

Supplemental file 1: homozygosity mapping

```
#!/usr/bin/env python
import subprocess
import os
import glob
import shutil
import homozygosity_mapping_ub26

##parameters
delim = '\t'
thread_number = '6'

##working dir --- add location of fastq files and a working directory
# working_dir = '/data/atimms/timon_0317'
working_dir = ''
fastq_dir = ''
os.chdir(working_dir)

##programs and files
convert_2_annovar = '/home/atimms/programs/annovar/convert2annovar.pl'
table_annovar = '/home/atimms/programs/annovar/table_annovar.pl'
fasta = '/data/atimms/references/mm10/mm10.fa'
bwa = '/home/atimms/programs/bwa-0.7.12/bwa'
samtools = '/home/atimms/programs/samtools-1.3/samtools'
bcftools = '/home/atimms/programs/bcftools-1.3/bcftools'
picard = '/home/atimms/programs/picard-tools-1.139/picard.jar'
delly = '/home/atimms/programs/delly_v0.7.6_CentOS5.4_x86_64bit'
##files etc
##fastq dictionary, include name of sample and name of fastq files
fq_dict = {}
post_bwa_bam = '.bwa.bam'
sorted_bam = '.bwa_sorted.bam'
mkdup_bam = '.bwa_mkdup.bam'
bamslis_file = 'bams.list'
##name of samtools vcf and input files for annovar
st_vcf = 'timon_0317.vcf.gz'
st_avinputs = []
delly_exclude_regions =
'/data/atimms/references/annovar/mm10/mouse.mm10.excl.tsv'
bedtools = '/home/atimms/programs/bedtools2-master/bin/bedtools'

##annovar parameters
av_genome = 'mm10'
av_buildver = ['-buildver', av_genome]
av_ref_dir = ['/data/atimms/references/annovar/' + av_genome]
##annotate with in house controls and public databases
av_protocol = ['-protocol',
'refGene,rmsk,genomicSuperDups,snp142,generic,generic,generic,generic,ge
neric,generic,generic,generic,generic',
'-genericdbfile',
'pleather_sc_st.K416.avinput,daredevil.avinput,bub.avinput,J308.st.avinput,J318.
```

```

st.avinput,J320.st.avinput,J327.st.avinput,J328.st.avinput,J329.st.avinput,mgp.v
5.merged.snps_all.dbSNP142.avinput']
av_operation = ['-operation', 'g,r,r,f,f,f,f,f,f,f,f,f,f,f,f,f']
av_options = ['-otherinfo', '-remove', '-arg', '-splicing 10 ,,,,,,,,,,,,,,']
#av_options = ['-otherinfo', '-remove', '-vcfinput']

acceptable_chr = ['chr1', 'chr2', 'chr3', 'chr4', 'chr5', 'chr6', 'chr7',
'chr8',
                    'chr9', 'chr10', 'chr11', 'chr12', 'chr13', 'chr14',
'chr15',
                    'chr16', 'chr17', 'chr18', 'chr19', 'chr20', 'chr21',
'chr22',
                    'chr23', 'chr24', 'chr25', 'chrX', 'chrY', 'chrX',
'chrY',
                    '1', '2', '3', '4', '5', '6', '7', '8', '9', '10', '11',
'12',
                    '13', '14', '15', '16', '17', '18', '19', '20', '21',
'22', '23',
                    '24', '25', 'x', 'y', 'X', 'Y']

##filtering and hom mapping parameters
##filtering
col_exon = 6
exon_definition = ['exonic', 'splicing']
col_function = 9
syn_definition = 'synonymous SNV'
zygosity_col = 24
cov_col = 26
cov_definition = 5
qual_col = 25
qual_definition = 30
##het snp mapping
genome_fai = '/data/atimms/references/mm10/mm10.fa.fai'
window_size = [1000000,5000000,2000000]
# window_size = [10000000]
step_size = 1000000
info_col = 36
naf_values = [0.8,0.9,0.95]

##check if 's' is a number and return boolean
def is_number(s):
    try:
        float(s)
        return True
    except ValueError:
        return False

##dictionary used to convert operator to .operator command
def operator_dict(op):
    ops = {"==": operator.eq, "!=": operator.ne, ">=": operator.ge, "<=":
operator.le, ">": operator.gt, "<": operator.lt, "in": operator.contains}
    return ops[op]

```

```

def filter_ann(working_dir, test_type, input_file, output_file, col_number, op,
comparison):
    """
    method for filtering files by operator and a value, need to provide:
    working directory,
    test type i,e, 'and' or 'or',
    input filename,
    output filename.
    parameters for tests i.e;
    column number
    operator (must be in dict)
    comparison (what to compare value in col number with)
    """
    os.chdir(working_dir)
    print "filtering variants from the file '%s' in an '%s' fashion" %
(input_file,test_type)
    ##variables
    row_number, variants_kept = 0,0 #keep track of
number of tests and variants kept
    delim = '\t'
    ##open files for reading and writing
    infile = open(input_file, 'rb')
    outfile = open(output_file, 'wb')
    writer = csv.writer(outfile, delimiter=delim)
    ##check row by row
    for row in csv.reader(infile, delimiter=delim):
        row_number += 1
        if row_number == 1:
            writer.writerow(row) #writes
header to out file
            header = row #writes
header to header
        else:
            tests_correct = 0 #keep
track of number of correct tests and reset
            for i in range(len(comparison)):
                op_func = operator_dict(op[i])
#queries operator dictionary so can perform comparisons
                col = col_number[i] -1 #adjust
column number so starts at 1 not 0

                if row_number == 2: #print
info on test completed
                    print "test %r: '%s %s' in column '%s'" % (i,
op[i], comparison[i], header[col])

                    if is_number(comparison[i]): #if
comparison is a number
                        # print row[col]
                        if row[col] == 'NA' or row[col] == '.' or row[col]
== '': #if using na as 'empty' if can't compare number to text so say test
is true
                            tests_correct += 1
                            elif row[col] == '.': # in
gatk sometime coverage = 0 but printed as .

```

```

        pass
        elif op_func(float(row[col]), comparison[i]):
#proper comparison
            tests_correct += 1
        else:
            if op_func(row[col], comparison[i]):           #if
comparison is not a number
                tests_correct += 1
                ##looks if a 'and' a 'or' or a 'once' test type, and writes
into new file
                if test_type == "or":                       #if any
test correct per row
                    if tests_correct > 0:
                        writer.writerow(row)
                        variants_kept += 1
                    elif test_type == "and":               #if all
tests correct per row
                        if tests_correct == len(comparison):
                            writer.writerow(row)
                            variants_kept += 1
                        elif test_type == "once":          #if one
test correct per row
                            if tests_correct == 1:
                                writer.writerow(row)
                                variants_kept += 1
                    else:
                        print "incorrect test type!!"
                ##print overall numbers
                print "%s variants kept out of %s checked" % (variants_kept, row_number -
1)
                print "filtered variants in file:", output_file
                print "" #empty line
                infile.close()
                outfile.close()

##align with bwa
def align_with_bwa(sample_dict):
    for sample in sample_dict:
        r1_fq = sample_dict[sample][0]
        r2_fq = sample_dict[sample][1]
        print sample, r1_fq, r2_fq
        rg = '@RG\tID:' + sample + '_RG\tSM:' + sample + "\tPL:ILLUMINA"
        pe_bam = sample + post_bwa_bam
        sort_bam = sample + sorted_bam
        pic_dup_bam = sample + mkdup_bam
        bwa_pe = subprocess.Popen([bwa, 'mem', '-M', '-t', '20', '-R', rg,
fasta, r1_fq, r2_fq], stdout=subprocess.PIPE)
        st_sam_bam_pe = subprocess.Popen([samtools, 'view', '-q', '20', '-
b', '-@', '5', '-o', pe_bam, '-'], stdin=bwa_pe.stdout)
        st_sam_bam_pe.wait()
        st_sort_pe = subprocess.Popen([samtools, 'sort', '-O', 'bam', '-o',
sort_bam, '-T', sample, '-@', '10', '-m', '10G', pe_bam])
        st_sort_pe.wait()

```

```

        ##mark duplicates
        picard_md = subprocess.Popen(['java', '-Xmx80g', '-jar', picard,
'MarkDuplicates', 'VALIDATION_STRINGENCY=SILENT', 'CREATE_INDEX=true', 'INPUT='
+ sort_bam, 'OUTPUT=' + pic_dup_bam, 'METRICS_FILE=' + sample + '.metrics',
'TMP_DIR=' + working_dir])
        picard_md.wait()

##make list of all bam files to be analyzed
def make_list_of_bams(sample_dict, bam_suffix, bamlist_file):
    with open(bamlist_file, "w") as outf:
        for sample in sample_dict:
            bam = sample + bam_suffix
            outf.write(bam + '\n')

##call samtools on bamfiles
def variant_calling_samtools(bamlist, final_vcf):
    vcf_temp1 = 'temp_st.vcf.gz'
    vcf_temp2 = 'temp_fb2.vcf.gz'
    stmp = subprocess.Popen([samtools, 'mpileup', '-ug', '-t', 'DP,DV,DPR', '-
q', '20', '-C', '50', '-f', fasta, '-b', bamlist], stdout=subprocess.PIPE)
    bcft = subprocess.Popen([bcftools, 'call', '-vm0', 'z', '-V', 'indels', '-
o', vcf_temp1], stdin=stmp.stdout)
    bcft.wait()
    bcf_index = subprocess.Popen([bcftools, 'index', vcf_temp1])
    bcf_index.wait()
    #split multi-allelic variants calls in separate lines
    bcf_norm1 = subprocess.Popen([bcftools, 'norm', '-m-both', '-o',
vcf_temp2, vcf_temp1])
    bcf_norm1.wait()
    bcf_norm2 = subprocess.Popen([bcftools, 'norm', '-f', fasta, '-O', 'z', '-
o', final_vcf, vcf_temp2])
    bcf_norm2.wait()
    bcf_index = subprocess.Popen([bcftools, 'index', final_vcf])
    bcf_index.wait()

##make avinput files
def convert_to_annoar(vcf):
    con_ann = subprocess.Popen([convert_2_annoar, '-format', 'vcf4', vcf, '-
includeinfo', '-withzyg', '-allsample', '-outfile', 'temp'])
    con_ann.wait()
    temp_files = glob.glob('temp*.avinput')
    for temp_file in temp_files:
        real_file = temp_file[5:]
        os.rename(temp_file, real_file)
        shutil.copy(real_file, str(av_ref_dir[0]))

def run_table_annoar(avinputs):
    for avinput in avinputs:
        av_prefix = avinput.rsplit('.',1)[0]
        command = [table_annoar] + av_builderver + [avinput] + av_ref_dir +
av_protocol + av_operation + av_options + ['-out', av_prefix]
        annoar = subprocess.Popen(command)
        annoar.wait()

```

```

def multianno_to_annotated(avinputs):
    head = ['Chr', 'Start', 'End', 'Ref', 'Alt', 'Func.refGene',
'Gene.refGene', 'GeneDetail.refGene', 'ExonicFunc.refGene', 'AChange.refGene',
'rmsk', 'genomicSuperDups',
'snp142', 'pleather', 'daredevil', 'bub', 'J308', 'J318', 'J320', 'J327', 'J328', 'J329',
'mgp.v5.snps', 'Zygotity', 'Qual', 'Coverage', 'Chr', 'Pos', 'Filter', 'Ref2',
'Alt2', 'Qual', 'Filter', 'GT_info', 'Format', 'Info']
    head_out = delim.join(head + ['\n'])
    for avinput in avinputs:
        av_prefix = avinput.rsplit('.',1)[0]
        multianno = av_prefix + '.mm10_multianno.txt'
        annotated = av_prefix + '.annotated.txt'
        with open(multianno, "r") as multi, open(annotated, "w") as final:
            final.write(head_out)
            line_count = 0
            for line in multi:
                line_count += 1
                if line_count > 1:
                    final.write(line)

def make_bed_from_ann_txt(infile, outfile):
    with open(outfile, "w") as out_fh, open(infile, "r") as in_fh:
        line_count = 0
        for line in in_fh:
            line_count += 1
            if line_count > 1:
                line = line.rstrip().split(delim)
                out_fh.write(delim.join(line[:3] + ['\n']))

##edit bedfile to remove non standard chromosomes
def remove_nonStd_chr(infile, outfile):
    fh = open(infile, 'r')
    outfh = open(outfile, 'w')
    for line in fh:
        line = line.split(delim)
        if line[0] in acceptable_chr:
            outfh.write(delim.join(line))
    fh.close()
    outfh.close()

##divide genome into windows
def make_windows(working_dir, genome_fai, window_size, step_size):
    os.chdir(working_dir)
    genome = genome_fai.split('/')[-1].split('.')[0]
    genome_window_bed = genome + '_' + str(window_size / 1000) + 'kb_' +
str(step_size / 1000) + 'kb' + '.bed'
    print 'bed file %s created from file: %s with window size %r and step size
%r' % (genome_window_bed, genome_fai, window_size, step_size)
    print ''
    ##run bedtools makewindows without pybedtools installed
    command = bedtools + ' makewindows -g ' + genome_fai + ' -w ' +
str(window_size) + ' -s ' + str(step_size) + ' > ' + 'bed.temp'
    subprocess.call(command, shell=True)

```



```

        #print var_caller, info, non_ref_allele, total_alleles, naf

        if naf > 0.1:                #prevents very low frequency SNPs and
those we've filtered
            zygosity = line [zygosity_col -1]
            line_out = [line [0], str(int(line [1]) - 1), line [2],
zygosity, str(naf), '\n']
            outfh.write(delim.join(line_out))
            kept_count += 1
            #print 'kept'
        #else:
            #print info, naf
print 'bed file %s created from %s' % (outfile, filename)
print '%r variants checked and %r kept' % (line_count -1, kept_count)
print ''
fh.close()
outfh.close()

##take bedfile with zygosity in 4th col and write a het and hom temp file
def split_het_hom(working_dir, filename):
    fh = open(filename, 'r')
    het_fh = open('het.temp', 'w')
    hom_fh = open('hom.temp', 'w')
    line_count = 0
    het_count = 0
    hom_count = 0
    for line in fh:
        line_count += 1
        line = line.split(delim)
        if line[3] == 'het':
            het_fh.write(delim.join(line))
            het_count += 1
        elif line[3] == 'hom':
            hom_fh.write(delim.join(line))
            hom_count += 1
        else:
            print 'issue with snp on line', line_count
    fh.close()
    het_fh.close()
    hom_fh.close()
    print 'for file %s we have %r snps, of which %r are het and %r are hom' %
(filename, line_count, het_count, hom_count)
    print ''

##combines multiple beds
##'chr:start-end' must match and be sorted in same way
##will only add last col, so need to adjust if need more
def combine_multiple_bed.bed_list, outfile):
    results = []
    no_of_bed = 0
    ##loop through bedfiles and add to lists
    for bed in bed_list:
        fh = open(bed, 'r')
        no_of_bed += 1

```



```

line_number = -1
for line in fh:
    line_number += 1
    line = line.strip('\n').split(delim)
    ##if first file add all columns
    if no_of_bed == 1:
        results.append(line)
    ##if not first file check 'chr:start-end' and add last column
    if no_of_bed >1:
        if results[line_number][:3] == line[:3]:
            results[line_number].append(line[-1])

    fh.close()
##write to outfile
outfh = open(outfile, 'w')
for i in range(len(results)):
    outfh.write(delim.join(results[i]) + '\n')
outfh.close()
print "combined the %r bedfiles %s" % (no_of_bed, ','.join.bed_list))
print "checked %r and printed %r lines" % (line_number + 1 , len(results))

##make het and hom count bedgraphs for specified window size
def het_and_hom_bed(working_dir, genome_and_window, filename):
    os.chdir(working_dir)
    print 'making het and hom bedfiles for:', filename
    split_het_hom(working_dir, filename)
    sample = filename.split(".", 1)[0]
    window_bed = genome_and_window + '.bed'
    hom_count_bed = sample + '_' + genome_and_window + '_hom_count.bedgraph'
    het_count_bed = sample + '_' + genome_and_window + '_het_count.bedgraph'
    ##pbt version
    # a = pbt.BedTool(genome_and_window + '.bed')
    # b = pbt.BedTool('hom.temp')
    # c = pbt.BedTool('het.temp')
    # a_with_b = a.intersect(b, c=True).moveto(hom_count_bed)
    # a_with_c = a.intersect(c, c=True).moveto(het_count_bed)
    ##bedtools version
    #bedtools intersect -a A.bed -b B.bed -c
    with open(hom_count_bed, "w") as hom_fh:
        hom_bt_intersect = subprocess.Popen([bedtools, 'intersect', '-a',
window_bed, '-b', 'hom.temp', '-c'], stdout=hom_fh)
        hom_bt_intersect.wait()
    with open(het_count_bed, "w") as het_fh:
        hom_bt_intersect = subprocess.Popen([bedtools, 'intersect', '-a',
window_bed, '-b', 'het.temp', '-c'], stdout=het_fh)
        hom_bt_intersect.wait()
    return hom_count_bed, het_count_bed

##count hom and het, and hom percentage in specified window
def count_and_percentage(working_dir, genome_and_window, filename):
    os.chdir(working_dir)
    print 'calculating hom count, het count and hom percentage for:', filename
    hom_count_bed, het_count_bed = het_and_hom_bed(working_dir,
genome_and_window, filename)
    combine_multiple_bed([hom_count_bed, het_count_bed], 'hom_het.temp')
    fh = open('hom_het.temp', 'r')

```

```

sample = filename.split(".", 1)[0]
outfile = sample + '_' + genome_and_window + '_hom_percentage.bedgraph'
outfh = open(outfile, 'w')
for line in fh:
    line = line.strip('\n').split(delim)
    if line[3] == '0':
        hom_percentage = 0
    else:
        hom_percentage = float(line[3]) / (float(line[3]) +
float(line[4]))
    outfh.write(delim.join(line[:3]) + delim + str(hom_percentage) +
'\n')
fh.close()
outfh.close()
print 'generated files:'
print 'hom_count:', hom_count_bed
print 'het_count:', het_count_bed
print 'hom_percentage:', outfile

##novel allele frequency within windows
def naf_in_window(working_dir, genome_and_window, bed_file):
    os.chdir(working_dir)
    print 'calculating novel allele freq within windows for file:', bed_file
    sample = bed_file.split(".", 1)[0]
    ##bedtools intersect window bed and sample bed
    # a = pbt.BedTool(genome_and_window + '.bed')
    # b = pbt.BedTool(bed_file)
    # a_with_b = a.intersect(b, wa=True, wb=True).moveto(sample + '_naf.temp')
    ##bedtools version
    window_bed = genome_and_window + '.bed'
    #bedtools intersect -a A.bed -b B.bed -c
    with open(sample + '_naf.temp', "w") as naf_fh:
        hom_bt_intersect = subprocess.Popen([bedtools, 'intersect', '-a',
window_bed, '-b', bed_file, '-wa', '-wb'], stdout=naf_fh)
        hom_bt_intersect.wait()
    ##make dictionary: key = chr:start-end and value a list of naf values
    naf_dict = {}
    fh = open(sample + '_naf.temp', 'r')
    for line in fh:
        line = line.strip('\n').split(delim)
        chr_start_end = delim.join(line[:3])
        naf_value = float(line[-1])
        if naf_dict.has_key(chr_start_end):
            naf_dict[chr_start_end].append(naf_value)
        else:
            naf_dict[chr_start_end] = []
            naf_dict[chr_start_end].append(naf_value)
    fh.close()

    ##make list of lists containing chr,start,end,average_naf, snp#
    list_of_lists = []
    for cse in naf_dict.keys():
        split_cse = cse.split(delim)
        average_naf = [sum(naf_dict[cse])/float(len(naf_dict[cse]))]

```

```

        total_snps = [len(naf_dict[cse])]
        outlist = split_cse + average_naf + total_snps
        outlist[1] = int(outlist[1])
        outlist[2] = int(outlist[2])
        list_of_lists.append(outlist)
list_of_lists.sort(key = operator.itemgetter(0, 1))

##print bed file with average_naf
outfh = open(sample + '_' + genome_and_window + '_naf.bedgraph', 'w')
for line in list_of_lists:
    for i in range(len(line)):
        line[i] = str(line[i])
    outfh.write(delim.join(line) + '\n')
outfh.close()

##total snp number within windows (for testing)
def total_snp_in_window(working_dir, genome_and_window, bed_file):
    os.chdir(working_dir)
    print 'calculating total snp number within windows for file:', bed_file
    sample = bed_file.split(".", 1)[0]
    ##bedtools intersect window bed and sample bed
    # a = pbt.BedTool(genome_and_window + '.bed')
    # b = pbt.BedTool(bed_file)
    # a_with_b = a.intersect(b, wa=True, wb=True).moveto(sample + '_naf.temp')
    ##bedtools version
    window_bed = genome_and_window + '.bed'
    #bedtools intersect -a A.bed -b B.bed -c
    with open(sample + '_naf.temp', "w") as naf_fh:
        hom_bt_intersect = subprocess.Popen([bedtools, 'intersect', '-a',
window_bed, '-b', bed_file, '-wa', '-wb'], stdout=naf_fh)
        hom_bt_intersect.wait()

    ##make dictionary: key = chr:start-end and value a list of naf values
    naf_dict = {}
    fh = open(sample + '_naf.temp', 'r')
    for line in fh:
        line = line.strip('\n').split(delim)
        chr_start_end = delim.join(line[:3])
        naf_value = float(line[-1])
        if naf_dict.has_key(chr_start_end):
            naf_dict[chr_start_end].append(naf_value)
        else:
            naf_dict[chr_start_end] = []
            naf_dict[chr_start_end].append(naf_value)
    fh.close()

    ##make list of lists containing chr,start,end,total_snp, snp#
    list_of_lists = []
    for cse in naf_dict.keys():
        split_cse = cse.split(delim)
        total_snps = [len(naf_dict[cse])]
        outlist = split_cse + total_snps
        outlist[1] = int(outlist[1])
        outlist[2] = int(outlist[2])
        list_of_lists.append(outlist)

```

```

list_of_lists.sort(key = operator.itemgetter(0, 1))

##print bed file with total snp number
outfh = open(sample + '_' + genome_and_window + '_snp_number.bedgraph',
'w')
for line in list_of_lists:
    for i in range(len(line)):
        line[i] = str(line[i])
    outfh.write(delim.join(line) + '\n')
outfh.close()

##method to combine hom mapping data i.e. bedgraphs
##suply working dir, file prefix name, and genome and window information
def combine_bedgraphs_for_r(working_directory, prefix, genome_window):
    chr_wanted = ['chr1', 'chr2', 'chr3', 'chr4', 'chr5', 'chr6', 'chr7',
'chr8', 'chr9', 'chr10',
                'chr11', 'chr12', 'chr13', 'chr14', 'chr15', 'chr16', 'chr17',
'chr18', 'chr19', 'chr20',
                'chr21', 'chr22', 'chr23', 'chr24', 'chr25', '1', '2', '3', '4', '5',
'6', '7', '8', '9',
                '10', '11', '12', '13', '14', '15', '16', '17', '18', '19', '20',
'21', '22', '23', '24', '25']
    snp_count = prefix + '_' + genome_window + '_snp_number.bedgraph'
    het_count = prefix + '_' + genome_window + '_het_count.bedgraph'
    hom_count = prefix + '_' + genome_window + '_hom_count.bedgraph'
    hom_perc = prefix + '_' + genome_window + '_hom_percentage.bedgraph'
    naf_ph = prefix + '_' + genome_window + '_naf.bedgraph'
    combined_r_file = prefix + '_' + genome_window +
'_combined_hom_mapping.txt'
    with open(combined_r_file, "w") as r_file:
        r_file.write(delim.join(['chr', 'start', 'end', 'value',
'chromosome', 'analysis', '\n']))
        with open(hom_count, "r") as homc:
            for line in homc:
                line = line.strip('\n').split(delim)
                line = [line[0].replace('chr', '')] + line[1:] #removes
chr from start of line
                ##remove chromosomes we're not interested in
                if line[0] in chr_wanted:
                    start = int(line[1])
                    end = int(line[2])
                    midpoint = start + ((end - start) / 2)
                    r_file.write(delim.join(line[:4] + [str(midpoint),
'hom count', '\n']))
        with open(hom_perc, "r") as homp:
            for line in homp:
                line = line.strip('\n').split(delim)
                line = [line[0].replace('chr', '')] + line[1:] #removes
chr from start of line
                if line[0] in chr_wanted:
                    start = int(line[1])
                    end = int(line[2])
                    midpoint = start + ((end - start) / 2)

```

```

        r_file.write(delim.join(line[:4] + [str(midpoint),
'hom percentage', '\n']))
    with open(naf_ph, "r") as naf:
        for line in naf:
            line = line.strip('\n').split(delim)
            line = [line[0].replace('chr','')] + line[1:] #removes
chr from start of line
            if line[0] in chr_wanted:
                start = int(line[1])
                end = int(line[2])
                midpoint = start + ((end - start) / 2)
                r_file.write(delim.join(line[:4] + [str(midpoint),
'average naf', '\n']))
##call methods, samtools, annovar and format multianno

##map with bwa and process with samtools etc
align_with_bwa(fq_dict)
make_list_of_bams(fq_dict, mkdup_bam, bamslst_file)
variant_calling_samtools(bamslst_file, st_vcf)
convert_to_annovar(st_vcf)
run_table_annovar(st_avinputs)
multianno_to_annotated(st_avinputs)

##filter vars and hom mapping

#filter variants for candidates snps
for sample in fq_dict:
    ##exonic_variants
    filter_ann(working_dir, "or", sample + '.annotated.txt' , sample +
"_1.temp", [col_exon, col_exon], ['==','=='],
[exon_definition[0],exon_definition[1]])
    ##remove synonymous
    filter_ann(working_dir, "and", sample + "_1.temp", sample + "_2.temp",
[col_function], ['!='], [syn_definition])
    ##remove if in dbsnp, sanger, or other mouse line
    filter_ann(working_dir, "and", sample + "_2.temp", sample + "_3.temp",
[13,14,15,16,17,18,19,20,21,22,23],
['==','==','==','==','==','==','==','==','==','==','=='],
['','','','','','','','','','',''])
    ##keep if hom
    filter_ann(working_dir, "and", sample + "_3.temp", sample +
'.hom_exonic_rare.xls', [zygosity_col], ['=='], ['hom'])
    ##filter variants by coverage and quality
    filter_ann(working_dir, "and", sample + '.hom_exonic_rare.xls', sample +
'.hom_exonic_rare_qual_filtered.xls', [cov_col,qual_col], ['>','=','>='],
[cov_definition,qual_definition])

##homozygosity mapping
for ws in window_size:
    for sample in fq_dict:
        ##remove if in dbsnp, sanger, other ped or rmsk
        filter_ann(working_dir, "and", sample + '.annotated.txt', sample +
"11.temp", [11,12,13,14,15,16,17,18,19,20,21,22,23],

```

```

['==','==','==','==','==','==','==','==','==','==','==','==','=='],
['','','','','','','','','','','','','','','','',''])
# filtering_annotated.filter(working_dir, "and", sample +
'.annotated.txt', sample + "11.temp", [11,12,23], ['==','==','=='], ['','',''])

##filter variants by coverage and quality
filter_ann(working_dir, "and", sample + "11.temp", sample +
'.hom_temp.txt', [cov_col,qual_col], ['>=','>='],
[cov_definition,qual_definition])

#make bed file with windows and returns genome name and window size
variable
genome_and_window = make_windows(working_dir, genome_fai, ws,
step_size).split('.')[0]

##make bed file from variants
make_bed_from_ann(working_dir, 'samtools', sample +
'.hom_temp.txt', zygoty_col, info_col)
##hom and het count and hom percentage
count_and_percentage(working_dir, genome_and_window, sample +
'.bed')

##naf
naf_in_window(working_dir, genome_and_window, sample + '.bed')
##total snp number
total_snp_in_window(working_dir, genome_and_window, sample + '.bed')

##combine bedgraphs for graphing in r
combine_bedgraphs_for_r(working_dir, sample, genome_and_window)

```

Supplemental Table 1. Recombination map of the ENU-induced *fosse* locus on chromosome 8.

Embryo ID	Sex	<i>Ddx60</i>	<i>Colgalt1</i>	<i>Jak3</i>	<i>Ces1e</i>	<i>Hsf4</i>	Embryo Phenotype
K407.0003.4	F	MUT/MUT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i>
K407.0009.7	M	MUT/MUT	MUT/MUT	N/A	WT/WT	WT/WT	<i>fosse</i>
K421.0003.1	M	MUT/MUT	MUT/MUT	MUT/MUT	WT/MUT	WT/MUT	<i>fosse</i>
K421.0003.5	M	WT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i>
K421.0018.8	F	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	WT/MUT	<i>fosse</i>
K421.0035.7	F	WT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i>
K422.0002.5	M	MUT/MUT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i>
K422.0002.6	F	MUT/MUT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i>
K421.0009.4	F	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K421.0018.7	M	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K421.0019.5	M	WT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K421.0022.3	F	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K421.0022.4	M	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K421.0035.8	F	WT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with cleft palate
K433.0006.4	M	WT/WT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i> with cleft palate
K433.0014.2	F	WT/WT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i> with cleft palate
K421.0009.5	F	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with exencephaly
K421.0019.4	M	MUT/MUT	MUT/MUT	MUT/MUT	N/A	WT/MUT	<i>fosse</i> with exencephaly
K421.0028.3	M	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	MUT/MUT	<i>fosse</i> with exencephaly
K421.0060.3	F	MUT/MUT	MUT/MUT	MUT/MUT	WT/MUT	WT/MUT	<i>fosse</i> with exencephaly
K433.0006.5	M	WT/WT	MUT/MUT	MUT/MUT	WT/WT	WT/WT	<i>fosse</i> with exencephaly
K407.0032.4	M	WT/WT	WT/WT	WT/WT	WT/WT	WT/WT	<i>ugli</i>
K407.0034.2	M	WT/MUT	WT/MUT	WT/MUT	WT/WT	WT/WT	<i>ugli</i>
K407.0034.7	F	WT/WT	WT/MUT	WT/WT	WT/WT	WT/WT	<i>ugli</i>

All sample ID #s in bold type were included in WGS analysis

Supplemental Table 2. Genotyping data for *ugli* candidate variants.

Embryo ID	Sex	<i>Pnpla3</i> Chr 15	<i>Gm15800</i> Chr 5	<i>Plod3</i> Chr 5	Phenotype
K407.0032.4	M	?	MUT/MUT	MUT/MUT	<i>ugli</i> exencephaly
K407.0034.2	M	MUT/MUT	WT/MUT	MUT/MUT	<i>ugli</i> exencephaly
K407.0034.7	F	WT/MUT?	MUT/MUT	MUT/MUT	<i>ugli</i> exencephaly
K407.0003.4	F	WT/MUT?	MUT/MUT	WT/MUT	<i>fosse</i>
K407.0009.7	M	WT/WT	WT/WT	WT/WT	<i>fosse</i>
K421.0009.5	F	WT/WT	WT/WT	WT/WT	<i>fosse</i> exencephaly
K421.0019.4	M	WT/WT	WT/WT	WT/WT	<i>fosse</i> exencephaly
K421.0028.3	M	WT/WT	WT/MUT	WT/WT	<i>fosse</i> exencephaly
K421.0060.3	F	WT/WT	WT/WT	WT/WT	<i>fosse</i> exencephaly

All sample ID #s in bold type were included in WGS analysis

Supplemental Table 3. Genotyping of *daredevil* and *bub* candidate variants from affected and unaffected littermates.

<i>daredevil</i> candidate genotyping				
Embryo ID	<i>Pgm5</i>	<i>Kif20b</i>	-----	Embryo Phenotype
K416_0006_5	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K416_0007_9	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K416_0036_1	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0013_5	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0024_8	WT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0024_9	WT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0024_10	WT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0032_3	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0044_1	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0044_2	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0051_1	WT/MUT	MUT/MUT	-----	<i>daredevil</i>
K417_0062_1	MUT/MUT	MUT/MUT	-----	<i>daredevil</i>
K416_0006_4	WT/WT	WT/WT	-----	Unaffected
K416_0007_4	WT/MUT	WT/MUT	-----	Unaffected
K417_0032_4	WT/WT	WT/MUT	-----	Unaffected
K417_0044_4	WT/MUT	WT/MUT	-----	Unaffected
<i>bub</i> candidate genotyping				
Embryo ID	<i>Dact1</i>	<i>Myh6</i>	<i>Tgds</i>	Embryo Phenotype
K402_0014_1	MUT/MUT	MUT/MUT	MUT/MUT	<i>bub</i>
K402_0014_5	WT/MUT	WT/WT	MUT/MUT	<i>bub</i>
K402_0014_6	WT/MUT	MUT/MUT	MUT/MUT	<i>bub</i>
K402_0015_3	WT/WT	MUT/MUT	MUT/MUT	<i>bub</i>
K402_0015_4	MUT/MUT	WT/WT	MUT/MUT	<i>bub</i>
K402_0035_1	WT/MUT	MUT/MUT	MUT/MUT	<i>bub</i>
K402_0035_2	MUT/MUT	MUT/MUT	MUT/MUT	<i>bub</i>
K402_0040_10	WT/WT	WT/WT	MUT/MUT	<i>bub</i>
K402_0014_3	WT/WT	WT/MUT	WT/WT	Unaffected
K402_0015_2	WT/MUT	WT/MUT	WT/MUT	Unaffected
K402_0035_5	WT/MUT	MUT/MUT	WT/MUT	Unaffected
K402_0035_6	WT/WT	WT/WT	WT/WT	Unaffected

All sample ID #s in bold type were included in WGS analysis

Supplemental Table 4. Genotyping of *timon* mutants across two critical intervals on chromosomes 7 and 18.

Chr7	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
Gene ID																		
<i>Ccp110</i>	HET	WT	HET	HET	HET	HET	WT	HET	MUT	HET	MUT	MUT	MUT					
<i>Sult1a1</i>	HET	WT	HET	HET	HET	HET	HET	HET	MUT	HET	MUT	MUT	MUT					
Chr18																		
Gene ID																		
<i>Chst9/Chd2</i>	WT	WT	MUT	MUT	MUT	WT	HET	HET	MUT	MUT	HET	HET	MUT					
<i>Pcdhb10</i>	WT	WT	MUT	MUT	MUT	WT	HET	HET	MUT	MUT	HET	MUT	MUT					
<i>1700011I03Rik</i>	WT	WT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT
<i>Fbn2</i> Deletion	WT	WT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT
<i>Pdgfrb</i>	WT	WT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT
<i>miR466p</i>	WT	WT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT	MUT
						SEQ			SEQ				SEQ					

Chromosome 7 critical interval = 32.9 Mb

Chromosome 18 critical interval = 52 Mb

Deletion on chromosome 18 = 58,012,626-58,014,322, < 400 kb from the *1700011I03Rik* variant, 3 Mb from *Pdgfrb* variant

SEQ: Genomic DNA was submitted for WGS