

# Low Resting Membrane Potential and Low Inward Rectifier Potassium Currents Are Not Inherent Features of hiPSC-Derived Cardiomyocytes

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## SUMMARY

Human induced pluripotent stem cell (hiPSC) cardiomyocytes (CMs) show less negative resting membrane potential (RMP), which is attributed to small inward rectifier currents ( $I_{K1}$ ). Here,  $I_{K1}$  was measured in hiPSC-CMs (proprietary and commercial cell line) cultured as monolayer (ML) or 3D engineered heart tissue (EHT) and, for direct comparison, in CMs from human right atrial (RA) and left ventricular (LV) tissue. RMP was measured in isolated cells and intact tissues.  $I_{K1}$  density in ML- and EHT-CMs from the proprietary line was similar to LV and RA, respectively.  $I_{K1}$  density in EHT-CMs from the commercial line was 2-fold smaller than in the proprietary line. RMP in EHT of both lines was similar to RA and LV. Repolarization fraction and  $I_{K,ACh}$  response discriminated best between RA and LV and indicated predominantly ventricular phenotype in hiPSC-CMs/EHT. The data indicate that  $I_{K1}$  is not necessarily low in hiPSC-CMs, and technical issues may underlie low RMP in hiPSC-CMs.

## INTRODUCTION

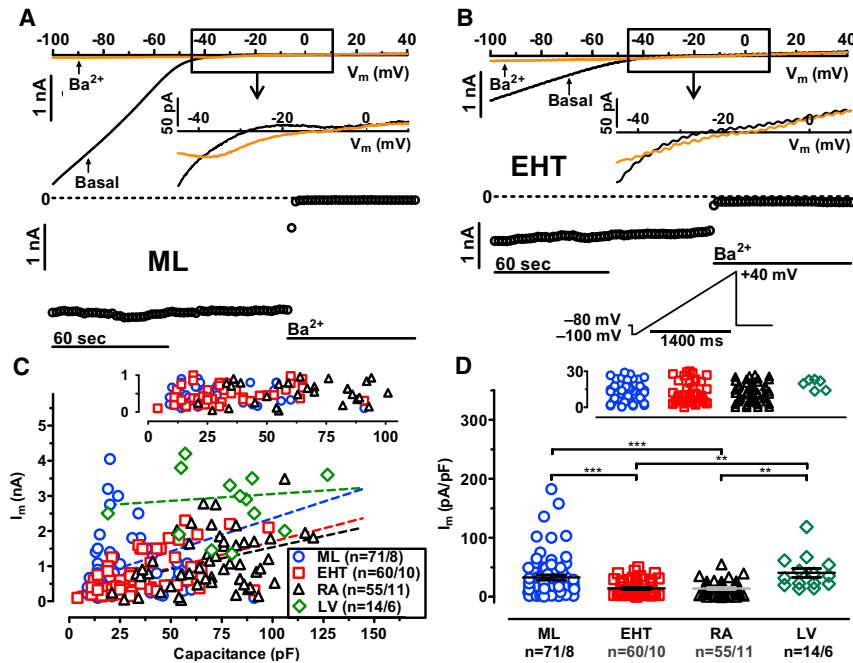
Human induced pluripotent stem cell-derived cardiomyocytes (hiPSC-CMs) are promising tools for cardiac repair (Weinberger et al., 2016), disease modeling (Burridge et al., 2011; Itzhaki et al., 2011; Liang et al., 2013, 2016; Moretti et al., 2010; Yazawa et al., 2011; Zhang et al., 2014), and cardiovascular drug testing (Eder et al., 2014; Kijlstra et al., 2015; Liang et al., 2013; Lu et al., 2015; Ma et al., 2011; Maddah et al., 2015). However, there is concern that hiPSC-CMs differ in their electrophysiological properties from human adult CMs. Studies with patch-clamp electrodes consistently reported low resting membrane potentials (RMPs) in hiPSC-CMs compared with adult atrial or ventricular myocardium (Chen et al., 2017; Davis et al., 2012; Doss et al., 2012; Herron et al., 2016; Karakikes et al., 2015; Liang et al., 2013, 2016; Ma et al., 2011; Vaidyanathan et al., 2016). This is an alarming finding since correct RMP is mandatory for excitability and refractoriness. One of the possible explanations for a slightly negative RMP reported in hiPSC-CMs is related to the inward rectifier  $K^+$  current ( $I_{K1}$ ). This current maintains the stable RMP in adult CMs (Hibino et al., 2010). In line with this assumption, current density of  $I_{K1}$  were reported to be low, or almost absent, in hiPSC-CM cell lines (Doss et al., 2012; Herron et al., 2016; Ma et al., 2011). Consequently, sophis-

ticated electronic approaches based on dynamic patch clamping or overexpression of channel subunits of  $I_{K1}$  were proposed to compensate low current density in hiPSC-CMs (Meijer van Putten et al., 2015; Vaidyanathan et al., 2016). On the other hand, capacitance of hiPSC-CMs is smaller compared with adult CMs, and patch-clamp-based determination of the membrane potential in small cells has been associated with methodological problems (Amos et al., 1996; Wilson et al., 2011). These findings raise the hypothesis that part of the reported differences between hiPSC-CM and adult human heart electrophysiology are in fact methodically in nature.

## RESULTS

### hiPSC-CMs Show Robust Inward Rectifier Potassium Currents

To detect inward rectifier currents, a classical ramp protocol was applied (Figure 1B, inset). High extracellular potassium concentration (20 mM) was used to evoke large inward currents even at moderately negative test pulse potentials, but also to facilitate larger outward currents (Figures 1A and 1B) (Anumonwo and Lopatin, 2010). It is also reported that applying higher external  $K^+$  concentration can reduce leakage and improve the stability of the measurements



**Figure 1. Cell Size and Inward Rectifier Potassium Current in hiPSC-CMs from ML, EHT, RA, and LV**

(A and B) Original traces of inward rectifier currents ( $I_{K1}$ ) and time courses of current at  $-100$  mV in hiPSC-CMs obtained from ML and EHT exposed to  $1$  mM  $Ba^{2+}$ . Outward component of  $I_{K1}$  and voltage protocol is given as inset in (B).

(C)  $Ba^{2+}$ -sensitive  $I_{K1}$  current amplitudes measured at  $-100$  mV plotted against cell capacitance; dotted lines indicate linear regression fit ( $R^2$  values were  $0.1$  for ML,  $0.28$  for EHT,  $0.04$  for iCell-EHT,  $0.15$  for RA, and  $0.007$  for LV). The upper scale shows data points with lower current amplitude.

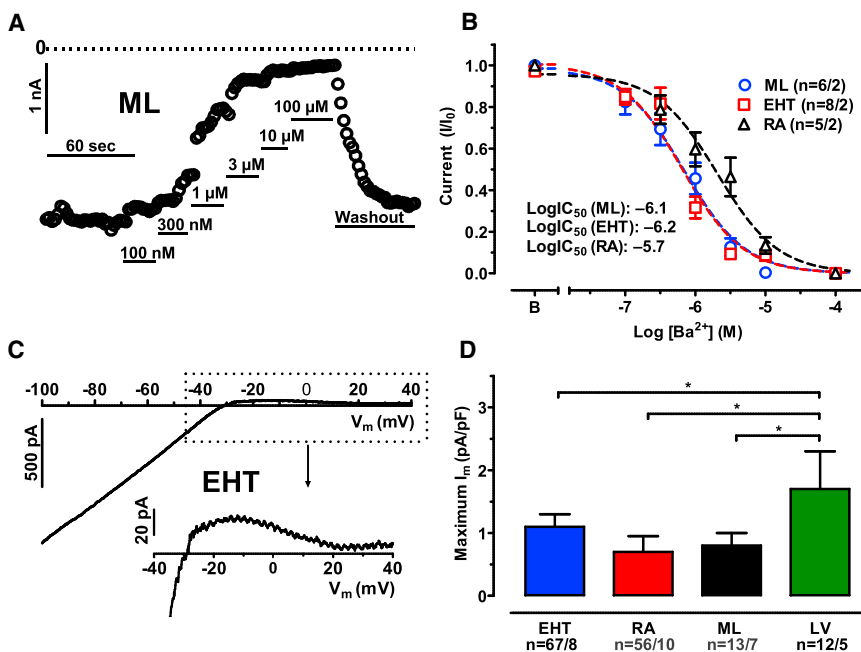
(D) Individual data points and respective mean values  $\pm$  SEM for  $Ba^{2+}$ -sensitive  $I_{K1}$  current densities measured at  $-100$  mV in hiPSC-CMs from ML, EHT, and in CMs from RA and LV. The upper scale shows data points with lower current density.  $**p < 0.01$ ,  $***p < 0.001$  (one-way ANOVA followed by Bonferroni test). n/n, number of experiments/number of isolations in hiPSC-CMs and number of experiments/number of patients in RA and LV.

(Wilson et al., 2011). We found clear evidence for inwardly rectifying currents not only in every adult CM (left ventricular [LV] and right atrial [RA]), but also in every hiPSC-CM from both monolayer (ML) and engineered heart tissue (EHT) (Figure 1C). There was a large variability in current size, with some currents close to zero, not only in ML and EHT, but also in RA (Figure 1C). However, we found in every cell from RA, LV, ML, and EHT an instantaneous block of inward current in response to  $Ba^{2+}$  ( $1$  mM). To investigate whether larger cells show a more mature phenotype with larger  $I_{K1}$ , we plotted individual inward current amplitudes versus cell size (Figure 1C). As shown before (Liang et al., 2013; Uzun et al., 2016), hiPSC-CMs, isolated from ML or EHT, were smaller than RA and LV CMs ( $30.8 \pm 1.5$  pF,  $n = 167$  in ML,  $47.0 \pm 1.7$  pF,  $n = 260$  in EHT,  $73.6 \pm 4.0$  pF,  $n = 55$  in RA,  $88.8 \pm 13.2$  pF,  $n = 15$  in LV). However, there was only a weak positive association between current amplitude and cell membrane capacitance. The steepness of the regression lines were low in all groups, and did not differ between hiPSC-CMs and human adult CMs (Figure 1C,  $p = 0.6$ , one-way ANOVA). Statistical significance for slope different from zero was only determined in ML, EHT, and RA ( $R^2$  values were  $0.09$  for ML,  $0.28$  for EHT, and  $0.14$  for RA). This argues against the assumption that smaller hiPSC-CMs represent a more immature phenotype with smaller  $I_{K1}$  current amplitude. To facilitate comparison with other publications, we present current amplitudes

normalized to cell size. Current densities in ML were not significantly smaller than in LV (Figure 1D; Table S1). Current density of  $I_{K1}$  in EHT was smaller than in ML and LV, but still reached the values of RA (Figure 1D; Table S1). Thus,  $I_{K1}$  current densities in hiPSC-CMs were not lower than in human adult CMs, when identical patch-clamp protocols were applied.

### $I_{K1}$ in hiPSC-CMs Is Conducted by Highly $Ba^{2+}$ -Sensitive Kir Channels

The  $I_{K1}$  conducting channel exists as tetramer, assembled from different  $\alpha$  subunits (Kir2.1–2.4; for review see Hibino et al., 2010). Different channel-forming subunits of  $I_{K1}$  show different sensitivity to  $Ba^{2+}$  (Liu et al., 2001; Schram et al., 2003). Heart muscle expresses Kir2.1, 2.2, and 2.3, which show high sensitivity to  $Ba^{2+}$  (in the low  $\mu$ M range) (Schram et al., 2003). Kir2.4 exhibits lower  $Ba^{2+}$  sensitivity and is expressed in neuronal tissue only (Liu et al., 2001). To elucidate whether  $I_{K1}$  in hiPSC-CMs is conducted by the cardiac subunits, we measured concentration-response curves for  $Ba^{2+}$  block on the inward  $I_{K1}$  current (Figure 2A). We observed a monophasic concentration-response curve, arguing against the contribution of Kir2.4 (Figure 2B).  $I_{K1}$  in hiPSC-CMs from both culture conditions showed higher  $Ba^{2+}$  sensitivity than in RA: the  $\log IC_{50}$  values for  $Ba^{2+}$  were  $-6.09$  (95% confidence interval [CI]:  $-6.27$  to  $-5.91$ ) in ML, versus  $-6.15$  (CI:  $-6.32$  to  $-5.99$ ) in



**Figure 2. Ba<sup>2+</sup> Sensitivity and Outward Component of I<sub>K1</sub> in hiPSC-CMs and Human Adult CMs**

(A) Time course of inward currents measured at -100 mV in a hiPSC-CMs from ML exposed to increasing concentrations of Ba<sup>2+</sup>.

(B) Concentration-response curves for Ba<sup>2+</sup> on inward current at -100 mV in hiPSC-CMs from ML, EHT, and in CMs from RA. Mean values ± SEM.

(C) Ba<sup>2+</sup>-sensitive current trace from hiPSC-CMs from ML. Outward component of current given on an extended scale at the bottom.

(D) Mean values ± SEM of maximum outward peak currents in hiPSC-CMs from ML, EHT, and in CMs from RA and LV. \*p < 0.05 (one-way ANOVA followed by Bonferroni test) n/n = number of experiment/number of isolations in hiPSC-CMs and number of experiment/number of patients in RA and LV.

EHT, versus -5.66 in RA (CI: -5.92 to -5.41, p < 0.01, F test for ML versus RA and for EHT versus RA; Figure 2B). The qRT-PCR data confirmed that the cardiac isoforms of the Kir channels (2.1–2.3) are expressed in hiPSC-CMs (Figures S1A–S1C), while Kir2.4 is not (Figure S1D).

### I<sub>K1</sub> in hiPSC-CMs Shows Small Outward Contribution

In cardiac myocytes, the outward component of I<sub>K1</sub> contributes to the late phase of the repolarization. There is no simple relationship between the size of inward and outward currents: the relative outward contribution of I<sub>K1</sub> is larger in ventricular than in atrial CMs from human hearts (Amos et al., 1996; Koumi et al., 1995; Varró et al., 1993; Wang et al., 1998). The outward component of I<sub>K1</sub> is generally larger in rabbit and canine than in human (Amos et al., 1996; Jost et al., 2013; Koumi et al., 1995; Major et al., 2016; Varró et al., 1993; Wang et al., 1998). Overall, contribution of I<sub>K1</sub> to overall repolarization in human heart is small and restricted to the late phase of action potential (AP) (Jost et al., 2013). Therefore, we measured the maximum outward current density of I<sub>K1</sub> during ramp pulses (Figure 2C). As shown before, in human CMs, outward currents conducted via I<sub>K1</sub> were small compared with inward currents and smaller in human atrial than in ventricular CMs (0.7 ± 0.25 pA/pF versus 1.7 ± 0.6 pA/pF, n = 16 and 13; Figure 2D). Outward currents in hiPSC-CMs were as small as in RA (1.1 ± 0.2 pA/pF, n = 67 in ML, 0.8 ± 0.2 pA/pF, n = 56 in EHT; Figure 2D). Thus, hiPSC-CMs from both ML and EHT showed small outward contribution of I<sub>K1</sub> as known for human adult CMs. The small contribution of I<sub>K1</sub> to repolarization

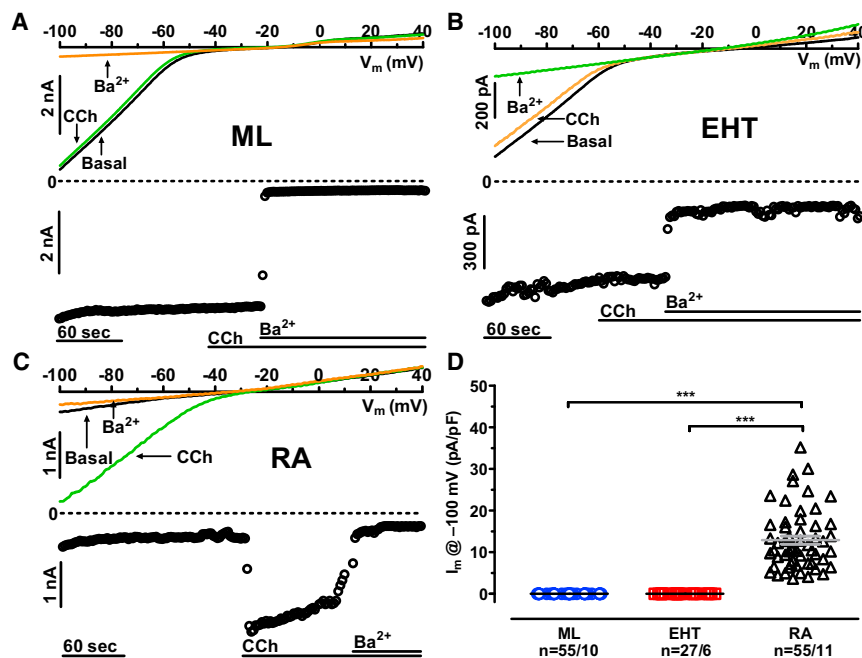
may favor stronger action potential duration (APD) prolongation upon block of I<sub>Kr</sub> (Jost et al., 2013). Therefore, prediction of QT prolongation in human by hERG blockers could be more meaningful, when hiPSC-CMs are used instead of canine or rabbit. Further studies are needed to investigate whether hiPSC-CMs quantitatively reflect the repolarization reserve in human heart.

### hiPSC-CMs Do Not Express Acetylcholine-Activated Potassium Currents

The existence of acetylcholine-activated potassium currents (I<sub>K,ACh</sub>) is a hallmark of atrial tissue (Dobrzynski et al., 2001). To assess the specification of hiPSC-CMs, we applied the muscarinic (M2) receptor agonist carbachol ([CCh], 2 μM; Figure 3A). In RA CMs, CCh activated a large inward current (Figure 3C), which was absent in all hiPSC-CMs (ML and EHT; Figures 3A and 3C). On the transcript level, we found large expression of Kir3.1 in RA, but not in LV, ML, or EHT (Figure S1). Interestingly, Kir3.4 was expressed in hiPSC-CMs under both culture conditions at least as high as in RA (Figure S1). The latter finding implies that expression of Kir3.4 in hiPSC-CMs alone is not sufficient to generate I<sub>K,ACh</sub> (Krapivinsky et al., 1995). The lack of I<sub>K,ACh</sub> indicates that hiPSC-CMs did not exhibit an atrial phenotype.

### RMP and AP Measurements in Single Cells and Intact Tissues

To decide whether the relatively normal I<sub>K1</sub> densities observed in hiPSC-CMs result in physiological RMP, we measured action potential in isolated CMs by patch-clamp



### Figure 3. Lack of Carbachol Effect on Inward Rectifiers in hiPSC-CMs

(A and B) Original traces of inward rectifier currents and time courses of current at  $-100 \text{ mV}$  in hiPSC-CMs obtained from ML and EHT exposed to carbachol ( $[\text{CCh}], 2 \mu\text{M}$ ). (C) Effect of CCh ( $2 \mu\text{M}$ ) on  $I_{K, \text{ACh}}$  in CMs from RA.

(D) CCh effects expressed as absolute current change in response to CCh ( $2 \mu\text{M}$ ) in ML, EHT, and in CMs from RA. Individual data points and mean values  $\pm$  SEM. LV \*\*\*  $p < 0.001$  (one-way ANOVA followed by Bonferroni test). n/n = number of experiment/number of isolations in hiPSC-CMs and number of experiment/number of patients in RA.

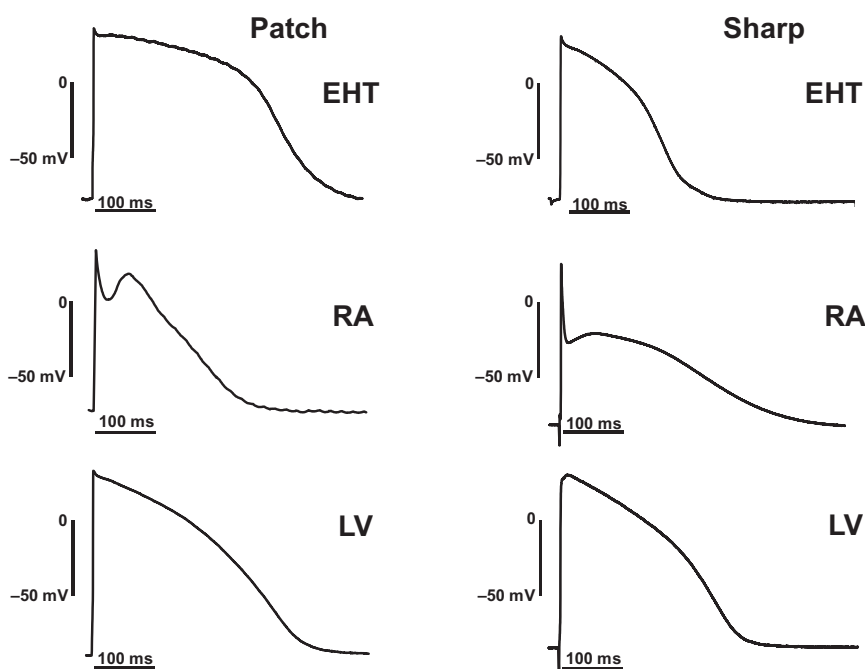
technique (Figure 4, left column). RMP measured by patch electrodes in hiPSC-CMs was low ( $-39.3 \pm 6.1 \text{ mV}$ ,  $n = 12$ ). Application of holding currents was necessary to elicit stable AP in most of the hiPSC-CMs (40 out of 41 in EHT). The amount of holding current was in the range of  $0.2 \text{ nA}$ . AP could be elicited from a relatively low RMP in EHT ( $-59.7 \pm 1.2 \text{ mV}$ ,  $n = 41$ ). In contrast, AP could be recorded in human adult CMs from RA and LV without any holding current, and, in both, RMP was significantly more negative than in EHT. Respective RMP amounted to  $-74.4 \pm 0.5 \text{ mV}$  in RA ( $n = 49$ ) and to  $-75.9 \pm 1.1 \text{ mV}$  in LV ( $n = 10$ , Figure S2B, upper panel; Table S3).

The sharp microelectrode technique represents the gold standard to measure AP in multicellular preparations. To determine whether the apparent discrepancy between  $I_{K1}$  density and RMP in hiPSC-CMs could be related to methodological issues, we compared data from EHT with a larger number of recordings measured in intact RA and LV preparations (Figures 4, right column, 5C, and 5D). The majority of data was collected over many years at the Department of Pharmacology and Toxicology at the Medical Faculty of Dresden University of Technology. All EHTs were beating spontaneously, slightly slower than  $60 \text{ bpm}$ , which allowed us to record stimulated AP at  $1 \text{ Hz}$ . RMP in sharp microelectrode recordings was slightly, but significantly less negative in RA than in LV ( $-73.3 \pm 0.3 \text{ mV}$ ,  $n = 220$  versus  $-75.9 \pm 0.7 \text{ mV}$ ,  $n = 57$ ;  $p < 0.001$ , Figure S2B, lower panel; Table S3). RMP in intact EHT was in between ( $-74.6 \pm 1.2 \text{ mV}$ ,  $n = 24$ ) not significantly different from RA and LV (Figure S2B, lower panel; Table S3). At this point,

it should be emphasized that  $I_{K1}$  density alone does not determine RMP but the relation of all conductances present near RMP. Two other parameters, action potential duration at 90% repolarization ( $\text{APD}_{90}$ ) and repolarization fraction – calculated as  $(\text{APD}_{90} - \text{APD}_{50})/\text{APD}_{90}$  – were recently proposed as a possible approach to distinguish between atrial- and ventricular-like hiPSC-CMs (Du et al., 2015). It should be noted that impact of  $I_{K1}$  on  $\text{APD}_{90}$  in human LV is very small and repolarization fraction dominated by transient potassium outward currents. Therefore, both parameters are probably rather independent of  $I_{K1}$  (Jost et al., 2013). Nevertheless, we aimed to determine if adult human cardiac tissue from RA and LV can be classified correctly and, more importantly, whether hiPSC-CMs represent atrial- or ventricular-like phenotype or a mixture of both.

When measured by patch electrodes,  $\text{APD}_{90}$  was shorter in RA than in LV ( $220 \pm 16 \text{ ms}$ ,  $n = 41$  versus  $434 \pm 39 \text{ ms}$ ,  $n = 10$ ;  $p < 0.001$ ) and even shorter in hiPSC-CMs ( $119 \pm 17 \text{ ms}$ ,  $n = 41$ ;  $p < 0.001$  versus RA  $p < 0.01$  versus LV; Figure S2A, upper panel; Table S2). However, in these two parameters, there was substantial overlap of individual data points between RA and LV (Figures 5A and 5B).

Using sharp microelectrodes,  $\text{APD}_{90}$  scatter was much lower than in patch-clamp recordings (Figures S2A and 5C). The mean  $\text{APD}_{90}$  was shorter in RA than in LV ( $317 \pm 3 \text{ ms}$ ,  $n = 220$  versus  $334 \pm 6 \text{ ms}$ ,  $n = 57$ ;  $p < 0.05$ ; Figure S2A, lower panel; Table S2). Again,  $\text{APD}_{90}$  was clearly shorter in EHT ( $271 \pm 11.4 \text{ ms}$ ,  $n = 24$ ;  $p < 0.001$  versus RA and  $p < 0.001$  versus LV; Figure S2A, lower panel; Table S2). Next we retrospectively evaluated whether differences in



**Figure 4. Original AP Recordings Taken from EHT, RA, and LV**

Action potential recorded by patch-clamp (left) and sharp microelectrode (right) techniques. The patch-clamp recordings were measured from isolated CMs, while the sharp microelectrode recordings were measured from intact muscle preparations.

repolarization fraction would be a useful parameter to differentiate between LV and RA. As shown in [Figure S2C](#) (bottom panel), a narrow distribution of repolarization fraction was found in AP measurements from intact muscle preparations. There was almost no overlap between LV and RA, indicating the usefulness of the approach ([Figure 5D](#)). Repolarization fraction of intact EHTs was similar to LV and differed significantly from RA (EHT:  $0.32 \pm 0.01$ ,  $n = 24$  versus RA:  $0.54 \pm 0.01$ ,  $n = 220$  versus LV:  $0.28 \pm 0.01$ ,  $n = 57$ ; [Figure S2C](#), bottom panel; [Table S4](#)). In contrast to sharp microelectrode recordings, measurements of repolarization fraction in individual CMs showed a wide range of distribution ([Figures S2C](#), upper panel, and [5B](#)).

#### Robust RMP in Our hiPSC-CMs: A Peculiarity of a Single-Cell Line?

We were concerned that normal RMP may be a peculiarity of our proprietary hiPSC-CMs. Therefore we measured  $I_{K1}$  densities and AP characteristics in a commercially available cell line (iCell, Cellular Dynamics International, Madison, WI, USA; [Figures 6A](#) and [6B](#)). Capacitance of cells was not different from EHT cast from C25 ( $41.4 \text{ pF} \pm 3.7 \text{ pF}$ ,  $n = 32$  in iCell-EHT). However, inward  $I_{K1}$  density was significantly smaller ( $7.8 \pm 1.8 \text{ pA/pF}$ ,  $n = 32$ ;  $p < 0.05$ ). Nevertheless, by applying sharp microelectrodes, we found a physiological RMP similar to EHT from C25 line ( $-74.3 \pm 0.1 \text{ mV}$ ,  $n = 8$  for iCell-EHT versus  $-74.6 \pm 1.2 \text{ mV}$ ,  $n = 24$  for C25-EHT). The data suggest that  $I_{K1}$  densities may differ in CMs obtained from different hiPSC lines, but this does not necessarily result in a less negative RMP. Repolarization

fraction of iCell-EHT was even lower than LV ( $0.2 \pm 0.01$ ,  $n = 8$  in iCell-EHT versus  $0.28 \pm 0.01$ ,  $n = 57$ ;  $p < 0.05$  in LV).

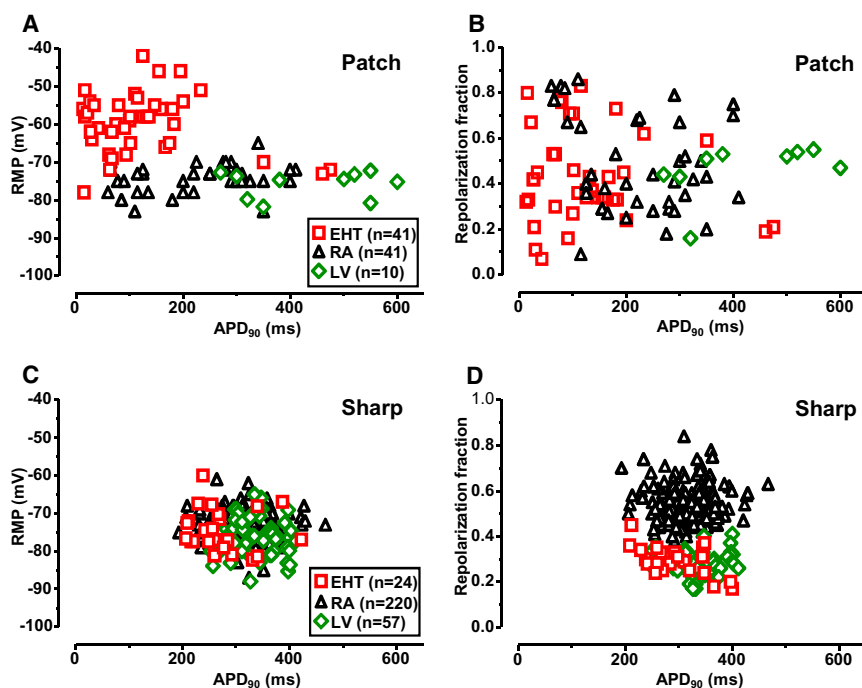
## DISCUSSION

In this study, we directly compare  $I_{K1}$  density, RMP, and AP properties in hiPSC-CMs and human atrial and ventricular CMs/tissues under identical experimental conditions. The main findings are (1) that inward  $I_{K1}$  current densities and RMP were similar in hiPSC-CMs and human CMs/tissues, (2) that  $\sim 2$ -fold differences in inward  $I_{K1}$  density between CMs from a proprietary and commercial hiPSC line did not translate in significant differences of RMP, and (3) that sharp microelectrode measurements (of intact 3D heart muscle preparations) may provide more reliable data on RMP and repolarization fraction than patch-clamp recordings (of isolated cells). As a side result, in sharp microelectrode recordings repolarization fraction differentiated much better between atrial and ventricular tissue than RMP or APD.

#### $I_{K1}$ in hiPSC-CMs and in Adult Human Atrial and Ventricular CMs

$I_{K1}$  density is known to be larger in ventricular than in atrial CMs ([Amos et al., 1996](#); [Koumi et al., 1995](#); [Varró et al., 1993](#); [Wang et al., 1998](#)) and very low or absent in nodal cells ([Guo et al., 1997](#); [Schram et al., 2002](#); [Tamargo et al., 2004](#)). We found the inward component of  $I_{K1}$  to be 2.5-fold larger in LV than in RA CMs. The difference is





**Figure 5. Individual Distribution of APD<sub>90</sub>, RMP, and Repolarization Fraction in EHT, RA, and LV**

(A and C) Individual data points of APD<sub>90</sub> plotted versus respective RMP values measured by patch clamping of isolated CMs from EHT, RA, and LV (A) and by sharp microelectrodes in intact tissues from EHT, RA, and LV (C).

(B and D) Individual data points of repolarization fraction versus APD<sub>90</sub> in isolated CMs: from EHT, RA, and LV (B); and in intact tissues: EHT, RA, and LV (D).

smaller than reported in some studies (Koumi et al., 1995; Wang et al., 1998), but well in line not only with the report (Varró et al., 1993; small sample size), but also with the largest study published to date (Amos et al., 1996). The observation that our proprietary hiPSC-CMs showed similar inward I<sub>K1</sub> densities as human CMs was unexpected, given that earlier publications reported low I<sub>K1</sub> in hiPSC-CMs (Doss et al., 2012; Herron et al., 2016; Ma et al., 2011). Yet, several data argue that true cardiac I<sub>K1</sub> current was measured. (1) Our data in native human CMs closely reflect reference values as stated above. (2) High Ba<sup>2+</sup> sensitivity and normal transcript levels of the cardiac ion channel subunits indicate normal I<sub>K1</sub>. (3) The small outward contribution and the absence of a CCh response in hiPSC-CMs (as well as in LV CMs) argues against contribution of I<sub>K,ACh</sub>, an atrial-specific potassium current. (4) We could reproduce relatively low I<sub>K1</sub> values in the commercially available iCells measured previously (Ma et al., 2011). Thus, we are confident that the data reflect cardiac I<sub>K1</sub> in hiPSC-CMs.

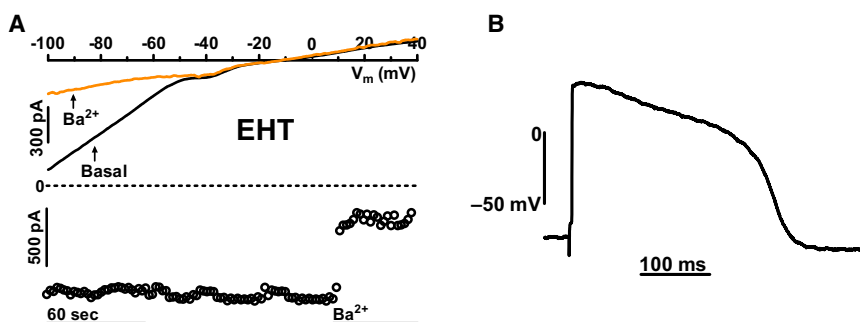
I<sub>K1</sub> density in hiPSC-CMs from ML was as high as in human LV CMs. In hiPSC-CMs isolated from EHT, the current density only reached the lower values of RA. As culture of hiPSC-CMs in EHT leads to signs of advanced maturation, this was an unexpected finding, while it was shown before that culturing hiPSC-CMs on different platforms (for example, polydimethylsiloxane) can increase inward I<sub>K1</sub> density (Herron et al., 2016; Lemoine et al., 2017; Manhardt et al., 2016). The reasons are not clear at present.

### RMP in hiPSC-CMs and in Adult Human Atrial and Ventricular CMs

In line with previous publications we found less negative RMP in isolated hiPSC-CMs, measured by patch-clamp electrodes (Chen et al., 2017; Davis et al., 2012; Doss et al., 2012; Herron et al., 2016; Karakikes et al., 2015; Liang et al., 2013; Ma et al., 2011; Vaidyanathan et al., 2016). In contrast, RMP reached the physiological range when measured in intact EHT by sharp microelectrodes. This discrepancy raised the question whether RMP measurements with patch-clamp electrodes would give a systematic error. The reliability of patch-clamp recordings critically depends on seal resistance (in the range of 1–10 GΩ). The remaining leak current is expected to reduce the actual membrane voltage (Amos et al., 1996; Schneider and Chandler, 1976). The corrected membrane potential (V<sub>CM</sub>) can be calculated from the actual seal resistance (R<sub>seal</sub>), the membrane resistance (R<sub>M</sub>), and the membrane potential, measured during the experiment (V<sub>mM</sub>):

$$V_{CM} = V_{mM} + V_{mM} * R_M/R_{seal}$$

If R<sub>seal</sub> is considered in series with R<sub>M</sub>. R<sub>seal</sub> in our experiments was between 1 and 10 GΩ, both in adult CMs and in hiPSC-CMs. R<sub>M</sub> at the RMP is determined by conductivity via I<sub>K1</sub>. To get an estimate of R<sub>M</sub> generated by I<sub>K1</sub> under the same experimental conditions used for AP recordings, we measured the barium-sensitive I<sub>K1</sub> (1 mM) at physiological [K<sup>+</sup>]<sub>ext</sub> (5.4 instead of 20 mM) and at physiological temperature (37°C) in hiPSC-CMs from EHT. As expected,



**Figure 6. Inward Rectifier Potassium Current and Action Potential in hiPSC-CMs from iCell-EHT**

(A) Original traces of inward rectifier currents ( $I_{K1}$ ) and time courses of current at  $-100$  mV in hiPSC-CMs obtained from iCell-EHT exposed to  $1$  mM  $Ba^{2+}$ .

(B) Action potential recorded from iCell-EHT by sharp microelectrode technique. The recordings were measured from intact EHT preparations at  $37^{\circ}C$ .

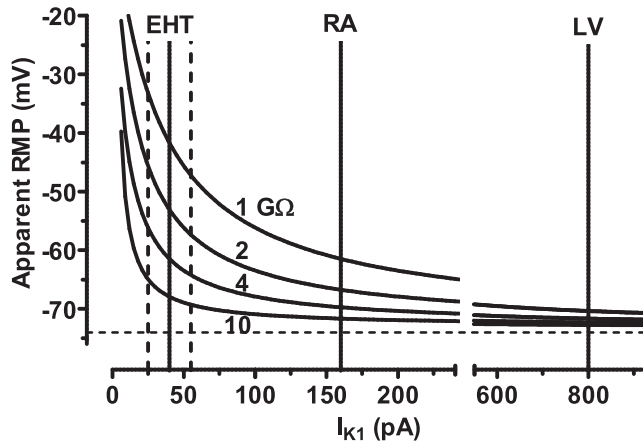
current density of  $I_{K1}$  was smaller at lower than at higher external  $K^+$  concentration ( $1.5 \pm 0.7$  pA/pF,  $n = 12$  at  $5.4$  mM  $K^+_{[ext]}$  versus  $14.1 \pm 12.0$  pA/pF,  $n = 60$  at  $20$  mM  $K^+_{[ext]}$ ;  $p < 0.05$ ). Reversal potential shifted to about  $-70$  mV under those conditions. Dividing the actual driving force for potassium ( $\sim 30$  mV) by the  $Ba^{2+}$ -sensitive absolute current amplitude measured at  $-100$  mV ( $41.1 \pm 13.7$  pA,  $n = 12$ ) gives a membrane resistance of around  $0.75$  G $\Omega$ . If we assume a true RMP around  $-73$  mV and a typical seal resistance of  $3$  G $\Omega$ , membrane voltage recorded by patch-clamp electrodes will therefore be reduced to a value of  $-58$  mV (according to the equation above). We do not have data for  $I_{K1}$  in LV CMs at  $5.4$  mM  $K^+$  and at  $37^{\circ}C$ . Therefore we refer to published data. Mean  $I_{K1}$  current densities were  $1.1$  and  $2.8$  pA/pF in RA and LV, respectively. Multiplying the values with the capacitances given in that study ( $145$  and  $285$  pF) we estimated absolute amplitudes of  $\sim 160$  pA in atrial and  $\sim 800$  pA in ventricular CMs under these conditions (Amos et al., 1996). Plotting  $V_{cm}$  versus  $I_{K1}$  amplitudes reveals that, even at the same seal resistances in hiPSC-CMs and human adult CMs, errors in hiPSC-CMs will be larger because of their small cell size. Even the 2.5 times higher  $I_{K1}$  density in ML may not be sufficient to leave the error zone, since absolute current amplitudes calculates to only  $79$  pA. For methodological reasons, we cannot provide seal resistance values after getting access to the cells. We found hiPSC-CMs rather fragile, and are afraid that seal resistance could drop down during an experiment. Importantly, in cells with low  $I_{K1}$  conductivity, due to a combination of low current density and small cell size, even small changes in seal resistance can have drastic effects on apparent RMP. In addition, our calculations are based on mean values for cell size and  $I_{K1}$ . Due to large variability both in cell size and  $I_{K1}$  density (Figure 1C), underestimation of RMP may be much larger in an individual cell (Figure 7). Therefore, given the present limitations, measurements of AP with patch-clamp pipettes are prone to error and not well suited as an indirect parameter of  $I_{K1}$  density. Small cell size is not necessarily an issue, since AP measurements are feasible even in much smaller cells, such as pancreatic cells (Rizzetto et al., 2015), whose size

are in the range of  $5$  pF (Rorsman Patrik, 1986). Very high seal resistance up to  $10$  G $\Omega$  and very large potassium conductance may facilitate AP recording in those cells (Keizer and Magnus, 1989). Sharp microelectrodes can be used to measure AP even in isolated cardiac myocytes (Szentandrassy et al., 2015) as well as in clusters of hiPSC-CMs (Christoforou et al., 2013; Cordeiro et al., 2015; Doss et al., 2012; Zhang et al., 2009). We applied this technique in single hiPSC-CMs, but were able to measure AP in only one single cell from  $85$  trials (Figure S3). In most cases we had problems to impale the very flat hiPSC-CMs without touching the bottom of the recording chamber.

#### Discrimination of Atrial and Ventricular Cells by $I_{K,ACh}$ , $I_{K1}$ , RMP, $APD_{90}$ , and Repolarization Fraction

Earlier reports suggested standard hiPSC-CM cultures contain a mixture of ventricular-, atrial-, and nodal-like CMs (Itzhaki et al., 2011; Liang et al., 2013, 2016; Ma et al., 2011, 2013; Moretti et al., 2010). Classification in these studies was based on differences of AP parameters. Here we have used a different approach based on the presence or absence of  $I_{K,ACh}$ . Only atrial and nodal CMs express  $I_{K,ACh}$  (Heidbüchel et al., 1987; Sakmann et al., 1983; Yamada et al., 1998). We could not find a single hiPSC-CM responding to CCh with a change of inwardly rectifying current. The absence of  $I_{K,ACh}$  cannot be explained by a lack of muscarinic receptors, since CCh reverses positive inotropic effects of isoprenaline in the same hiPSC-EHTs (Mannhardt et al., 2016), the classical accentuated antagonism (Levy, 1971). Thus, the data argue against an atrial phenotype of the hiPSC-CMs studied herein.

As outlined above, different  $I_{K1}$  densities might also discriminate between atrial and ventricular CMs. However, we found mean  $I_{K1}$  densities to differ only moderately between human RA and LV (2.5-fold) and, importantly, to scatter largely and substantially overlap between the groups (Figure 1D). Thus, our data do not support recent suggestions that  $I_{K1}$  density should be used to discriminate between atrial and ventricular phenotype in hiPSC-CMs (Giles and Noble, 2016).



**Figure 7. Influence of Seal Resistance and Absolute  $I_{K1}$  Amplitude on Apparent RMP in Patch-Clamp Experiments**

Curves give calculated apparent RMP as a function of absolute  $I_{K1}$  amplitudes and seal resistance. It is assumed that inward current amplitudes are determined by  $I_{K1}$  only. True RMP was assumed to be  $-73$  mV, as indicated by the dotted horizontal line. Vertical lines indicate absolute  $I_{K1}$  amplitudes at  $-100$  mV. Dotted vertical lines illustrate 95% CI of  $I_{K1}$  measured in EHT. The apparent RMP was calculated by the equation  $V_{cm} = V_{mm} + V_{mm} * R_M/R_{seal}$ . Please note data for EHT are taken from experiments independent from those presented in Figure 1 (numbers given in the Discussion). Data for RA and LV are taken from the literature (Amos et al., 1996). Numbers near the fit curves indicates seal resistance used for calculation. For details, see Discussion.

A third parameter differing between atrial and ventricular CMs is RMP. In line with the higher  $I_{K1}$  density in atrium, mean RMP was more negative in LV than in RA. However, the 2.5-fold higher  $I_{K1}$  related to only a small difference in mean RMP and, both in patch-clamp and sharp microelectrode measurements in intact tissue, individual RMP values largely overlapped between LV and RA. Thus, the power of RMP to discriminate atrial from ventricular CMs was modest. Tissues with an RMP negative to approximately  $-82$  mV had an 80% probability to be correctly classified as ventricles. However, they represent less than 20% of ventricular preparations. Mean RMP values in EHT were similar to RA and LV. Together with the atrial-like  $I_{K1}$  densities, the atrial-like RMP would suggest an electrophysiological phenotype close to human RA. Yet, this interpretation is at odds with the lack of  $I_{K_{ACh}}$  in EHT from C25, and smaller  $I_{K1}$  density and more negative RMP in iCells, indicating that these cells exhibit a mixed phenotype. RMP in EHT was almost exactly between LV and RA, but not significantly different from either. Therefore, RMP may not be a useful parameter to decide if EHT may possess atrial or ventricular  $I_{K1}$ . In addition, it should be emphasized that  $I_{K1}$  density alone does not determine RMP, but the relation of all conductances present, such as

background sodium and calcium currents, NCX, and  $Na^+/K^+$ -ATPase (Maleckar et al., 2009), many of them not yet studied in detail in hiPSC-CMs.

It is common use in the stem cell field to classify hiPSC-CMs as atrial or ventricular like according to their AP duration (Doss et al., 2012; Liang et al., 2013; Ma et al., 2011, 2013; Moretti et al., 2010; Vaidyanathan et al., 2016). Such assumptions are based on data from different studies reporting  $APD_{90}$  values for human heart (Chandler et al., 2009; Dobrev et al., 2001; Drouin et al., 1998; Jakob et al., 1989; Li et al., 1998; Verkerk et al., 2007a, 2007b; Wang et al., 1993), which are difficult to compare for methodological reasons. Here, we present a large number of sharp microelectrode data, which were obtained under identical recording conditions. While mean  $APD_{90}$  values were indeed shorter in RA than in LV, individual values again largely overlapped, questioning whether measurements of  $APD_{90}$  are indeed helpful to discriminate between LV and RA in individual recordings.

Other approaches were used for a more precise discrimination between atrial- and ventricular-like APs such as  $APD_{90}/APD_{50}$  (Chen et al., 2017; Dorn et al., 2015; Liang et al., 2013; Zhang et al., 2012). The early phase of repolarization in human heart is dominated by transient potassium outward currents. In contrast to ventricular CMs, the transient outward currents in atrial CMs exhibit a long-lasting component, which results in a larger repolarization fraction (Amos et al., 1996). We found a wide scattering of repolarization fraction in individual RA CMs. This finding is in line with the reported wide heterogeneity of transient outward currents in RA CMs (Amos et al., 1996). The repolarization fraction measured by sharp microelectrode in tissues likely integrates different AP shapes from individual cells and therefore the scatter is smaller. This interpretation is supported by the observation that mean values did not differ between individual isolated RA CMs and intact RA trabeculae. The same holds true for EHT. The smaller scatter in intact tissue allowed almost perfect discrimination between RA and LV and, according to this parameter, EHT resembled LV more than RA.

### Limitations

The present results have been obtained from two hiPSC lines (C25, iCell) and may differ in from other hiPSC lines. The points we want to make with this paper are (1) that patch clamping of hiPSC-CMs can underestimate the “true” RMP (as substantiated by sharp microelectrode data) and (2) that hiPSC-CMs can express relatively normal (human RA-like)  $I_{K1}$ . The differences in  $I_{K1}$  density compared with earlier data are likely explained by donor-dependent differences between the hiPSC lines and/or by different culture conditions such as different media, culture time, or extracellular matrix as suggested previously





(Doss et al., 2012; Herron et al., 2016). A more systematic comparison of different cell lines and different differentiation protocols and culture conditions is warranted. For technical reasons,  $I_{K1}$  was measured in isolated CMs, but RMP,  $APD_{90}$ , and repolarization fraction in intact tissues/3D EHTs. Thus, we cannot exclude that the physiological RMP is a consequence of the EHT culture format and does not reflect that of ML-cultured hiPSC-CMs. Another question is how reliable patch-clamp protocols are and if they reflect physiology. Here we have used protocols applied previously for many years in human atrial CMs (Dobrev et al., 2001). The human CMs used in this study were isolated from heart tissues from patients undergoing open heart surgery for various cardiac diseases. While most RA samples were from patients with normal ventricular function, the LV samples necessarily were from patients with severe forms of heart failure. Thus, we cannot exclude that the measurements are influenced by the heart failure phenotype. Finally, RMP is not exclusively determined by  $I_{K1}$  conductance, because it was reported that over 70% block of  $I_{K1}$  (10  $\mu$ M BaCl<sub>2</sub>) does not influence RMP in human LV tissue using sharp microelectrode technique (Jost et al., 2013). Therefore, we cannot rule out that culture-related changes in other background currents contribute to the physiological RMP measured in EHT.

## Conclusions

hiPSC-CMs from the C25 line possess robust  $I_{K1}$  currents, which, in ML, reached values of human LV CMs and, in EHT, that of RA CMs.  $I_{K1}$  density was 2-fold smaller in iCells than in C25, but was still associated with RA-like RMP in EHT. Technical issues related to patch clamping of small cells probably contribute to the reported low RMP in hiPSC-CMs. hiPSC-CMs exhibit features of both an atrial ( $I_{K1}$  and RMP) and ventricular phenotype (absence of  $I_{K,ACH}$  and low repolarization fraction). Low  $I_{K1}$  and depolarized RMP are not inherent characteristics of hiPSC-CMs.

## EXPERIMENTAL PROCEDURES

### Differentiation of hiPSC-CM and EHT Generation

The undifferentiated hiPSC line C25 (a kind gift from Alessandra Moretti, Munich, Germany) was expanded in FTDA medium (Frank et al., 2012) and differentiated in a three-step protocol based on growth factors and a small-molecule Wnt inhibitor DS07 (a kind gift from Dennis Schade, Dortmund, Germany), as published previously (Lanier et al., 2012; Mannhardt et al., 2016). Details are given in the Supplemental Information.

### Dissociation of hiPSC-CMs from ML and EHT

After culturing hiPSC-CMs in ML and EHT for 28 days, cells were isolated with collagenase type II (200 U/mL, Worthington Biochemical, Lakewood, NJ, USA, LS004176 in HBSS minus

Ca<sup>2+</sup>/Mg<sup>2+</sup>, Gibco 14175-053 and 50  $\mu$ M CaCl<sub>2</sub>) for 3 hr (ML) and 5 hr (EHT). At least three different batches of hiPSC-CMs were used. Details are given in the Supplemental Information.

### Human Samples

Myocardial tissue was obtained from patients undergoing cardiac surgery at the University Heart Center Hamburg. The study followed the declaration of Helsinki. All patients gave written informed consent. Atrial and ventricular CMs were isolated and prepared as described previously (Dobrev et al., 2001) (Figures S4D and S4E). Details are given in the Supplemental Information.

### Voltage Clamp Recordings (K<sup>+</sup> Currents)

Inwardly rectifying K<sup>+</sup> currents were measured at room temperature, using the *whole-cell* configuration of the patch-clamp technique. The Axopatch 200B amplifier (Axon Instruments, Foster City, CA, USA) and ISO2 software were used for data acquisition and analysis (MFK, Niedernhausen, Germany). Details are given in the Supplemental Information.

### Current Clamp Recordings

Action potentials were recorded using the perforated patch (amphotericin B) configuration of the patch-clamp technique. The Axopatch 200B (Axon Instruments) was set to current-clamp mode and the experiments performed at 37°C, 1 Hz. Details are given in the Supplemental Information.

### Sharp Microelectrode Recordings

Sharp microelectrodes were used to record action potentials in RA and LV trabeculae and in intact EHT. The experiments were performed at 37°C. Details are given in the Supplemental Information.

### Drugs

All drugs and chemicals were obtained from Sigma-Aldrich (St. Louis, MO, USA).

## SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, four figures, and four tables and can be found with this article online at <https://doi.org/10.1016/j.stemcr.2018.01.012>.

## AUTHOR CONTRIBUTIONS

A. H., A.U.U., I.M., M.D.L., A.L., C.N., and K.B. performed the research. A. H., A.U.U., A. H., A.L., N.J., T.E., and T.C. planned the experiments. A. H., M.D.L., I.M., K.B., C.N., A.L., and E.W. analyzed the results. A. H., T.E., and T.C. wrote the manuscript. All authors approved the final version of the manuscript.

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**Stem Cell Reports, Volume 10**

**Supplemental Information**

**Low Resting Membrane Potential and Low Inward Rectifier Potassium Currents Are Not Inherent Features of hiPSC-Derived Cardiomyocytes**

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## Supplemental information

### Supplemental Data

Figure S1. Related to Figure 1, 2 and 3

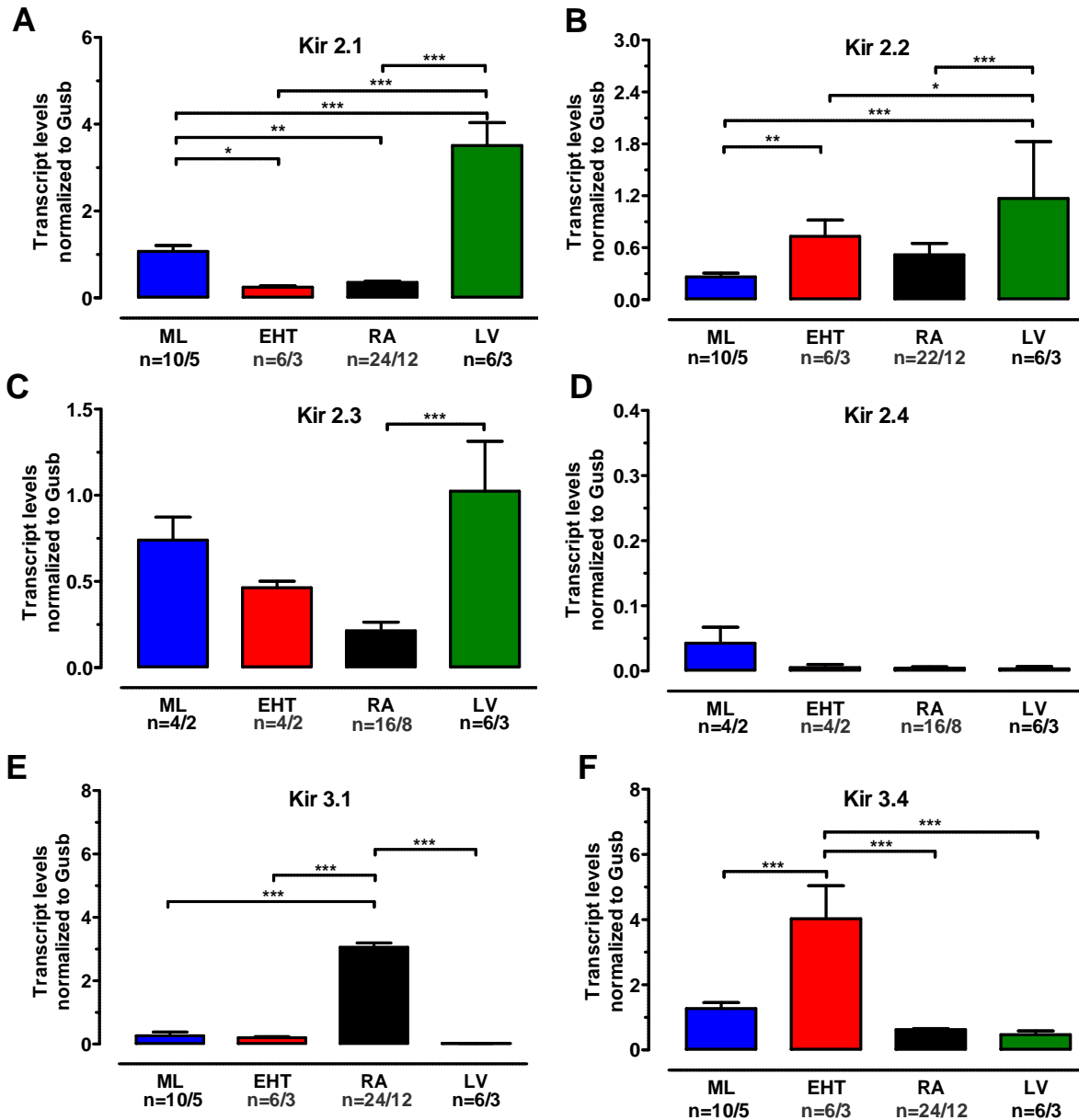
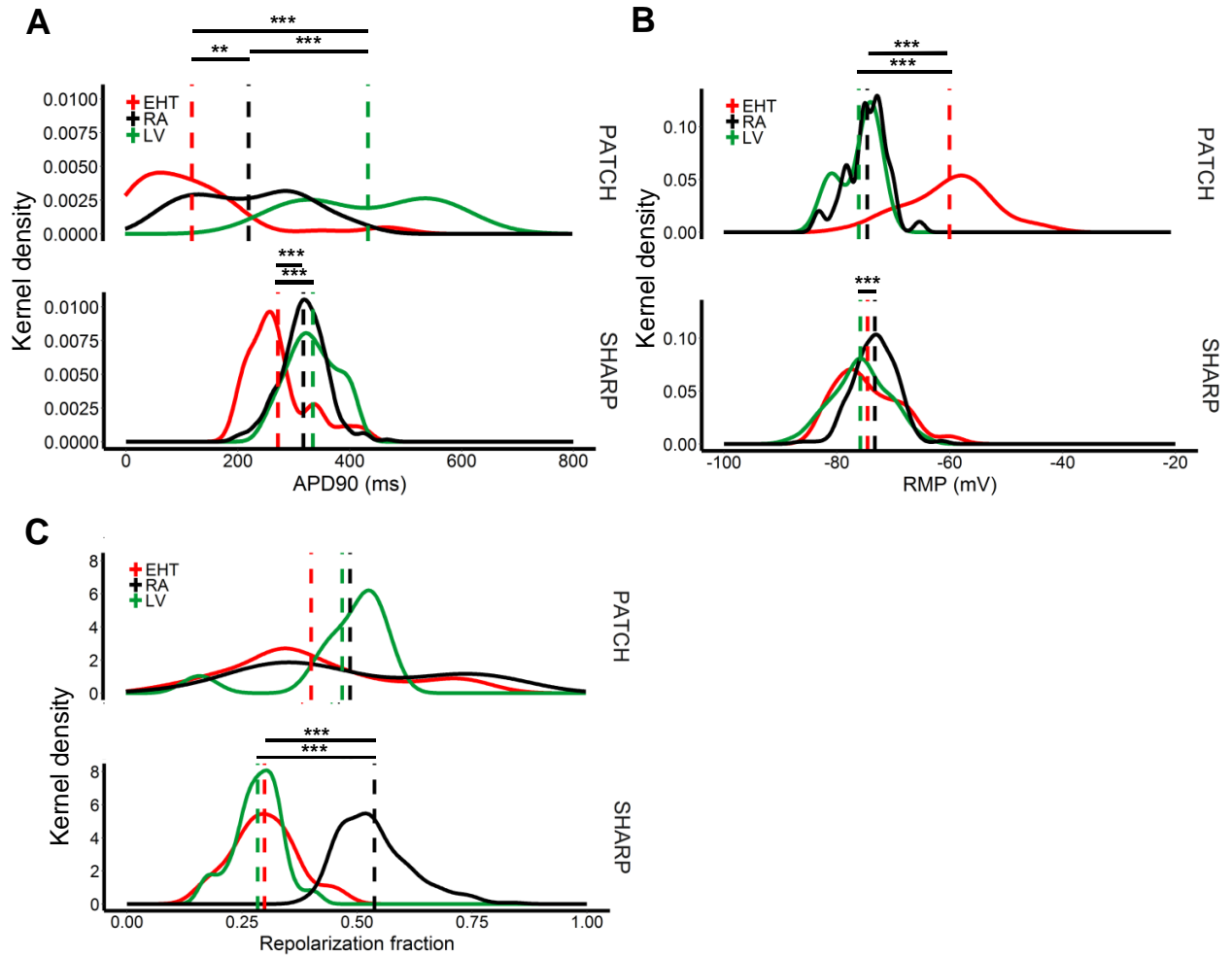


Figure S1. Expression of Kir channels in hiPSC-CM and human adult CM (Related to Figure 1, 2 and 3).

Transcript levels of  $I_{K1}$  channel-forming subunits Kir 2.1, 2.2, 2.3 and Kir2.4 and transcript levels of  $I_{K_{ACh}}$  channel-forming subunits Kir 3.1 and Kir 3.4 were normalized to housekeeping gene (GUSB). \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  (One-way Anova followed by Bonferroni test). n/n=number of experiment/number of isolations in hiPSC-CM and number of experiment/number of patients in RA and LV.

**Figure S2. Related to Figure 5.**



**Figure S2.** Frequency distribution of APD<sub>90</sub>, RMP and repolarization fraction in EHT, RA and LV.

Frequency distribution expressed as kernel density and mean values for APD<sub>90</sub> (A) RMP (B) and repolarization fraction (C). Data obtained by patch clamping of isolated CM from EHT, RA and LV are given in the upper panels, measured by sharp microelectrodes in intact tissues from EHT, RA and LV are given in the lower panels. \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 (One-way ANOVA followed by Kruskal-Wallis test).

Figure S3.

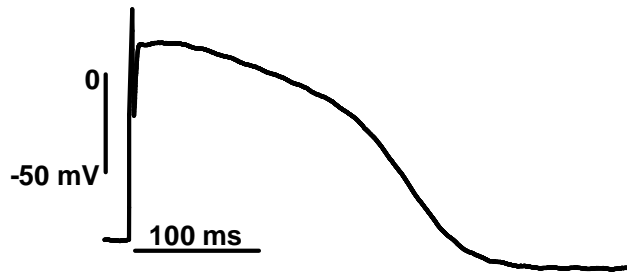
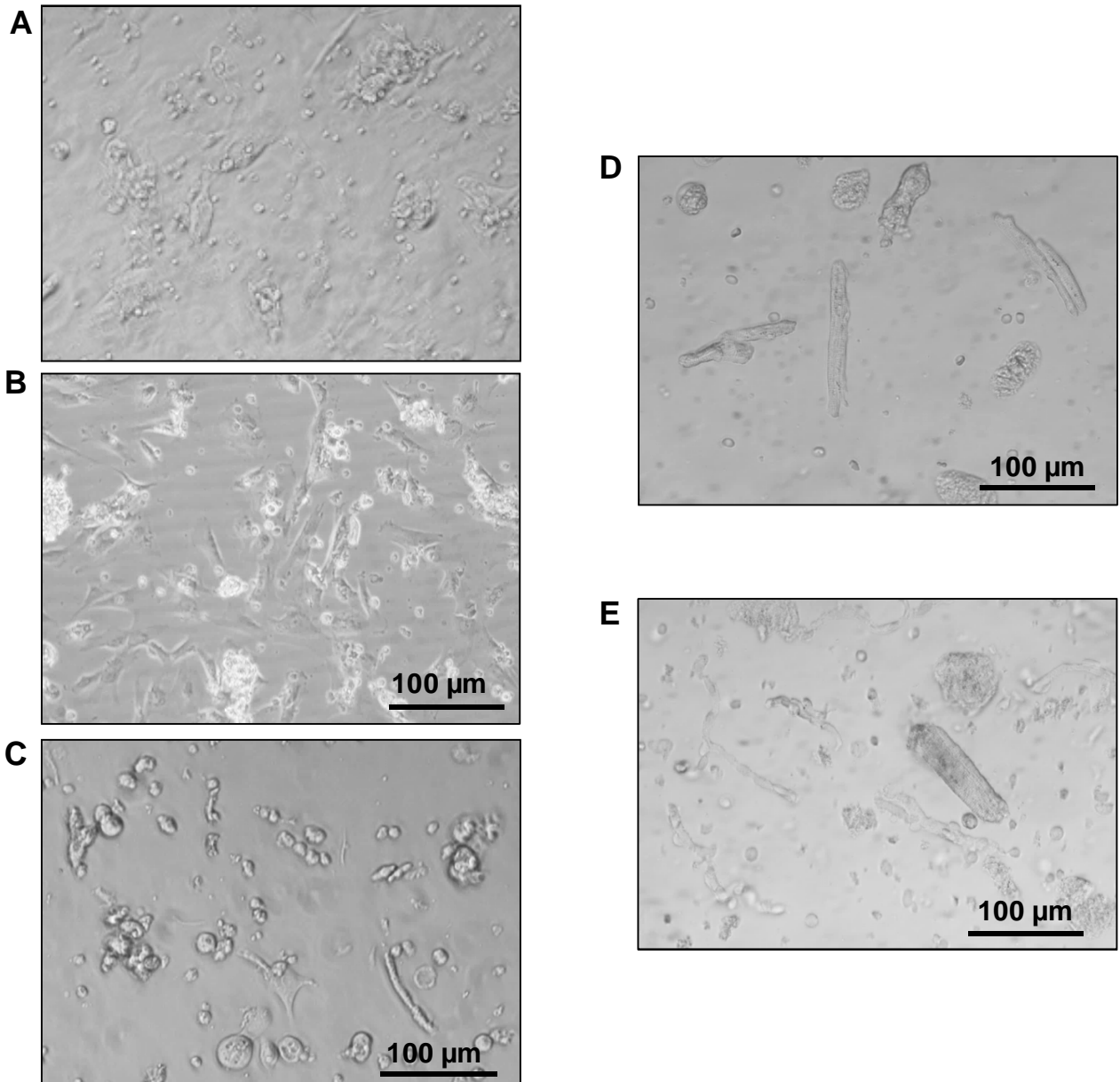


Figure S3. Original recording from a single hiPSC-CM using sharp microelectrode.

Figure S4.





**Figure S4.** Images of isolated hiPSC-CM and adult CM.

A: C25-ML, B: C25-EHT, C: CDI-EHT, D: RA, E: LV

## 2. Supplemental Tables

**Table S1.  $I_{K1}$  densities in pA/pF, descriptive values.**

	<b>n</b>	<b>mean</b>	<b>SD</b>	<b>median</b>	<b>min</b>	<b>max</b>	<b>skew</b>	<b>kurtosis</b>
<b>ML</b>	71	32.7	36.2	19.70	0.4	182	2.12	4.824
<b>EHT</b>	60	14.1	12.0	8.98	0.0	50	1.20	0.418
<b>RA</b>	55	14.0	11.0	11.85	0.6	55	1.17	1.916
<b>LV</b>	14	40.5	28.2	33.52	13.5	118	1.40	1.413

ML: Monolayer, EHT: Engineered Heart Tissue, RA: Right Atrium, LV: Left Ventricle

**Table S2.  $APD_{90}$  in ms, descriptive values.**

	<b>n</b>	<b>mean</b>	<b>SD</b>	<b>median</b>	<b>min</b>	<b>max</b>	<b>skew</b>	<b>kurtosis</b>
<b>EHT_Patch</b>	41	119.06	108.04	100.0	12.5	475.0	1.72	3.05
<b>RA_Patch</b>	41	220.68	102.68	220.0	60.0	410.0	0.07	-1.22
<b>LV_Patch</b>	10	434.00	122.04	440.0	270.0	600.0	-0.02	-1.89
<b>EHT_Sharp</b>	24	271.45	56.07	259.3	205.9	421.7	0.53	1.22
<b>RA_Sharp</b>	220	316.91	41.98	319.0	193.0	467.0	0.01	0.84
<b>LV_Sharp</b>	57	333.77	43.66	331.5	250.0	411.2	0.02	-0.99

EHT-Patch: Cardiomyocytes from Engineered Heart Tissue measured with patch clamp technique, RA-Patch: Cardiomyocytes from right atrium measured with patch clamp technique, LV-Patch: Cardiomyocytes from left ventricle measured with patch clamp technique, EHT-Sharp: Engineered Heart Tissue measured with sharp microelectrode technique, RA-Sharp: right atrium measured with sharp microelectrode technique, LV-Sharp: left ventricle measured with sharp microelectrode technique.

**Table S3. RMP in mV, descriptive values.**

	<b>n</b>	<b>mean</b>	<b>SD</b>	<b>median</b>	<b>min</b>	<b>max</b>	<b>skew</b>	<b>kurtosis</b>
<b>EHT_Patch</b>	41	-59.66	7.77	-58.00	-78.00	-42.00	-0.13	-0.25
<b>RA_Patch</b>	41	-74.37	3.65	-75.00	-83.00	-65.00	-0.30	0.31
<b>LV_Patch</b>	10	-75.87	3.54	-74.58	-81.78	-72.27	-0.62	-1.48
<b>EHT_Sharp</b>	24	-74.58	5.69	-76.75	-82.20	-60.00	0.60	0.18
<b>RA_Sharp</b>	220	-73.31	3.78	-73.00	-87.00	-61.00	-0.16	0.74

LV\_Sharp 57 -75.87 4.96 -76.00 -88.00 -65.00 -0.09 -0.42

Same abbreviations are used as in Table S2.

**Table S4. Repolarization fraction descriptive values.**

	<b>n</b>	<b>mean</b>	<b>SD</b>	<b>median</b>	<b>min</b>	<b>max</b>	<b>skew</b>	<b>kurtosis</b>
<b>EHT_Patch</b>	41	0.40	0.18	0.36	0.07	0.77	0.46	-0.56
<b>RA_Patch</b>	41	0.49	0.21	0.43	0.09	0.86	0.26	-1.23
<b>LV_Patch</b>	10	0.47	0.12	0.52	0.16	0.55	-1.74	1.93
<b>EHT_Sharp</b>	24	0.32	0.07	0.29	0.17	0.45	0.71	0.15
<b>RA_Sharp</b>	220	0.54	0.08	0.52	0.38	0.84	0.92	0.82
<b>LV_Sharp</b>	57	0.28	0.05	0.29	0.17	0.41	-0.21	0.12

Same abbreviations are used as in Table S2.

### 3. Supplemental Experimental Procedures

#### 3.1. Differentiation of hiPSC-CM and EHT generation

In brief, confluent undifferentiated cells were dissociated (0.5 mM EDTA; 10 min) and cultivated in spinner flasks (30\*10<sup>6</sup> cells/100 ml; 40 rpm) for embryoid body formation overnight (Zweigerdt et al., 2011). Mesodermal differentiation was initiated in embryoid bodies over three days in suspension culture with growth factors (BMP-4 [R&D Systems, 314-BP], activin-A [R&D Systems, 338-AC], FGF2 [PeproTech, 100-18B]). Cardiac differentiation was performed either in adhesion or in suspension culture with Wnt-inhibitor DS07 (Lanier et al., 2012). Cells were cultured in a humidified temperature and gas-controlled incubator (37 °C, 5% CO<sub>2</sub>, 5% O<sub>2</sub>; 21% O<sub>2</sub> for final cardiac differentiation). At day 14 the spontaneously beating hiPSC-CM were dissociated with collagenase II (Worthington, LS004176; 200 U/ml, 3.5 h) and either cultured in ML or EHT format. For three-dimensional culture EHT were generated as previously described with 1x10<sup>6</sup> cells / 100 µl EHT (Schaaf et al., 2014). EHT as well as ML were cultured in a 37 °C, 7% CO<sub>2</sub>, 40% O<sub>2</sub> humidified cell culture incubator with a medium consisting of DMEM (Biocrom; F0415), 10% heat-inactivated horse serum (Gibco 26050), 1% penicillin/streptomycin (Gibco 15140), insulin (10 µg/ml; Sigma I9278) and aprotinin (33 µg/ml; Sigma A1153). For further comparability, experiments were performed in parallel from the same batch of cells.

#### 3.2 Quantification of transcript levels

The RNeasy® Plus Mini Kit (Qiagen, Venlo, The Netherlands) was used to isolate total RNA from human heart tissue, cardiomyocytes cultured in a ML and dissociated EHT. For quantification of transcript levels, cDNA was generated using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems). Quantitative PCR was performed with the 5x HOT FIREPol® EvaGreen® qPCR Mix Plus (ROX) (Solis BioDyne, Tartu, Estonia) on ABI PRISM 7900HT Sequence detection system (Applied Biosystems, Foster City, California, USA). Relative transcript levels were calculated using the  $\Delta$ CT-method with glucuronidase beta (Gusb) as a housekeeping gene.

#### 3.3 Dissociation of hiPSC-CM from ML and EHT

Cells were plated on gelatine-coated (0.1%) glass coverslips (12 mm diameter, Carl Roth GmbH+Co, Karlsruhe, Germany) for 24–48 h before patch clamp experiments were performed (Figure S4A, B, C). To avoid changes in morphology and ion currents due to culture conditions as much as possible, the cells were investigated within 48 hours.

#### 3.4 Voltage clamp recordings (K<sup>+</sup> currents)

Heat-polished pipettes were pulled from borosilicate filamented glass (Hilgenberg, Malsfeld, Germany). Tip resistances were 2.5–5 M $\Omega$ , seal resistances were 2–4 G $\Omega$ . The cells were investigated in a small perfusion chamber placed on the stage of an inverse microscope. To reduce the liquid junction potential an agarose bridge was placed into a solution containing high KCL. The experiments were performed with the following bath solution (in mM): NaCl 120, KCl 20, HEPES 10, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1 and glucose 10 (pH 7.4, adjusted with NaOH)(Dobrev et al., 2005). Contaminating Ca<sup>2+</sup> currents were suppressed with the selective L-type calcium channel blocker nifedipine (10  $\mu$ M). For I<sub>K1</sub> and I<sub>K,ACh</sub> the internal solution included (in mM): DL-Aspartate potassium salt 80, KCl 40, NaCl 8, HEPES 10, Mg-ATP 5, Tris-GTP 0.1, EGTA 5 and CaCl<sub>2</sub> 2, pH 7.4, adjusted with KOH (Dobrev et al., 2005). Inward Current amplitudes were determined as currents at –100 mV. A single concentration (2  $\mu$ M) of the muscarinic receptor agonist carbachol (CCh) was used to evoke I<sub>K,ACh</sub>.

### 3.5 Current clamp recordings

The Axopatch 200B (Axon Instruments, Foster City, CA, USA) was set to current clamp mode and the experiments performed at 37 °C, 1 Hz, with the following bath solution (in mM): NaCl 120, KCl 5.4, HEPES 10, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1 and glucose 10 (pH 7.4, adjusted with NaOH). The internal solution was (in mM): DL-aspartate potassium salt 80, KCl 40, NaCl 8, HEPES 10, Mg-ATP 5, Tris-GTP 0.1, EGTA 5 and CaCl<sub>2</sub> 2 (pH 7.4, adjusted with KOH, Dobrev et al., 2001). The liquid junction potential was calculated by using the JPCalcW software (SDR Scientific, Sydney, Australia) to –12 mV, for both voltage and current clamp recordings. The tips of the pipettes were filled with the same solution, but supplemented with 20  $\mu$ g/ml Amphotericin B to allow formation of perforated patch configuration within a few minutes. In action potential recordings the repolarization fraction was calculated by the following way: (APD<sub>50</sub>–APD<sub>90</sub>)/APD<sub>90</sub>(Du et al., 2015).

### 3.6 Sharp microelectrode recordings

Microelectrode tip resistances were 20 to 50 M $\Omega$  when filled with 3 mM KCl). Action potentials were elicited by field stimulation at 1 Hz: 0.5 ms stimulus, 50% above threshold intensity. The following bath solution was used (in mM): NaCl 125, KCl 5.4, MgCl<sub>2</sub> 0.6, CaCl<sub>2</sub> 1, NaH<sub>2</sub>PO<sub>4</sub> 0.4, NaH<sub>2</sub>CO<sub>3</sub> 22, and glucose 5.5 and was equilibrated with O<sub>2</sub>–CO<sub>2</sub> (95:5).

### 3.7 Sharp microelectrode recordings in single hiPSC-CM

Sharp microelectrodes were used to record action potentials in isolated single hiPSC-CM. Microelectrode tip resistances were 20 to 50 M $\Omega$  when filled with 3 mM KCl). Action potentials were elicited by field stimulation at 1 Hz: 0.5 ms stimulus. The following bath solution was used (in mM): NaCl 120, KCl 5.4, HEPES 10, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1 and glucose 10 (pH 7.4, adjusted with NaOH). The experiments were performed at 37 °C.

### 3.8 Statistics

Results are presented as mean values $\pm$ SEM. Curve fitting was performed by using the GraphPad Prism Software 5.02 (GraphPad Software, San Diego, CA, USA). Statistical differences were evaluated by using the Student's t-test (paired or unpaired) or repeated measures ANOVA, followed by Bonferroni test, where appropriate. A value of p<0.05 was considered to be statistically significant. Linear correlations, concentration-response curves were fitted to mean data points and –logIC<sub>50</sub> values were compared by F-test using the GraphPad Prism Software 5.02. Analyses of frequency distribution were performed using R (ver. 3.3.1) (R Core Team, 2013). Please note that the statistical term “Kernel density estimation” is used in Fig. S2, with a “kernel” defined as a probability density function which must possess the following properties: even non-negative, real-valued and its definite integral over its support set must equal to 1. Therefore, “kernel density estimation” is a non-parametric method, which allows estimating the probability density function of a random variable (<http://scikit-learn.org/stable/modules/density.html>). The kurtosis describes the tail shape of the data distribution (<http://www.r-tutor.com/elementary-statistics/numerical-measures/kurtosis>). The skewness is a measure of symmetry (<http://www.r-tutor.com/elementary-statistics/numerical-measures/skewness>).

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