4 Å molecular sieves and *N*-methylmorpholine-*N*-oxide (82 mg, 1.70 mmol), and the mixture was stirred at 50 °C overnight. The reaction mixture was washed with brine and extracted with ethyl acetate, and the organic phase was concentrated. After evaporating the solvents, the residue was purified by silica gel flash chromatography to give the desired product as a white solid (123 mg, 82%): ¹H NMR (400 MHz, CDCl₃) δ 10.22 (s, 2H), 8.26 (s, 2H), 3.97 (s, 3H).

Methyl 3,5-dibromo-4-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)benzoate (S12): Tetrazole S12 was synthesized using a similar procedure as tetrazole 2 with 45% yield: ¹H NMR (400 MHz, CDCl₃) δ 8.33 (s, 2H), 8.12 (d, *J* = 9.0 Hz, 2H), 7.07 (d, *J* = 9.0 Hz, 2H), 3.98 (s, 3H), 3.90 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.02, 162.73, 160.81, 134.78, 133.98, 132.72, 130.22, 125.49, 121.56, 114.77, 55.70, 52.96; HRMS calcd for C₁₆H₁₂Br₂N₄O₃ 466.9349 [M+H⁺], found 466.9344.

(3,5-Dibromo-4-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)phenyl)methanol (S13) Tetrazole S13 was synthesized using the same procedure as tetrazole 17 and used directly without further purification.

Di*tert***-butyl** 2,2'-(5-(hydroxymethyl)-2-(2-(4-methoxyphenyl)-2*H*-tetrazol-5-yl)-1,3-phenylene) bis(1*H*-pyrrole-1-carboxylate) (S14): To a round bottom flask containing a solution of tetrazole S14 (30 mg, 0.068 mmol) in DMF (1.5 mL) and H₂O (100 µL) was added (1-(*tert*-butoxycarbonyl)-1*H*-pyrrol-2yl)boronic acid (58 mg, 0.273 mmol), K₂CO₃ (38 mg, 0.273 mmol), and bis(triphenylphosphine)palladium (II) dichloride (5 mg, 0.007 mmol) under argon. The reaction was stirred at 80 °C under argon overnight. The reaction mixture was then concentrated under reduced pressure, and the residue was purified by silica gel flash chromatography to give the title compound as a white solid (29 mg, 70%): ¹H NMR (400 MHz, CDCl₃) δ 7.84 (d, *J* = 9.1 Hz, 2H), 7.48 (s, 2H), 7.29 (s, 2H), 6.96 (d, *J* = 9.1 Hz, 2H), 6.08 (d, *J* = 3.2 Hz, 2H), 5.95 (s, 2H), 4.81 (s, 2H), 3.84 (s, 3H), 1.26 (s, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 160.08, 149.30, 141.13, 135.92, 131.90, 130.31, 128.86, 127.63, 121.11, 120.90, 114.46, 110.21, 83.11, 64.49, 55.53, 27.44; HRMS calcd for C₃₃H₃₆N₆O₆ 613.2769 [M+H⁺], found 613.2764.

4-((**3**,**5**-**Bis**(**1**-(*tert*-**butoxycarbony**])-**1***H*-**pyrrol-2-y**])-**4**-(**2**-(**4**-**methoxypheny**])-**2***H*-**tetrazo**]-**5**-**y**])**benzy**]) **oxy**)-**4**-**oxobutanoic acid** (**30**): To a solution of tetrazole **S14** (20 mg, 0.033 mmol) in dioxane was added succinic anhydride (16.3 mg, 0.163 mmol), triethylamine (46 μL, 0.327 mmol), and DMAP (2.0 mg, 0.016 mmol). The mixture was stirred at 70 °C under argon for 5 hours until TLC showed complete disappearance of the starting materials. The reaction mixture was cooled down and concentrated, and the residue was purified by silica gel flash chromatography to give the desired product as a white solid (20 mg, 86%)): ¹H NMR (500 MHz, CD₃OD) δ 7.82 (d, *J* = 10.0 Hz, 2H), 7.49 (s, 2H), 7.26 (s, 2H), 7.06 (d, *J* = 10.0 Hz, 2H), 6.10 (d, *J* = 5.0 Hz, 2H), 5.95 (s, 2H), 3.86 (s, 2H), 2.69 (d, *J* = 5.0 Hz, 2H), 2.64 (d, *J* = 5.0 Hz, 2H), 1.27 (s, 18H); ¹³C NMR (126 MHz, CD₃OD) δ 172.54, 160.73, 148.95, 137.25, 131.40, 129.86, 120.80, 120.72, 114.35, 109.96, 83.20, 65.18, 54.76, 28.75, 28.51, 26.41; HRMS calcd for C₃₇H₄₀N₆O₉ 713.2930 [M+H⁺], found 713.2918.

Scheme S10



Methyl 3,5-dibromo-4-(2-(4-hydroxyphenyl)-2*H*-tetrazol-5-yl)benzoate (S15): To a solution of S12 (210 mg, 0.45 mmol) in DCM (30 mL) cooled with an ice-water bath was slowly added boron tribromide (430 µL, 4.5 mmol), and the mixture was stirred at room temperature overnight. After cooling the reaction mixture with an ice-water bath, 1 mL methanol was slowly added and the solution was concentrated. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (128 mg, 63%): ¹H NMR (400 MHz, acetone- d_6) δ 8.34 (s, 2H), 8.06 (d, *J* = 9.0 Hz, 2H), 7.13 (d, *J* = 9.0 Hz, 2H), 3.98 (s, 3H), 2.95 (s, 1H); ¹³C NMR (100 MHz, acetone- d_6) δ 163.42, 159.26, 134.69, 132.50, 125.08, 121.70, 116.38, 52.50; HRMS calcd for C₁₅H₁₀Br₂N₄O₃ 452.9192 [M+H⁺], found 452.9184.

4-(5-(2,6-Dibromo-4-(hydroxymethyl)phenyl)-2H-tetrazol-2-yl)phenol (S16): To a solution of **S15** (130 mg, 0.29 mmol) in anhydrous THF in a round-bottom flask cooled in a water-ice bath was added diisobutylaluminium hydride (1 M in hexane; 1.43 mL, 1.43 mmol). The reaction mixture was stirred at room temperature with TLC monitoring. The Rochelle salt solution was added after TLC showed complete disappearance of the starting materials. The mixture was stirred while ethyl acetate was added. The organic layer was collected and concentrated. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (98 mg, 80%): ¹H NMR (400 MHz, CD₃OD) δ 8.00 (d, *J* = 9.0 Hz, 2H), 7.78 (s, 2H), 7.01 (d, *J* = 9.0 Hz, 2H), 4.69 (s, 2H); ¹³C NMR (100 MHz, CD₃OD) δ 163.08, 159.37, 148.12, 129.49, 128.96, 128.62, 124.51, 121.31, 115.86, 61.79; HRMS calcd for C₁₄H₁₀Br₂N₄O₂ 424.9243 [M+H⁺], found 452.9241.

tert-Butyl (2-(4-(5-(2,6-dibromo-4-(hydroxymethyl)phenyl)-2*H*-tetrazol-2-yl)phenoxy)ethyl) carbamate (S17): To a solution of S16 (90 mg, 0.21 mmol) in acetone was added K₂CO₃ (146 mg, 1.06 mmol) and *tert*-butyl (2-iodoethyl) carbamate (92 mg, 0.34 mmol), and the mixture was refluxed under argon overnight. Then the mixture was filtered through Celite after cooling down and washed with ethyl acetate. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel flash chromatography to give the desired product as a white solid (100 mg, 83%): ¹H NMR (500 MHz, CDCl₃) δ 8.12 (d, *J* = 9.1 Hz, 2H), 7.65 (s, 2H), 7.06 (d, *J* = 9.1 Hz, 2H), 5.07 (s, 1H), 4.76 (s, 2H), 4.10 (t, *J* = 5.1 Hz, 2H), 3.70–3.44 (m, 3H), 1.46 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 163.18, 159.72, 147.08,

130.44, 129.29, 128.62, 125.01, 121.59, 115.24, 67.59, 62.75, 28.35; HRMS calcd for $C_{21}H_{23}Br_2N_5O_4$ 568.0190 [M+H⁺], found 568.0178.

Di*tert***-butyl** 2,2'-(2-(4-(2-aminoethoxy)phenyl)-2*H*-tetrazol-5-yl)-5-(hydroxymethyl)-1,3-phenylene)bis(1*H*-pyrrole-1-carboxylate) (S18): To a solution of S17 (95 mg, 0.167 mmol) in DCM (6 mL) was added 2,2,2-trifluoroacetic acid (4 mL), and the mixture was stirred at 0 °C until all starting materials were consumed. The solvent was removed to obtain the crude tetrazole-amine intermediate. To a solution of tetrazole-amine in 2 mL DMF and 300 μ L H₂O was added 1-(*tert*-butoxycarbonyl)-1*H*-pyrrol-2-yl)boronic acid (187 mg, 0.887 mmol), K₂CO₃ (162 mg, 1.17 mmol) and bis(triphenylphosphine)palladium(II) dichloride (16 mg, 0.022 mmol), and the mixture was stirred at 80 °C overnight under argon. The reaction mixture was cooled down and concentrated. The residue was purified by silica gel flash chromatography to give the desired product as a white solid (27 mg, 25%): ¹H NMR (400 MHz, CD₃OD) δ 7.84 (d, *J* = 9.0 Hz, 2H), 7.45 (s, 2H), 7.23 (s, 2H), 7.15 (d, *J* = 9.1 Hz, 2H), 6.08 (s, 2H), 5.94 (s, 2H), 4.75 (s, 2H), 4.37–4.26 (m, 2H), 3.43–3.35 (m, 2H), 3.08 (s, 1H), 1.25 (s, 18H); ¹³C NMR (100 MHz, CD₃OD) δ 159.20, 158.89, 157.20, 142.55, 135.79, 131.98, 130.66, 128.40, 120.79, 120.68, 115.16, 109.96, 83.13, 64.49, 62.88, 38.82, 26.41; HRMS calcd for C₃₄H₃₉N₇O₆ 642.3035 [M+H⁺], found 642.3024.

Sodium 1-(6-((2-(4-(5-(2,6-bis(1-(*tert*-butoxycarbonyl)-1*H*-pyrrol-2-yl)-4-(hydroxymethyl)phenyl)-2*H*-tetrazol-2-yl)phenoxy)ethyl)amino)-6-oxohexyl)-3,3-dimethyl-2-((1*E*,3*E*,5*E*)-5-(1,3,3-trimethyl-5-sulfonatoindolin-2-ylidene)penta-1,3-dien-1-yl)-3*H*-indol-1-ium-5-sulfonate (S19): To a solution of S18 (2.28 mg, 0.0038 mmol) in 500 μ L DMF was added *N*,*N*-diisopropylethylamine (2 μ L, 0.0078 mmol) and sulfo-Cy5-NHS ester (2.0 mg, 0.0026 mmol), and the mixture was stirred at room temperature overnight. The product mixture was concentrated and the residue purified by reverse-phase HPLC using acetonitrile/H₂O as eluent to give the title compound as a green solid (1.35 mg, 30%): HRMS calcd for C₆₆H₇₄N₉NaO₁₃S₂ 1264.4853 [M-Na⁻], found 1264.4860.

Sodium 1-(6-((2-(4-(5-(2,6-bis(1-(*tert*-butoxycarbonyl)-1*H*-pyrrol-2-yl)-4-((sulfooxy)methyl)phenyl)-2*H*-tetrazol-2-yl)phenoxy)ethyl)amino)-6-oxohexyl)-3,3-dimethyl-2-((1*E*,3*E*,5*E*)-5-(1,3,3-trimethyl-5-sulfonatoindolin-2-ylidene)penta-1,3-dien-1-yl)-3*H*-indol-1-ium-5-sulfonate (Tetrazole-Cy5; 31): To a solution of **S19** (1.00 mg, 0.0008 mmol) in 200 μ L DMF was added SO₃-pyridine complex (1.64 mg, 0.0103 mmol), and the mixture was stirred until MS (LCQ) showed disappearance of the starting materials. The solvent was removed and dried thoroughly under vacuum to give the title compound as a green solid with 95% yield: HRMS calcd for C₆₆H₇₄N₉NaO₁₆S₃ 1344.4421 [M-Na⁻], found 1344.4456.

Computational Details:

All calculations were carried out with the Gaussian 09 program package.^[S6] Geometry optimizations were performed with the B3LYP^[S7] functional with Grimme's DFT-D3 dispersion correction,^[S8] the 6-31+G(d) basis set, and the SMD^[S9] solvation model with water as the solvent. Frequency analysis was conducted at the same level of theory to verify the stationary points to be minima or saddle points and to obtain zero-point energy (ZPE) and thermal energy corrections at 298.15 K. Single-point energy calculations on the B3LYP-D3-optimized geometries were performed with the ω B97X-D^[S10] functional, using 6-311++G(d,p) basis set and the SMD solvation model with water as the solvent. Computed structures are illustrated using CYLView.^[S11]

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NMR spectra:



















































S62



N=N S8



-OMe



S63





































120 110 100 90 80 70 f1 (ppm) 160 150 140 130 -10






















S79

















Appendix

Table S1, entry 1:



Based on the peak integration (red line, 254 nm), the ratio of glutathione addition product over the acrylamide cycloaddition product was 0.51:0.42. In the presence of acrylamide, the acetic acid addition product was not detected.

Table S1, entry 2:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was 1:1.

Table S1, entry 3:



Based on the chromatogram, the intramolecular addition product was the major product. The acrylamide cycloaddition product and the glutathione addition product were not observed.

Table S1, entry 4:



Based on the integration, only the intramolecular addition product was detected when acrylamide and glutathione were present.



The intramolecular addition product was confirmed by $^1\mathrm{H}$ and $^{13}\mathrm{C}$ NMR spectra.

Table S1, entry 5:



Based on the integration, the ratio of glutathione addition products over the acrylamide cycloaddition product was determined to be 1:1.

Table S1, entry 6:



Based on the integration, the ratio of the glutathione addition product over the acrylamide cycloaddition product was determined to be 0.45:0.55.

Table S1, entry 7:



Based on chromatograms, only the intramolecular addition and nucleophilic addition products were observed.

Table S1, entry 8:



Based on HPLC chromatograms, only the intramolecular addition product was detected.

Table S1, entry 9:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 1:1.

Table S1, entry 10:



Based on HPLC chromatogram, the glutathione addition products were the major products.

Table S1, entry 11:



Based on HPLC chromatograms, both the glutathione addition and the acrylamide cycloaddition were blocked.

Table S1, entry 12:



Based on HPLC chromatograms, both the glutathione addition and the acrylamide cycloaddition were blocked.

Table S1, entry 13:



Based on HPLC chromatograms, both the glutathione addition and the acrylamide cycloaddition were blocked.

Table S1, entry 14:



Based on HPLC chromatograms, both the glutathione addition and the acrylamide cycloaddition were blocked.

Table S1, entry 15:



Based on HPLC chromatograms, both the glutathione addition and the acrylamide cycloaddition were blocked.

Table S1, entry 16:



Based on HPLC chromatograms, the solvent quenching products were significant.

Table S1, entry 17:



Based on HPLC chromatograms, the acrylamide cycloaddition were significantly blocked. The glutathione addition product was the major product.

Table S1, entry 18:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.58:0.37.

Table S1, entry 19:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.56:0.36.

Table S1, entry 20:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.39:0.58.

Table S1, entry 21:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.41:0.56.

Table S1, entry 22:



Based on HPLC chromatogram, only the intramolecular addition product was detected.

Table S1, entry 23:



Based on HPLC chromatograms, only trace amounts of acrylamide cycloaddition and glutathione addition products were detected.

Table S1, entry 24:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.59:0.30.
Table S1, entry 25:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.63:0.27.

Table S1, entry 26:



Based on HPLC chromatogram, only trace amount of glutathione addition product was detected. The major product was the acrylamide cycloaddition product (>90%).

Table S1, entry 27:



Based on HPLC chromatograms, neither acrylamide cycloaddition product nor glutathione addition product was detected.

Table S1, entry 28:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.20:0.78.

Table S1, entry 29:



Based on the integration, the ratio of glutathione addition product over the acrylamide cycloaddition product was determined to be 0.24:0.74.

Table 1.





Based on the peak integrations, the glutathione addition products were dominant for most dipolarophiles except dimethyl fumarate. Sph and TCO gave the cycloaddition products at \sim 60% yield.





Based on HPLC chromatograms, small amounts of glutathione addition product were detected when 1,3-Cp, styrene, norbornene and 3,3-Cp were used as a dipolarophile. The glutathione addition products were not detected when Sph, dimethyl fumarate, and TCO-amine was used as a dipolarophile.

Cartesian Coordinates

spirohexene

b3lyp-d3 SCF energy in solution: -272.66162297 a.u. b3lyp-d3 enthalpy in solution: -272.507054 a.u. b3lyp-d3 free energy in solution: -272.546370 a.u. wb97xd SCF energy in solution: -272.62411063 a.u. wb97xd enthalpy in solution: -272.469542 a.u. wb97xd free energy in solution: -272.508858 a.u.

Cartesian coordinates

ATOM	Х	Y	Z
С	-0.224107	1.100113	-0.046043
С	-1.255362	0.000019	-0.447651
С	-0.224263	-1.100211	-0.046035
С	0.855448	-0.000144	0.042395
Н	-0.048626	1.906180	-0.769750
Н	-0.454341	1.548266	0.930898
Н	-1.409921	0.000033	-1.533880
Н	-0.048926	-1.906341	-0.769711
Н	-0.454553	-1.548307	0.930917
С	-2.599196	0.000097	0.268673
Н	-3.191796	-0.887099	0.007867
Н	-2.457929	0.000097	1.358017
Н	-3.191679	0.887366	0.007851
С	2.182870	0.000103	0.736288
Н	2.708463	-0.000138	1.683325
С	2.222813	-0.000035	-0.567811
Н	2.800091	0.000293	-1.484428

sme-anion

b3lyp-d3 SCF energy in solution: -438.22587504 a.u. b3lyp-d3 enthalpy in solution: -438.185307 a.u. b3lyp-d3 free energy in solution: -438.211902 a.u. wb97xd SCF energy in solution: -438.22526151 a.u. wb97xd enthalpy in solution: -438.184693 a.u. wb97xd free energy in solution: -438.211288 a.u.

ATOM	Х	Y	Z
S	0.000000	0.00000	0.717176
С	0.000000	0.00000	-1.140203
Н	0.000000	1.021442	-1.544532
Н	0.884595	-0.510721	-1.544532
Н	-0.884595	-0.510721	-1.544532

nitrile imine-1

b3lyp-d3 SCF energy in solution: -834.96607471 a.u. b3lyp-d3 enthalpy in solution: -834.708431 a.u. b3lyp-d3 free energy in solution: -834.768640 a.u. wb97xd SCF energy in solution: -834.78917537 a.u. wb97xd enthalpy in solution: -834.531532 a.u. wb97xd free energy in solution: -834.591741 a.u.

ATOM	Х	Y	Z
С	5.662651	-0.259102	-0.118023
С	4.374384	-0.791337	-0.194202
С	3.254792	0.036523	-0.002000
С	3.446748	1.403428	0.265149
С	4.737033	1.929688	0.340438
С	5.849125	1.101620	0.149367
Н	6.520658	-0.908715	-0.268448
Н	4.237185	-1.847489	-0.402892
Н	2.589712	2.052345	0.415995
Н	4.873815	2.987438	0.548633
Н	6.853022	1.513496	0.209051
Ν	0.780552	0.190503	0.054305
Ν	-0.165541	-0.736502	-0.085568
Ν	0.319601	-1.958179	-0.294944
Ν	1.627685	-1.841651	-0.287249
С	1.902228	-0.524184	-0.079177
С	-1.563731	-0.456884	-0.019493
С	-2.439363	-1.436092	0.465909
С	-2.038420	0.785000	-0.439484
С	-3.799414	-1.162418	0.521732
Н	-2.060023	-2.393430	0.806109
С	-3.404916	1.063829	-0.374189
Н	-1.352562	1.531981	-0.824270
С	-4.289756	0.087509	0.106669
Н	-4.495347	-1.904447	0.900817
Н	-3.759051	2.031805	-0.706980
0	-5.643248	0.258362	0.212711
С	-6.207306	1.522772	-0.172345
Н	-7.278172	1.434095	0.015810
Н	-5.790895	2.335797	0.432962
Н	-6.032771	1.718929	-1.236389

nitrile imine-26

b3lyp-d3 SCF energy in solution: -1944.67711127 a.u. b3lyp-d3 enthalpy in solution: -1944.015954 a.u. b3lyp-d3 free energy in solution: -1944.129252 a.u. wb97xd SCF energy in solution: -1944.27325314 a.u. wb97xd enthalpy in solution: -1943.612096 a.u. wb97xd free energy in solution: -1943.725394 a.u.

Cartesiar	n coordinate	es	
ATOM	Х	Y	Z
С	1.721800	0.425994	0.543684
С	3.027702	0.003082	0.201507
С	1.530970	1.239373	1.688014
С	3.343883	-0.800956	-0.998925
С	4.115859	0.412168	0.986740
С	0.205962	1.671499	2.186960
С	2.639341	1.609456	2.463516
Ν	2.911668	-2.111655	-1.290106
С	4.106691	-0.385557	-2.063884
Н	5.112621	0.082165	0.710281
С	3.926898	1.207627	2.114378
N -	-0.733655	2.489127	1.527070
С –	-0.358113	1.305516	3.386086
Н	2.479181	2.233609	3.337351
С	2.163418	-3.020223	-0.518505
С	3.391909	-2.469882	-2.552634
Н	4.561488	0.593590	-2.142983
С	4.129270	-1.429251	-3.045255
Н	4.777357	1.510265	2.718848
С –	-0.651584	3.150339	0.288722
С –	-1.880261	2.586132	2.318443
С –	-1.670977	1.874862	3.467287
Н	0.114883	0.653196	4.108913
0	1.889313	-2.534612	0.677164
0	1.847553	-4.117701	-0.960856
Н	3.177869	-3.447508	-2.955683
Н	4.636213	-1.409387	-4.001360
0	0.562675	3.071780	-0.220562
0 -	-1.621231	3.723702	-0.191209
н –	-2.716848	3.182157	1.988161
н –	-2.374899	1.767925	4.282531
С	1.096772	-3.295531	1.698196
С	0.949889	3.729377	-1.510934
С	0.953088	-2.264972	2.815114
С	1.919160	-4.499300	2.146982
С –	-0.268940	-3.678866	1.131705
С	2.371807	3.210460	-1.709570

С	0.924021	5.239670	-1.300079
С	0.037391	3.267344	-2.645140
Н	1.935142	-1.919565	3.154902
Н	0.430543	-2.721713	3.661860
Н	0.375688	-1.402406	2.473237
Н	2.053525	-5.220686	1.337115
Н	1.396766	-4.998877	2.970379
Н	2.902838	-4.178485	2.506963
Н	-0.759988	-2.807222	0.689216
Н	-0.198489	-4.468527	0.381314
Н	3.002449	3.465319	-0.851265
Н	2.801334	3.667918	-2.606734
Н	2.372236	2.124219	-1.836515
Н	-0.090082	5.599475	-1.106438
Н	1.297757	5.731090	-2.205165
Н	1.569315	5.521689	-0.461045
Н	-0.051991	2.177630	-2.643093
Н	0.489622	3.572484	-3.595284
Н	-0.959109	3.707781	-2.579851
Н	-0.893788	-4.040172	1.955943
С	0.570865	0.023996	-0.281396
N	-0.661452	-0.165405	0.200238
N	-1.356190	-0.476891	-0.888918
N	0.604792	-0.187799	-1.628740
N	-0.616120	-0.497348	-1.997569
С	-2.758070	-0.735409	-0.868602
С	-3.562652	-0.033576	0.028225
С	-3.303384	-1.679922	-1.745315
С	-4.936725	-0.275521	0.059614
Н	-3.124095	0.704679	0.691767
С	-4.672481	-1.913195	-1.721063
Н	-2.663585	-2.230503	-2.426804
С	-5.494653	-1.215650	-0.819451
Н	-5.552188	0.278025	0.758292
Н	-5.119520	-2.643644	-2.388298
0	-6.827449	-1.521779	-0.873249
С	-7.724770	-0.827092	0.007888
Н	-8.716702	-1.222101	-0.215856
Н	-7.476425	-1.027137	1.056427
Н	-7.703816	0.251450	-0.185108

b3lyp-d3 SCF energy in solution: -998.06351660 a.u. b3lyp-d3 enthalpy in solution: -997.663709 a.u. b3lyp-d3 free energy in solution: -997.739871 a.u. wb97xd SCF energy in solution: -997.89369749 a.u. wb97xd enthalpy in solution: -997.493890 a.u. wb97xd free energy in solution: -997.570052 a.u. Imaginary frequency: -353.7511 cm-1

ATOM	Х	Y	Z
0	5.044498	-1.476987	-1.103621
С	1.431687	-1.006955	0.967700
С	1.475891	-1.493210	-0.354088
Н	0.555389	-1.757412	-0.866645
С	2.693049	-1.638153	-1.010033
Н	2.728382	-2.014860	-2.028638
С	2.636739	-0.689329	1.609696
Η	2.612729	-0.315351	2.629716
С	3.893097	-1.299508	-0.365826
С	3.862266	-0.823469	0.951929
Н	4.773586	-0.557346	1.474932
С	6.298633	-1.175145	-0.477660
Н	6.352951	-0.117226	-0.194717
Н	7.062198	-1.393330	-1.226185
Н	6.458083	-1.805645	0.405152
С	-1.906373	-0.509304	0.714490
Ν	-0.847060	-0.974051	1.065296
Ν	0.242509	-0.800847	1.700083
С	0.089124	1.750374	-0.716856
С	-0.065073	3.199508	-1.266763
С	-0.234593	3.721832	0.192035
С	-0.421793	2.258901	0.652790
Н	-0.495525	0.964640	-1.207447
Н	1.139181	1.432945	-0.681393
Н	-1.009998	3.299677	-1.815868
Н	-1.068056	4.409009	0.386819
Н	0.690208	4.171374	0.581156
С	1.080531	3.765519	-2.094710
Н	0.925268	4.829673	-2.318080
Н	2.032006	3.671185	-1.554077
Н	1.183087	3.233117	-3.049901
С	-0.321949	1.688792	2.024596
Н	0.272209	1.669808	2.926990
С	-1.541241	1.716996	1.494566
Н	-2.590491	1.864839	1.715228
С	-3.163619	-0.920377	0.142512
С	-4.362931	-0.281670	0.508116
С	-3.184319	-1.941547	-0.832598
С	-5.567539	-0.676911	-0.076345
С	-4.391787	-2.315886	-1.419115

Н	-6.491286	-0.187285	0.219671
С	-5.586871	-1.689177	-1.041938
Н	-4.400896	-3.102046	-2.169432
Н	-6.525703	-1.987234	-1.500602
Н	-2.255745	-2.425719	-1.120846
Н	-4.349278	0.509673	1.250497

b3lyp-d3 SCF energy in solution: -2107.78624098 a.u. b3lyp-d3 enthalpy in solution: -2106.982750 a.u. b3lyp-d3 free energy in solution: -2107.108579 a.u. wb97xd SCF energy in solution: -2107.48401284 a.u. wb97xd enthalpy in solution: -2106.680522 a.u. wb97xd free energy in solution: -2106.806351 a.u. Imaginary frequency: -309.9561 cm-1

ATOM	Х	Y	Z
0	6.059802	-2.474338	-0.701904
С	1.993409	-2.182673	0.242947
С	2.622288	-1.275703	-0.629841
H	2.052652	-0.465777	-1.067349
С	3.972379	-1.403717	-0.925550
Н	4.453667	-0.706693	-1.606207
С	2.752267	-3.216235	0.807570
Н	2.274400	-3.920202	1.483570
С	4.729655	-2.434918	-0.347350
С	4.115307	-3.343760	0.525666
Н	4.677261	-4.149230	0.984265
С	6.868434	-3.535014	-0.175811
Н	6.917716	-3.487459	0.918465
Н	7.864707	-3.381185	-0.593949
Н	6.484375	-4.512957	-0.489542
С	-0.777886	-0.232965	0.240764
N	-0.059110	-1.193207	0.061601
Ν	0.633028	-2.103523	0.615266
С	2.077514	0.814963	1.987887
С	2.345785	1.825608	3.142220
С	1.606981	0.872485	4.127432
С	1.039660	0.122168	2.902331
Н	1.716608	1.214212	1.034790
Н	2.955158	0.185443	1.794887
Н	1.752862	2.735776	2.993377
Н	0.877563	1.321020	4.814231
Н	2.307244	0.255134	4.708272

С	3.792194	2.192194	3.441707
Н	3.871608	2.803227	4.350951
Н	4.398429	1.288278	3.589620
Н	4.236665	2.762023	2.614564
С	0.365772	-1.204830	2.846488
Н	0.402106	-2.209719	3.242476
С	-0.409568	-0.139932	2.631932
Н	-1.377140	0.285822	2.846703
С	-1.687275	0.592536	-0.520291
C	-2,928890	0.995217	0.036052
C	-1.297011	1.083374	-1.792585
C	-3.417247	0.482892	1.324856
C	-3.709700	1,939791	-0.641849
C	-0 076436	0 613196	-2 487486
C	-2.107478	2.021798	-2,442391
N	-3 457869	-0 874011	1 701203
C	-3 803828	1 213665	2 427596
н	-4 658431	2 241440	-0 207798
C	-3 294719	2.241440	-1 863124
N	1 101002	1 375527	-2 627280
C	0 078997	-0 504576	-3 271211
с ц	-1 796469	2 398956	-3 /12/70
C	-3 336409	-2 03/271	0 901390
C	-3 844023	-0.961166	3 037884
U U	-3.824335	2 205232	2 466630
п С	-3.024333	2.295252	2.400030
U U	-2.002010	2 204747	-2 275120
п	-3.908910	J.ZU4/4/ 2 562006	-2.375120
C	1.402/13	2.303000	-1.903923
C	1 260200	0./11041	-3.490303
	1.300200	-0.446106	-3.090022
п	-0.6/044/	-1.2/409/	-3.397733
0	-3.543650	-1./536//	-0.370608
0	-3.106485	-3.126216	1.399309
H T	-3.95/466	-1.926468	3.50/610
H O	-4.388123	0.566350	4.504366
0	0.689123	2.770920	-0.91/546
0	2.394/9/	3.253686	-2.354223
H	2.952286	1.120646	-3.6908/3
H	1./99650	-1.186495	-4.55/84/
C	-3.492685	-2.783954	-1.461400
С	0.649900	4.069407	-0.172559
С	-3.744468	-1.928942	-2.700229
C	-4.617836	-3./88922	-1.234858
C	-2.113591	-3.433129	-1.509293
С	-0.379136	3.769818	0.915164
С	0.155348	5.151632	-1.127043
С	2.022881	4.379850	0.419214

Н	-4.731876	-1.457524	-2.651052
Н	-3.703256	-2.560901	-3.593287
Н	-2.985326	-1.145471	-2.788822
Н	-4.435336	-4.408292	-0.353475
Н	-4.684355	-4.443867	-2.110547
Н	-5.575974	-3.271329	-1.117142
Н	-1.333123	-2.677760	-1.626779
Н	-1.908944	-4.023924	-0.614214
Н	-1.347940	3.518614	0.471325
Н	-0.502029	4.653888	1.549050
Н	-0.054181	2.932768	1.540599
Н	0.862086	5.318805	-1.944728
Н	0.045532	6.089515	-0.571643
Н	-0.819011	4.877402	-1.545283
Н	2.437560	3.498713	0.914851
Н	1.910980	5.173080	1.166562
Н	2.726621	4.720388	-0.342943
Н	-2.078853	-4.099964	-2.377918

b3lyp-d3 SCF energy in solution: -1163.63005119 a.u. b3lyp-d3 enthalpy in solution: -1163.344957 a.u. b3lyp-d3 free energy in solution: -1163.415201 a.u. wb97xd SCF energy in solution: -1163.49894422 a.u. wb97xd enthalpy in solution: -1163.213850 a.u. wb97xd free energy in solution: -1163.284094 a.u. Imaginary frequency: -50.0117 cm-1

ATOM	Х	Y	Z
0	5.743244	-1.037619	-0.283678
С	1.942065	0.692025	0.217317
С	-2.425819	-0.777862	-0.105612
С	-2.213059	-1.905667	0.717589
С	-3.615001	-0.671118	-0.848955
С	2.111466	-0.660767	-0.147835
H	1.243191	-1.293866	-0.310257
С	3.386285	-1.199425	-0.303818
H	3.510749	-2.241287	-0.587503
С	3.091649	1.470902	0.426319
Н	2.980262	2.514457	0.709685
С	4.524864	-0.407930	-0.099924
С	-4.577888	-1.676132	-0.769020
Н	-5.496392	-1.585251	-1.342454
С	4.373363	0.934826	0.269977

H	5.234318	1.572524	0.436764
С	-3.180421	-2.908211	0.783347
Н	-3.012371	-3.774996	1.416910
С	-4.364304	-2.796529	0.043545
Н	-5.117015	-3.578275	0.101338
С	6.929174	-0.239714	-0.189913
Н	6.913820	0.573990	-0.925438
Н	7.758838	-0.914915	-0.407598
Н	7.049817	0.173987	0.818729
С	-1.391161	0.211304	-0.215938
Ν	-0.317435	0.598080	0.101887
Ν	0.692774	1.325913	0.389303
Н	-1.293448	-1.986192	1.289889
Н	-3.770929	0.210393	-1.461289
S	-3.098776	2.852177	-0.298509
С	-4.617217	2.174977	0.518745
Н	-5.107579	2.934465	1.141723
Н	-5.354902	1.826267	-0.215458
Н	-4.375257	1.322781	1.167246

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b3lyp-d3 SCF energy in solution: -1163.62995247 a.u. b3lyp-d3 enthalpy in solution: -1163.344867 a.u. b3lyp-d3 free energy in solution: -1163.414341 a.u. wb97xd SCF energy in solution: -1163.49781786 a.u. wb97xd enthalpy in solution: -1163.212732 a.u. wb97xd free energy in solution: -1163.282206 a.u. Imaginary frequency: -97.6315 cm-1

ATOM	Х	Y	Z
0	-5.415362	-1.171807	0.628080
С	-1.704067	0.489711	-0.443961
С	2.683200	-0.762734	-0.042681
С	2.309863	-2.085324	-0.374176
С	4.014430	-0.487569	0.315624
С	-1.806403	-0.675084	0.346978
Н	-0.907315	-1.176354	0.695497
С	-3.053203	-1.196570	0.683018
Н	-3.127018	-2.094389	1.291211
С	-2.890410	1.102525	-0.879587
Н	-2.829616	2.001469	-1.487842
С	-4.229065	-0.574507	0.240514
С	4.959369	-1.514319	0.329433
Н	5.988010	-1.291325	0.600498

С	-4.143970	0.582192	-0.545363
Н	-5.034951	1.086934	-0.901344
С	3.260964	-3.104542	-0.345364
Н	2.966481	-4.118946	-0.601493
С	4.588631	-2.823918	0.002237
Н	5.327386	-3.620850	0.017352
С	-6.639451	-0.599702	0.152552
Н	-6.765911	0.425925	0.520371
Н	-7.435811	-1.230149	0.552349
Н	-6.679590	-0.608243	-0.943642
С	1.667168	0.260863	-0.039938
N	0.560099	0.504996	-0.401332
N	-0.489132	1.094127	-0.828019
Н	1.280640	-2.298945	-0.648160
Н	4.282254	0.533958	0.565453
S	3.054121	2.923245	-0.031331
С	1.530739	3.527601	0.829158
Н	1.019470	2.705253	1.344679
Н	1.773685	4.289885	1.580870
Н	0.815702	3.972721	0.125658

b3lyp-d3 SCF energy in solution: -2273.34900771 a.u. b3lyp-d3 enthalpy in solution: -2272.660544 a.u. b3lyp-d3 free energy in solution: -2272.779766 a.u. wb97xd SCF energy in solution: -2273.08420149 a.u. wb97xd enthalpy in solution: -2272.395738 a.u. wb97xd free energy in solution: -2272.514960 a.u. Imaginary frequency: -127.9307 cm-1

ATOM	Х	Y	Z
0	-6.443980	-2.433525	-0.080547
С	-2.971459	-0.399108	1.152670
С	-3.008604	-1.190246	-0.015361
Н	-2.120069	-1.289793	-0.628483
С	-4.175254	-1.847063	-0.394720
Н	-4.192083	-2.454487	-1.296376
С	-4.143008	-0.305045	1.923857
Н	-4.134729	0.295400	2.830195
С	-5.337566	-1.740799	0.381807
С	-5.316876	-0.965484	1.548932
Н	-6.198134	-0.864633	2.172624
С	-7.629912	-2.412474	0.722120
Н	-7.439254	-2.831337	1.717844

Н	-8.355572	-3.035610	0.196048
Н	-8.025600	-1.394017	0.818842
С	0.314636	0.305565	0.495564
N	-0.825003	0.270970	0.827508
N	-1.835002	0.279169	1.620108
С	1.324927	0.653121	-0.458263
С	2.091883	1.838271	-0.308710
С	1.539094	-0.192315	-1.573814
С	1.847190	2.795203	0.781505
С	3.105442	2.109051	-1.235105
С	0.671756	-1.366167	-1.820650
С	2.532386	0.135455	-2.502567
N	0.593933	3.372994	1.074823
С	2.722744	3.227716	1.748615
H	3.691650	3.015756	-1.115992
С	3.332387	1.264101	-2.323285
N	0.771827	-2.615540	-1.173886
С	-0.401144	-1.442207	-2,675010
н	2 684366	-0 516384	-3 357850
C	-0.568179	3 437831	0 277939
C	0.713509	4 141877	2 234453
н	3 754023	2 908575	1 826684
C	2 012315	4 072756	2 661075
ч	4 116091	1 496809	-3 038467
C	1 807247	-3 118319	-0 359718
C	-0 25/722	-3 113112	-1 635575
C	-0 982665	-2 7/8807	-2 561108
U U	-0.902000	-2.740007	-2.301100
	-0.752770	-0.024043	-0.000550
0	-0.298703	3 767249	-0.980559
U	-1.047900	1 604752	2 6069914
п	-0.133012	4.094732	2.000004
п	2.41002/	4.0000009	0.201542
0	2.01/904	-2.273497	-0.291343
0	1.724419	-4.221002	0.164121
H	-0.344045	-4.453/83	-1.20/355
н С	-1.850003	-3.119441	-3.091904
	-1.346973	3.044307	-2.04/456
C	4.150727	-2.641900	0.292903
C	-0.506838	2.679096	-3.268495
C	-2.020434	4.402836	-2.222357
C	-2.326584	1.928635	-1.698//1
С	4.960532	-1.374005	0.034939
C	4.715417	-3.827305	-0.485288
С	4.007492	-2.912363	1.786333
H	0.208906	3.475667	-3.498715
H	-1.164456	2.539480	-4.132575
Н	0.045741	1.750841	-3.093765

Н	-2.675765	4.645384	-1.383164
Н	-2.623918	4.378614	-3.136510
Н	-1.268227	5.192189	-2.328844
Н	-1.799389	0.980339	-1.574360
Н	-2.888498	2.146184	-0.787691
Н	5.039428	-1.173871	-1.038493
Н	5.968582	-1.499359	0.443544
Н	4.491414	-0.511497	0.519037
Н	4.159081	-4.746935	-0.288866
Н	5.755513	-3.983358	-0.178664
Н	4.700559	-3.622033	-1.561350
Н	3.550188	-2.050131	2.282840
Н	5.007478	-3.070888	2.206345
Н	3.405969	-3.803033	1.979425
S	1.709603	-0.023087	3.005915
С	0.876829	-1.673119	3.076620
Н	1.523077	-2.438050	3.526415
Н	-0.049427	-1.635984	3.664780
Н	0.611085	-2.023917	2.071064
Н	-3.038181	1.819204	-2.524737