

	360	370	380	390	400	410	
800039	R	N	L	S	P	E	A
800041	R	N	L	S	P	E	A
900044	R	N	L	S	P	E	A
900046	R	N	L	S	P	E	A
800024	R	N	L	S	P	E	A
800025	R	N	L	S	P	E	A
800044	R	N	L	S	P	E	A
800036	R	N	L	S	P	E	A
800037	R	N	L	S	P	E	A
900038	R	N	L	S	P	E	A
800035	R	N	L	S	P	E	A
800038	R	N	L	S	P	E	A
800043	R	N	L	S	P	E	A
900039	R	N	L	S	P	E	A
900047	R	N	L	S	P	E	A
900043	R	N	L	S	P	E	A
800030	R	N	L	S	P	E	A
900040	R	N	L	S	P	E	A
800022	R	N	L	S	P	E	A
800023	R	N	L	S	P	E	A
800040	R	N	L	S	P	E	A
900041	R	N	L	S	P	E	A
800026	R	N	L	S	P	E	A
AY12	R	D	L	L	S	E	A
AY11	R	D	L	L	S	E	A
DQ	R	D	L	L	S	E	A
Clustal Consensus	*	:	*	*	*	*	*

AY12 = AY282412; AY11= AY282411; DQ = DQ060522

Fig S1: Multiple sequence alignment of squalene epoxidase of terbinafine resistant strains of *Trichophyton rubrum* demonstrating F397L non-synonymous mutation in some of the resistant strains. Only partial sequence is shown here.

Table S1: Reports on terbinafine susceptibility of *T. rubrum* and *T. interdigitale*

S L no	Yea r	Author (referenc e no.)	Country	Patien t sampl e	Growth of dermatophyt e	Species (No.)	Terbinafine MIC in mg/L			Onychomycosi s cases only	Nail, hair and skin involvemen t
						Organism	MIC ₅₀	MIC ₉₀	GM		
1	2017	Yamada <i>et al.</i> (1)	Japan	2056	2056	<i>T. rubrum</i> (1644)	0.1-	-	-	-	Yes
						<i>T. interdigitale</i> (412)	12.8 [#]	3.2*	-		
2	2016	Baghi <i>et al.</i> , (2)	Iran	100	100	<i>T. rubrum</i> (29)	0.063	0.25	0.06	-	Yes
						<i>T. interdigitale</i> (52)	0.063	0.25	0.007		
3	2016	Mohd Nizam <i>et al.</i> (3)	Malaysia	11	11	<i>T. rubrum</i> (3)	-	2-4 [#]	3.17	-	Yes
						<i>Trichophyton</i> spp (7)	-	0.06- 0.12 [#]	0.08		
4	2011	Ansar <i>et al.</i> (4)	Iran	316	316	<i>T. interdigitale</i> (156)	0.016	0.125	0.017	-	Yes
						<i>T. rubrum</i> (60)	0.016	0.063	0.017		
5	2013	Adimi <i>et al.</i> (5)	Iran	320	320	<i>T. rubrum</i> (89)	0.031	16	0.172	-	Yes
						<i>T. mentagrophyte</i> s (136)	2	0.031	16	0.142	
6	2014	Silva <i>et al.</i> (6)	Brazil	216	70	<i>T. interdigitale</i> (24)	<0.03	<0.031	0.03	-	Yes
						<i>T. rubrum</i> (37)	1	<0.03	<0.031	0.06	
							1				

7	2010	Magagnin <i>et al.</i> (7)	Brazil	26	26	<i>T. interdigitale</i> (4)	0.06*	0.04	-	Yes
						<i>T. rubrum</i> (3)	0.5*	0.07		
						<i>T.</i> <i>mentagrophyte</i> s (8)	16*	0.04		
8	2010	Zalacain <i>et al.</i> (8)	Spain	100	70	<i>T. rubrum</i> (35)	0.032	0.125	0.039	Yes
						<i>T.</i> <i>mentagrophyte</i> s (29)	0.125	0.5	0.082	-
9	2010	Beuno <i>et</i> <i>al.</i> (9)	Colombia	103	30	<i>T. rubrum</i> (10)	0.03	0.06	0.026	Yes
						<i>T.</i> <i>mentagrophyte</i> s (18)	0.015	0.03	0.014	-
10	2009	Rodrigues <i>et al.</i> (10)	Brazil	60	60	<i>T. rubrum</i> (27)	0.125	0.25	-	-
						<i>T.</i> <i>mentagrophyte</i> s (14)	0.06	0.25	-	Yes
11	2007	da Silva <i>et al.</i> (11)	Brazil	100	100	<i>T. rubrum</i> (50)	<0.00 7	0.007	-	Yes
						<i>T.</i> <i>mentagrophyte</i> s (50)	0.007	0.015		-
12	2007	Singh <i>et</i> <i>al.</i> (12)	Canada	63	63	<i>T. rubrum</i> (16)	-	0.006*	-	-
						<i>T.</i> <i>mentagrophyte</i> s (16)	-	0.004*		Yes
13	2006	Barros <i>et</i> <i>al.</i> (13)	Brazil	100	100	<i>T. rubrum</i> (50)	-	0.007- 0.031 [#]	-	Yes
						<i>T.</i> <i>mentagrophyte</i> s (50)	-	<0.007 -0.015 [#]	-	-

14	2005	Esteban <i>et al.</i> (14)	Spain	59	59	<i>T. rubrum</i> (10) 8	0.007 0.01	0.01 0.03	0.008 0.015	-	Yes
						<i>T. mentagrophytes</i> (9)					
15	2004	Karaca <i>et al.</i> (15)	Turkey	56	56	<i>T. rubrum</i> (50) T. mentagrophytes (3)	0.002 0.001	0.008 -	0.02 0.001	-	Yes

* Minimum inhibitory concentration in µg/ml, # Range of Minimum inhibitory concentration in µg/ml, AFST= Antifungal susceptibility testing, GM= geometric mean

References

- Yamada T, Maeda M, Alshahni MM, Tanaka R, Yaguchi T, Bontems O, Salamin K, Fratti M, Monod M. 2017. Terbinafine resistance of *Trichophyton* clinical isolates caused by specific point mutations in the squalene epoxidase gene. *Antimicrob Agents Chemother* 61.
- Baghi N, Shokohi T, Badali H, Makimura K, Rezaei-matehkolaee A, Abdollahi M, Didehdar M, Haghani I, Abastabar M. 2016. In vitro activity of new azoles luliconazole and lanoconazole compared with ten other antifungal drugs against clinical dermatophyte isolates 1–7.
- Mohd Nizam T, Binting RAA, Mohd Saari S, Kumar TV, Muhammad M, Satim H, Yusoff H, Santhanam J. 2016. In Vitro Antifungal Activities against Moulds Isolated from Dermatological Specimens. *Malays J Med Sci* 23:32–9.
- Ansar A, Farshchian M, Nazeri H, Ghiasian SA. 2011. Clinico-epidemiological and mycological aspects of tinea incognito in Iran: A 16-year study. *Japanese J Med Mycol* 52:25–32.
- Adimi P, Hashemi SJ, Mahmoudi M, Mirhendi H, Shidfar MR, Emmami M, Rezaei-Matehkolaee A, Gramishoar M, Kordbacheh P.

2013. In-vitro Activity of 10 Antifungal Agents against 320 Dermatophyte Strains Using Microdilution Method in Tehran. *Iran J Pharm Res IJPR* 12:537–45.
6. Silva LB, de Oliveira DBC, da Silva B V, de Souza RA, da Silva PR, Ferreira-Paim K, Andrade-Silva LE, Silva-Vergara ML, Andrade AA. 2014. Identification and antifungal susceptibility of fungi isolated from dermatomycoses. *J Eur Acad Dermatol Venereol* 28:633–40.
 7. Magagnin CM, Stopiglia CDO, Vieira FJ, Heidrich D, Machado M, Vetoratto G, Lamb FM, Scroferneker ML. Antifungal susceptibility of dermatophytes isolated from patients with chronic renal failure. *An Bras Dermatol* 86:694–701.
 8. Zalacain A, Obrador C, Martinez JP, Viñas M, Vinuesa T. 2010. Characterization of the antimicrobial susceptibility of fungi responsible for onychomycosis in Spain. *Med Mycol* 49:1–5.
 9. Bueno JG, Martinez C, Zapata B, Sanclemente G, Gallego M, Mesa a C. 2010. In vitro activity of fluconazole, itraconazole, voriconazole and terbinafine against fungi causing onychomycosis. *Clin Exp Dermatol* 35:658–63.
 10. Rodrigues C, Mota A, Miranda KC, Lemos JDA, Costa CR, Kioko L, Passos XS, Meneses H, Rodrigues R. 2009. Comparison of in vitro activity of five antifungal agents against dermatophytes, using the agar dilution and broth microdilution methods. *Rev da Soc Bras Med Trop* 42:250–254.
 11. da Silva Barros ME, de Assis Santos D, Hamdan JS. 2007. Evaluation of susceptibility of *Trichophyton mentagrophytes* and *Trichophyton rubrum* clinical isolates to antifungal drugs using a modified CLSI microdilution method (M38-A). *J Med Microbiol* 56:514–8.
 12. Singh J, Singh J, Zaman M, Gupta AK. 2007. Evaluation of microdilution and disk diffusion methods for antifungal susceptibility testing of dermatophytes. *Med Mycol* 45:595–602.
 13. Barros ME da S, Santos D de A, Hamdan JS. 2006. In vitro methods for antifungal susceptibility testing of *Trichophyton* spp. *Mycol Res* 110:1355–60.
 14. Esteban A, Abarca ML, Cabañas FJ. 2005. Comparison of disk diffusion method and broth microdilution method for antifungal susceptibility testing of dermatophytes. *Med Mycol* 43:61–6.

15. Karaca N, Koç AN. 2004. In vitro susceptibility testing of dermatophytes: comparison of disk diffusion and reference broth dilution methods. *Diagn Microbiol Infect Dis* 48:259–64.

