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# **Supporting Information**

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In Situ Monitoring of MicroRNA Replacement Efficacy and Accurate Imaging-Guided Cancer Therapy through Light-Up Inter-Polyelectrolyte Nanocomplexes

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#### Supporting Information

In Situ Monitoring of MicroRNA Replacement and Accurate Imaging-Guided Cancer Therapy through Light-Up Inter-Polyelectrolyte Nanocomplexes

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Figure S1. High performance liquid chromatography (HPLC) analysis of purified S-Arg<sub>4</sub> peptides with reverse phase C18 column.



Figure S2. Liquid chromatography-mass spectrometry (LC-MS) analysis of S-Arg<sub>4</sub> peptides prepared by solid phase peptide synthesis. The major peak (MW=865.7) is consistent with the estimated product molecular weight of 865.06.



Scheme S1. Synthesis process of carboxymethyl dextran (CMD).



Figure S3. FT-IR spectra of dextran and carboxymethy dextran (CMD).



Figure S4. Optimization and characterization of blank nanocomplexes (BNs) composed of various mass ratios of S-Arg<sub>4</sub> and CMD. (A) Hydrodynamic size, (B) polydispersity index (PDI) and (C) surface zeta potential determined by DLS analysis. (D) Production yield of the BNs.

S-Arg <sub>4</sub> :CMD:MiR-	1.1:1:	1.1:1:	1.1:1:	1.1:1:	1.1:1:
34a:ICG(mass rano)	0.01:0.01	0.025:0.025	0.05:0.05	0.075:0.075	0.1:0.1
Embedding efficiency	93.1	87.2	75.2	51.2	41.1
of miR-34a (EE) %					
Loading content of miR- 34a (LC) %	0.66	1.41	2.55	2.60	2.79
Embedding efficiency of ICG (EE) %	90.3	82.5	70.2	45.2	38.3
Loading content of miR- 34a (LC) %	0.63	1.40	2.45	2.30	2.61

**Table S1.** Effects of mass ratio of materials on embedding efficiencies (EE) and loading content (LC) of miR-34a and ICG



Figure S5. Average sizes and zeta potentials of MINs upon embedding with different amounts of miR-34a and ICG.



Figure S6. Size and zeta potential of uncross-linked MINs and cross-linked CMINs determined by DLS measurements.



Figure S7. Post-formulation colloidal stability of CMINs dispersed in different solvent environments at 0, 1, 3, 5 and 7 days based on the average hydrodynamic diameters.



Figure S8. Bovine serum albumin (BSA) adsorption assays on the CMINs after different time co-incubation at 37 °C.



Figure S9. TEM images of CMINs in PBS of pH 7.4 in the presence of 2  $\mu$ M GSH for 2 h (A) and 4 h (B).



Figure S10. Linear correlation between the ratios of ICG release and miR-34a release.



Figure S11. UV/Vis spectra of free ICG solution, CMINs and CMINs incubated with 10 mM GSH for 4 h.



Figure S12. Cell cytotoxicity. HepG-2 incubated with different concentrations of scramble miRNA-encapsulating CMINs (Scr-CMINs). Error bars represent standard deviations of three dependent experiments.



Figure S13. Normalized cell number at the endpoint of wound healing and transwell assays by CCK-8 assays.



Figure S14. The expression of Bcl-2 and Notch-1 was quantitatively detected by qRT-PCR upon the treatment of lipo/miR-34a and CMINs in the presence of different concentrations of serum.



Figure S15. Hemolysis assay with different concentrations of CMINs (negative control: PBS; positive control: water).



Figure S16. Fluorescence Images and relative fluorescence intensity of CMINs (100  $\mu$ L, 0.2  $\mu$ g of ICG) at 0.5 h, 2 h and 4 h in mice blood with or without 10 mM GSH.







Figure S18. QRT-PCR was used to measure the Bcl-2 and Notch-1 expression at transcriptional level in tumors upon treatment of CMINs with different doses of miR-34a.



Figure S19. Body weights measurements. No significant differences in body weights were found in two tumor mouse models upon treatment with saline, GSH-CMINs, CMINs (2 mg/kg of miR-34a) and CMINs (4 mg/kg of miR-34a).