# Supplementary information

for Guan et al.

Gene refashioning through innovative shifting of reading frames in mosses

Organism	Blast Name	Score	Number of Hits	Description
cellular organisms			1688	
. Eukaryota	eukaryotes		997	
Viridiplantae	green plants		10	
Embryophyta	land plants		8	
Marchantia polymorpha subsp. ruderalis	liverworts	246	1	Marchantia polymorpha subsp. ruderalis hit
Selaginella moellendorffi	club-mosses	230	4	Selaginella moellendorffii hits
Physcomitrella patens	mosses	196	1	Physcomitrella patens hits
Quercus suber	eudicots	107	2	Quercus suber hits
Monoraphidium neglectum	green algae	90.1	2	Monoraphidium neglectum hits
Sordaria macrospora k-hell	ascomycetes	157	2	Sordaria macrospora k-hell hits
Phellinus noxius	basidiomycetes	152	1	Phellinus noxius hits
Thielavia terrestris NRRL 8126	ascomycetes	150	2	Thielavia terrestris NRRL 8126 hits
Spizellomyces punctatus DAOM BR117	chytrids	148	6	Spizellomyces punctatus DAOM BR117 hit
Madurella mycetomatis	ascomycetes	147	2	Madurella mycetomatis hits
Hydnomerulius pinastri MD-312	basidiomycetes	146	2	Hydnomerulius pinastri MD-312 hits
<ul> <li>Neurospora tetrasperma FGSC 2508</li> </ul>	ascomycetes	146	2	Neurospora tetrasperma FGSC 2508 hits
Neurospora tetrasperma FGSC 2509	ascomycetes	146	1	Neurospora tetrasperma FGSC 2509 hits
Exophiala spinifera	ascomycetes	146	2	Exophiala spinifera hits
Neurospora crassa OR74A	ascomycetes	145	2	Neurospora crassa OR74A hits
Neurospora crassa	ascomycetes	145	1	Neurospora crassa hits
Rhizopogon vinicolor AM-OR11-026	basidiomycetes	144	1	Rhizopogon vinicolor AM-OR11-026 hits
<ul> <li>Saitoella complicata NRRL Y-17804</li> </ul>	ascomycetes	145	4	Saitoella complicata NRRL Y-17804 hits
Rhizopogon vesiculosus	basidiomycetes	144	1	Rhizopogon vesiculosus hits
Tulasnella calospora MUT 4182	basidiomycetes	144	2	Tulasnella calospora MUT 4182 hits
Ramazzottius varieornatus	tardigrades	144	3	Ramazzottius varieomatus hits
Chaetomium globosum CBS 148.51	ascomycetes	142	2	Chaetomium globosum CBS 148.51 hits
Pisolithus microcarpus 441	basidiomycetes	142	1	Pisolithus microcarpus 441 hits
Fomitiporia mediterranea MF3/22	basidiomycetes	142	2	Fomitiporia mediterranea MF3/22 hits
Kwoniella heveanensis BCC8398	basidiomycetes	141	1	Kwoniella heveanensis BCC8398 hits
Kwoniella heveanensis CBS 569	basidiomycetes	141	1	Kwoniella heveanensis CBS 569 hits
<ul> <li>Paxillus involutus ATCC 200175</li> </ul>	basidiomycetes	141	1	Paxillus involutus ATCC 200175 hits

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Organism	Blast Name	Score	Number of Hits	Description
cellular organisms			1727	
. Eukaryota	eukaryotes		718	
Viridiplantae	green plants		18	
Chlorophyceae	green algae		11	
Monoraphidium neglectum	green algae	544	6	Monoraphidium neglectum hits
Gonium pectorale	green algae	118	1	Gonium pectorale hits
<u>Tetrabaena socialis</u>	green algae	114	1	Tetrabaena socialis hits
Chlamydomonas reinhardtii	green algae	110	1	Chlamydomonas reinhardtil hits
Volvox carteri f. nagariensis	green algae	107	2	Volvox carteri f. nagariensis hits
Selaginella moellendorffi	club-mosses	93.1	4	Selaginella moellendorffii hits
Quercus suber	eudicots	91.9	2	Quercus suber hits
Physcomitrella patens	mosses	83.4	1	Physcomitrella patens hits
Colletotrichum chlorophyti	ascomycetes	120	1	Colletotrichum chlorophyti hits
Coniochaeta ligniaria NRRL 30616	ascomycetes	120	1	Coniochaeta ligniaria NRRL 30616 hits
Colletotrichum orbiculare MAFF 240422	ascomycetes	117	1	Colletotrichum orbiculare MAFF 240422 hits
. Allomyces macrogynus ATCC 38327	blastocladiomycetes	116	2	Allomyces macrogynus ATCC 38327 hits
Trichoderma reesei QM6a	ascomycetes	115	4	Trichoderma reesei QM6a hits
Trichoderma reesei RUT C-30	ascomycetes	115	2	Trichoderma reesei RUT C-30 hits
Trichoderma parareesei	ascomycetes	114	2	Trichoderma parareesei hits
Coniosporium apollinis CBS 100218	ascomycetes	111	2	Coniosporium apollinis CBS 100218 hits
Podospora anserina S mat+	ascomycetes	110	9	Podospora anserina S mat+ hits
Hypoxylon sp. EC38	ascomycetes	110	3	Hypoxylon sp. EC38 hits
Pyronema omphalodes CBS 100304	ascomycetes	110	1	Pyronema omphalodes CBS 100304 hits
Fusarium pseudograminearum CS3096	ascomycetes	109	2	Fusarium pseudograminearum CS3096 hits
. Fusarium pseudograminearum CS3220	ascomycetes	109	1	Fusarium pseudograminearum CS3220 hits
. Fusarium pseudograminearum CS3427	ascomycetes	109	1	Fusarium pseudograminearum CS3427 hits
. Fusarium pseudograminearum CS3487	ascomycetes	109	1	Fusarium pseudograminearum CS3487 hits
. Elaphomyces granulatus	ascomycetes	113	1	Elaphomyces granulatus hits
Spizellomyces punctatus DAOM BR117	chytrids	108	6	Spizellomyces punctatus DAOM BR117 hits
. Fusarium oxysporum f. sp. melonis 26406	ascomycetes	107	7	Fusarium oxysporum f. sp. melonis 26406 hits
Schizosaccharomyces pombe	ascomycetes	107	2	Schizosaccharomyces pombe hits
Schizosaccharomyces pombe 972h-	ascomycetes	107	1	Schizosaccharomyces pombe 972h- hits

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**Supplementary Figure 1. (a)** Partial taxonomy distribution generated from BLAST search of the NCBI non-redundant (*nr*) protein sequence database using hemerythrin from *Marchantia polymorpha* (JGI ID: Mapoly0042s0056.1). Hits from only one green alga *Monoraphidium neglectum* were found. (b) Partial taxonomy distribution generated from BLAST search of nr database using the *Monoraphidium* sequence identified above, indicating all green algal hits were from Chlorophyceae. In both searches, E-value = 1e-8 was used as cutoff; land plant and algal hemerythrin sequences were more similar to fungal homologs than to each other.

hypothetical protein M407DRAFT\_241621 [Tulasnella calospora MUT 4182] Sequence ID: <u>KIO31868.1</u> Length: 224 Number of Matches: 1

						- aprille									
Score		E	xpect	Met	thod					Ide	ntities	Po	sitives		Gaps
95.4 b	its(310	) 2	e-22	Con	nposit	ional	mat	rix a	djust.	55/1	20(46%)	) 75,	/120(62	%)	7/120(5%
Query	2	IIL	EVKHDI	HAEL	EECFQ	RYKK	AHSK		EARNL	FNQF	WEISRHS	VSEE	LILYPM	MDL-	- 60
Sbjct	41	ISE	EITQDI	IREL	EEYYG	NYRT	ATS-	EF	EKTQW	ANQF	CWELARHS	VGEE	LVLYPA	FEKI	I 97
Query	61	LGD LG	RGKEL	ADQS	REDHH R +H	RTKE K+	ILAE +L E	LQT-	-ISDP ++DP	SLFE	KRLNIMMA L +M	ELRI	HMKMEE	EEDI E DI	118
Sbict	98	LGP	EGKQI	ADTD	RAEHL	EAKK	LLYE	LEGT	HVNDP	N-FP	TILKKLME	ELRE	HMKSEE	ENDI	156

b

с

putative hemerythrin-like protein [Monoraphidium neglectum] Sequence ID: <u>XP 013901669.1</u> Length: 253 Number of Matches: 1 See 1 more title(s)

Score		Ex	pect	Meth	bd		Identities	Positives	Gaps
45.6 b	its(1	40) 0.0	02	Comp	ositional mat	trix adjust.	38/123(31%)	68/123(55%)	7/123(5%)
Query	1	DIIL	EVKHD	HAELEI	ECFQRYKKAHS	SKGQDHEAR-	NLFNQFVWEISR	HSVSEELILYPMM	D 59
Sbjct	75	NIVD.	AITDD	HKRL-	FSYCDQLRG	GOLGHDQRE	HLRHQLVHDLAVI	HSLSEEEVLYPA-	- 129
Query	60	LLGD	G E+	ADQSRI	EDHHRTKEILA	AELQTI-SDF	SLFEKRLNIMMA	ELRDHMKMEEEED	L 118
Sbjct	130	LKNA	CGNEM	RNHVL	DEHTTLKHILM	IDIDKMDADE	PNFNPRLDELVH	ELRHHVQEEEEKV	L 189
Query	119	AYL	121						
Sbjct	190	PMF	192						
·Consei	nsust	romCor	tig3	8372-a	abinit-gene	2-0.1			
Conser Score	= 44	fromCor Length	tig38 = 182	8372-a 2 03), I	abinit-gene Expect = 86	e-0.1	ethod: Compos	itional matr:	ix adjust.
Conser Score Ident:	nsusi I = 44 ities	fromCor Length 1.3 bit 3 = 34/	tig33 = 182 s (10 115	8372-a 2 03), I (29%),	abinit-gene Expect = 8e . Positives	e-0.1 e-06, Me s = 64/115	ethod: Compos 5 (55%), Gaps	itional matr: = 7/115 (6%)	ix adjust.
Conser Score Ident: Query:	nsusi I = 44 ities 6	FromCor Length 1.3 bit 5 = 34/ VKHDH# ++ DH+	tig33 = 182 s (10 115 ELEE( ++E	8372-a 2 03), F (29%), CFQRYF + R+F	abinit-gene Expect = 8e Positives (KAHSKGQDHE C SK + 4	2-0.1 2-06, Me 5 = 64/115 EARNLFNQF' + R NQF4	ethod: Compos 5 (55%), Gaps WEISRHSVSEEL + ++S+HSV+EE+	itional matr: = 7/115 (6%) ILYPMMDLLGDRG + YP ++ (	ix adjust. ) 3 65 3
Conser Score Ident: Query:	nsusi I = 44 ities 6 43	FromCor Length 4.3 bit 5 = 34/ VKHDHA ++ DH4 LQKDHS	tig3 = 18 :s (10 115 ELEE( ++E FIKE)	8372-a 2 03), F (29%), CFQRYF + R+F DYNRWF	Abinit-gene Expect = 86 Positives KKAHSKGQDHE C SK + 4 ASKNAEEP	e-0.1 s = 64/115 EARNLFNQFV + R NQF+ KIR-WANQFJ	ethod: Compos 5 (55%), Gaps /WEISRHSVSEEL + ++S+HSV+EE+ LLDVSQHSVAEEI	itional matr: = 7/115 (6%) ILYPMMDLLGDRG + YP ++ ( VYYPEIEKRQKGG	ix adjust. ) 3 65 3 3 99
Conser Score Ident: Query: Sbjct: Query:	nsusi I = 44 ities 6 43 66	FromCor Length 4.3 bit 5 = 34/ VKHDHA ++ DH4 LQKDHS KELADQ +	etig30 = 182 ss (10 115 ELEE0 ++E FIKED SFIKED SREDD ++)	8372-a 2 03), F (29%), CFQRYF + R+H DYNRWF HHRTKE H K	Abinit-gene Expect = 86 Positives (KAHSKGQDHE (SK + 4 KA-SKNAEEF SILAELQTI +L L +	e-0.1 e-06, Me s = 64/115 EARNLFNQF4 + R NQF4 KIR-WANQF3 -SDPSLFEKH +DP ++	ethod: Compos 5 (55%), Gaps WEISRHSVSEEL ++S+HSV+EE+ LLDVSQHSVAEEI RLNIMMAELRDHM L + + L+ H+	itional matr: = 7/115 (6%) ILYPMMDLLGDRG + YP ++ ( VYYPEIEKRQKGC KMEEEEDL 118 K EEE D	ix adjust. ) 3 65 3 3 99

**Supplementary Figure 2.** Land plant hemerythrins are more similar to fungal homologs than to those from chlorophyte green algae and glaucophytes. Figures show pairwise comparisons of the conserved HHE domain between liverwort *Marchantia polymorpha* (Mapoly0042s0093.1) and homologs from fungi, chlorophytes, and glaucophytes, respectively. Fungal and green algal sequences here are top hits from BLASTP search of nr database. (a) Sequence alignment between *Marchantia polymorpha* and the fungus *Tulasnella calospora*. (b) Sequence alignment between *Marchantia polymorpha* and chlorophyte *Monoraphidium neglectum*. (c) Sequence alignment between *Marchantia polymorpha* and the glaucophyte *Cyanophora paradoxa*.

a

### >Partial sequence of the Isoete yunguiensis mosaic gene derived from fusion of oleosin and Hr

CCTCTCGCGATGCTGGGGACGAGCCTAACCATAGGAACCCTAATTCTGGTAATATCTTCGCCGCTGTTACTCCTTTTTAGCCCAATTCTAGTACC TCTAGGGTTTGTGCTTTTCATGGCCGCGGCAGCCTTCGCGGCTATGGTGGCAGCAGCAGCAGCGAGGCATGGATCTACCGCTACAAAAAAGG CCGCCACCCATGGGATCCGATAAACTTGATGCAGCAGCAATCCACATGTTCATACCCAGAAGAGATATCATCGACCTCGTTAGGGAGGAGAACATGC TAAACTGGAAGAGGATTACGGTAATTACAAGAGCGCCAGCCGTAGAGGAGAGCACTACGAGGCCAGAAAATGGTTCAACCAGTTTGTGTGGGG AAATTTCCAGACATTCTGTCTCCGAAGAGCTTGTTATGTATCCCCTCCTTGATGGGCTGGGCCCAAAAGGCCGAGATTTGGCATATCAATCCCG TGCAGATCATCACAAGATCAAGGAATTGCTCACGGAATTGCAGCAGCACAACACTGATTCTGAGGATTTTGATTCCAGAATGGAGACCATGATGCC CAATCTCAGAGACCATATTAAGTTGGAAGAAAAGCGCGATGGAGATCTGGCTTGTTTGCGTGATAATATGGATCAGCAGGCTCGCGAGGCTGC CGGAGCTACATTTGCACTGGGCAAGAATCT

#### >Translated peptide sequence of the Isoete yunguiensis mosaic gene

PLAMLGTSLTIGTLILVISSPLLLLFSPILVPLGFVLFMAAAAFAAMVAAGNAVAWIYRYKKGRHPMGSDK LDAAIHMFIPRRDIIDLVREDHAKLEEDYGNYKSASRRGDHYEARKWFNQFVWEISRHSVSEELVMYPLL DGLGPKGRDLAYQSRADHHKIKELLTELQHNTDSEDFDSRMETMMSNLRDHIKLEEKRDGDLACLRD NMDQQAREAAGATFALGKN



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**Supplementary Figure 3.** (a) Partial nucleotide sequence and translated peptide sequence of the *oleosin* and *Hr* mosaic gene in the lycophyte *Isoetes yunguiensis* (GenBank accession number MG254883). (b) CDD comparison with *I. yunguiensis* protein sequence, indicating fusion of oleosin and HHE domain. Although the same gene fusion was also found in the liverwort *Treubia lacunosa* (1KP ID: FITN-2089742), its nucleotide sequence shared 100% identity with PYHZ-2006808, pointing to a possibility of contamination.







**Supplementary Figure 4.** Molecular phylogeny of hemerythrin (Hr) protein sequences. LG and JTT substitution models were used in maximum and distance analyses, respectively, and fourgamma rate categories plus invariant sites were used in the analyses. Numbers above branches show bootstrap values from maximum likelihood and distance analyses, respectively. Asterisks indicate values lower than 50%. Nodes without values indicate that bootstrap support values from both analyses are lower than 50%. Taxonomic affiliations are after species names. (a) Phylogenetic analyses with a larger taxonomic sampling, including multiple angiosperm and seedless vascular plant sequences from 1KP (underlined in red). (b) Phylogenetic analyses with sequences of known scaffold information, mostly from genome sequence projects. The sequence of glaucophyte *Cyanophora paradoxa* was removed from the analyses because of its much shorter length. Inclusion of the *Cyanophora* sequence provided a similar topology, with much lower branch support values. >Chr21: 12755680..12757359

12757359	cgcatget cagcacctgt cgagcacacgcccgt accagt catcgt cct ccccaat ccacctt tcgcct cctget t caatcgt catcgt catc
12757279	$\begin{tabular}{lllllllllllllllllllllllllllllllllll$
12757199	C T L S D L Q P L Y H S L S V I L Y I T H K A L V V H
12757119	R       E       A       L       L       P       A       D       H       T       V       H       H       G       R       S       R       I       H       P       L       E       R       C       R         CCGTGAAGCCGAAGCACTTCTCCCAGCAGATCACACTGTCCACCATGGCCGCAGGCGCGCGC
12757039	F R E V G N H P S Q L Q Q C G L P A Q R G H C C V Q <u>GCTTCCGCGAGGTTGGCAACCACCCAAGCCAGCCAGCCAG</u>
12756959	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$
12756879	${\tt GATTGGATGGCATGTGAACAAGTGTAGTGCGATTGCTGAGCTCGGGAACCAAGTCCATGGCGAGCATGGGTGTGGTTCCT}$
12756799	GAGCTGAGCAGGAAGTGGGGACGTGAAATTTGGCCGCGGCCAGGGAGTGTGGGCTTGTTGCAATTGTGGGTGTTTGAGAGGGAGTGTGGGGTGTTTGAGAGGGAGTGTGGGGTGTTGGAATTGTGGGTGTTTGAGAGGGAGTGTGGGGTGTTGGGGTGTTTGGGGTGTTTGGAGAGGGGGG
12756719	$\begin{array}{c cccc} V & G & E & P & V & L & R \\ CGATGACAATGGATGTTTGGTGCTCTAACGTGAAAAACATGTTGTGTCTGGATCGCAGCAGTCGGCGAGCCAGTTCTGCG \\ s & a & s & q & f & c & g \end{array}$
12756639	R         P         R         H         F         L         Q         Q         Q         Q         Q         P         G         R         E           GCGTCCCCGTCACTTTCTCCAACAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAG
12756559	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
12756479	$\begin{array}{llllllllllllllllllllllllllllllllllll$
12756399	E T G R V D Q M L R H S E E F G G G GGGGGTGGGGGATGGGTTTGAGCAG <mark>AGGAAACAGGGCGGGTCGATCAGATGCTCCGCCACTCAGAAGAGTTTGGAGGTGG</mark>
12756319	R R H H R Q S E A G P Q G A G G G V L Q L Q E V P Q A GGGCCGACATCATCGACAAAGTGAAGCAGGACCACAAGGAGCTGGAGGAGGCGTACTCCAATTACAAGAAGTTCCACAAG
12756239	G Q R G R G R I M V Q P V R V G D L S P C R H R G A CAGGGCAACGAGGAAGAGGCCGGATAATGGTTCAACCAGTTCGTGTGGGAGATCTCTCGCCATGCCGTCACCGAGGAGCT
	*
12756159	G V V P V D R V A G R Q G P E A G G P V A R R A P E D GGTGTTGTACCCGTTGATCGCGTCGCAGGGGCGACAAGGGGCCAGAAGCTGGCGGACCAGTCGCGCGACGAGCACCAGAAGA
12756079	E G H A G G D P G H R G R R P V * CGAAGGACATGCTGGTGGAGATCCAGGGCATCGAGGACGACGACGACGTGTTTGAGAAGAAGTTCGACGCCATCATGGACGTG
12755999	CTGCGAGAGCACGTGCAAAAAGGAGGAGTCCGGAGGATCTGGAGTACTTGAGGCAGAACGTGGACCAGGCAGCCCGCGAGGCCGCGCGCGCCGC
12755919	AGCGGGAACCACATTCGCGCTGGGCAAGAACATCGTGCCGACGAAGCCCCATGCGGCAGTGCCGAACAGGTCGGCGATAG
12755839	TGGAGGGCGCGCTGGGGCTGTTCGCGACTCCCTTGGACAAGCTGCGCGACGTGTTCAGGCCGTATCCCAAGGACAACTAG
12755759	aggagcaatttgtagctttttgtacatcagcccccgtcccgcgagtttgttgttgcaaataggtggctccgggatggat
	ATCG YAN
	ATCG AIt YAN
	ATCG Hr

**Supplementary Figure 5.** Sequences of *YAN*, *AltYAN*, and *Hr*. Black boxes indicate exons of *YAN*. Red letters show exons of *AltYAN*. Blue letters show the pseudogenized *Hr* region. Capital letters above the nucleotide sequences indicate the amino acids encoded by *YAN*, while lowercase letters below indicate those encoded by *AltYAN*.



**Supplementary Figure 6.** RT-PCR confirmation of *YAN*, *AltYAN*, and *Hr*. (**a**) Schematic diagram of primers. To confirm the mRNA completeness of this locus, seven forward primers were designed, with F1-F5 annealing to the upstream and F6-F7 annealing to exons. Each forward primer cooperated with a reverse primer R2 for RT-PCR reactions. To further confirm the CDS sequences of *YAN* and *AltYAN*, two primer pairs, (F6, R1) and (F7, R3), were used to amplify the CDSs of *YAN* and *AltYAN*, respectively. Amplified CDSs were then sequenced. (**b**) Gel result of RT-PCR. F1-F5 cooperating with R2 failed to amplify the appropriate product, but F6 and F7 cooperating with R2 could amplify the appropriate product. The lengths of *YAN* and *AltYAN* CDS sequences were correct.

>Physcomitrium sp. YAN and Hr annotation
MYNMCSLGRSPPYINONSFCL
pisavapariatiqassisaasqp
GIGCOGGCATIGITGCGITCAAGCGAGCTGCCACGCCAICTIGCCIG
sagıvar kraat psci
A V G E P V L R R P R H F L Q Q Q Q Q Q P G R
q s a s q ī c g v p v t ī s n s s s s s s i v e
AGGAAACAGGGCGGGCGCGACACACACACACACACAGAGGTTGGGGCGCCCCGACATCATCGAC
r k q g g s i r c s a t q k s l e v g a d i i d
Q S E A G P Q G A G G G V L Q L Q E V P Q A G Q
AAAGTGAAGCAGGACCACAAGGAGCTGGAGGAGGCGTACTCCAATTACAAGAAGTTCCACAAGCAGGGCAA
kv kqd h kelee ays ny kkf h kqg n
R G R G R K M V Q P V R V G D L A P C R H R G A
CGAGGAAGAGGCCGGAAAATGGTTCAACCAGTTCGTGTGGGAGATCTCGCGCCATGCCGTCACCGAGGAGCT
e e e ag kwfnq fvwe isrhavte e l
G V V P V D R V A G R Q G P E A G G P V A R R A
GGTGTTGTACCCGTTGATCGCGTCGCAGGGCGACAAGGGCCAGAAGCTGGCGGACCAGTCGCGCGACGAGC
vlyplia sąg dkg ąkladąs rde
PEDEGHAGGDPGHRGRRPV*
ACCAGAAGACGAAGGACATGCTGGTGGAGATCCAGGGCATCGAGGACGACGACCTGTTTGAGAAGAAGTTC
hqktkdmlveiqgiedddlfekkf
GACGCAATCATGGACGTGCTGCGAGAGCACGTGCAAAAGGAGGAGTCGGAGGATCTGGAGTACTTGAGGCA
daim dvl rehvqkeesedleyl rq
GAACGTGGAACAGGCAGCCCGCGAGGCAGCGGGAACCACATTCGCGCTGGGCAAGAACATCGTGCCGACGA
n veq aare aag ttfalg knivp t
AGCCCCATGCGGCAGTGCCGAACAGGTCGGCGATAGTGGAGGGCGCCCTGGGGCTGTTCGCGACTCCCTTGG
k p h a a v p n r s a i v e g a l g l f a t p l
ACAAGCTGCGCGACGTGTTCACGCCGTATCCCAAGGACAACTAGAGGAGCAATTTGTAGCTTTTTGTACATCA
d k l r d v f t p y p k d n *
ATCG YAN

ATCG YAN ATCG Hr

## b

	1	10	20	30	40	50	60	70	80 90	100	110	120	130	
Physconitriun_sp_for Sphfalx0048s0034,1 Consensus	MAAVAYII	PLSAVAPA	RLATTQASSSN	AASQPSAGI	VAFKRAATPSC	LQSASQFCGVF	VTFSNSSSSS	SLVERKQGGSI	ERCSATOKSLEVG HSSHVTTTHHVP rcSatoksheVg	ADIIDKVKQDH GDLHDEIKHDH aDiiDe!KqDH	KELEERYSNYKK REFEEFYTNYKK rE1EEaYsNYKr	KFHKOGNEEEF RAYAAGDEKEF ahaaG#EeEf	1GKHFNQF 1ekhfnql 1ekhfnq1	
	131 :	140	150	160	170	180	190	200 2	210 220	230	240	250	260	
Physconitriun_sp_for Sphfalx0048s0034,1 Consensus	VHEISRH VHEISRH VHEISRH	AVTEELVL HVGEELVV AVgEELV1	YPLIASQGDKG YPLLDANGPEG YPLiaagGdeG	QKLADQSRD KEMADKDRA iqe\$ADqdRa	EHQKTKDMLVE Dhqkvkkelee #Hqktkdelee	IQGIEDDDLFE IHRLSDPGLFE IgrieDddLFE	KKFDAIMDVL ATLDRIMQDL aklDaIM#dL	REHVQKEE <mark>s</mark> ed Rohtnikeetoo R#HL#Kees#D	DLEYLRQNVEQAA DLEYLKQHADKKS DLEYLrQna#gaa	REAAGTTFALG LENAAKAFKYG rEaAakaFalG	KNIVPTKPHAA\ KKHAPTRPHPGI KniaPTrPHaa	/PNRSAIVEG IPNRSAALEAF PNRSAalEaf	ILGLFATP ILAFFVTP ILa1FaTP	
Physconitriun_sp_for Sphfalx0048s0034.1 Consensus	261 LDKLRDVI LDKLRDVI LDKLRDMI LDKLRDmi	270 FTPYPKDN FTPFPKEE FTP%PK##	280 + HVPYQPKQEEH	289 1 AKK										
c														
Physconitriun_sp Pp3c21_19720V3.2	1   hynhc9 hynhc9	10 SLGRSPPY SLGRSPPY	20 INQNSFCLCT INQNPFCLCT	30 LSDLQPLYH LSDLQPLYH	40 SLSVILYITH SLSVILYITH	50 IKALVVHREAE IKALVVHREAE	60 ALLPADHTVH ALLPADHTVH	70 HGRSRIHPLE HGRSRIHPLE	80 RCRSREVGNHP	90 SQLQQCGLPAC SOLOOCGLPAC	100 CGHCCYQASCH	110 IAILPAVGEP IAILPAVGEP	120 VLRRPRHFL	130 I 0000000
Consensus	HYNHCS	SLGRSPPY	INQNPFCLCT	LSDLQPLYH	SLSVILVITH	IKALYYHREAE	ALLPADHTYH	HGRSRIHPLE	RCRFREVGNHP	SQLQQCGLPA	<b>T</b> GHCCVQASCH	IAILPAYGEP	VLRRPRHFL	aqqqqqq
	131	140	150	160	170	180	190	200	210	220	230	240	248	
Physconitriun_sp Pp3c21_19720V3.2 Consensus	PGREET PGREET PGREET	FGRYDOML FGRYDOML FGRYDOML	RHSEEFGGGR RHSEEFGGGR RHSEEFGGGR	RHHRQSEAG Rhhrqseag Rhhrqseag	PQGAGGGYLC PQGAGGGYLC PQGAGGGYLC	)LQEVPQAGQR )LQEVPQAGQR )LQEVPQAGQR	GRGRKHVQPV GRGRIMVQPV GRGRIMVQPV	RVGDL <mark>A</mark> PCRH RVGDLSPCRH RVGDLaPCRH	irgagyypydryi Irgagyypydryi Irgagyypydryi	AGRQGPEAGGI Agrqgpeaggi Agrqgpeaggi	PVARRAPEDEGH PVARRAPEDEGH PVARRAPEDEGH	iaggdpghrg iaggdpghrg iaggdpghrg	RRPV RRPV RRPV	

**Supplementary Figure 7**. (a) Sequence and annotation of RT-PCR product of the HHE region in *Physcomitrium sp*. (GenBank accession number MG254884). Living materials were collected from Xinping, Yunnan, China. Amino acids encoded by the *Hr* gene are shown by lower case characters below the nucleotide sequence. Upper case characters above the nucleotide sequence indicate amino acids encoded by *YAN*. Boxes show exons of *YAN* (GenBank accession number MG254882). (b) Alignment of putative Hr protein sequences from *Physcomitrium sp*. and *Sphagnum fallax*. (c) Alignment of putative YAN protein sequences from *Physcomitrium sp*. and *Physcomitrella patens*.





### FPKM Coefficient of variance: 3.342

(across samples below)



Note: FPKM from DESeq2 analysis.

b

	Gene models – Pp3c21_19720	
	Gene models	
	Transcripts: 2 Exons: 5 Introns: 3 CDSs: 2	
	Expression	
	Pearson Correlated Expression	
	There are 3 genes with correlated expression > 0.85	
Gene	Defline	Correlation
Pp3c16_15390	(1 of 4) PTHR24096:SF219 - LONG CHAIN ACYL-COA SYNTHETASE 8	0.8926
Pp3c19_19049		0.86176
Pp3c8_4710		0.85877

**Supplementary Figure 9.** Transcription data of *YAN/AltYAN* from Phytozome (https://phytozome.jgi.doe.gov/phytomine/report.do?id=237468538). (a) *YAN/AltYAN* is strongly induced by both dehydration and rehydration. (b) *YAN/AltYAN* is co-expressed with multiple other genes, including long-chain acyl-CoA synthetase 8 (*LACS 8*).



**Supplementary Figure 10.** Generation of *yan/altyan* knockout mutants and molecular identification. (a) Schematic diagram of *YAN/AltYAN* gene disruption construct. The positions of the primers used for PCR analyses of wild-type plants and mutants are indicated with arrows. (b) The transformants were tested by PCR on genomic DNA using primer pairs shown in schematic diagram. (c) *YAN* and *AltYAN* transcriptions were detected in wild-type plants and *yan/altyan* mutants using semi-quantitative RT-PCR. Primers specific to *ACTIN* were used as the control gene. As shown in the figure, *YAN* and *AltYAN* transcriptions were abolished in *yan/altyan* knockout mutants.



**Supplementary Figure 11**. Chromosome ploidy analyses. The chromosome ploidy of 2-week old plants was detected using flow cytometry.



**Supplementary Figure 12.** Nitric oxide (NO) and superoxide anion accumulation in wild-type plants and *yan/altyan* knockout mutants. Two-week old *P. patens* plants were dehydrated for 20 hours and transferred into sterile water for 1 hour. Plants were then transferred onto standard medium for recovery. Superoxide anions were detected following two days of recovery. (a) Toxic NO accumulation in wild-type plants and *yan/altyan*. More toxic NO can be observed in *yan/altyan* mutants than wild-type plants under water stress. NO (green), chlorophyll (red). Scale bar = 1 mm. (b) Quantitative analyses of NO fluorescence in *yan/altyan* mutants and wild-type plant using image J software. Values show means and standard deviations of three biological replicates for the wild-type and two independent mutants. Asterisks indicate a statistically significant difference compared with wild-type plants based on a two-tailed student's *t*-test (p<0.05). Scale bar = 2 mm. (c) Comparison of superoxide anion accumulation in *yan/altyan* mutants and wild-type plants under dehydration stress. Dark brown areas show superoxide anions.



**Supplementary Figure 13.** Genotyping of *YAN-OE* and *AltYAN-OE* lines. (**a**) Genomic PCR was performed using DNA templates of *YAN-OE*, wild-type (negative control), and *pPOG1-YAN* (positive control). A primer pair designed to amplify full-length CDS of *YAN* was used. (**b**) Quantitative PCR was used to detect the expression level of *YAN* in *YAN-OE* lines. Values show means and standard deviations of three biological replicates for the wild-type and three *YAN-OE* lines. Asterisks indicate a statistically significant difference compared with wild-type plants based on a two-tailed student's *t*-test (p<0.05). (**c**) Genomic PCR was used. (**d**) Quantitative PCR was used to detect the expression level. (**d**) Quantitative PCR was used to detect the expression level of *AltYAN-OE* lines. Nature pair designed to amplify full-length CDS of *AltYAN* was used. (**d**) Quantitative PCR was used to detect the expression level of *AltYAN-OE* lines. Values show means and standard deviations of three biological replicates for the wild-type plants based to detect the expression level of *AltYAN* in *AltYAN-OE* lines. Values show means and standard deviations of three biological replicates for the wild-type plants based on a two-tailed student's *t*-test (p<0.05).



**Supplementary Figure 14.** Quantification of fatty acids in wild-type, *AltYAN-OE* and *YAN-OE* lines. Ten-week old plants were used to detect the composition and total amount of fatty acids. Values show means and standard deviations of three biological replicates for the wild-type, *AltYAN-OE-11#* and *YAN-OE-11#*. Asterisks indicate a statistically significant difference compared with wild-type plants based on a two-tailed student's *t*-test (p<0.05). (**a**) Total fatty acid content in the wild-type, *AltYAN-OE* and *YAN-OE* lines. (**b**) Comparison of fatty acid amount and composition in wild-type, *AltYAN-OE* and *YAN-OE* lines.



**Supplementary Figure 15.** Sporophytes of the wild-type and mutants. Plants were grown for 8 weeks at 25 °C before they were transferred to a short-day regime at 15 °C. After 4 weeks of sporophyte induction, visible sporophytes were photographed. Scale bar =  $100 \,\mu$ m.



Supplementary Figure 16. Germination of spores from wild-type and mutant plants of *Physcomitrella patens*. Mature sporangia were sterilized using 10% sodium hypocholorite solution for 10 minutes, then washed 5 times using sterilized water. The sporangia were crushed using the tip of a pipet. The released spores were spread on BCDAT medium. The germinated spores were photographed after 4 days of incubation. Scale bar =  $50 \mu m$ .





**Supplementary Figure 17**. Quantification of wax in wild-type, *AltYAN-OE* and *YAN-OE* lines. Ten-week old plants were used to detect the composition of wax. Values show means and standard deviations of three biological replicates for the wild-type, *AltYAN-OE-11#* and *YAN-OE-11#*. Asterisks indicate a statistically significant difference compared with wild-type plants based on a two-tailed student's *t*-test (p<0.05).

Tor10224 Gametophyte rehydration Library Syntrichia ruralis cDNA, mRNA sequence. Sequence ID: <u>CN200553.1</u> Length: 501 Number of Matches: 1

Score         Expect Method         Identities         Positives         Gaps           192 bits(489)         7e-60         Compositional matrix adjust.         84/148(57%)         110/148(74%)         0/148(0%)           Query         47         KWFNQLVWELSRHWVGEELVVYPLLDAMGPEGKEMADKDRADHQKVKKELEEIHRLSDPG         106           +WFNQ         VWE+SRH V EELV+VPL++A G G E+A++ R DEQ+ K LEE+ R +DP         106           sbjct         1         RWFNQVWEISRHAVSEELVLYPLEAKQORGSELAUNOSRTDHOQOTKDILEELQRTTDP         180           Query         107         LFEATLDRIMQDLRQHTNKEETQDLEYLKQHADKKSLENAAKAFKYGKKMAPTRPHPGIP         166           LFE         +D++M         LEE++DL <ylk+h< td="">         E A AF GK         AMAPTRPH +P           Sbjct         181         LFEQRMDKMMAELREHIRKEESEDLVYLKHENVSQDGREAAGTAFSLGKTMAPTRPHASVP         360           Query         167         NRSAALEAALAFFVTPLDKLRDMFTPFP         194           N+S         A+EAAL         +TP+DKLRD+FTPFP         194</ylk+h<>	Range 1: 1 to 444 GenBank Graphics Vext Match 🛦 Previous Match									
192 bits(489)       7e-60       Compositional matrix adjust. 84/148(57%)       110/148(74%)       0/148(0%)         Query       47       KWFNQUWELSRHNVGEELVVYPLLDAMGPEGKEMADKDRADHQKVKKELEEIHRLSDPG       106         +WFNQ       VWE+SRH V       EELV+YPL++A G       G E+A++ R       DR0+ K       LEE+ R +DP         Sbjot       1       RWFNQFVWEISRHAVSEELVLYPLIEAKGQRGSELANQSRTDHQQTKDILEELQRTDPD       180         Query       107       LFEATLDRINQDLRQHTKKEETQDLEVLKQHADKSLENAAKAFKYGKKMAPTRPHBGIP       166         LFE       +D++M       LRE++DL       YLK+H       E       A AF GK       MAPTRPH +P         Sbjot       181       LFEQRMDKMMAELREHIRKEESEDLVILKHENVSQDGREAAGTAFSLGKTMAPTRPHASVP       360         Query       167       NRSAALAAFLAFYTPLDKLRDMFTPFP       194         N+S       A+EAAL       TP+DKLRD+TPFP       194	Frame	Gaps	Positives	Identities	od	ct Metho	Expect		Score	
Query     47     KWFNQLVWELSRHNVGEELVVYPLLDAMGPEGKEMADKDRADHQKVKKELEEIHRLSDPG     106       +WFNQ     VWE+SRH V EELV+YPL++A G G E+A++ R DBQ+ K LEE+ R +DP       Sbjct     1     RWFNQFVWEISRHAVSEELVLYPLIEAKGQRGSELANQSRTDHQQTKDILEELQRTDPD     180       Query     107     LFEATLDRINQDLRQHTKKEETQDLEVLKQHADKSLENAAKAFKYGKKMAPTRPHPGIP     166       LFE     +D++M +LR+H     KEE++DI YLK+H     E A AF GK MAPTRPH +P       Sbjct     181     LFEQRMDKMMAELREHIRKEESEDLVYLKHVSQDGREAAGTAFSLGKTMAPTRPHASVP     360       Query     167     NRSAALAAFLYPLDKLRDMFTPFP     194       N+S A+EAAL     TP+DKLRD+TPFP     194	) +1	0/148(0%)	110/148(74%)	84/148(57%)	ositional matrix adjust.	Compo	7e-60	s(489)	192 bit	
Sbjct     1     RWFNÖFVWEISRHAVSEELVLYPLIEAKGORGSELANOSRTDHÖOTKDILEELORTTDPD     180       Query     107     LFEATLDRIMQDLRQHTNKEETQDLEYLKQHADKKSLENAAKAFKYGKKMAPTRPHPGIP     166       LFE     +D++M     +LFE++DL     YLK+H     +E     A     AF     GK     MAPTRPH     +P       Sbjct     181     LFEORMDKMMAELREHIKEESEDLVYLKEHVSQDGREAAGTAFSLGKTMAPTRPHASVF     360       Query     167     NRSAALAAALAFVTPLDKLRDMFTPFP     194       N+S     AFEAAL     +TP+DKLRD+FTPFP		106	KKELEEIHRLSDPG K LEE+ R +DP	EMADKDRADHQKV E+A++ R DHO+	MVGEELVVYPLLDAMGPEGK V EELV+YPL++A G G	VWELSRHM	KWFNQLV	47	Query	
Query 107 LFEATLDRIMQDLRQHTNKEETQDLEYLKQHADKKSLENAAKAFKYGKKMAPTRPHPGIP 166 LFE +D++M +LR+H KEE++DL YLK+H + E A AF GK MAPTRPH +P Sbjct 181 LFEQRMDKMMAELREHIRKEESEDLVYLKEHVSQDGREAAGTAFSLGKTMAPTRPHASVP 360 Query 167 NRSAALAAFVTPLDKLRDHFTPFP 194 N+S A+EAAL +TP+DKLRD+FTPFP		180	KDILEELQRTTDPD	ELANQSRTDHQQT	AVSEELVLYPLIEAKGORGS	VWEISRHA	RWFNQFV	1 1	Sbjct	
Sbjct     181     LFEQRMDKMMAELREHIRKEESEDLVYLKEHVSQDGREAAGTAFSLGKTMAPTRPHASVP     360       Query     167     NRSAALEAALAFFVTPLDKLRDMFTPFP     194       N+S     A+EAAL     +TP+DKLRD+FTPFP		166	GKKMAPTRPHPGIP GK MAPTRPH +P	KKSLENAAKAFKY + E A AF	RQHTNKEETQDLEYLKQHAD R+H KEE++DL YLK+H	DRIMQDLR	LFEATLDI	107	Query	
Query 167 NRSAALEAALAFFVTPLDKLRDMFTPFP 194 N+S A+EAAL +TP+DKLRD+FTPFP		360	GKTMAPTRPHASVP	ODGREAAGTAFSL	REHIRKEESEDLVYLKEHVS	DKMMAELR	LFEQRMD	181	Sbjct	
					VTPLDKLRDMFTPFP 194 +TP+DKLRD+FTPFP	EAALAFFV	NRSAALEA N+S A+EA	167	Query	
Sbjct 361 NKSVAIEAALGLLLTPMDKLRDIFTPFP 444					LTPMDKLRDIFTPFP 444	EAALGLLL	NKSVAIE	361	Sbjct	

b

EST00039 mRNP Lambda ZapII Express Library Syntrichia ruralis cDNA clone mRNP39 3', mRNA sequence. Sequence ID: <u>Al305074.1</u> Length: 323 Number of Matches: 1

Range 1: 10 to 108 GenBank Graphics V Next Match 🛕 Previous Match Score **Expect Method** Identities Positives Gaps Frame 42.7 bits(99) 0.014 Compositional matrix adjust. 18/33(55%) 24/33(72%) 0/33(0%) +1 
 Query
 150
 FKYGKKMAPTRPHPGIPNRSAALEAALAFFVTP
 182

 + G++
 PT+PH
 +PN+
 AALEAAL
 FVTP

 Sbjct
 10
 VRVGEEDVPTKPHAAVPNKPAALEAALGLFVTP
 108
 LIBRARY LIBEST\_001531 mRNP Lambda ZapII Express Library Lib Name: Organism: Syntrichia ruralis Tissue type: Desiccated Gametophytes Vector: pBluescriptII R. Site 1: EcoRI R. Site 2: XhoI mRNA derived from the polysomal mRNP fraction of desiccated Description: Tortula ruralis gametophytes, directionally cloned cDNA (lambda ZAPII) and in vivo excised to pBluescript (Stratagene)

С

SLA006H09\_34729 An expressed sequence tag (EST) collection from the resurrection plant Selaginella lepidophylla Selaginella lepidophylla cDNA clone SLA006H09 5, mRNA sequence. Sequence ID: BM402674.1 Length: 590 Number of Matches: 1

 Range 1: 249 to 590 GenBank Graphics
 Next Match & Previous Match

 Score
 Expect Method
 Identities
 Positives
 Gaps
 Frame

 125 bits (313) 7e-33
 Compositional matrix adjust. 58/114(51%) 82/114(71%) 0/114(0%) +3

 Frame

 Frame

An expressed sequence tag (EST) collection from the resurrection plant Selaginella lepidophylla

Identifiers	BioSample: SAMN001	170085; EST: LIBEST_010176					
Organism	Salaginella tepidophylla cellular organisma; Eukaryota; Viridiplantae; Streptophyta; Streptophytina; Embryophyta; Tracheophyta; Lycopodiopaida; Selaginellales; Selaginellaceae; Selaginella						
Attributes	development stage	adult					
	tissue	microphyll fronds undergoing desiccation for 2.5 h					
	re_1	EcoRI					
	v_type	Phagemid					
	re_2	Xhol					
	vector	Lambda Uni-Zap XR, Bluescript SK-					
Description	Library construction w vector and cDNA synt	as performed according to manufacture's (Stratagene, Inc.) recommended protocol for the Lambda UniZapXR hesis kit.					
Submission	University of Nevada,	Cushman JC; 2002-01-22					
Accession: SAN	IN00170085 ID: 170085						

a

**Supplementary Figure 18**. *Hr* genes are induced by dehydration and rehydration in two resurrection plants, the twisted moss *Syntrichia ruralis* and the spikemoss *Selaginella lepidophylla*. The Hr protein sequence from *Sphagnum fallax* (Phytozome identifier Sphfalx0048s0034.1) was used as query to perform TBLASTN search of NCBI dbEST database. (a) Alignment of *Sphagnum* Hr protein sequences with a translated sequence of *S. ruralis* rehydration transcript. (b) Alignment of *Sphagnum* Hr protein sequences with the translated sequence of another *S. ruralis* transcript, with the associated biosample record indicating that the sample was under desiccation. (c) Alignment of *Sphagnum* Hr protein sequences with a translated with a translated sequence of *S. lepidophylla* EST, with the associated biosample record indicating that the sample was under desiccation for 2.5 hours.

Supplementary	Table 1.	TBLASTN	result of	f 1KP	database	using a	a <i>Sphagnum</i>	Hr	protein
sequence as quer	y; Query=	Sphfalx0048	s0034.1 p	peptide	(212 ami	no acids	s); E-value cu	toff	= 0.01.

	Score	Е
Sequences producing significant alignments:	(Bits)	Value
scaffold-GOWD-2080086-Sphagnum_lescurii	285	7e-95
scaffold-RD00-2010340-Racomitrium_varium	227	4e-70
scaffold-WOGB-2015424-Andreaea_rupestris	219	4e-68
scaffold-FFPD-2009361-Ceratodon_purpureus	221	9e-68
scaffold-RD00-2116375-Racomitrium_varium	218	3e-67
scaffold-VMXJ-2128859-Leucobryum_albidum	220	7e-67
scaffold-KEFD-2005771-Encalypta_streptocarpa	214	3е-бб
<pre>scaffold-ABCD-2074154-Niphotrichum_elongatum</pre>	213	1e-65
scaffold-RD00-2116980-Racomitrium_varium	211	5e-64
scaffold-HRWG-2013098-Buxbaumia_aphylla	207	1e-63
scaffold-BPSG-2019772-Scouleria_aquatica	205	3e-62
scaffold-WOGB-2092690-Andreaea_rupestris	204	8e-62
scaffold-LGDQ-2041893-Selaginella_apoda	199	2e-61
scaffold-TXVB-2071911-Lunularia_cruciata	199	4e-60
scaffold-GRKU-2004786-Syntrichia_princeps	193	3e-59
scaffold-JKAA-2008718-Selaginella_wallacei	199	3e-59
scaffold-ABCD-2010707-Niphotrichum_elongatum	196	4e-59
scaffold-JKAA-2008719-Selaginella_wallacei	199	6e-59
scaffold-NGTD-2098183-Dicranum_scoparium	196	1e-58
scaffold-JMXW-2003398-Bryum_argenteum	187	1e-56
scaffold-ZYCD-2004286-Selaginella_acanthonota	196	1e-56
scaffold-JMXW-2003397-Bryum_argenteum	187	3e-56
scaffold-JMXW-2003396-Bryum_argenteum	186	6e-56
scaffold-ZZOL-2052205-Selaginella_stauntoniana	185	5e-55
scaffold-ZYCD-2004285-Selaginella_acanthonota	196	8e-55
scaffold-ORKS-2011502-Philonotis_fontana	191	5e-53
scaffold-GRKU-2012644-Syntrichia_princeps	179	7e-53
scaffold-PKOX-2012929-Isoetes_tegetiformans	181	1e-52
scaffold-GRKU-2012643-Syntrichia_princeps	180	1e-52
scaffold-VMXJ-2124089-Leucobryum_albidum	173	3e-51
scaffold-RD00-2109788-Racomitrium_varium	169	9e-51
scaffold-FITN-2089742-Treubia_lacunose	177	9e-51
scaffold-PYHZ-2006808-Isoetes_sp	177	2e-50
scaffold-PKOX-2096679-Isoetes_tegetiformans	175	4e-50

scaffold-TMAJ-2173065-Neckera_douglasii	161	2e-47
scaffold-YPIC-2134561-Microstegium_vimineum	149	5e-42
scaffold-RXRQ-2128249-Phaeoceros_carolinianus-sporophyte	145	5e-41
scaffold-WCZB-2013633-Phaeoceros_carolinianus-gametophyte	146	5e-41
scaffold-JMXW-2040177-Bryum_argenteum	146	6e-41
scaffold-WNGH-2006956-Aulacomnium_heterostichum	145	3e-40
scaffold-OFTV-2128708-Barbilophozia_barbata	144	1e-39
scaffold-CMEQ-2082481-Orthotrichum_lyellii	144	2e-39
scaffold-JBGU-2011382-Amaranthus_palmeri	143	2e-39
<pre>scaffold-YEPO-2071721-cfPhyscomicromitrium_sp</pre>	140	6e-39
scaffold-WCZB-2114770-Phaeoceros_carolinianus-gametophyte	142	4e-38
scaffold-RXRQ-2022078-Phaeoceros_carolinianus-sporophyte	144	4e-38
scaffold-IRBN-2152420-Scapania_nemorosa	139	1e-37
scaffold-RGKI-2054671-Leucobryum_glaucum	134	4e-37
scaffold-JKAA-2174841-Selaginella_wallacei	137	4e-37
scaffold-RXRQ-2132940-Phaeoceros_carolinianus-sporophyte	136	3e-36
scaffold-FFPD-2008086-Ceratodon_purpureus	131	4e-36
scaffold-FAJB-2053787-Paraphymatoceros_hallii	137	8e-36
scaffold-RXRQ-2018681-Phaeoceros_carolinianus-sporophyte	136	2e-35
scaffold-GRKU-2004785-Syntrichia_princeps	132	4e-35
scaffold-YBQN-2122714-Odontoschisma_prostratum	130	9e-35
scaffold-JHFI-2105756-Pellia_neesiana	129	2e-34
scaffold-CMEQ-2081739-Orthotrichum_lyellii	129	5e-34
scaffold-TGKW-2009714-Frullania	126	3e-33
scaffold-LNSF-2063071-Hypnum_subimponens	125	4e-33
scaffold-KJZG-2059461-Asplenium_platyneuron	129	5e-33
scaffold-LNSF-2008214-Hypnum_subimponens	127	7e-33
scaffold-IRBN-2150099-Scapania_nemorosa	125	8e-33
scaffold-NOKI-2092089-Lindsaea_linearis	128	1e-32
scaffold-ABCD-2011017-Niphotrichum_elongatum	129	1e-32
scaffold-NWQC-2149828-Plagiochila_asplenioides	126	2e-32
scaffold-IRBN-2150967-Scapania_nemorosa	125	2e-32
scaffold-JKAA-2177860-Selaginella_wallacei	127	3e-32
scaffold-GSXD-2062693-Myriopteris_eatonii	124	5e-32
scaffold-OFTV-2123041-Barbilophozia_barbata	121	9e-32
scaffold-MZLD-2123763-Ligustrum_sinense	124	1e-31
scaffold-DCDT-2072974-Cheilanthes_arizonica	125	1e-31
scaffold-KJZG-2058634-Asplenium_platyneuron	124	2e-31
scaffold-HNDZ-2129005-Cystopteris_utahensis	122	3e-31
scaffold-JBGU-2121651-Amaranthus_palmeri	123	3e-31
scaffold-LNSF-2008215-Hypnum_subimponens	122	4e-31

scaffold-JKAA-2170971-Selaginella_wallacei	119	8e-31
scaffold-OCZL-2010080-Homalosorus_pycnocarpos	122	1e-30
scaffold-ABCD-2009871-Niphotrichum_elongatum	122	1e-30
scaffold-IGUH-2161180-Schwetschkeopsis_fabronia	120	1e-30
scaffold-WNGH-2085370-Aulacomnium_heterostichum	121	3e-30
scaffold-NWQC-2005566-Plagiochila_asplenioides	120	4e-30
scaffold-NWQC-2005567-Plagiochila_asplenioides	118	9e-30
scaffold-CQPW-2020236-Anemia_tomentosa	119	3e-29
scaffold-CPKP-2097340-Lophophora_williamsii	114	1e-28
scaffold-WFBF-2000168-Podophyllum_peltatum	114	2e-28
scaffold-JKAA-2167480-Selaginella_wallacei	111	5e-28
scaffold-NRWZ-2091968-Metzgeria_crassipilis	110	1e-27
scaffold-XDDT-2012577-Argyrochosma_nivea	112	2e-27
scaffold-IRBN-2142735-Scapania_nemorosa	110	2e-27
scaffold-RDOO-2109148-Racomitrium_varium	106	4e-26
scaffold-WOGB-2081836-Andreaea_rupestris	103	1e-25
scaffold-OSHQ-2044949-Sarcandra_glabra	106	2e-25
scaffold-ZTHV-2000232-Atrichum_angustatum	103	2e-24
scaffold-NRWZ-2009763-Metzgeria_crassipilis	99.4	2e-23
scaffold-WTDE-2023700-Johnsonia_pubescens	99.8	2e-23
scaffold-IGUH-2154130-Schwetschkeopsis_fabronia	99.4	2e-23
scaffold-SALZ-2065380-Pittosporum_resiniferum-fruit	100	8e-23
scaffold-WCZB-2102726-Phaeoceros_carolinianus-gametophyte	96.3	2e-22
scaffold-CHJJ-2127518-Lejeuneaceae_sp	96.7	2e-22
scaffold-CIEA-2049265-Juncus_inflexus	98.6	2e-22
scaffold-NGTD-2090279-Dicranum_scoparium	96.3	3e-22
scaffold-IRBN-2139892-Scapania_nemorosa	95.9	5e-22
scaffold-VQYB-2010950-Neurachne_lanigera	95.5	2e-21
scaffold-ABCD-2064447-Niphotrichum_elongatum	93.6	2e-21
scaffold-OCWZ-2051463-Dioscorea_villosa	97.1	5e-21
scaffold-PSHB-2008862-Lantana_camara	94.0	7e-21
scaffold-KEFD-2050257-Encalypta_streptocarpa	91.7	9e-21
scaffold-KEFD-2054814-Encalypta_streptocarpa	92.8	1e-20
scaffold-VMXJ-2107202-Leucobryum_albidum	89.0	5e-20
scaffold-YWNF-2008347-Hedwigia_ciliata	90.9	1e-19
scaffold-RGKI-2054107-Leucobryum_glaucum	88.2	2e-19
scaffold-TMAJ-2001356-Neckera_douglasii	92.0	2e-19
scaffold-YWNF-2008346-Hedwigia_ciliata	91.7	4e-19
scaffold-RTMU-2149348-Calypogeia_fissa	86.7	7e-19
scaffold-HATH-2079510-Aristida_stricta	87.0	9e-19
scaffold-ZXJO-2012394-Hemionitis_arifolia	87.0	3e-18

scaffold-YPIC-2132251-Microstegium_vimineum	86.3	4e-18
scaffold-WVMY-2126909-Phlox_drummondii	86.3	4e-18
scaffold-ABIJ-2031254-Selaginella_lepidophylla	83.2	9e-18
scaffold-MVSE-2025297-Griselinia_littoralis	85.5	9e-18
scaffold-RD00-2015219-Racomitrium_varium	85.5	1e-17
scaffold-IRBN-2122784-Scapania_nemorosa	80.9	7e-17
scaffold-VMXJ-2115109-Leucobryum_albidum	81.6	7e-17
scaffold-UEEN-2146356-Forestiera_segregata	80.5	1e-16
scaffold-YPIC-2127709-Microstegium_vimineum	81.3	1e-16
scaffold-NWQC-2113770-Plagiochila_asplenioides	80.1	1e-16
scaffold-IRBN-2155004-Scapania_nemorosa	85.1	2e-16
scaffold-WNGH-2012483-Aulacomnium_heterostichum	83.2	2e-16
scaffold-YPIC-2115414-Microstegium_vimineum	79.0	3e-16
scaffold-POPJ-2014900-Pteris_vittata	80.5	4e-16
scaffold-PKOX-2008033-Isoetes_tegetiformans	59.7	9e-16
scaffold-IGUH-2105402-Schwetschkeopsis_fabronia	77.0	1e-15
scaffold-QMWB-2052150-Anomodon_attenuatus	78.6	1e-15
scaffold-NGTD-2089101-Dicranum_scoparium	79.0	1e-15
scaffold-BGXB-2069679-Plagiomnium_insigne	78.6	1e-15
scaffold-OCWZ-2048666-Dioscorea_villosa	79.3	2e-15
scaffold-IRBN-2002456-Scapania_nemorosa	78.6	2e-15
scaffold-TVSH-2057737-Bituminaria_bituminosa	77.0	3e-15
scaffold-XWHK-2033525-Funaria	75.9	3e-15
scaffold-NGTD-2072576-Dicranum_scoparium	75.9	3e-15
scaffold-NDUV-2084566-Vittaria_appalachiana	75.9	6e-15
scaffold-TMAJ-2176774-Neckera_douglasii	78.6	1e-14
scaffold-WTDE-2120279-Johnsonia_pubescens	75.5	1e-14
scaffold-WPUV-2050890-Oenothera_grandis-MTJ_135	74.7	3e-14
scaffold-ZENX-2075599-Neurachne_alopecuroidea	73.6	4e-14
scaffold-VMXJ-2101389-Leucobryum_albidum	72.8	6e-14
<pre>scaffold-XWDM-2024866-XWDM-Cystopteris_fragilis-2_samples_combined</pre>	62.0	9e-14
scaffold-CPKP-2083294-Lophophora_williamsii	72.0	2e-13
scaffold-CVEG-2117220-Azolla_cfcaroliniana	72.4	2e-13
scaffold-RTMU-2146607-Calypogeia_fissa	72.0	3e-13
scaffold-PYHZ-2058919-Isoetes_sp	65.9	3e-13
scaffold-TMAJ-2146025-Neckera_douglasii	70.1	8e-13
scaffold-UPMJ-2080149-Pseudolycopodiella_caroliniana	73.6	1e-12
scaffold-ZTHV-2078155-Atrichum_angustatum	59.3	1e-12
scaffold-RTMU-2082020-Calypogeia_fissa	68.6	1e-12
scaffold-CMEQ-2067654-Orthotrichum_lyellii	68.9	2e-12
scaffold-RTMU-2159248-Calypogeia_fissa	70.9	2e-12

<pre>scaffold-KEFD-2052864-Encalypta_streptocarpa</pre>	70.5	2e-12
scaffold-CMEQ-2063716-Orthotrichum_lyellii	67.4	4e-12
scaffold-YPIC-2024478-Microstegium_vimineum	70.1	5e-12
scaffold-HATH-2073870-Aristida_stricta	68.2	6e-12
scaffold-PMRP-2036296-Cannabis_sativa-stem	70.1	1e-11
scaffold-BJSW-2066960-BJSW-Cannabis_sativa-4_samples_combined	70.1	1e-11
<pre>scaffold-IMZV-2080047-IMZV-Oenothera_grandis-5_samples_combined</pre>	67.0	2e-11
scaffold-RXRQ-2131384-Phaeoceros_carolinianus-sporophyte	69.7	2e-11
scaffold-OCWZ-2044044-Dioscorea_villosa	66.2	3e-11
scaffold-CHJJ-2130531-Lejeuneaceae_sp	68.2	3e-11
scaffold-GKAG-2079692-Huperzia_lucidula	65.1	5e-11
scaffold-IGUH-2160559-Schwetschkeopsis_fabronia	68.6	6e-11
scaffold-CHJJ-2012733-Lejeuneaceae_sp	68.6	6e-11
scaffold-KRUQ-2055797-Porella_navicularis	63.2	1e-10
scaffold-VMXJ-2081240-Leucobryum_albidum	63.5	1e-10
scaffold-JKAA-2154702-Selaginella_wallacei	64.3	1e-10
scaffold-JSAG-2054274-Masdevallia_yuangensis	62.8	2e-10
scaffold-FITN-2084753-Treubia_lacunose	65.9	2e-10
scaffold-GRKU-2069920-Syntrichia_princeps	63.5	3e-10
scaffold-ABCD-2067800-Niphotrichum_elongatum	64.3	5e-10
scaffold-IGUH-2157080-Schwetschkeopsis_fabronia	64.7	6e-10
scaffold-RTMU-2014569-Calypogeia_fissa	46.6	6e-10
scaffold-NRWZ-2079339-Metzgeria_crassipilis	62.4	8e-10
scaffold-NRWZ-2092900-Metzgeria_crassipilis	64.3	1e-09
scaffold-LHLE-2056411-Cystopteris_fragilis	61.6	1e-09
scaffold-HATH-2079179-Aristida_stricta	63.2	1e-09
scaffold-UUHD-2077091-Porella_pinnata	60.5	1e-09
scaffold-LNSF-2055857-Hypnum_subimponens	62.0	2e-09
scaffold-FFPD-2029090-Ceratodon_purpureus	59.7	2e-09
scaffold-TMAJ-2012211-Neckera_douglasii	62.8	2e-09
scaffold-MIRS-2073569-Climacium_dendroides	61.2	3e-09
scaffold-HPXA-2104167-Ptilidium_pulcherrimum	60.5	3e-09
scaffold-CHJJ-2000052-Lejeuneaceae_sp	63.2	4e-09
scaffold-GRKU-2071783-Syntrichia_princeps	60.8	5e-09
scaffold-VMXJ-2123744-Leucobryum_albidum	63.2	5e-09
scaffold-HVBQ-2112238-Tetraphis_pellucida	60.5	6e-09
scaffold-QMWB-2055650-Anomodon_attenuatus	62.0	6e-09
scaffold-HPXA-2127478-Ptilidium_pulcherrimum	62.0	7e-09
scaffold-WOGB-2084446-Andreaea_rupestris	60.5	7e-09
scaffold-JKAA-2078739-Selaginella_wallacei	57.8	1e-08
scaffold-AEXY-2130519-Blasia_sp	60.8	1e-08

scaffold-JKAA-2165817-Selaginella_wallacei	60.5	1e-08
scaffold-PSHB-2043956-Lantana_camara	58.2	1e-08
scaffold-NGTD-2089411-Dicranum_scoparium	60.1	1e-08
scaffold-JMXW-2034222-Bryum_argenteum	58.5	2e-08
scaffold-TMAJ-2003433-Neckera_douglasii	58.9	2e-08
scaffold-NRWZ-2085378-Metzgeria_crassipilis	58.9	2e-08
scaffold-KEFD-2044179-Encalypta_streptocarpa	57.4	4e-08
scaffold-GRRW-2048048-Grevillea_robusta	57.8	4e-08
scaffold-ABCD-2058843-Niphotrichum_elongatum	57.4	5e-08
scaffold-PSHB-2039572-Lantana_camara	55.5	9e-08
scaffold-XNXF-2001698-Dendrolycopodium_obscurum	53.5	1e-07
scaffold-IRBN-2103858-Scapania_nemorosa	55.5	2e-07
scaffold-TMWO-2076697-Papaver_bracteatum-bulb	54.7	2e-07
scaffold-SSDU-2123337-SSDU-Papaver_bracteatum-3_samples_combined	54.7	2e-07
scaffold-VMXJ-2120453-Leucobryum_albidum	57.4	2e-07
scaffold-JHCN-2084608-Oxalis_sp	57.0	3e-07
scaffold-NRWZ-2088588-Metzgeria_crassipilis	56.2	3e-07
scaffold-UAXP-2066247-Gyrostemon_ramulosus	55.5	3e-07
scaffold-IGUH-2005101-Schwetschkeopsis_fabronia	57.4	3e-07
scaffold-NRWZ-2089194-Metzgeria_crassipilis	56.2	4e-07
scaffold-RTMU-2146256-Calypogeia_fissa	55.5	4e-07
scaffold-VMXJ-2077554-Leucobryum_albidum	53.9	5e-07
scaffold-OFTV-2003852-Barbilophozia_barbata	57.4	5e-07
scaffold-ZXJO-2005224-Hemionitis_arifolia	55.8	6e-07
scaffold-ABIJ-2034612-Selaginella_lepidophylla	55.5	8e-07
scaffold-YEPO-2030533-cfPhyscomicromitrium_sp	52.8	8e-07
scaffold-WOGB-2008726-Andreaea_rupestris	55.8	9e-07
scaffold-CHJJ-2116743-Lejeuneaceae_sp	53.9	1e-06
scaffold-IRBN-2005941-Scapania_nemorosa	56.2	2e-06
scaffold-IRBN-2005942-Scapania_nemorosa	56.2	2e-06
scaffold-IRBN-2076572-Scapania_nemorosa	52.0	2e-06
scaffold-ABCD-2014511-Niphotrichum_elongatum	51.2	3e-06
scaffold-OFTV-2087677-Barbilophozia_barbata	42.0	3e-06
scaffold-HPXA-2008638-Ptilidium_pulcherrimum	38.9	3e-06
scaffold-RDOO-2089861-Racomitrium_varium	52.0	4e-06
scaffold-IRBN-2155117-Scapania_nemorosa	55.1	5e-06
scaffold-IGUH-2005100-Schwetschkeopsis_fabronia	53.9	5e-06
scaffold-YPIC-2124849-Microstegium_vimineum	52.4	6e-06
scaffold-HPXA-2105951-Ptilidium_pulcherrimum	51.6	7e-06
scaffold-MROH-2036845-Thelypteris_acuminata	52.0	7e-06
scaffold-ZSAB-2180975-Hoheria_angustifolia	52.0	9e-06

scaffold-YCKE-2055091-Notholaena_montieliae	52.8	9e-06
scaffold-TMAJ-2120109-Neckera_douglasii	50.8	9e-06
scaffold-LRTN-2087772-Monotropa_uniflora	51.6	1e-05
scaffold-GRKU-2044556-Syntrichia_princeps	49.7	1e-05
scaffold-CHJJ-2119525-Lejeuneaceae_sp	51.2	2e-05
scaffold-NGTD-2066643-Dicranum_scoparium	49.7	2e-05
scaffold-CMEQ-2082860-Orthotrichum_lyellii	53.1	3e-05
scaffold-WOGB-2051835-Andreaea_rupestris	48.5	3e-05
scaffold-MVSE-2190933-Griselinia_littoralis	49.3	4e-05
scaffold-PSHB-2043518-Lantana_camara	43.1	5e-05
scaffold-HATH-2002294-Aristida_stricta	49.7	5e-05
scaffold-JKAA-2053839-Selaginella_wallacei	48.1	5e-05
scaffold-YWNF-2020517-Hedwigia_ciliata	47.8	5e-05
scaffold-YWNF-2020516-Hedwigia_ciliata	47.8	5e-05
<pre>scaffold-FROP-2070730-FROP-Epifagus_virginiana-2_samples_combined</pre>	49.3	6e-05
scaffold-WCZB-2085267-Phaeoceros_carolinianus-gametophyte	48.5	6e-05
<pre>scaffold-BJSW-2012126-BJSW-Cannabis_sativa-4_samples_combined</pre>	50.1	7e-05
scaffold-WFBF-2003972-Podophyllum_peltatum	51.2	7e-05
scaffold-WOGB-2008722-Andreaea_rupestris	49.3	8e-05
scaffold-VMXJ-2036090-Leucobryum_albidum	47.0	9e-05
scaffold-OQWW-2161174-Davallia_fejeensis	47.4	1e-04
scaffold-YPIC-2134240-Microstegium_vimineum	50.1	2e-04
scaffold-KRUQ-2090857-Porella_navicularis	47.8	2e-04
scaffold-NGTD-2062919-Dicranum_scoparium	47.0	2e-04
scaffold-OXYP-2078939-Sideroxylon_reclinatum	46.6	2e-04
scaffold-KEFD-2052641-Encalypta_streptocarpa	48.5	2e-04
scaffold-XRCX-2095148-Aster_tataricus	48.5	2e-04
scaffold-CHJJ-2127168-Lejeuneaceae_sp	48.5	3e-04
scaffold-JKAA-2061455-Selaginella_wallacei	45.8	3e-04
scaffold-CVEG-2094112-Azolla_cfcaroliniana	46.2	4e-04
scaffold-OQWW-2161093-Davallia_fejeensis	45.8	4e-04
scaffold-JKAA-2008453-Selaginella_wallacei	47.8	4e-04
scaffold-LNSF-2045802-Hypnum_subimponens	46.2	5e-04
scaffold-NRWZ-2086705-Metzgeria_crassipilis	47.4	5e-04
scaffold-FAJB-2050601-Paraphymatoceros_hallii	47.8	6e-04
scaffold-NDUV-2080635-Vittaria_appalachiana	45.8	7e-04
scaffold-VMXJ-2028561-Leucobryum_albidum	44.7	7e-04
scaffold-TGKW-2021476-Frullania	44.3	0.001
scaffold-EWXK-2077938-Thyrsopteris_elegans	44.7	0.001
scaffold-RD00-2025299-Racomitrium_varium	43.9	0.001
scaffold-KEFD-2043942-Encalypta_streptocarpa	45.1	0.001

scaffold-VMXJ-2028149-Leucobryum_albidum	43.5	0.002
scaffold-VBMM-2023838-Anomodon_rostratus	43.5	0.002
scaffold-JKAA-2129507-Selaginella_wallacei	43.9	0.002
scaffold-PMRP-2006102-Cannabis_sativa-stem	45.4	0.003
scaffold-NRWZ-2069162-Metzgeria_crassipilis	43.9	0.003
scaffold-NDUV-2096264-Vittaria_appalachiana	45.8	0.004
scaffold-WOGB-2064377-Andreaea_rupestris	43.1	0.005
scaffold-NRWZ-2047801-Metzgeria_crassipilis	42.0	0.009
scaffold-CHJJ-2131097-Lejeuneaceae_sp	44.7	0.009

**Supplementary Table 2.** Taxonomic report of TBLASTN search of *nr* database using a translated charophyte sequence (ISGT-2038756) as query. The sequence was retrieved from 1KP and translated into six frames. The match to Hr protein sequences was used as query to search the nr databases. E-value cutoff = 1.

Organism	Blast Name	Score	Number of Hits	Description
cellular organisms			167	
Bacteria	bacteria		159	
Drotochastoria	protochastoria		1/2	
Alphaprotophatoria	a-protochastoria		143	
Alphaproteobacteria	a-proteobacteria		99	
Rhodospirillales	a-proteobacteria		11	
Rhodospirillaceae	a-proteobacteria		4	
Rhodospirillaceae bacterium	a-proteobacteria	50 4	1	Rhodospirillaceae bacterium
CCH5-H10 Phaeospirillum fulvum	a-proteobacteria	46 6	± 1	CCH5-H10 hits Phaeospirillum fulvum hits
Phaeospirillum fulvum MGU-	a-proteobacteria	39.7	1	Phaeospirillum fulvum MGU-K5
K5 Dongia gp UBHE0060	a proteobacteria	27 /	± 1	hits
Rhodospirillales bacterium	a proteobacteria	47 8	4	Rhodospirillales bacterium
URHD0017 Revranella massiliensis	a-proteobacteria	40 4	1	URHD0017 hits Revranella massiliensis hits
Rhodospirillales bacterium	a-proteobacteria	40 0	1	Rhodospirillales bacterium SCN
SCN 65-16 Enhydrobacter aerosaccus	a-proteobacteria	37 0	± 1	65-16 hits Enhydrobacter aerosaccus hits
Sphingobium glosgoo	a proteobacteria	47 0	2	Sphingobium glosgoo hitg
	a-proceobacceria	47.0	2	Splitligoblum cloacae litts
Sphingobium baderi	a-proteobacteria	46.6	2	Sphingobium baderi hits
Sphingobium chungbukense	a-proteobacteria	46.2	2	Sphingobium chungbukense hits
Sphingobium japonicum	a-proteobacteria	46.2	2	Sphingobium japonicum hits
Sphingobium japonicum UT26S	a-proteobacteria	46.2	<u>1</u>	hits
Sphingobium	a-proteobacteria	46.2	2	Sphingobium hits
Sphingobium chinhatense IP26	<u>a-proteobacteria</u>	46.2	<u>1</u>	hits
Sphingobium sp. HDIP04	a-proteobacteria	46.2	1	Sphingobium sp. HDIP04 hits
Sphingobium lucknowense F2	a-proteobacteria	46.2	1	Sphingobium lucknowense F2 hits
Sphingomonadaceae	a-proteobacteria	46 2	2	Sphingomonadaceae hits
Sphingomonag gp PUC-1	a proteobacteria	46.2	1	Sphingomonag gp PUC-A hitg
Spiringomonas sp. BHC-A	a-proceobacceria	40.2	<u>+</u>	Splitligomonas sp. BHC-A litts
Sphingobium indicum B90A	a-proteobacteria	46.2	1	Sphingobium indicum B90A hits
Sphingobium yanoikuyae	a-proteobacteria	44.7	4	Sphingobium yanoikuyae hits
Sphingobium sp. YBL2	a-proteobacteria	44.7	1	Sphingobium sp. YBL2 hits
Sphingobium sp. AM	a-proteobacteria	44.7	1	Sphingobium sp. AM hits
Sphingohium sp. 22B	a-proteobacteria	44 7	1	Sphingohium sp 22B hits
Sphingobium bp. 225	<u>a proceobacceria</u>	44 7	-	Chimesen and the hite
Sphingomonas paucimobilis Sphingomonas sp. MEA3-1	a-proteobacteria a-proteobacteria	44.7 44.7	3	Sphingomonas paucimobilis nits Sphingomonas sp. MEA3-1 hits
alpha proteobacterium LLX12A	a-proteobacteria	43.1	<u>1</u>	alpha proteobacterium LLX12A hits
Novosphingobium sp. Rr 2-17	a-proteobacteria	43.1	2	Novosphingobium sp. Rr 2-17
~ ] ] · · · · · · · · · · · · · · · · ·		40.1	_	nits
Caulobacter sp. K31	a-proteobacteria	43.1	2	Caulobacter sp. K31 hits
Sphingobium sp. DC-2	a-proteobacteria	42.4	1	Sphingobium sp. DC-2 hits
Caulobacter sp. Root343	a-proteobacteria	42.4	2	Caulobacter sp. Root343 hits
Caulobacter henricii	a-proteobacteria	42.4	1	Caulobacter henricii hits
Brevundimonas viscosa	a-proteobacteria	42.4	2	Brevundimonas viscosa hits
Sphingomonas koreensis	a-proteobacteria	42 0	3	Sphingomonas koreensis hits
Cohingohium guigguiliorum	a protochactoria	12.0	1	Cohingohium guigguiliorum hita
Sphingobium duisduillarum	a-proceobacceria	42.0	1	Sphingobium quisquiliarum P25
Sphingopium quisquillarum P25	a-proteobacteria	42.0	<u>_</u>	hits
Sphingobium sp. M11205	a-proteobacteria	41.2	2	Sphingobium sp. MI1205 hits
Sphingomonas sp. Root241	a-proteobacteria	41.2	2	Sphingomonas sp. Root241 hits
Novosphingobium panipatense	<u>a-proteobacteria</u>	41.2	<u>1</u>	Novosphingobium panipatense hits
Sphingobium herbicidovorans	a-proteobacteria	41.2	<u>1</u>	Sphingobium herbicidovorans
Sphingobium herbicidovorans	a-proteobacteria	41.2	<u>1</u>	Sphingobium herbicidovorans NBRC 16415 hits
Sphingomonas pruni	a-proteobacteria	40.8	1	Sphingomonas pruni hits
Sphingohium sp. VL23	a-proteobacteria	40 0	1	Sphingohium sp VI.23 hits
Cophingomonog Mala 00	a proteobacteria	40.0	± 1	Cabingomonog ga Make 00 bil
Spningomonas sp. Ndbn-20	a-proteopacteria	40.0	1	Spuingomonas sp. Ndbn-20 hits
Sphingomonas sp. URHD0057	a-proteobacteria	40.0	1	Sphingomonas sp. URHD0057 hits
Sphingobium sp. SYK-6	a-proteobacteria	40.0	2	Sphingobium sp. SYK-6 hits
Sphingomonas	a-proteobacteria	39.7	1	Sphingomonas hits
Sphingomonas sp Root1294	a-proteobacteria	39 7	1	Sphingomonas sp Root1294 hits
Sphingomonag an Bootto	a-protochastoria	20.7		Sphingomonag gp BootE0 hit-
Spiringomonas sp. Kootsu	a-proceobacteria	39./	1	Sphingomonas sp. Rootsu hits
Spningomonas sp. Root/20	a-proteobacteria	39.7	1	spningomonas sp. Root/20 hits
Sphingomonas sp. MCT13	a-proteobacteria	39.7	2	Sphingomonas sp. MCT13 hits

Sphingobium sp. MP6037 Methylogytis sp. SC2 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 39.3 a-proteobacteria 38.5 1 Sphingobium sp. XP657 Sphingobium japonicum BiD2 Sphingobium sp. Roct1423 Sphingomonas sp. Roct1423 Sphingomonas mali Sphingobium sp. XP49 Sphingobium sp. XP49 Paraburkholderia sediminicla Paraburkholderia sediminicla Paraburkholderia seturicola Sphingobium sp. XP27 Paraburkholderia seturicola Sphingobium sp. XP27 Paraburkholderia sp. NC214 Paraburkholderia sp. NC214 Paraburkholderia sp. NC214 Roseateles depolymerans Roseateles depolymerans Roseateles depolymerans Streptomyces sp. T-3 Streptomyces sp. T-3 Streptomyces sp. TP-A0356 Streptomyces sp. TP-C2 1982 Roseateles terree Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356 Streptomyces sp. TP-A0356	Sphindomonas sp .T.T-A5	a-proteobacteria	29 2	2	St
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<ul> <li>Abovsphingobium sp. MD-1</li> <li>a-proteobacteria</li> <li>a-proteobacteri</li></ul>	UNC305MECol5 2	<u>a-proteobacteria</u>	38.1	1	h
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<ul> <li>Brevundimonas sp. Root1423 a-proteobacteria 37.0 1</li> <li>Brevundimonas sp. Root608 a-proteobacteria 37.0 2</li> <li>Sphingomonas mengshuiensis a-proteobacteria 37.0 1</li> <li>Sphingomonas mali a-proteobacteria 47.4 1</li> <li>Paraburkholderia terricola b-proteobacteria 47.4 1</li> <li>Paraburkholderia terrae BSO01 b-proteobacteria 46.6 2</li> <li>Burkholderia sp. Y281 b-proteobacteria 46.6 2</li> <li>Burkholderia sp. Y281 b-proteobacteria 46.6 2</li> <li>Burkholderia sp. Y277 b-proteobacteria 40.8 2</li> <li>Mitsuaria sp. 7 b-proteobacteria 40.8 4</li> <li>Nitrosomonas sp. Nm166 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W277 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W177 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W177 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W177 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W277 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W177 b-proteobacteria 40.8 2</li> <li>Burkholderia sp. W177 b-proteobacteria 30.7 2</li> <li>Rosateles depolymerans b-proteobacteria 30.7 2</li> <li>Rosateles depolymerans b-proteobacteria 30.7 2</li> <li>Resateles terrae b-proteobacteria 30.7 2</li> <li>Resateles terrae b-proteobacteria 30.7 1</li> <li>Streptomyces sp. F-A0356 high GC Gram+ 40.0 1</li> <li>Phormidium ambiguum IAM M-71 cyanobacteria 38.1 1</li> <li>Oscillatoria sp. PCC 10802 cyanobacteria 38.5 1</li> <li>Mocardia africana high GC Gram+ 30.7 1</li> <li>Streptomyces sp. PF-A0356 high GC Gram+ 30.7 1</li> <li>Streptomyces sp. PF-A0356 high GC Gram+ 36.2</li></ul>	Brevundimonas	a-proteobacteria	37.0	1	B
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The many hemerythrin hits from flowering plants raises a significant concern about contamination in 1KP data. Other than chlorophycean sequences, which can be verified through the scaffold information of their complete genomes, other algal sequences are only sporadically found in various lineages, including red algae (Glaucosphera vacuolata, 1KP IDs: RSOF scaffold 2008604, RSOF\_scaffold\_2041944, RSOF\_scaffold\_2004035, RSOF\_scaffold\_2032925), glaucophytes (Glaucocystis nostochinearum, 1KP IDs: POOW\_scaffold\_2004542, POOW\_scaffold\_2053347) and charophytes (Hormidiella sp., 1KP IDs: ISGT-scaffold\_2038756, ISGT-scaffold\_2001291; Coleochaete irregulares, 1KP IDs: OPDY-scoffold 2002876, OPDY-scaffold 2002877), and other chlorophytes (Chlorella IKP ID: MWAN\_scaffold\_2004366; Parachlorella kessleri, IKP ID: AKCR\_scaffold\_2002030; Ignatius tetrasporus, 1KP ID: KADG\_scaffold\_2037975; *Botryococcus\_sudeticus*, IKP ID: VJDZ\_scaffold\_2007822). Because the 1KP database only contains plant sequence data, we further searched the *nr* database using the above sequences as query to investigate whether they were indeed similar to chlorophycean and land plant hemerythrins. These analyses showed that the red algal sequences were significantly more similar to fungal homologs than to chlorophycean and land plant sequences. Not hits to red algae were identified, even though multiple red algal genomes (e.g., Cyanodioschyzon merolae, Chondrus crispus, Galdieria sulphuraria, Porphyridium purpureum, Porphyra umbilicalis, and Pyropia yezoensis) have been sequenced and deposited in NCBI. Similarly, the glaucophytes were significantly more similar to fungal and bacterial sequences than to chlorophycean and land plant sequences (in both sequence coverage and identity).

Of the four charophyte sequences, three (1KP IDs: ISGT-scaffold\_2038756, QPDYscaffold\_2002876 and QPDY-scaffold\_2002877) shared higher or similar percent identities with bacterial and fungal homologs (see Supplementary Table 2 above), and one (1KP ID: ISGTscaffold 2001291) was slightly more similar to chlorophycean sequences. Although many other green algae outside Chlorophyceae (e.g., the charophyte Klebsormidium nitens, chlorophytes Ostreococcus lucimarinus, Ostreococcus tauri. Micromonas pusilla, *Cymbomonas* tetramitiformis, Coccomyxa subellipsoidea, Chlorella variabilis, Chlorella vulgaris, Auxenochlorella protothecoides, Picochlorum sp., Helicosporidium sp.) have been sequenced and their data have been deposited in NCBI, no hits from these taxa were detected. Except for Parachlorella kessleri, all other chlorophyte sequences also shared a higher sequence coverage and identity with bacterial or fungal homologs, and again no hits from green algae outside Chlorophyceae were detected. The *Parachlorella kessleri* sequence was more similar to a protein sequence of Chlorella variabilis that was translated from mRNA data (NCBI accession number: XP 005847154), which in turn was more similar to fungal and bacterial sequences.

**Supplementary Note 2.** We searched the *Nitella hyalina* transcriptomic data at NCBI (https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR064326), but only two matches of 19-32 nucleotides were found; these two matches were mapped onto different regions (separated by about 470 nucleotides) of the *Hr* CDS (see figure below, top panel). Search of the transcriptomic data of the charophyte <u>Closterium peracerosum-strigosum-littorale complex</u> at NCBI (https://www.ncbi.nlm.nih.gov/bioproject?term=PRJNA296352) provided a similar result (also see below, lower panel).



BLASTN search of transcriptomic data of charophytes *Nitella hyalina* (top panel) and *Closterium peracerosum-strigosum-littorale* complex (lower panel) at NCBI. Query is the CDS of *Marchantia polymorpha Hr* (JGI Id: Mapoly0042s0093). Results indicate that only short fragments are mapped to the query.

Because the assessment of sequence homology is more difficult at the level of nucleotide sequences, lack of sufficient coverage to the query in the above search does not necessarily suggest the absence of Hr in the two charophytes. Therefore, we further investigated whether the 2-3 charophyte hits were specific to Hr (i.e., the HHE domain). To this end, we performed the BLASTN search of NCBI non-redundant nucleotide sequence database (nt) using the same Marchantia polymorpha Hr (JGI Id: Mapoly0042s0093) as query. Indeed, the results included hits corresponding to those from the two charophytes (Nitella hyalina and Closterium peracerosum-strigosum-littorale complex) (see figure below, top panel). However, further inspections of these hits indicated that they were not particular to the Hr gene. For instance, the hits to the 670-600 bp region of the query were annotated as part of the genes encoding small integral membrane protein 10-like protein 2A, peroxidase 7-like protein, and myomegalin-like protein; they were found in both flowering plants (Lupinus angustifolius, Vitis venifera, and Cucurbita maxima) and animals (Crocodylus porosus, Oncorhynchus mykiss, and Columba livia). Similarly, hits to the 150-200 bp region of the query were annotated as genes encoding erythrocyte membrane protein, RP1 like 1 (rp111) protein, and retrotransposon Gag like 5 (Rtl5) protein; they could be found in animals (e.g., Oryzias latipes, Heterocephalus glaber, Labrus *bergylta*) and apicomplexan parasites (*Plasmodium falciparum*). On the other hand, the most significant hits, which also had the longest query coverages, were from Selaginella moellendorffii, the fungus Fusarium verticillioides and Physcomitrella patens (see figure below, lower panel). The former two were part of the Hr gene, whereas the later (i.e. P. patens) evolved directly from Hr. These data suggest that the hits from the two charophytes (Nitella hyalina and Closterium peracerosum-strigosum-littorale complex) might be associated with genes other than Hr.

### Job title: Mapoly0042s0093.1 CDS

RID	<u>3MTGYX9M015</u> (Expires on 12-21 20:31 pm)
Query ID	Icl]Query_54899
Description	Mapoly0042s0093.1 CDS
Molecule type	nucleic acid
<b>Query Length</b>	795

 Database Name
 nr

 Description
 Nucleotide collection (nt)

 Program
 BLASTN 2.7.1+ ▷ Citation

Other reports: > Search Summary [Taxonomy reports] [Distance tree of results] [MSA viewer]

### Graphic Summary



1	Alignments Download Y GenBank Graphics Distance tree of results						0
	Description	Max score	Total score	Query cover	E value	Ident	Accession
	Selaginella moellendorffii hypothetical protein partial mRNA	123	123	18%	6e-24	78%	XM 002981430.1
	Selaginella moellendorffii hypothetical protein partial mRNA	116	116	15%	9e-22	80%	XM 002969959.1
	Fusarium verticillioides 7600 hypothetical protein (FVEG 10078), partial mRNA	53.6	53.6	17%	0.009	69%	XM 018899141.1
	Physcomitrella patens subsp. patens predicted protein (PHYPADRAFT 151693) mRNA, partial cds	53.6	53.6	19%	0.009	68%	XM 001782780.1

The following includes uncropped gel images for corresponding Supplementary Figures above.



Uncropped gel of supplementary figure 6b



Uncropped gel of supplementary figure 10b



Uncropped gel of supplementary figure 10b



Uncropped gel of supplementary figure 10c



Uncropped gel of supplementary figure 13a



Uncropped gel of supplementary figure 13a