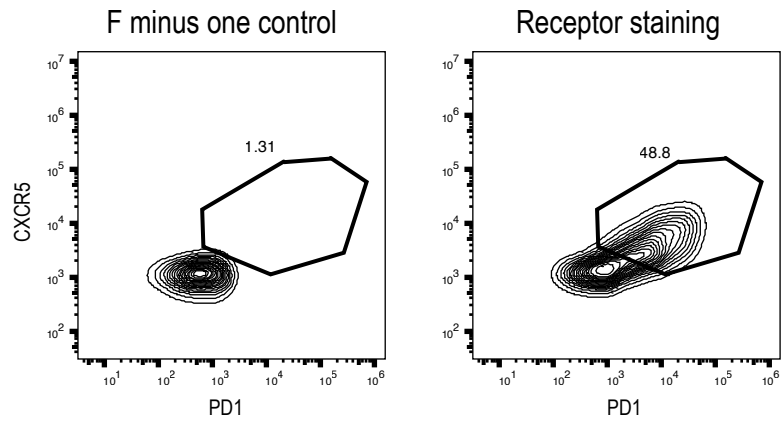
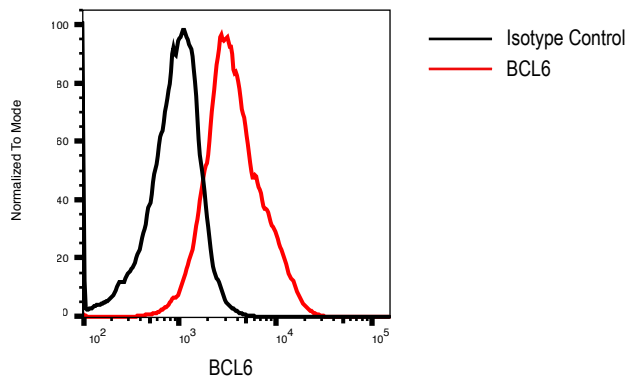


# Supplementary Figure 1

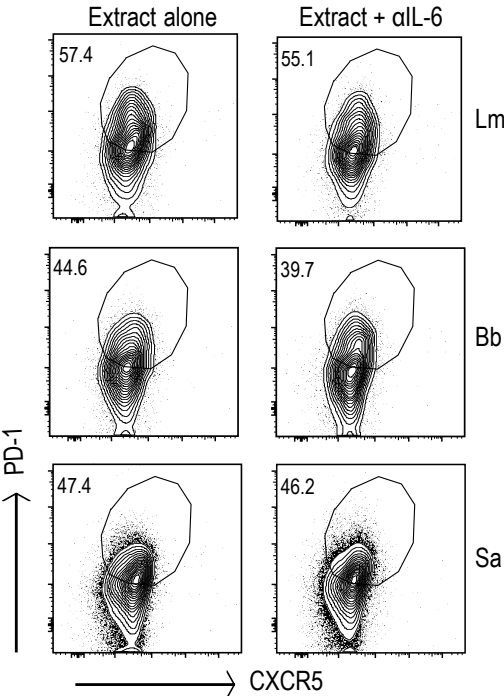
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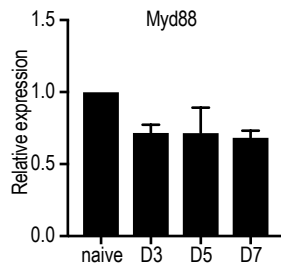
B



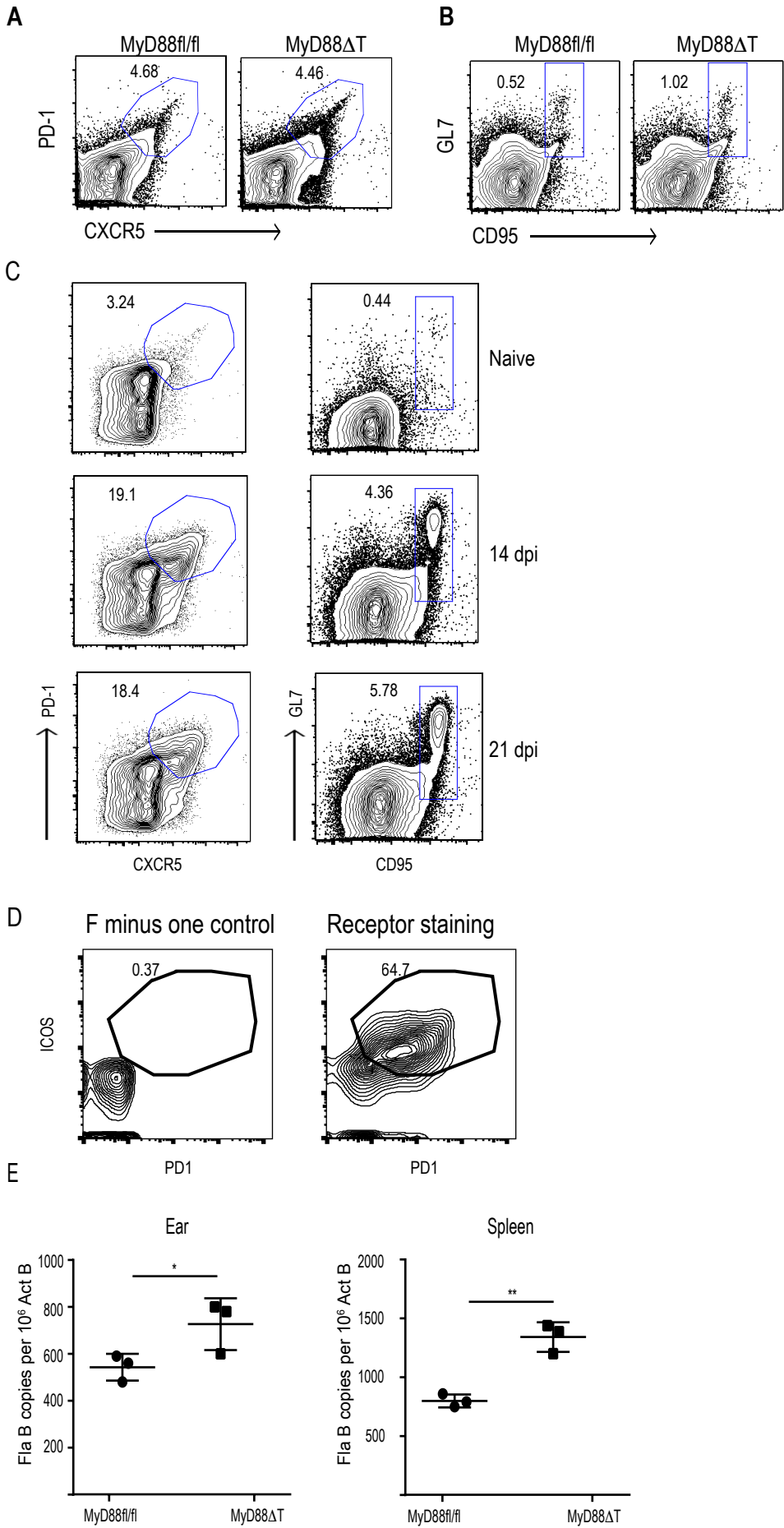
# Supplementary Figure 2



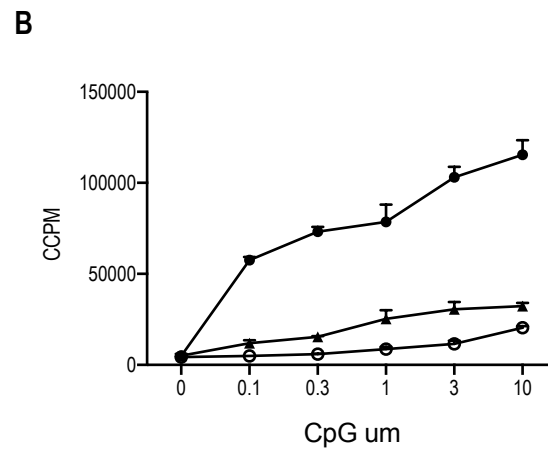
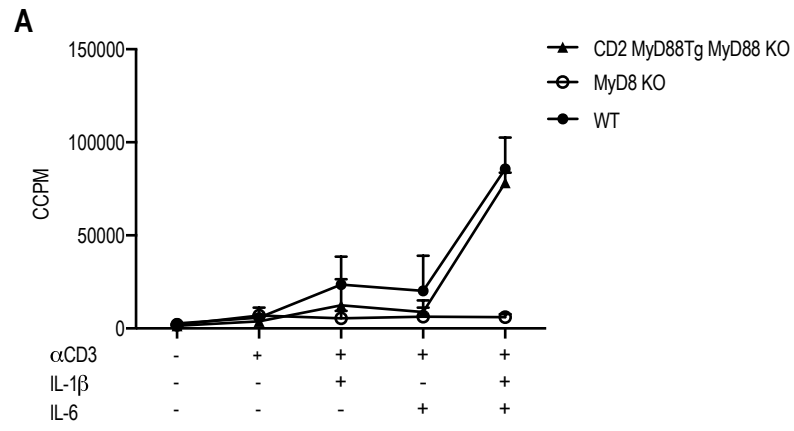
### Supplementary Figure 3



# Supplementary Figure 4



## Supplementary Figure 5



## Supplementary Figure Legends

### **Figure S1. Gating strategy for PD-1 and CXCR5 expression and BCL6 expression in activated CD4 T cells.**

(A) Cells are gated on CD4<sup>+</sup> T cells from Peyer's patches from a naïve mouse. (B) Sorted naïve WT OT-II cells (CD45.1) were transferred into congenic (CD45.1) immunocompetent mice ( $1 \times 10^5$  cells/mouse). The next day, mice were immunized with OVA+LPS emulsified in IFA subcutaneously in the hind footpads. Histograms show intracellular expression of BCL6 in donor CD4 T cells on day 7 post-immunization.

### **Figure S2. Neutralization of IL-6 does not affect the generation of Tfh lineage cells.**

Naïve CD4 T cells from WT mice were co-cultured with splenic DCs in the presence of *Lm* bacterial extract (10 $\mu$ g/ml) for 5 days in the presence or absence of  $\alpha$ IL-6 neutralizing antibody (10 $\mu$ g/ml). Flow plots show expression of PD-1 and CXCR5 on CFSE<sup>lo</sup> CD4<sup>+</sup> T cells. Data are representative of two independent experiments.

### **Figure S3. MyD88 expression in CD4 T cells during antigen specific differentiation**

WT naïve CD4 T cells were CFSE labelled and co-cultured with WT CD11c<sup>+</sup> splenic DCs in the presence of 10 $\mu$ g/ml of *Lm* extract. CFSE<sup>lo</sup> CD4 T cells were sorted on day 3, 5 and 7 post stimulation and relative expression of MyD88 was quantified by qPCR in. Data is normalized to 18s. Data are representative of two independent experiments. Error bars represent SEM.

**Figure S4. Intradermal infection of mice by *Borrelia Burdgorferi* leads to measurable and sustained Tfh and GC responses in the spleen that fail to confer protection in the absence of MyD88 in T cells.**

(A and B) Splens from MyD88<sup>fl/fl</sup> and MyD88ΔT littermate controls were harvested, processed, and analyzed for steady state proportions of (A) CD4<sup>+</sup> CXCR5<sup>+</sup> PD-1<sup>+</sup> Tfh cells and (B) CD19<sup>+</sup> CD95<sup>+</sup> GL7<sup>+</sup> GC B cells by flow cytometry. (C) WT mice were infected with low passage *Bb* strain 296 (10<sup>5</sup>/mice) intradermally. CXCR5<sup>+</sup> PD-1<sup>+</sup> and ICOS<sup>+</sup> Tfh cells and GL7<sup>+</sup> CD95<sup>+</sup> GC B cells in the spleen of infected mice at given time points. (D) Gating strategy for PD1 and ICOS expression on CD4<sup>+</sup> T cells from Peyer's patches from a naïve mouse. (E) MyD88<sup>fl/fl</sup> and MyD88ΔT mice were infected with *Bb* (10<sup>5</sup> CFU/mice) intradermally. Quantification of *Bb* burden in MyD88<sup>fl/fl</sup> and MyD88ΔT by qPCR in the ears and spleen.

**Figure S5. Functional characterization of CD2-MyD88 Tg mice on MyD88 deficient background.**

(A) Naive CD4 T cells from indicated strains were stimulated with plate bound anti-CD3ε and anti-CD28 in the presence or absence of IL-1β (20ng/ml) and IL-6 (20ng/ml) for 72 hours. After 60 hours 3H-thymidine was added to the cultures and cell proliferation was measured using a beta-counter. (B) Purified CD19<sup>+</sup> B cells were cultured in the presence of titrating concentrations of CpG ODNs for 72 hours and proliferation was measured by a 3H thymidine incorporation assay.