

Chronic β 2AR stimulation limits CFTR activation in human airway epithelia

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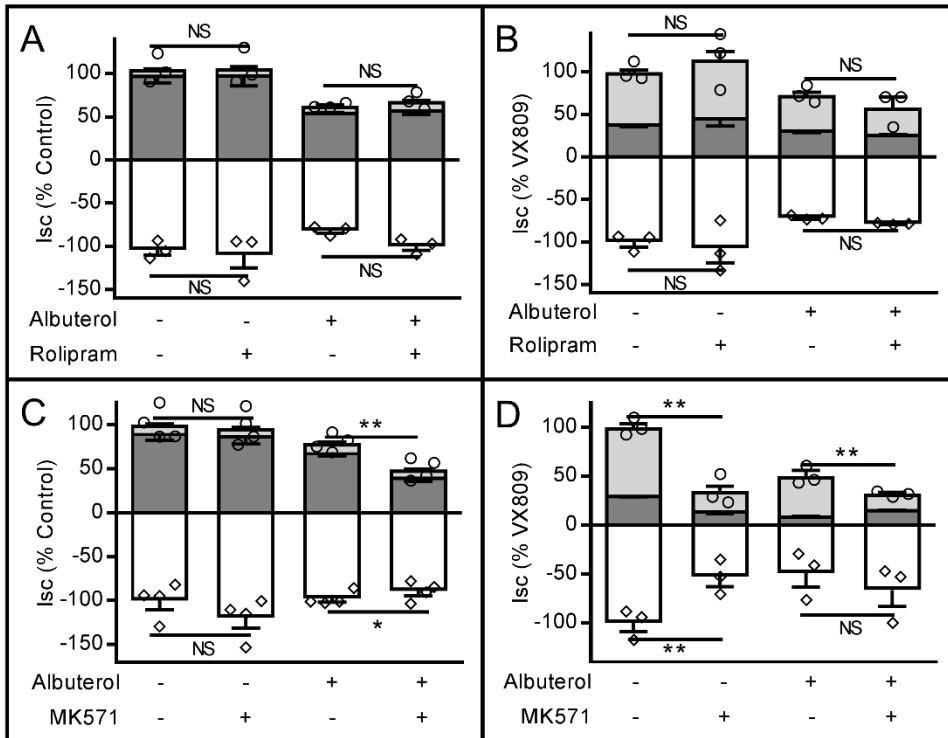


Figure S1:

Chronic albuterol-induced CFTR impairment is not rescued by functional inhibition of the key spatial cAMP regulators Phosphodiesterase, isoform 4 (PDE-4) and the multidrug resistance pump 4 (MRP4). Wild-type and VX809-corrected F508del CFTR+ CFBE41o- cells were exposed to albuterol (10 μ M) in media for 72 hours, then mounted in Ussing chambers and exposed to inhibitors of PDE-4 (rolipram, 100 μ M) or MRP4 (MK571, 20 μ M). CFTR function was then quantified under voltage clamp conditions. The addition of rolipram (100 μ M, added after forskolin/IBMX) to further inhibit PDE-4 did not rescue albuterol-induced CFTR dysfunction in either wtCFTR+ (**Panel A**, n=3 inserts/condition; circles represent total CFTR activity [cAMP+genistein], diamonds represent inhibited CFTR currents [Inh172]) or VX809-corrected F508del CFTR+ cells (**Panel B**, 3 inserts/condition). Similarly, the albuterol-induced reductions in CFTR-dependent I_{sc} in pretreated wtCFTR+ (**Panel C**, n=4 inserts/condition) and VX809-corrected F508del CFTR+ (**Panel D**, n=3 inserts/condition) CFBE41o- cells were not rescued following exposure to the MRP4 inhibitor MK571 (20 μ M, added before amiloride). All studies are internally normalized as indicated to allow for comparisons. Stimulation protocol was as follows: Amiloride (100 μ M, not shown), cAMP (forskolin 10 μ M/IBMX 100 μ M – dark grey bars), CFTR potentiator (genistein 50 μ M for wtCFTR+; VX770 1 μ M for F508del CFTR+ cells – light grey bars), and CFTR inhibition (Inh172 10 μ M – white bars). All experiments are representative of studies repeated in duplicate or triplicate with similar results. Data presented represent mean \pm SEM. *p<0.05; **p<0.01; NS: non-significant by two-way ANOVA with Tukey's multiple comparisons test.

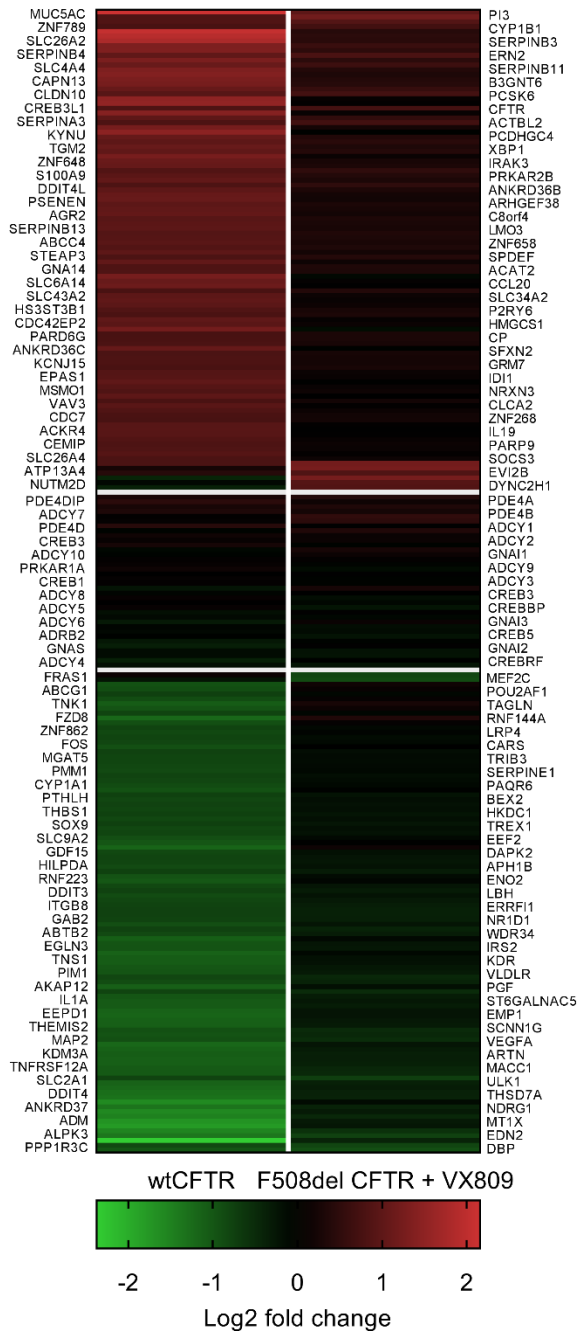


Figure S2:

Transcriptional changes in primary human airway epithelial cells (HAECs) following 72 hours of albuterol exposure.

Heatmap displaying differentially expressed genes in wtCFTR+ (left column) and VX809-corrected F508del CFTR+ primary HAECs following chronic albuterol exposure; each experimental group is compared to its same-donor baseline (untreated wtCFTR+ on the left, VX809-treated F508del CFTR+ on the right). The top 100 significantly up- or down-regulated genes are listed in the top and bottom groups, respectively; expression of genes in these columns was statistically changed with albuterol exposure, though not all reach a 2-fold change. The center group represents key genes of interest (AC isoforms, *CREB*, etc.); no genes in the center row had a significant change in expression. Key genes are noted to the left or right of their respective row (left/right placement is for spacing only and does not carry significance).

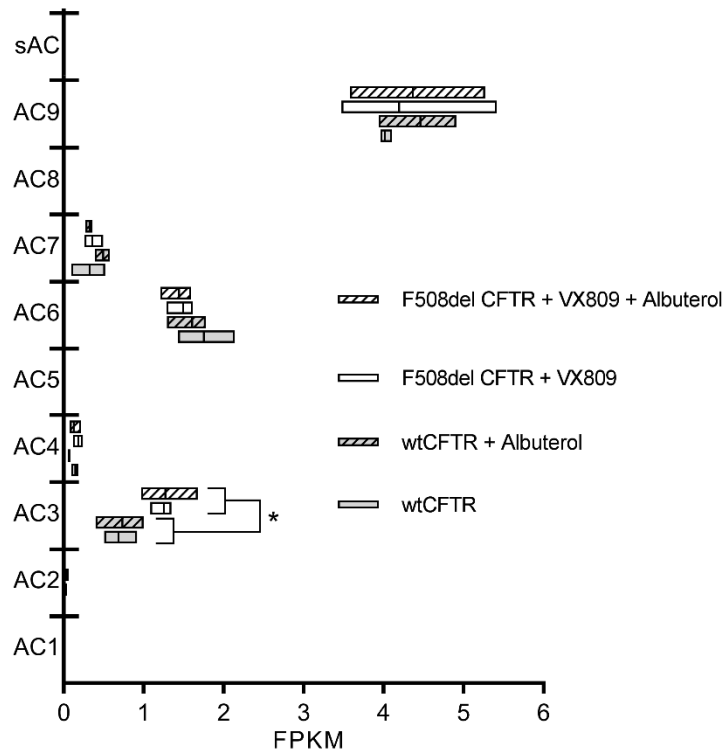


Figure S3:

Expression patterns of Adenylyl Cyclase (AC) isoforms are similar between wild-type and F508del CFTR+ human airway epithelial cell (HAEC) donors, and are not modified by albuterol pre-treatment.

Expression levels, as measured in Fragments per Kilobase of transcript per Million mapped reads (FPKM), of the nine transmembrane AC isoforms (AC1-AC9) and soluble AC (sAC) in albuterol-treated and untreated wt and VX809-corrected F508del CFTR+ primary HAECs. The isoforms of highest expression include AC3, AC6, AC7, and AC9, and expression levels are unchanged by albuterol exposure. Expression of AC3 was slightly higher in VX809-corrected F508del CFTR+ cells compared to wtCFTR+ cells; all other isoforms were similarly expressed in both cell lines.

* $p < 0.05$ by two-way ANOVA with Tukey's multiple comparisons test.

Supplemental Tables and Table Legends

Primary Antibody	Secondary Antibody
Rabbit anti-CFTR (Sigma C7491); 1:100	FitC Goat anti-Rabbit IgG (Invitrogen 656111); 1:500
Alexa Fluor 568 Phalloidin (Life Technologies A12380); 1:250	N/A
Chicken anti- β 2AR (Abcam ab13989); 1:250	Goat anti-Chicken IgY (Abcam ab150171); 1:500

Table S1:

Antibodies and fluorophores used for immunofluorescence.