

APPENDIX

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Appendix Figure S1

Representative transmission electron microscopy images from serial cross sections of the hermaphrodite amphid sensory pore in WT, *rabs-5(ok1513)* and *vps-45(tm0246)* worms. The entire pore is shown in the large panels; individual cilia are shown at higher magnifications in the small panels. Red outline denotes the PCMCs. The schematics show the the section positions (colour coded) and the ultrastructure phenotypes observed. For simplicity, the cartoons of the pores in longitudinal orientation only show 3 out of the 10 channel axonemes. PCMC; periciliary membrane compartment. sMT; singlet microtubules.

Appendix Figure S2

A. Representative transmission electron microscopy images from serial cross sections of the male-specific cephalic (CEP and CEM) sensory pore in WT, *rabs-5(ok1513)* and *vps-45(tm0246)* worms. CEM and CEP cilia outlined in pink and yellow, respectively. Neighbouring OLQ cilium outlined in green. PCMC; periciliary membrane compartment. Left images show the entire pore; smaller right-hand images show indicated cilia at higher magnifications. Blue arrowheads denote vesicles in the PCMC of CEM cilia. Yellow arrowheads denote extracellular vesicles in the sensory pore lumen. Schematics show the

CEP, CEM and OLQ cilia and associated pores in longitudinal and cross section, and indicate the ultrastructural phenotypes observed, as well as the section positions (colour coded). Note that whilst the proximal and middle (doublets and singlets; (Silva et al. Curr Biol. 2017 Apr 3; 27(7):968-980)) portions of the mutant CEM cilia are normal, we cannot tell from our images if the distal portion is normal or not in the mutants.

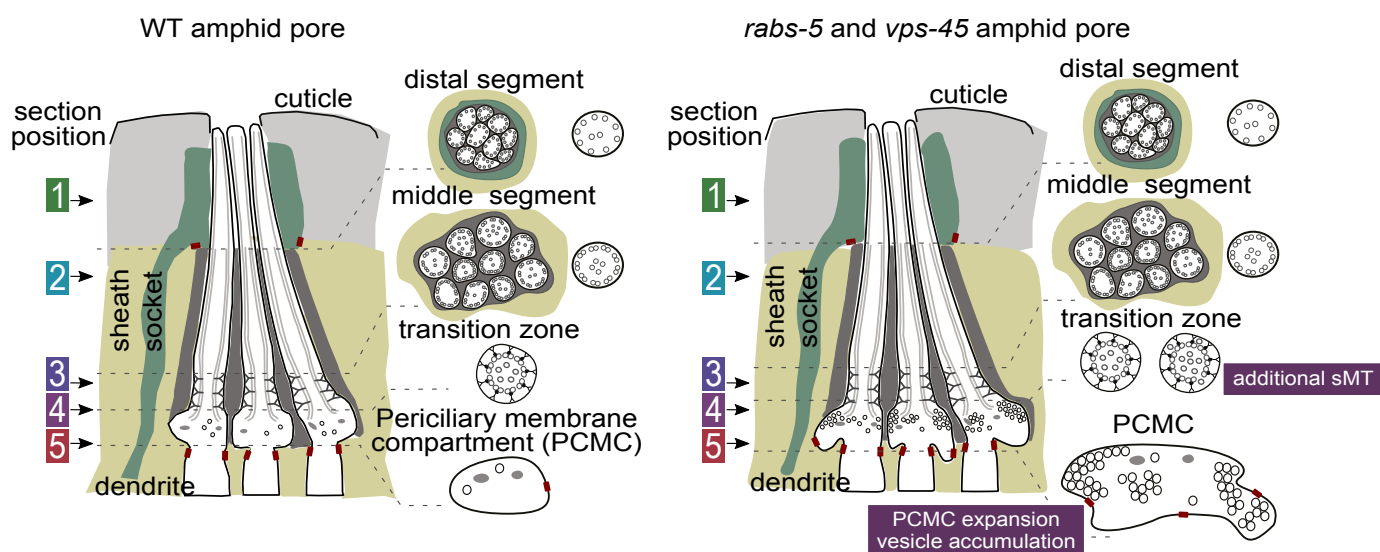
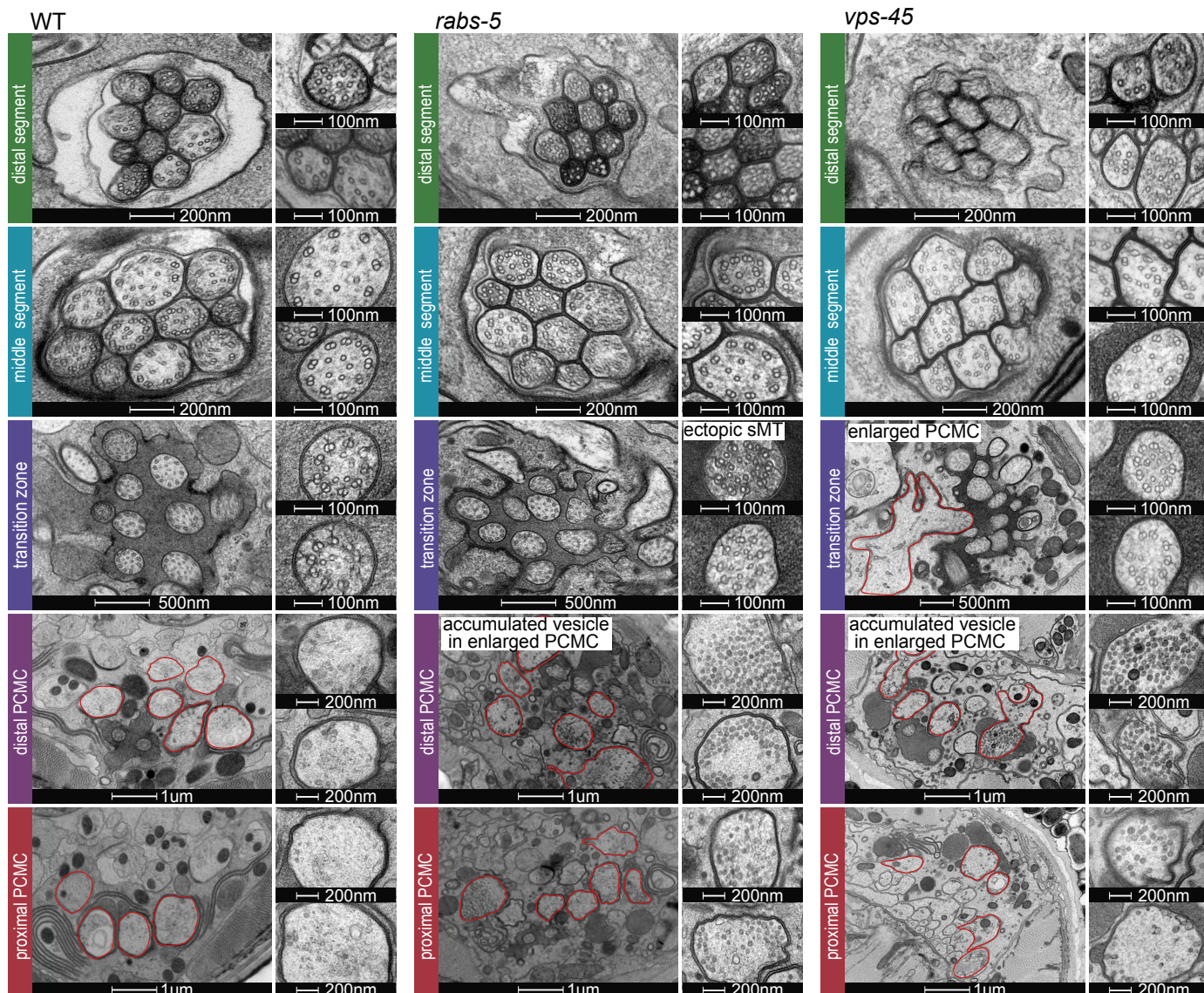
B. Representative transmission electron microscopy images from serial cross sections of the hermaphrodite inner labial sensory pore in WT, *rabs-5(ok1513)* and *vps-45(tm0246)* worms. IL1 cilium outlined in dark blue. IL2 cilium outlined in light blue.

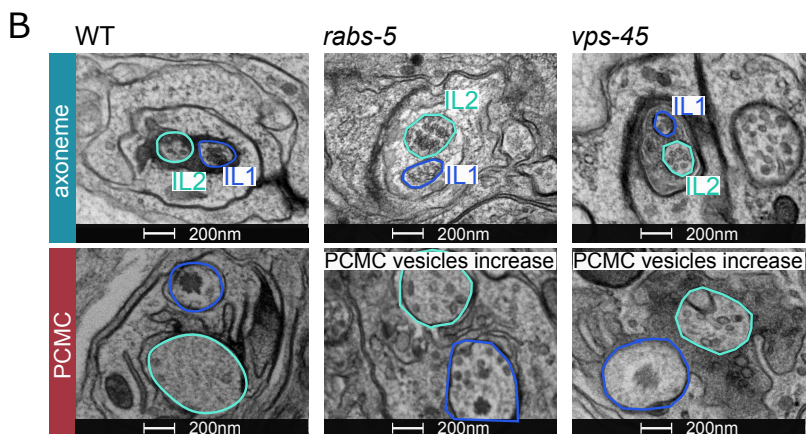
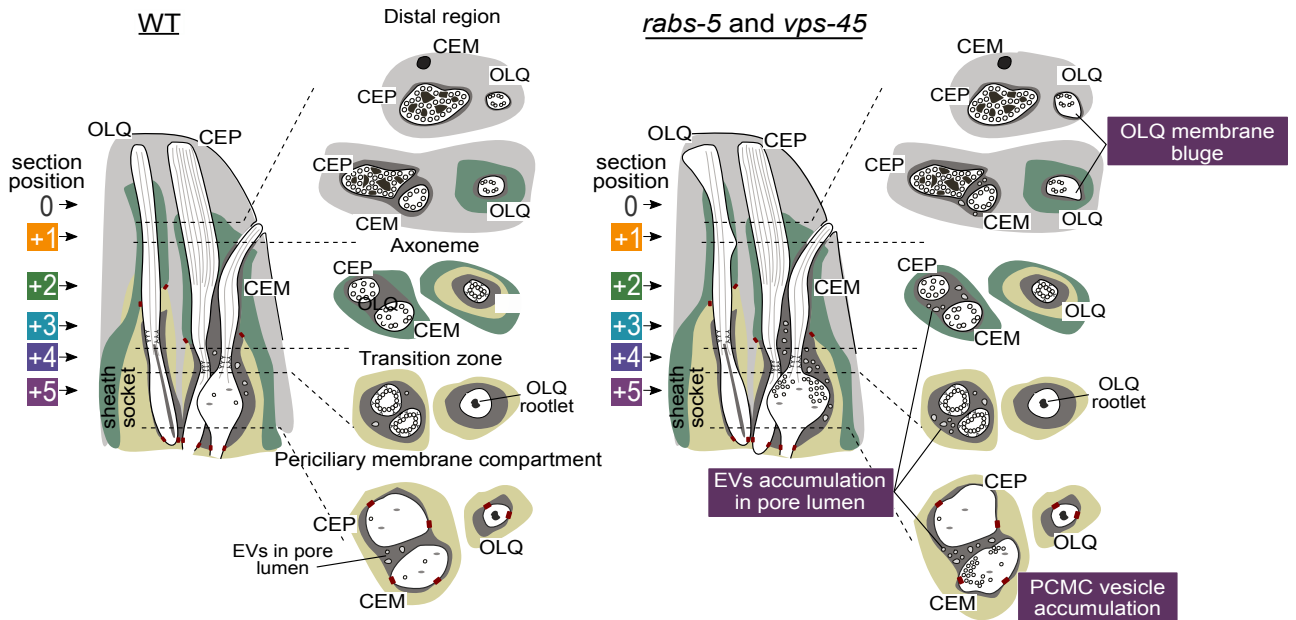
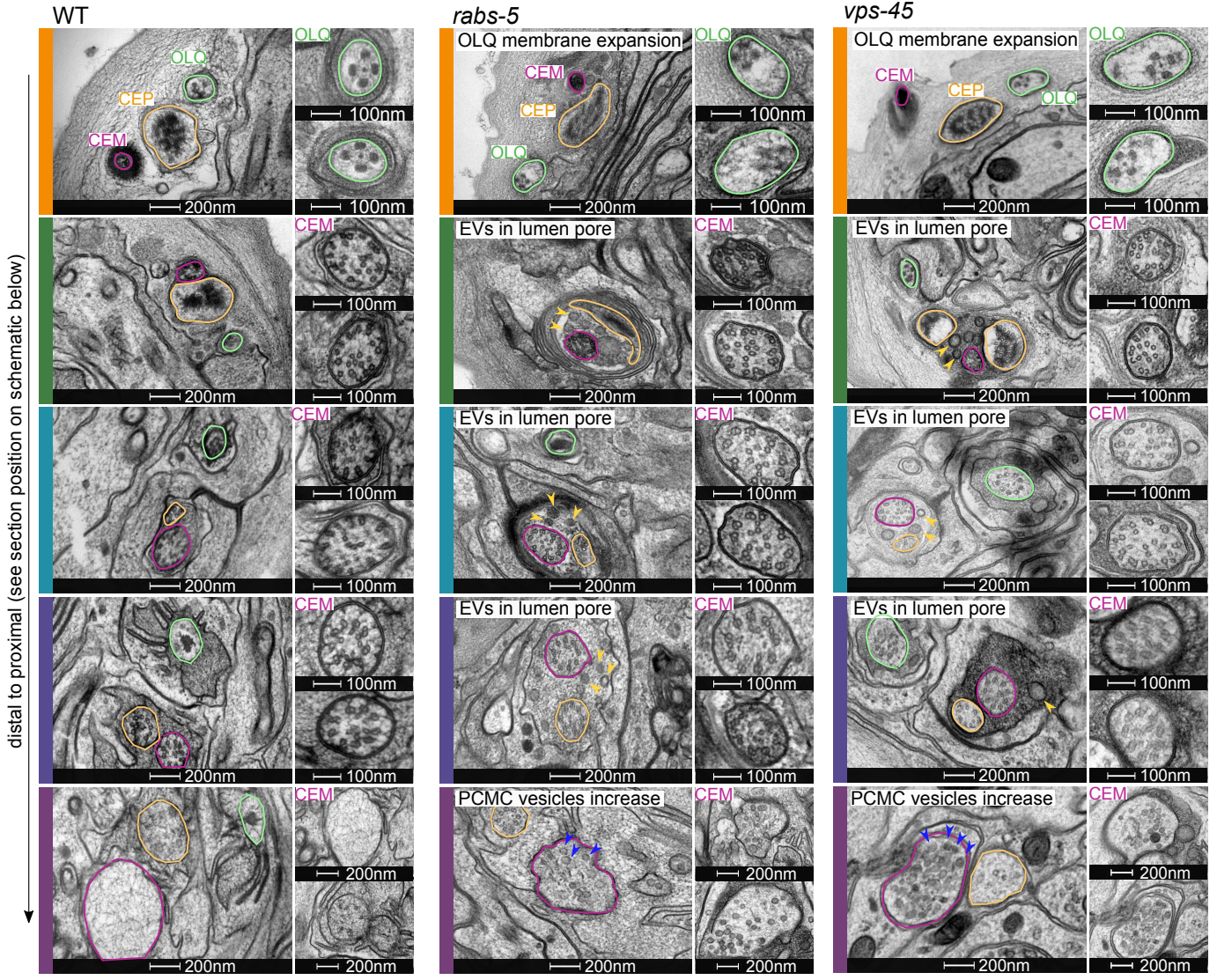
Appendix Table S1 List of *C. elegans* genes and corresponding alleles screened for cilium structure and function defects (**Fig 1A**).

Appendix Table S2 *C. elegans* strains used in this study.

Appendix Table S3. Primer used in this study to genotype by PCR the indicated *C. elegans* mutant alleles.

Appendix Figure S1, related to Figure 4





Appendix Figure S2, related to Figure 4

Appendix Table S1:List of *C. elegans* genes and corresponding alleles screened for cilium structure and function defects

<i>C. elegans</i> gene	allele	human orthologue
<i>arf-1.2</i>	<i>ok796</i>	ARF1 ADP ribosylation factor 1
<i>asm-2</i>	<i>gk293929</i>	SMPD1 sphingomyelin phosphodiesterase 1
<i>cav-1</i>	<i>ok2089</i>	CAV1 caveolin 1
<i>cav-2</i>	<i>hc191</i>	CAV2 caveolin 2
<i>dgk-1</i>	<i>ok1462</i>	DGKQ diacylglycerol kinase theta
<i>eea-1</i>	<i>ok1040</i>	EEA1 early endosome antigen 1
<i>gta-1</i>	<i>ok517</i>	GABAT 4-aminobutyrate aminotransferase
<i>hyl-1</i>	<i>ok976</i>	CERS6 ceramide synthase 6
<i>hyl-2</i>	<i>ok1766</i>	CERS4 ceramide synthase 4
M01E5.2	<i>ok2552</i>	MTG2 mitochondrial ribosome associated GTPase 2
<i>mec-2</i>	<i>e75</i>	STOM stomatin
<i>mtm-6</i>	<i>ok330</i>	MTMR6 myotubularin related protein 6
<i>mtm-9</i>	<i>ok3523</i>	MTMR9 myotubularin related protein 9
<i>mtr-4</i>	<i>ok2642</i>	SKIV2L2 Ski2 like RNA helicase 2
<i>nex-1</i>	<i>gk148</i>	ANX13 annexin A13
<i>nex-2</i>	<i>ok764</i>	ANX7 annexin A7
<i>nex-3</i>	<i>gk385</i>	ANX5 annexin A5
<i>nex-4</i>	<i>gk102</i>	ANX6 annexin A6
<i>num-1</i>	<i>bc365</i>	NUMBL NUMB like, endocytic adaptor protein
<i>prmn-1</i>	<i>gk545</i>	PROM1 prominin 1
<i>rab-10</i>	<i>ok1494</i>	RAB10 RAB10
<i>rabn-5</i>	<i>ok1555</i>	RABPT5 rabaptin-5, RAB GTPase binding effector protein 1
<i>rabs-5</i>	<i>ok1513</i>	RABSN rabenosyn, RAB effector
<i>rabx-5</i>	<i>tm1215</i>	RABGEF1 rabex5, RAB guanine nucleotide exchange factor 1
<i>ric-19</i>	<i>ok833</i>	ICA1 islet cell autoantigen 1
<i>ric-4</i>	<i>md1088</i>	SNAP25 synaptosome associated protein 25
<i>ric-8</i>	<i>md1909</i>	RIC8A RIC8 guanine nucleotide exchange factor A
<i>rme-1</i>	<i>b1045</i>	EHD1 EH domain containing 1
<i>rme-2</i>	<i>b1008</i>	LRP2 LDL receptor related protein 2
<i>rme-6</i>	<i>b1014</i>	GAPVD1 GTPase activating protein and VPS9 domains 1
<i>rme-8</i>	<i>b1023</i>	DNAJC13 DnaJ heat shock protein family (Hsp40) member C13
<i>sax-7</i>	<i>ok1244</i>	NRCAM neuronal cell adhesion molecule
<i>stn-2</i>	<i>ok2417</i>	SNTG2 syntrophin gamma 2
<i>syx-7</i>	<i>gk338314</i>	STX12 syntaxin 12
<i>tat-3</i>	<i>gk819359</i>	ATP10A ATPase phospholipid transporting 10A
<i>tat-6</i>	<i>ok1984</i>	ATP9B ATPase phospholipid transporting 9B
<i>tin-9.1</i>	<i>ok2194</i>	TIMM9 translocase of inner mitochondrial membrane 9
<i>tmem-135</i>	<i>ok1646</i>	TMEM135 transmembrane protein 135
<i>unc-1</i>	<i>e94</i>	STOM stomatin
<i>unc-24</i>	<i>e138</i>	STOML1 stomatin like 1
<i>unc-44</i>	<i>e362</i>	ANK1 ankyrin 1
<i>vps-45</i>	<i>tm0246</i>	VPS45 vacuolar protein sorting 45 homolog
<i>vps-54</i>	<i>ok1463</i>	VPS54, GARP complex subunit
<i>wdfy-2</i>	<i>ok3592</i>	WDFY2 WD repeat and FYVE domain containing 2

Appendix Table S2: *C. elegans* strains used in this study

N2 (Bristol)
him-5(e1490)
osm-5(p813)
bbs-8(nx77)
arf-1.2(ok796)
asm-2 (gk293929)
cav-1(ok2089)
cav-2(hc191)
dgk-1(ok1462)
eea-1(ok1040)
gta-1 (ok517)
hyl-1(ok976)
hyl-2(ok1766)
M01E5.2 (ok2552)
mec-2(e75)
mtm-6 (ok330)
mtm-9 (ok3523)
mtr-4 (ok2642)
nex-1(gk148)
nex-2(ok764)
nex-3(gk385)
nex-4(gk102)
num-1(bc365)
prmn-1(gk545)
rab-10(ok1494)
rabn-5(ok1555)
rabs-5(ok1513)
rabx-5(tm1215)
ric-19(ok833)
ric-4(md1088)
ric-8(md1909)
rme-1(b1045)
rme-2(b1008)
rme-6(b1014)
rme-8(b1023)
sax-7(ok1244)
stn-2(ok2417)
syx-7 (gk338314)
tat-3(gk819359)
tat-6(ok1984)
tin-9.1(ok2194)
tmem-135 (ok1646)
unc-1(e94)
unc-24(e138)
unc-44(e362)
vps-45(tm0246)
vps-54 (ok1463)
wdfy-2(ok3592)
N2; *oqEx306[rabs-5p::gfp + unc122 p::dsRed]*
rabs-5(ok1513; oqEx307[rabs-5p::rabs-5 + unc122 p::dsRed]
vps-45 (tm0246); oqEx308[arl-13p::vps45 + unc122 p::dsRed]
rabs-5(ok1513); oqEx401[arl-13p::rfp::rabs-5(WT) + unc122 p::dsRed]
rabs-5(ok1513); oqEx402[arl-13p::rfp::rabs-5(Δ ZN) + unc122 p::dsRed]
rabs-5(ok1513); oqEx403[arl-13p::rfp::rabs-5(Δ FYVE) + unc122 p::dsRed]
rabs-5(ok1513; oqEx404[arl-13p::rfp::rabs-5(Δ RBD) + unc122 p::dsRed]
N2; *oqEx405[arl-13p::rfp::rabs-5(WT) + unc122 p::gfp]*
N2; *oqEx406[arl-13p::rfp::rabs-5(Δ ZN) + unc122 p::gfp]*
N2; *oqEx407[arl-13p::rfp::rabs-5(Δ FYVE) + unc122 p::gfp]*

N2;oqEx408[arl-13p::rfp::rabs-5(Δ RBBD) + unc122 p::gfp]
N2;oqEx409[arl-13p::vps-45::gfp + unc122 p::dsRed]
N2;oqEx410[arl-13p::gfp::rab-5 + unc122 p::dsRed]
N2;oqEx411[arl-13p::wdfy-2::gfp + unc122 p::dsRed]
N2;oqEx412[arl-13p::rfp::eea-1 + unc122 p::gfp]
N2;oqEx413[arl-13p::dpy-23::gfp + unc122 p::dsRed]
N2;oqEx414[arl-13p::gfp::cav-1 + unc122 p::dsRed]
him-5(e1490);oqEx415[*pkd-2 p::rfp::rabs-5(WT)* + unc122 p::dsRed]
him-5(e1490);oqEx416[*pkd-2 p::rfp::cav-1* + unc122 p::dsRed]
him-5(e1490);oqEx417[*pkd-2 p::wdfy-2::rfp* + unc122 p::dsRed]
N2;oqEx418[*str-2 p::jbtS-14::gfp* + *str-2::rfp* + unc122 p::gfp]
N2;oqEx204[arl-13p::gfp::rab-5(Q78L) + *rol-6(su1006)*]
N2;oqEx205[arl-13p::gfp::rab-5(S33N) + *rol-6(su1006)*]
him-5(e1490);myls1[*pkd-2::gfp* + *unc-122p::gfp*] IV
N2;oqEx58[arl-13p::arl-13::gfp + *rol-6(su1006)*]
N2;oyEx[srbc-66::gfp + unc122 p::dsRed]
N2;nxEx[*tram-1::tdTomato* + *bbs-8p::mks-2::gfp* + *rol-6(su1006)*]
lin-15(n765);kyls141[*osm-9::gfp* + *lin-15(+)*]
N2;nxEx289[*rpi-2::gfp* + *xbx-1::tdTomato* + *rol-6(su1006)*]
N2;kyls104[*str-1 p::gfp*]
N2;mnls17[*osm-6 p::osm-6::gfp*]

Appendix Table S3_Primer used for genotyping the indicated mutant allele

primer name	primer sequence	allele genotyped
cav-1_F	AGGCGAAGTTACATTAGGCTG	<i>cav-1(ok2089)</i>
cav-1_R	GCACTCTTTCTACGCAATCG	<i>cav-1(ok2089)</i>
cav-1_Pn	TGTGTCAGTAGGGAAATCGC	<i>cav-1(ok2089)</i>
eea-1_F	ATTTAAGCGGAACAGCCTCA	<i>eea-1(b1040)</i>
eea-1_R	CTTCCAATTCTCTTGCTCG	<i>eea-1(b1040)</i>
eea-1_Pn	GAAATCAGAGGGAATCCGG	<i>eea-1(b1040)</i>
rabs-5_F	GATTGGAGCAACTGGTTCAG	<i>rabs-5(ok1513)</i>
rabs-5_R	ATGATCCGTTTGTATTGTATCATC	<i>rabs-5(ok1513)</i>
rabs-5_Pn	CGGTAAAATTTTCATCAGGAG	<i>rabs-5(ok1513)</i>
rabx-5_F	CGCTTAAGGAGAGTGGTGA	<i>rabx-5(tm1215)</i>
rabx-5_R	ATTCTTCGCAAAATCCTGAC	<i>rabx-5(tm1215)</i>
rabx-5_Pn	CTCGCCATGCTTATCTGAC	<i>rabx-5(tm1215)</i>
rme-1_F	CTTATCCGGAGAAAAGCAAA	<i>rme1(b1045)</i>
rme-1_R	TACCATGAAGCAGTCGACC	<i>rme1(b1045)</i>
rme-1_Pn	GAATGATCCCAAATAGACACG	<i>rme1(b1045)</i>
rme-6(WT)_F	GCAATCATCAGTCCGC	<i>rme-6(b1014)</i>
rme-6(b1014)_F	GGCAATCATCAGTCCGT	<i>rme-6(b1014)</i>
rme-6_R	TTTCTAAAAGTACGTCCCTC	<i>rme-6(b1014)</i>
vps-45_F	AATGCAAATTCCTCTGCTCT	<i>vps-45(tm0246)</i>
vps-45_R	TTCAACTGCAACAATAGCG	<i>vps-45(tm0246)</i>
vps-45_Pn	ACAGTGTCATGCGCATTT	<i>vps-45(tm0246)</i>
wdfy-2_F	ATGATAGATCCGTTTCGCCTG	<i>wdfy-2(ok3592)</i>
wdfy-2_R	AGAAGGGGCAGAGCAAAC	<i>wdfy-2(ok3592)</i>
wdfy-2_Pn	GAAAGTGGTCCGATTGAGAC	<i>wdfy-2(ok3592)</i>