Supporting Information

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Fig. S1. Tissue expression profile of human FAM49B and hyperactivation of FAM49B KO T cells. (A) RNA-seq data of human FAM49B generated from the Human Protein Atlas (https://www.proteinatlas.org/ENSG00000153310-FAM49B/tissue). The lymphoid tissues are color-coded in gray. (B) Schematic of deleting targeting genes by electroporating cas9-sgRNA-GFP into Jurkat T cells. (C) FACS analysis of CD69 levels in Jurkat cells expressing sgRNAs targeting NTC, c-Cbl, or FAM49B upon overnight anti-TCR stimulation. Data are representative of three independent experiments.



Fig. 52. Two different FAM49B KO clones have similar hyperactivation phenotypes. (A) FACS analysis of pERK phosphorylation in clone 1 and clone 2 described in Fig. 2B. (B) FACS analysis of CD69 levels in clone 1 and clone 2 upon different concentrations of anti-TCR C305 overnight activation. Data are representative of three independent experiments. (C) CD3 and CD28 surface expression levels of J.FAM49B.GFP and J.FAM49B.WT. (D) Western blot analysis of endogenous FAM49B and FAM49A expression level in Jurkat cells. GAPDH/β-actin were used as loading controls.

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Fig. S3. Endogenous Rac, but not THEMIS, interacted with C-tagged FAM49B. The samples described in Fig. 3C were assessed by immunobloting (IB) using anti-Rac and anti-THEMIS antibodies.

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Fig. 54. FAM49B and CYFIP1 share similar DUF1394 domain. (A) Schematic of DUF1394 domain in FAM49B or CYFIP1 and protein sequence alignment of DUF1394 domain in FAM49B and CYFIP1. (B) FACS analysis of intracellular myc level and GFP expression level in the samples described in Fig. 4B.



Fig. S5. J.FAM49B-WT and WT Jurkat cells had a similar PAK phosphorylation pattern and treatments of PAK inhibitor affected CD69 upregualtion. (A) The samples described in Fig. 5*B* and WT Jurkat cells were stimulated with anti-TCR for times indicated. Phosphorylation of PAK was assessed by immunoblotting. GAPDH was used as loading control. (*B*) FACS analysis of CD69 expression level of FAM49B deficient or sufficient cells. J.FAM49B cells were transduced with empty vector (GFP) or WT-FAM49B at an MOI of 0.5. These samples were activated upon anti-TCR stimulation in the presence of DMSO or PAK inhibitor (FRAX597, 1 μ M). CD69 expression was measured by flow cytometry. (C) CD69 inhibition index was calculated by MFI of CD69 in GFP⁻ population divided by that of GFP⁺ population. ****P* < 0.001.



Fig. S6. DRP1 phosphorylation in FAM49B-deficient and FAM49B-reconstituted T cells and FAM49 expression in mouse T cells. (A) J.FAM49B cells reconstituted with WT-FAM49B, GFP or FAM49B (R161D) were stimulated with anti-TCR for indicated time. Phosphorylation of DRP1(Ser161) were assessed by immunoblotting. GAPDH was used as loading control. (*B*) Both FAM49A and FAM49B are expressed in mouse T cells (https://www.ebi.ac.uk/arrayexpress/experiments/ E-MTAB-2582/).

Other Supporting Information Files

Dataset S1 (XLSX) Dataset S2 (XLSX) Dataset S3 (XLSX)

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