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Supplemental Information

Fc Effector Function Contributes

to the Activity of Human Anti-CTLA-4 Antibodies

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Biopsy/ **Mutational** Identifier Age Sex Subtype **Current therapy Previous therapy** Stage resection site status Melanoma **BRAF WT** Adjuvant MM1 53 Μ Cutaneous IIIc LN Nil ipilimumab/nivolumab NRAS mutant MM2 Μ Cutaneous IV – M1c Small bowel BRAF mutant Pembrolizumab Ipilimumab 52 **BRAF WT** MM3 76 F Cutaneous IV – M1c Small bowel Nil Nil **NRAS** mutant MM4 76 Μ Cutaneous IIIc LN **BRAF** mutant Nil Nil MM5 74 F Cutaneous IV – M1a LN **BRAF WT** Nil Nil MM6 61 Μ Cutaneous IV – M1a LN **BRAF WT** Nil Ipilimumab Nil MM7 42 Μ Cutaneous IIIc LN **BRAF** mutant Nil Paclitaxel + Trametinib **BRAF WT** MM8 49 Cutaneous LN Nil Ipilimumab Μ IV – M1c NRAS mutant Pembrolizumab NSCLC NSCLC1 Nil Nil 74 М Adenocarcinoma lla **Right lower lobe** _ (CRUK0394) NSCLC2 76 Illa **Right upper lobe** Nil Nil Μ Squamous (CRUK0400) NSCLC3 F Squamous lla Left upper lobe Nil Nil 53 (CRUK0381) NSCLC4 90 Μ Squamous lla Left lower lobe Nil Nil (CRUK0403) NSCLC5 60 Adenocarcinoma Left upper lobe Nil Nil Μ lla (CRUK0178) NSCLC6 IIb Left lower Lobe Nil Nil 57 Μ Adenocarcinoma (CRUK0009) NSCLC7 54 F Adenocarcinoma lla **Right upper lobe** Nil Nil (CRUK0230) NSCLC8 69 Μ Adenocarcinoma Ia Left lower lobe Nil Nil _

Table S1, related to Figure 1. Demographics and clinical characteristics of patients with advanced malignant melanoma (MM), non-small cell lung cancer (NSCLC) and renal cell carcinoma (RCC).

Identifier	Age	Sex	Subtype	Stage	Biopsy/ resection site	Mutational status	Current therapy	Previous therapy
RCC								
RCC1	44	М	Clear cell	111	-	-	Nil	Nil
RCC2	69	М	Clear cell	lla	-	-	Nil	Nil
RCC3	78	F	Mixed - clear cell/ tubulo-papillary	la	-	-	Nil	Nil
RCC4	65	F	Clear cell	IV	-	-	Nil	Nil
RCC5	50	F	Clear cell	IV	-	-	Nil	Nil
RCC6	60	F	Clear cell	IV	-	-	Nil	Nil
RCC7	19	F	Clear cell	IV	-	-	Nil	Nil
RCC8	59	М	Clear cell	IV	-	-	Nil	Nil

M, male; F, female; LN, lymph node; WT, wild type.



Figure S1, related to Figure 1. Expression profile of B7 and TNFR superfamily checkpoints on T cell subsets in murine and human tumors. (A) MFI of co-stimulatory and co-inhibitory molecules expressed on TIL subpopulations in murine tumors. (B) MFI of 4-1BB, PD-1 and TIM-3 expressed on TIL subpopulations in human tumors. Horizontal bars represent the mean and error bars show ± SEM; ns, no statistical significance.





Figure S2, related to Figure 2. Expression profile of hFcyRs in hFcyR mice and human tumors. (A) Quantification of the absolute number of tumor-infiltrating leukocyte subpopulations in B16, MC38 and MCA205 tumors in hFcyR mice. (B) Percentage of expression of individual FcyRs in tumor-infiltrating leukocyte subpopulations in each tumor model. (C) Percentage of expression of individual FcyRs in total CD45⁺ cells from different organs and tissues in tumor-bearing mice or samples from patients with melanoma (see Table S2). Error bars show \pm SEM.

Α

Table S2, related to Figure 2. Demographics and clinical characteristics of patients with advanced malignantmelanoma (MM) employed fresh for myeloid analyses.

Identifier	Age	Sex	Subtype	Stage	Biopsy site	Mutational status	Current therapy	Previous therapy
MM9	66	Μ	Cutaneous	IV M1c	LN	BRAF WT	Pembrolizumab	Ipilimumab
MM10	60	М	Ocular	IV M1c	Splenic metastasis	BRAF WT	Nil	Nil
MM11	76	М	Cutaneous	IV M1c	SC	BRAF mutant	Nil	Nil
MM12	86	F	Cutaneous	IV M1c	SC LN	BRAF WT	Nil	Nil
MM13	46	F	Cutaneous	IV M1c	SC	BRAF mutant	Nil	Dabrafenib/Trametinib
MM14	50	Μ	Cutaneous	IV M1c	Small bowel	BRAF mutant	Pembrolizumab	Ipilimumab
MM15	69	Μ	Cutaneous	IV M1c	SC	BRAF WT	Pembrolizumab	Ipilimumab
MM16	76	F	Cutaneous	IV M1c	Small bowel	BRAF WT	Nil	Nil
MM17	53	М	Cutaneous	IIIc	LN	BRAF WT	Nil	Adjuvant ipilimumab/ nivolumab

M, male; F, female; LN, lymph node; SC, subcutaneous; WT, wild type.



Figure S3, related to Figure 3. Expression pattern of Fc γ Rs in monocyte-derived human macrophages. Expression of human Fc γ Rs quantified by flow cytometry in CD14⁺ bead-sorted monocytes from healthy donor PBMCs (upper panel) and matched macrophages (M Φ) seven days after in vitro differentiation with human recombinant M-CSF (lower panel).



Figure S4, related to Figure 4. In vivo intra-tumoral Treg cell depletion by anti-CTLA-4-hlgG2 depends on CD32a. Percentage of CD4⁺FoxP3⁺ T cells of total CD4⁺ T cells in TIL and LN in hFcγR mice bearing MCA205 tumors and treated with different IgG variants of anti-CTLA-4 mAbs as described in Figure 4. In the *FCGR2A^{-/-}* group, mice expressed all human FcγRs except CD32a. IgG2_{Endos}, endoglycosidase-treated IgG2 mAb. Horizontal bars show the mean.



Frameshift Indel Load

A

Figure S5, related to Figure 5. Human FcγR polymorphisms impact upon response to ipilimumab in patients with advanced melanoma. (A) Anti-CTLA-4 response rate in the van Allen et al. and Snyder et al. patient cohorts based on indel mutational load and nsSNV neoantigen load combined with the presence (SNP⁺) or absence (SNP⁻) of the germline CD32a-H131R high affinity SNP. (B) Survival analysis of the Snyder et al. patient cohort based on nsSNV neoantigen burden and germline CD32a-H131R SNP. (C) Response rate of patients treated with anti-PD-1 from the Hugo et al. dataset based on indel mutational load with (SNP⁺) or without (SNP⁻) the CD16-V158F high affinity SNP.

0.75

1.00

0.50

0.00

0.25

nsSNV Neoantigen Load

1.00

1.00